Prevalence of Fascioliasis in Cattle Slaughtered in Sokoto Metropolitan Abattoir, Sokoto, Nigeria

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The prevalence of fascioliasis in cattle slaughtered in the Sokoto metropolitan abattoir was investigated. Faeces and bile samples were collected and processed using formal ether concentration technique. Gross lesions from 224 out of 1,313 slaughtered cattle were randomly selected and examined. Out of the 224 cattle examined, 95 (42.41%) were males and 129 (57.59%) were females. Out of 95 male cattle examined, 27 (28.42%) were infected and out of 129 females 35 (27.13%) were infected. Based on breed, infection rates were 31 (31.0%) and 31 (25.2%) for breeds of Sokoto Gudali and Red Bororo respectively. No infection was recorded in White Fulani breed. Lesions observed were more in males than in females and more in Red Bororo than in Sokoto Gudali. Overall, prevalence of infection with Fasciola was 27.68%. There was no statistically significant association between infection and breed and between infection and sex of the animals sampled (P > 0.05). Regular treatment of all animals with an effective flukicide, as well as snail habitat control, tracing source of animals, public enlightenment about the disease, proper abattoir inspection, adequate and clean water supply to animals, and payment of compensation of condemned tissues and organs infested with the parasite by government were suggested.

1. Introduction

Fascioliasis, a serious infectious parasitic disease infecting domestic ruminants and humans, tops all the zoonotic helminthes worldwide [1]. A large variety of animals, such as sheep, goats, cattle, buffalo, horses, donkeys, camels and, rabbits, show infection rates that may reach 90% in some areas [2]. According to a World Health Organization (WHO) report in 2007 [3], the infection was limited in the past to specific and typical geographical areas (endemiotopes) but is now widespread throughout the world, with human cases being increasingly reported from Europe, the Americas, and Oceania (where only F. hepatica is transmitted) and from Africa and Asia (where the two species overlap). Fascioliasis is endemic in 61 countries and has become a food-borne infection of public health importance in parts of the world such as the Andean Highlands of Bolivia, Ecuador, and Peru; the Nile Delta of Egypt; and Northern Iran. It is estimated that more than 180 million people are at risk of infection, and infection rates are high enough to make fascioliasis a serious public health concern [4].

Bovine fasciolosis is a parasitic disease of cattle caused by trematodes usually Fasciola gigantica and rarely Fasciola hepatica in the tropics. The life cycle of these trematodes involves snail as an intermediate host [5]. The disease is usually characterized by a chronic, sometimes acute or subacute inflammation of the liver and bile ducts, accompanied by submandibular oedema, anaemia, anorexia, general intoxication, and death [6]. Meats infected by these organisms are regularly condemned at inspection in abattoirs/slaughter slabs. It could be zoonotic while constituting a major economic problem by lowering the productivity of cattle, in addition
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to losses from condemnation of affected organs. Humans can
accidentally ingest the eggs/larvae and become infected [7].
Fasciolosis is enzootic in Nigeria and is of great economic
importance especially in Northern Nigeria where stagnant
water and fadamas are used as watering and grazing sites
in the dry season. *Fasciola gigantica* is estimated to have
diverged from *F. hepatica* approximately 17 million years ago
[8] and penetrated more tropical regions in Asia and the Far
East where it is the predominant parasitic disease of cattle
and water buffalo [9]. *Fasciola hepatica* infects more than
300 million cattle and 250 million sheep worldwide and,
一起, with *F. gigantica*, causes significant economic losses
to global agriculture estimated to be more than US$3 billion
annually through lost productivity [10]. In West Africa,
Nigeria importantly, economic losses associated with
fasciolosis are great expenses on anthelmintics for treatment;
 liver condemnation; production loss due to mortality; lower
production of meat, milk, and wool; reduced weight gain;
metabolic diseases and impaired fertility [11, 12]. The value
of the losses resulting from this disease runs into millions of
naira [6]. However, the estimation of economic losses due to
fasciolosis at national and regional level is limited by lack of
accurate estimation of prevalence of the disease [13]. Apart
from direct economic losses associated with the disease, other
nonquantifiable losses are also experienced.

The prevalence of fasciolosis in cattle in many parts of
the world has been reviewed [14]. In Africa [14] quoted
prevalence rates of 37% in Sudan, 45% in Cameroon, 30–
90% in Ethiopia, 16% in Uganda, 62% in the Central Africa
Republic, and 50% in Rwanda [15]. In Kenya, a retrospective
study covering a period of 10 years (1990–1999) was carried
out using postmortem meat inspection records [15]. A total
of 5,421,188 cattle were slaughtered in the seven provinces
of Kenya during the 10-year period and 427,931 (8%) of
these cattle were infected with *Fasciola*. The region with the
highest prevalence of fasciolosis was Western Province (16%) followed, in descending order, by Eastern Province (11%),
Nyanza Province (9%), Rift Valley Province (8%), Central Province (6%), Nairobi Province (4%), and Coast Province (3.5%). The total economic loss incurred by the country
during the 10-year period as a result of condemnation of the
infected livers was approximately US$2.6 million [15].

In Nigeria, prevalence of fascioliasis has been reported
from different parts of the country. There are also several
reports on the prevalence, regional incidence, and seasonal
variation for bovine fasciolosis [16, 17]. From slaughtered
cattle in Maiduguri, Biu et al. [7] reported a prevalence of
80%, from Zaria, Northwest, and a prevalence of 65.4% was
also reported by [18]. A study was conducted to evaluate the
number of cattle slaughtered, prevalence, seasonal variation,
and socioeconomic consequence of bovine fasciolosis at the
Ibadan municipal abattoir, Ibadan, Oyo State, Nigeria,
between 1994 and 2004, based on retrospective abattoir meat
inspection records and a perspective meat inspection survey
[19]. Of the 1,640,095 cattle slaughtered in 11 years, 37,828
livers were condemned due to fasciolosis, translating into a
prevalence of 2.31%. The incidence observed among males 172
(4.24%) was slightly higher than that noted for females 452
(3.73%) though significant at 0.05 level of error. Moreover,
the prevalence recorded in the dry season (October–March)
was 19,816 (2.58%) and 18,012 (2.07%) for the rainy season
(April–September) with a fairly strong positive correlation
(+0.76) occurring between the incidences of the disease in
the seasons [18]. Another study was carried out between July
and October 2012, aimed at determining the prevalence of
fascioliasis and the economic loss of condemned liver due to
*Fasciola* infection in cattle slaughtered at three abattoirs in
Eket Urban, Akwa Ibom State, Nigeria [20]. A total of 279
cattle consisting of 185 males and 94 females were examined.
The livers were examined for the adult flukes while the faecal
samples were examined for the eggs of *Fasciola*. The result
of the investigation showed that 38 (13.62%) of the cattle
were infected with fascioliasis. The prevalence rate recorded
for female cattle was 17.02% compared to the male with
prevalence rate of 11.89% [20].

This study would serve to recognize and appreciate the
magnitude of problems and losses posed by fascioliasis and
also fill the knowledge gaps (additional reference information)
and update existing information on bovine fascioliasis
in Sokoto State and Nigeria at large. This will eventually
stimulate the interest to intensify efforts on its monitoring
and surveillance towards the control and possible eradication
of the disease in Nigeria.

2. Materials and Methods

2.1. Study Area. Sokoto State is geographically located at the
North Western part of Nigeria between longitude 11° 30’ to
13° 50’ East and latitude 4° to 6° 40’ North. The state shares
common borders with Niger Republic to the North, Kebbi
State to the South, and Zamfara State to the East [21]. The
state falls in the dry Sahel surrounded by sandy Sudan type
Savannah [22].

The state, a major livestock producer lying in the arid
region of the country, covers a total land area of about
32,000 square km with an estimated human population of
3,696,999 million [21]. There are 23 local government areas
in the state and the populace is predominantly farmers.
Because of their spare arable land, a good percentage of the
populace is engaged in animal husbandry. The annual mean
livestock population of the former Sokoto State (including
Kebbi and Zamfara States which were carved out from the
former Sokoto State in 1991 and 1996, resp.) for cattle was
estimated to be 1,772,830 (17,290 density/km
2
), 2,466,484 (24,055 density/km
2
) for goats, 2,566,246 (25,028 density/km
2
) for sheep, 43,960 (0.429 density/km
2
) for camels, and 109,484 (1.068 density/km
2
) for dogs [22].
Rainfall starts late in April and ends early in September
with a mean rainfall ranging between 500 mm and 1,300 mm.
There are 2 main seasons in the state, namely, wet and dry
seasons. The dry season starts from October and lasts up to
April and the wet season begins in most parts of the state in
May and lasts up to September or October [22].

2.2. Sample Collection. The postmortem investigation was
conducted within the month of July 2010, by randomly
examining 224 out of 1,313 cattle slaughtered at the municipal
abattoir, Sokoto.
Faecal materials were collected into polythene bags directly from the rectum of each of the cattle being sampled after they have been slaughtered. Gloved hands were used in faecal sample collections. Bile of each of the cattle used for the study was also collected from the gall bladder. The samples (bile and faeces) per cattle were each placed in polythene bags and labeled for proper identification. The lesions and samples for corresponding cattle, as well as the breed, sex, were also recorded. Samples were preserved in a refrigerator and were processed within 24 hours.

3. Sample Processing

3.1. Faecal Sample. Two grams (2 g) of faeces was collected into labeled test tubes containing 3 mLs of distilled water. The faecal samples and the distilled water were strained to give a suspension. The suspension was strained through a tea strainer into a corresponding cleaned labeled Petri dish. The filtrate was poured into corresponding test tubes. One milliliter (1 mL) of 10% formalin was added into the test tubes which were allowed to stand for 5 minutes. Diethyl-ether (1 mL) was added in the test tubes after 5 minutes, using different 18-gauge hypodermic needle and syringe. The test tubes containing the suspension were then corked, shaken to mix, and centrifuged at 2000 rpm for 8 minutes. The eggs and cysts of the parasites sediment at the bottom of the mixture, while diethyl-ether with some fat comes up as debris becomes separated in a layer between the diethyl-ether and water. The supernatant was then decanted leaving few of it with the sediment. Drops (1-2) of the sediment were put on a glass slide, covered with cover slip, and viewed under microscope using ×100 magnification.

3.2. Bile Sample. From the gall bladder, 2 mL of the bile was collected using 18-gauge hypodermic needle. The bile sample was then poured into a labeled test tube in a test tube rack. 1 mL of 10% formalin was added using 18-gauge hypodermic needle and syringe into the bile sample and then allowed to stand for 5 minutes. Diethyl-ether (1 mL) was then added in the test tube after 5 minutes using a different 18-gauge hypodermic needle and syringe. The test tube containing the solution was then corked and shaken to mix the solution. The solution was then centrifuged at 2000 rpm for 10 minutes. The eggs/cysts of the parasites sediment at the bottom of the mixture, while diethyl-ether with some fat comes up as supernatant. The supernatant was decanted leaving few of it with the sediment. Drops (1-2) of the sediment were put on a glass slide and covered with a cover slip and viewed under microscope using ×100 magnification [23].

4. Results

Out of a total of 1,313 cattle slaughtered during the period of study, 224 were randomly selected. Out of the 224 sampled cattle, 62 (27.68%) were infected. Ninety-five (95) of these were males with infection rate of 27 (28.42%), while 129 were females with infection rate of 35 (27.13%) (Table 1).

Out of 100 Sokoto Gudali breeds examined 31 (31%) were infected, and out of 123 of Red Bororo breeds examined 31 (25.2%) were found infected, while 1 of White Fulani breed was free of the disease (Table 2).

Statistical analysis of the data revealed no statistically significant association (P > 0.05) between infection with the parasite and breed and sex of the sampled animals (Tables 1 and 2).

5. Discussion

The study has revealed the prevalence of fascioliasis in cattle slaughtered in the Sokoto metropolitan abattoir, Sokoto, Nigeria. Disease prevalence was found to be more in males than in females. This is in accordance with the findings of [24, 25]. The infection was again more in Sokoto Gudali than in Red Bororo breed and the White Fulani breed of cattle.

The disparity in susceptibility to helminthic infection between the 2 sexes could be attributed to the differences in the host intrinsic factors (genetics, physiology, and immunology) and extrinsic factors (environment and management practices). The lesions observed are more in males than in females and more common in Red Bororo than in Sokoto Gudali probably due to differences in the number of animals examined.

The prevalence of fascioliasis among cattle recorded in this research work can be attributed to the climatic conditions of this location which does not favour the survival of the intermediate hosts, the snail. This intermediate host prefers swampy areas with slowly moving water and small streams which also allow sufficient moisture for the survival of the infective metacercariae. In contrast, the study area, Sokoto, occupies low flat and naturally dry land (semiarid area). This probably explains the low percentage of infection with fascioliasis among cattle slaughtered in the abattoir.

The general picture of fascioliasis in the study area shows that there was no significant difference in infection between male and female cattle (P > 0.05). This is in sharp contrast with the studies earlier reported by [26, 27], who reported a higher infection rate among the male cattle than the female cattle from Gwagwalada abattoir, Federal Capital Territory, Abuja, Nigeria, and Jalingo abattoir, Taraba state, Nigeria.
respectively. The possible explanation could be that both sexes move together in search of food and water and therefore possibility for both sexes to be equally exposed to the risk of infection is high [26].

6. Conclusion

The prevalence of fascioliasis in cattle revealed by this work highlights the infection as being more common in male than in female and more common in Sokoto Gudali breed than in Red Bororo breed of cattle, with no statistically significant difference between rate of infection in males and in females and also between breeds.

Recommendations

Given the reported prevalence of fascioliasis in Sokoto metropolitan abattoir, a control program in this region appeared justified. The following recommendations are advocated for effective control program: public enlightenment about the disease and the role of snails in the life cycle of the parasite as well as associated health risks in animals and humans; artificial pasture land (rangeland system) that seems to be a good panacea in control of the disease in cattle; development of well-defined interval for deworming of cattle, especially newly purchased ones with effective drugs such as albendazole and praziquantel; proper meat inspection (with appropriate compensation for condemned animals or their parts) that should be revisited and properly enforced; abattoir record keeping that should be reviewed to provide information on livestock diseases.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References


