

## Supplementary file S1

### cDNA synthesis

#### **Table 1. Protocol of cDNA synthesis procedure.**

“Transcriptor First-Strand cDNA Synthesis Kit” Roche Diagnostics (Mannheim, Germany) (Cat. No.04896866001).

Components		Volume and concentration
Total RNA		10µL (1µg)
Anchored-oligo (dT) primer		1µL (2.5µM)
Random hexamer primer		2µL (60µM)
cDNA synthesis mix		Volume and concentration
Transcriptor Reverse Transcriptase		4µL (1×) (8mM MgCl <sub>2</sub> )
Protector RNase Inhibitor		0.5µL (20U)
Deoxynucleotide Mix, 10 mM		2µL (1mM)
Transcriptor Reverse Transcriptase		0.5µL (10U)

#### **Table 2. Cycling protocol for reverse transcription of mRNA.**

Steps	Preparation	
	Time	Temperature
Denaturation	15 min	65°C
	∞	4°C
cDNA synthesis		
Annealing	10 min	25°C
Elongation	30 min	55°C
Inactivation	5 min	85°C
	∞	4°C

### Quantitative real-time PCR (RT-qPCR).

#### **Table 3. Primers and UPL probe for target gene.**

Gene name/ accession number	Forward primers	Reverse primers	UPL probe	Amplicon (nt)
GADD45A NM_001924.3 variant 1	TTTGCAATATGACTTTGGAGGA	CATCCCCACCTTATCCAT	19	72
SARS NM_006513.2	TGGGCAAACCAAGAAGATG	GCAGATGGTACGGGTAGTGG	39	85
IFI6	CTGTGCCCATCTATCAGCAG	GGGCTCCGTCCTAGACCTT	41	75

NM_022873.2 variant 3	SPRY4	CCCCGGCTTCAGGATTTA	CTGCAAACCGCTCAATACAG	17	85
NM_030964.3 variant 1	GSTP1	TCTCCCTCATCTACACCAACTATG	AGGTCTTGCCTCCCTGGT	56	114
NM_000852.3					

**Table 4. Protocol of reaction mix using cDNAs and LightCycler480 Probes Master kit.**

Probe Master kit (Cat. No.04707494001) and UPL probe from Roche Diagnostics GmbH (Mannheim, Germany). White 96-well PCR-plates (Cat. No.04729692001). Amplification primers for candidate genes were purchased from TIB MOLBIOL Berlin.

	Volume
H <sub>2</sub> O	3.4µL
Upstream-Primer (10µM)	0.25µL
Downstream-Primer (10µM)	0.25µL
Probe (Universal Probe Library)	0.1µL
Master Mix (2 x conc.) <sup>1</sup>	5µL
cDNA template	1µL
Total volume	10µl

2× conc., ready-to-use hot-start PCR mix, contains FastStart Taq DNA Polymerase, reaction buffer, dNTP mix (with dUTP instead of dTTP), and 6.4 mM MgCl<sub>2</sub>.

**Table 5. Cycling protocol for relative quantification on LC480.**

Program	Temperature	Hold
Pre-incubation (1 cycle)	95°C	10:00 min
	95°C	00:10 sec
Amplification (45 cycles)	59°C	00:20 sec
	72°C	00:01 sec
Cooling (1 cycle)	40°C	00:30 sec

\* For each cycle Mono-color FAM fluorescence measurements with wave length of 483-533nm was used.

The analytical precision of the qPCRs (the standard deviation of the Ct values) was tested by intra-run (n=10) measurements for gene FABP6. Ct value ranged between 22.65 and 22.78, median-22.67, SD±0.044, SD% 0.19.

### **Reference genes.**

Commercially available mRNA-specific Roche Diagnostics GmbH Ready-to-Use reference genes assays were used to detect of expression of reference gene PBGD (Cat.No.05046149001), HPRT1 (Cat.No.05046157001) and TBP (Cat.No.05189284001). Further analyses for assessing their contribution as normalizer and compared suitability of housekeeping genes were done by geNorm<sup>PLUS</sup> software (Biogazelle, Belgium). In our case gene with the least expression level was TBP (M=0.6850) and the most stable one was PBGD (M=0.5950). Average stability level showed gene HPRT1 with M value 0.6225.