

TEMPERATURE INDUCED PROTEIN UNFOLDING AND FOLDING OF RNase A STUDIED BY TIME-RESOLVED INFRARED SPECTROSCOPY

H. GEORG^a, C. W. WHARTON^{b,*} and F. SIEBERT^a

^a *Institut für Biophysik und Strahlenbiologie, Albert-Ludwigs-Universität
Albertstr. 23, D-79104 Freiburg, Germany;*

^b *School of Biochemistry, University of Birmingham, Edgbaston,
Birmingham B152TT, UK*

(Received 6 April 1997)

When a protein finds its native three-dimensional structure from the unstructured amino-acid chain various processes spanning a large time range are relevant. To understand the mechanism of protein folding one needs to cover the entire folding/refolding ($U \leftrightarrow N$) reaction on a structural level. In the case of RNase A, the main structural changes occur in the ms time range, that can be monitored with rapid-scan-FTIR spectroscopy combined with rapid mixing techniques. To induce unfolding we inject aqueous protein solution into a hot IR cuvette and record the time course of the spectral changes. A lag phase is found when the unfolding conditions are relatively weak, suggesting an unfolding intermediate.

Keywords: Protein folding; time-resolved infrared spectroscopy; RNase A; stopped-flow

Unfolding studies on RNase A revealed three major kinetic phases. In addition to the main unfolding transition (ms) and slow proline isomerization steps (sec. to minutes), a very fast phase has been found by Phillips *et al.*, at pH* 5.7 [1], as well as by our group at pH* 2.0 [2]. Nevertheless, the medium phase seems to comprise the main unfolding transition and will be focused on in this study.

Deuterated Ribonuclease A (RNase A) was heated to defined temperatures and IR absorption spectra were recorded (insert in Fig. 1).

*Corresponding author.

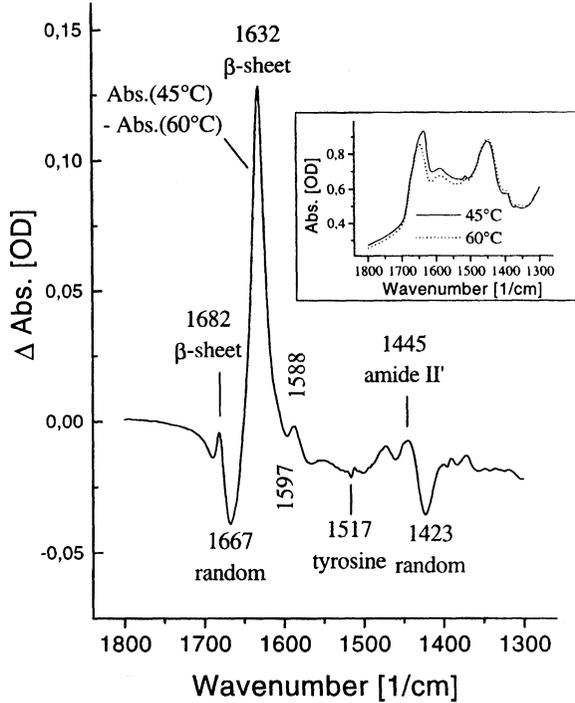
Unfolding spectrum of RNase A in D₂O

FIGURE 1 Temperature dependent IR absorption difference of 1 mM RNase A at pH* 3.9, Δ Abs. = Abs.(45°C)–Abs.(60°C); insert: IR absorption spectra at 45°C (solid line) and at 60°C (dotted line).

The calculated absorption difference is shown in Figure 1 and shows the assignment of distinct absorption bands to structural elements of RNase A. Injecting the native protein at 25°C by a stopped-flow apparatus into a hot ($T_{\text{final}} = 56(50)^{\circ}\text{C}$) IR-cuvette, a ms temperature jump is generated inducing the unfolding of the protein [3]. Rapid-scan-FTIR spectra are recorded every 16 ms. By rearranging the data the kinetic response of RNase A can be extracted for any wavenumber of interest. Figure 2 shows the unfolding kinetic under strong ($T_{\text{final}} = 56^{\circ}\text{C}$) or weak ($T_{\text{final}} = 50^{\circ}\text{C}$) unfolding conditions at 1632 cm^{-1} and 1667 cm^{-1} , representing the decay of β -sheet structure and the growth of “random” structure, respectively. Under the weak

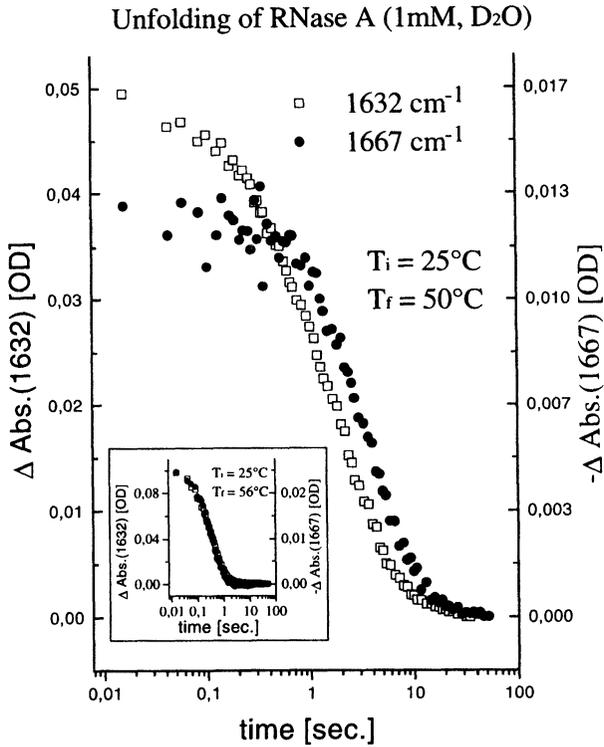


FIGURE 2 Temperature jump induced IR absorption changes of 1 mM RNase A at pH* 3.9 for 1632 cm^{-1} (open squares) and 1667 cm^{-1} (filled circles) and a final temperature of 50°C ; insert: Final temperature of 56°C .

unfolding condition the growth of the “random” structure relative to the decay of the β -sheet structure suggests an intermediate that accumulates under these conditions.

References

- [1] Phillips, C. M., Mizutani, Y. and Hochstrasser, R. M. (1995). *Proc. Natl. Acad. Sci., USA* 92, pp. 7292–7296.
- [2] Georg, H. and Siebert, F. (1995). Temperature-pulse induced protein unfolding and folding studied by time-resolved Infrared spectroscopy, *Conference proceedings of TRVS 7*.
- [3] White, A. J., Drabble, K. and Wharton, C. W. (1995). A stopped-flow apparatus for infrared spectroscopy of aqueous solutions, *Biochemical Journal*, **306**, 843–849.