A Study of Changes in Morphology of Osteoarthritic Articular Cartilage Using Computerized Image Analysis

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1. Introduction

Articular cartilage undergoes substantial structural and molecular changes with age, including surface fibrillation, alteration of structure and composition of collagen, and decrease in strength. Such changes increase the risk of synovial joint degeneration that leads to osteoarthritis (OA), which is a degenerative joint disease characterized by articular cartilage degeneration. It is the most common of the various articular disorders affecting man. Loss of articular cartilage is the major cause of joint dysfunction and disability in OA.

Since its early development, digital microscopic image analysis has offered the potential for improving the objectivity of microscopic observations. Substantial efforts have already been made to convert the evaluations of experienced pathologists into quantitative values in various research and diagnostic fields [1–3]. In a previous study, using computerized image analysis we observed the changes in the morphology of the non-OA femoral articular cartilage with age and only the lacunar size in zone 3 was found to correlate significantly with age. Despite this difference in lacunar size, normal histology, that is, four zones could be identified in all non-OA specimens though minor changes in the superficial zone, was observed in the ageing articular cartilage [4]. However, in other light microscopic studies [5, 6] we observed a completely altered histology throughout the thickness of the cartilage in OA. So the present study was conducted using computerized image analysis to quantitate the differences observed in OA articular cartilage.

2. Materials and Methods

Articular cartilage specimens were obtained from the most prominent area of the medial femoral condyle. The specimens were divided into 2 groups.
Table 1: Mean and standard deviation of the total thickness of articular cartilage in OA group.

<table>
<thead>
<tr>
<th>Age group</th>
<th>41–50 years</th>
<th>51–60 years</th>
<th>61–70 years</th>
<th>71–80 years</th>
<th>81–90 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total thickness (in mm)</td>
<td>2.18 ± 0.02</td>
<td>1.59 ± 0.54</td>
<td>2.29 ± 0.55</td>
<td>2.28 ± 0.14</td>
<td>1.85 ± 0.24</td>
</tr>
</tbody>
</table>

2.1. Group 1 (n = 23). This group consisted of OA specimens. The articular cartilage was obtained from patients (46–81 years) of primary OA undergoing total knee replacement in the Department of Orthopaedics, Postgraduate Institute of Medical Education and Research (PGIMER), Chandigarh, India. Patients suffering from rheumatoid arthritis and secondary OA were excluded from the study.

2.2. Group 2 (n = 12). This group (41–86 years) consisted of non-OA specimens. The articular cartilage was obtained from cadavers in the Department of Anatomy within 18 hrs after death. This group served as control group for comparing the primary OA changes as primary OA occurs after 40 years of age. Individuals suffering from any disease of knee joint such as tumor, any deformity, bony enlargement, or irregularities such as osteophytes were excluded from the study.

A 5 μm thick serial paraffin sections were stained with haematoxylin and eosin [7] to confirm the histological structure of the articular cartilage and to study the various features for morphometry.

The equipment for morphometric analysis consisted of an Olympus microscope fitted with a digital camera and computer loaded with Image-Pro Express software. In all specimens, every sixth section was observed. At least five sections of each specimen and five fields from each section were observed and photomicrographed. The image acquired on the computer was analyzed using the Image-Pro Express software for the following:

(i) total thickness of the articular cartilage,
(ii) area of lacunae in each zone,
(iii) area of subchondral cavities,
(iv) number of chondrocytes per 10,000 μm² area in each zone.

For total thickness of the articular cartilage, sections were photomicrographed at 4x magnification, the software was calibrated for 4x magnification, and thickness was measured in millimeters using “measure the distance” option of the software.

For area of lacunae and subchondral cavities, sections were photomicrographed at 40x magnification. The software was calibrated for 40x magnification, and the area was measured in square microns by drawing the area to be measured using the area of interest (AOI) option of the software. Area of minimum of 1000 cells was measured for every specimen.

For measuring the number of chondrocytes per 10,000 μm², photomicrographs at 40x magnification were used. The software was calibrated for 40x magnification and the area was measured in square microns by selecting the area. Number of cells in that given area was counted using the “tag points” option of the software.

3. Statistical Analysis

SPSS software package was used for statistical analysis. Data was expressed as mean ± standard deviation. Independent sample t-test (Student’s t-test) was used to determine the statistical significance between the means. Analysis of variance (ANOVA) was done followed by the Student-Newman-Keuls post hoc tests. Partial correlation of various parameters with age was computed using Pearson’s correlation test. For all statistical tests, probability levels of less than or equal to 5% (two-tailed P value < 0.05) were considered to be significant.

4. Results

In group 1, that is, the OA cartilage, the articular surface appeared to be very irregular and fibrillated in all the specimens. The cartilage did not show the normal zonation in any of the specimens. Various changes seen were different in all specimens and they were not related to age. The most common feature of the OA articular cartilage was the presence of large clusters of cells (Figure 1). Total thickness of the articular cartilage ranged from 0.52 to 3.17 mm with an average of 2.02 ± 0.56 mm (Table 1).

Comparison of the mean of the total thickness at various age groups using ANOVA showed no statistically significant difference in the mean of the total thickness in the OA cartilage specimens. No significant correlation of the total thickness with age was observed.

The area of the lacunae (Table 2) and density of cells (Table 3) were measured. The lacunae were classified as small sized (<100 μm²), medium sized (100–200 μm²), and large sized (>200 μm²) according to area.
Table 3: Mean and standard deviation of the cell density (number of cells per 10,000 µm² area) in all four zones in OA group.

<table>
<thead>
<tr>
<th>Age Group</th>
<th>Zone 1</th>
<th>Zone 2</th>
<th>Zone 3</th>
<th>Zone 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>41–50 years</td>
<td>4.58 ± 1.05</td>
<td>2.33 ± 0.45</td>
<td>3.82 ± 0.24</td>
<td>3.86 ± 1.36</td>
</tr>
<tr>
<td>51–60 years</td>
<td>5.52 ± 3.59</td>
<td>3.26 ± 2.01</td>
<td>3.39 ± 1.84</td>
<td>3.01 ± 1.39</td>
</tr>
<tr>
<td>61–70 years</td>
<td>5.53 ± 2.04</td>
<td>2.49 ± 0.50</td>
<td>2.55 ± 0.53</td>
<td>2.33 ± 0.45</td>
</tr>
<tr>
<td>71–80 years</td>
<td>2.92 ± 0.95</td>
<td>2.28 ± 0.76</td>
<td>2.23 ± 0.33</td>
<td>2.82 ± 0.43</td>
</tr>
<tr>
<td>81–90 years</td>
<td>1.75 ± 1.21</td>
<td>1.65 ± 0.41</td>
<td>2.05 ± 0.66</td>
<td>1.68 ± 0.21</td>
</tr>
</tbody>
</table>

4.1. Superficial/Tangential Zone (Zone 1). The area of the lacunae ranged from 34.77 to 165.19 µm² with an average of 78.71 ± 31.12 µm². A 77.89% of the lacunae were small sized (<100 µm²) in area, 20.60% were medium sized (100–200 µm²), and 1.51% of the lacunae were large sized (>200 µm²). The density of cells, that is, number of cells per 10,000 µm² area, ranged from 1.75 to 13.79 with an average of 4.94 ± 2.66.

4.2. Intermediate/Transitional Zone (Zone 2). The area of the lacunae ranged from 88.55 to 224.64 µm² with an average of 137.52 ± 32.70 µm². A 28.93% of the lacunae were small sized, 59.38% were medium sized, and 11.69% of the lacunae were large sized. The density of cells ranged from 1.47 to 7.77 with an average of 2.68 ± 1.29.

4.3. Radiate/Deep Zone (Zone 3). The area of the lacunae ranged from 77.09 to 254.29 µm² with an average of 167.09 ± 55.56 µm². A 22.79% of the lacunae were small sized, 53.12% were medium sized, and 24.09% of the lacunae were large sized. The density of cells ranged from 1.43 to 7.77 with an average of 2.89 ± 1.22.

4.4. Deepest/Calcified Zone (Zone 4). The area of the lacunae ranged from 63.39 to 384.48 µm² with an average of 199.77 ± 76.03 µm². A 17.29% of the lacunae were small sized, 47.89% were medium sized, and 34.82% of the lacunae were large sized. The density of cells ranged from 1.24 to 5.66 with an average of 2.73 ± 1.03.

4.5. Subchondral Cavities. The area of the subchondral cavities varied from 1,551.02 to 9,53,428.90 µm² with an average of 65,701.07 ± 204,963.67 µm².

Comparison of the mean of various parameters at different age groups using ANOVA showed no statistically significant difference in the various age groups in the OA articular cartilage specimens while the area of lacunae in zone 1 was found to correlate significantly with the age (P < 0.05). In group 2, normal histology was seen, that is, four zones could be identified in all specimens, though changes were observed in the superficial zone either in the form of horizontal or oblique splitting of collagen bundles or complete loss of the most superficial zone.

4.6. Comparison of Group 1 and Group 2. On comparing the means using independent sample t-test (Student’s t-test), the means of lacunar size in all four zones were found to differ significantly in the OA (group 1) and control (group 2) (P < 0.05). Size of the lacunae in zone 1 and zone 4 was more in OA group while the size of the lacunae in zone 2 and zone 3 was higher in the control group (Table 4). On applying ANOVA, means of lacunar size in all four zones were found to differ significantly (P < 0.05) in group 1 and group 2. On applying post hoc test (Student-Newman-Keuls test), significant difference was found in group 1 (OA group) from the control group.

5. Discussion

OA is often described as a chronic age-related degenerative disease and is considered by many as an inevitable feature of growing old. Rather than OA being a simple consequence of joint ageing and repeated wear and tear, the current concept is that ageing of musculoskeletal system increases the susceptibility to OA but alone does not cause it [8]. The present study also supports the same view point that OA is not an inevitable feature of ageing. In a previous study [4], using computerized image analysis we compared the morphology of non-OA articular cartilage in below 40 and above 40 years of age and we did not observe much difference in the structure of articular cartilage with age. While a completely altered histology of OA articular cartilage prompted us to compare the parameters from non-OA cartilage above 40 years of age.
age (control group) with OA articular cartilage as primary osteoarthritis occurs after 40 years of age.

In the present study, structure of the OA articular cartilage was found to be much different from that of control group. The thickness of the OA cartilage was less than that in the control group but this difference was not significant. Articular cartilage thickness has been extensively studied in the knee [9–12]. However, most of these have been performed using radiographic imaging studies, specifically magnetic resonance imaging (MRI). According to Eckstein et al. [10], MRI of strongly curved surfaces can overestimate cartilage thickness without appropriate derivations. Bentley and Hill [13] observed a decrease in cartilage thickness with the increasing severity of OA. They stated that initial changes during OA stimulate a reactive hypertrophy that actually results in an increase in cartilage thickness prior to degradation. After the initial thickening, the articular surface begins to degrade under the influence of degrading enzymes hence causing a final decrease in thickness of the cartilage.

In the present study, the area of lacunae in each zone was measured using image analysis software and it was found to be significantly different in the control and OA group. Significant increase in the size of the lacunae in zone 1 in OA group could actually be a false increase in size. The superficial zone of cartilage is lost during osteoarthritis so these lacunae with increased size could actually belong to zone 2. Further insights into this observation are required. While an increase in the size of lacunae in zone 4 can be explained as the cells of this zone undergo hypertrophy during OA, decrease in the size of lacunae in zone 2 and 3 in OA could be because the cells form clusters and the newly formed cells could be occupying the smaller lacunae.

No significant difference was observed in the density of chondrocytes in any of the four zones. It could be due to proliferation of chondrocytes in the OA articular cartilage seen in the form of clusters of cells. So the number of cells degenerating due to disease could be almost equal to the newly formed cells forming clusters. This observation signifies that the OA articular cartilage makes an effort to repair but perhaps the regeneration is not at par to compensate for the degenerative changes.

Cake et al. [14] studied the development of OA in ovine knee joints using computer-assisted histomorphometric methods and stated that computerized image analysis showed significant histological differences not detectable by traditional scoring methods. Albona et al. [15] also used computerized image analysis to study the effect of betamethasone on articular cartilage of rabbit knee joints. In the present study, we used the computerized image analysis technique to compare the OA and non-OA articular cartilage in humans.

In the present study, we observed numerous differences in the histological appearance and histomorphometric features of the OA cartilage when compared with ageing cartilage. The various differences observed in the morphology of the OA and age-matched control articular cartilages suggest that OA should be considered a specific process and not simply an inevitable feature of ageing and the diseased articular cartilage makes an effort to repair itself but these regenerative changes are not at par with the degeneration. A better understanding of the ageing process will help in revealing new therapeutic targets to slow the disease progression.

**Conflict of Interests**

The authors declare that they have no conflict of interests.

**References**


