Aerogels are a series of materials with porous structure and light weight which can be applied to many industrial divisions as insulators, sensors, absorbents, and cushions. In this study, cellulose-based aerogels (aerocelluloses) were prepared from cellulosic material (microcrystalline cellulose) in sodium hydroxide/water solvent system followed by supercritical drying operation. The average specific surface area of aerocelluloses was 124 m$^2$/g. The nitrogen gas (N$_2$) adsorption/desorption isotherms revealed type H1 hysteresis loops for aerocelluloses, suggesting that aerocelluloses may possess a porous structure with cylindrically shaped pores open on both ends. FTIR and XRD analyses showed that the crystallinity of aerocelluloses was significantly decreased as compared to microcrystalline cellulose and that aerocelluloses exhibited a crystalline structure of cellulose II as compared to microcrystalline cellulose (cellulose I). To perform cationic functionalization, a cationic agent, (3-chloro-2-hydroxypropyl) trimethylammonium chloride, was used to introduce positively charged sites on aerocelluloses. The cationized aerocelluloses exhibited a strong ability to remove anionic dyes from wastewater. Highly porous and low cost aerocelluloses prepared in this study would be also promising as a fast absorbent for environmental pollutants.

1. Introduction

Aerogels are materials having a remarkable porous structure with pores filled with air rather than liquid [1]. Typical aerogels generally present a three-dimensional network exhibiting extremely light weight and high surface area. Various materials can be used to prepare aerogels, one of which is a group of inorganic aerogels, such as SiO$_2$, Al$_2$O$_3$, TiO$_2$, ZrO$_2$, MgO, and their composites, which have been extensively studied [2, 3]. The preparation of aerogels involves a "sol-gel" process described by a series of successive steps including solvent dispersion of polymers or particles, gelation of material mixture under thermal or mechanical condition, and supercritical dehydration [4]. Aerogels have been broadly employed as thermal insulators, catalysts, sensors, absorbents, drug carriers, and other applications due to their appealing properties [5–9].

Cellulose can vary from 300 to 15,000 depending on the source of cellulose [10, 11]. Cellulose is stable, nontoxic, biodegradable, and inexpensive. Due to its high molecular weight and stable crystalline structure, cellulose is very difficult to dissolve in most common solvents. Only a few solvents, such as sodium hydroxide/urea/water, lithium chloride/N,N-dimethylacetamide (LiCl/DMAc), N-methyl-morpholine-N-oxide (NMMO), and ionic liquids (ILs), are able to swell and dissolve cellulose [12]. The effective dissolution of cellulose macromolecules is the predominant step to prepare cellulose-based aerogels (aerocelluloses) and develop value-added materials with well-defined architectures.

Aerocelluloses are one of the most valuable and environmentally friendly products that are derived from cellulosic materials. They were first developed using cellulose acetates dissolved in acetone followed by gelation and supercritical drying processes and the resultant aerocelluloses exhibited a fairly high specific surface area (~300 m$^2$/g) [13]. Other aerocelluloses were developed, form microcrystalline cellulose, by dissolving it in calcium thiocyanate, followed by...
being gelatinized and dehydrated by solvent exchange drying to achieve a porous structure with a specific surface area of 190 m$^2$/g [14]. By using supercritical drying method and the NMMO solvent, the aerocelluloses prepared from microcrystalline cellulose could achieve a higher specific surface area (400 m$^2$/g) [15]. Likewise, dissolving microcrystalline cellulose in NaOH/water solvent system led to aerocelluloses with a high specific surface area (300 m$^2$/g) and a low density (0.06 g/cm$^3$) [16]. In addition to supercritical drying, the freeze-drying was also used to prepare porous cellulose materials which can be denoted as cryogels. By using both calcium thiocyanate and LiCl/DMAC solvents, the surface area of cellulose cryogels could achieve around 160 m$^2$/g [17]. Both aerocelluloses and cryogels could exhibit a light weight ranging from 116 to 350 kg/cm$^3$ [18].

In this study, we prepared aerocelluloses using a “sol-gel” process by dissolving MCC in NaOH/water solvent system followed by gelation and supercritical drying operations. Furthermore, we employed (3-chloro-2-hydroxypropyl)trimethylammonium chloride to functionalize the as-prepared aerocelluloses to immobilize negatively charged dye molecules for wastewater clarification. Wastewater containing dye-contaminating molecules can lead to serious environmental problems [19]. Various methods have been developed to remove dye molecules from wastewater [20, 21]. Some biogenerated polymers have been also used as active reagents for wastewater treatment, such as starch-derived carbon aerogels, chitosan, and its derivatives [19, 22]. Due to the high surface area of aerocelluloses as well as the low cost, cationized aerocelluloses could be a candidate for wastewater clarification.

2. Experimental

2.1. Materials. Cellulose (Microcrystalline Cellulose, MCC, Avicel®, PH-102 NF) was obtained from FMC BioPolymer (Newark, DE, USA). Sodium hydroxide (NaOH, Cat. number S318-500) was purchased from Fisher Chemical (Fisher Scientific, USA) and sulfuric acid (H$_2$SO$_4$, Cat. number LC255503) was purchased from LabChem (Zelienople, PA, USA). CR-2000 (3-chloro-2-hydroxypropyl trimethylammonium chloride, CAS number 0003327-22-8) was obtained from Dow Chemical (Midland, MI, USA) and abbreviated as CHPTAC. Color Index Reactive Blue 19 (CI RB19) dye (Dye number 472308 1/0) was purchased from Organic Dyestuffs Corporation (East Providence, RI, USA). All chemicals were used without further purification except H$_2$SO$_4$, which was bone dried to ensure its purity of 99.9% with lower than 10 ppb of water prior to use.

2.2. Preparation of Aerocelluloses. Cellulose was dissolved in NaOH/water solvent system at different concentrations (5, 9, and 11%, w/w) as shown in Table 1. NaOH was first dissolved in deionized (DI) water and precooled at -12°C for 2 h and cellulose was dispersed in DI water and stirred at 5°C for 2 h. Next, the cellulose/water solution was mixed with NaOH solution and the mixture was placed in -12°C freezer. The mixture was stirred for 30 s every 5 min for 1 h to homogenize the mixture. The resulting cellulose/NaOH/water mixture was transferred to cylindrical molds with an approximate dimension of 16–20 mm diameter and 30–40 mm high, and placed in an oven at 50°C for 2 h to form a gel. The cylindrical gels were then regenerated in distilled water, neutralized with 0.1 M H$_2$SO$_4$, and rinsed with DI water to remove sulfuric acid. The solvent exchange of gels with pure acetone was performed once every 2 h for 48 h to ensure complete replacement of water with acetone. Afterwards, the carbon dioxide (CO$_2$) supercritical drying was used to dehydrate the gels in a Jumbo Critical Point Dryer (SPI Supplies, West Chester, PA, USA) equipped with a water jacket for cooling and heating (Thermo Scientific NesLab-RTE 10, Newington, NH, USA). The supercritical drying was first performed at 10°C under 5 MPa with liquid CO$_2$, and the excess acetone was purged intermittently for 30 s at the same pressure and temperature until the gels were filled with liquefied CO$_2$ instead of acetone. Next, the supercritical drying system was pressurized to 8 MPa and the temperature was increased to 35°C to dehydrate the gels for approximately 3 h. When the total amount of the interstitial liquefied CO$_2$ was removed, the system was slowly depressurized and the supercritical dryer was cooled down to collect the aerocelluloses.

2.3. Material Characterization. The aerocelluloses were characterized by Scanning Electron Microscopy (SEM) Surface Porous Profile Analysis, Fourier Transform Infrared Spectroscopy (FTIR), Thermogravimetric Analysis (TGA), and X-ray Diffraction (XRD). SEM observation of the aerocelluloses was performed in an environmental SEM (Hitachi TM-1000, Pleasanton, CA, USA). SEM samples were placed on carbon tape attached to the SEM sample stage. The cross sections of aerocelluloses prepared by 5%, 9%, and 11% cellulose concentrations were analyzed.

The pore size, the pore size distribution, and the specific surface area measurements were performed by Gemini 2390t (Micromeritics, Norcross, GA). All the data were obtained using Brunauer-Emmett-Teller (BET) surface area analysis and Barrett-Joyner-Halenda (BJH) pore size and volume analysis. The samples were ground in the mortar by a pestle, and then they were degassed at 90°C for 22 h. The data were analyzed based on the N$_2$ adsorption/desorption isotherms, surface area parameters, and curves of pore size versus pore volume of different aerocellulose samples. FTIR equipped with a Universal Attenuated Total Reflectance (ATR) accessory (Perkin Elmer Spectrum-400, Waltham, MA) was used to acquire the FTIR spectra of aerocellulose samples. Cellulose and aerocellulose samples were first placed in an environmentally controlled laboratory maintained at a relative humidity of 65 ± 2% and 21 ± 1°C at least 48 h prior to analysis. The FTIR spectra were

| Table 1: Mass ratio of cellulose and solvent system (w/w) used for dissolution. |
|---------------------------|-----------|-----------|-----------|
| Cellulose% | 5 | 9 | 11 |
| Avicel (g) | 5.0 | 9.0 | 11.0 |
| NaOH (g) | 7.6 | 7.6 | 7.6 |
| Water (g) | 87.4 | 83.4 | 81.4 |
Scheme 1: Schematic reaction route of cationic functionalization of aerocelluloses: (a) (3-chloro-2-hydroxypropyl) trimethylammonium chloride (CHPTAC); (b) 2,3-epoxypropyltrimethyl ammonium chloride (EPTAC).

then recorded at a spectra resolution of 4 cm$^{-1}$ with 32 coadded scans over the range from 650 to 4000 cm$^{-1}$. PerkinElmer software was used to perform spectra normalization, baseline corrections, and peak integration. FTIR spectra were also subjected to Principal Component Analysis (PCA) with leverage correction and mean-center cross validation boxes checked using Unscrambler V. 9.6 Camo Software AS (CAMO Software AS, Norway).

Thermogravimetric analysis of cellulose and aerocellulose samples were performed using Pyris1 TGA (PerkinElmer Sheltong, CT, USA). Samples were also placed in an environmentally controlled laboratory at relative humidity of 65 ± 2% and 21 ± 1°C at least 48 h prior to testing. Thermograms were recorded between 37°C and 600°C with a heating rate of 10°C/min in a flow of nitrogen at 20 mL/min. Pyris software was used to calculate the first derivatives of the thermograms (DTC), the percentages of weight loss, and the decomposition temperatures.

The crystalline structures of cellulose and aerocellulose samples were analyzed using SmartLab system (Rigaku Corporation, Model HD 2711N). XRD was performed using CuKα radiation ($\lambda = 0.15406$) generated at 40 kV and 44 mA. Scans were conducted between 10° and 50° two-theta in 2° stepwise per minute.

2.4. Cationic Functionalization of Aerocelluloses. Aerocellulose samples were placed in beakers with 30 mL of DI water containing 65% (w/w) of the cationic reagent (1.05 g CR-2000). The beaker was placed on the bunch at room temperature for 5 days to allow the cationic reagent to diffuse inside the aerocellulose network. Next, 0.5 g of NaOH was added to the solution containing aerocelluloses and the cationic reagent, and the mixture was stirred until it was homogenous and remained overnight. The cationized aerocelluloses were rinsed with DI water to ensure complete removal of unreacted cationic reagent and then air-dried at room temperature. The schematic route of cationic functionalization of aerocelluloses is shown in Scheme 1. To test the absorbability of dye, the aerocellulose and cationized aerocelluloses prepared from 5% and 11% cellulose concentrations were placed in 20 mL of DI water containing 0.1 g/L of CI RB19 dye. The absorbance of the solution was measured using Lambda 650 UV/Vis Spectrometer (PerkinElmer, USA) at 650 nm.

2.5. Statistical Analysis. Experimental runs of all samples of cellulose and aerocelluloses were repeated at least for three times. The significant differences between sample groups were calculated using One-Way ANOVA (LSD, Least Significant Difference) ($p < 0.05$ confident interval; IBM SPSS Statistical Software; Release 19.0.0).

3. Results and Discussion

3.1. Preparation and Material Characterization of Aerocelluloses. Aerocelluloses prepared from different cellulose concentrations are illustrated in Figure 1. The aerocellulose prepared from 5% cellulose concentration appears to be fluffier than aerocelluloses prepared from 9% and 11% cellulose concentrations, suggesting that 5% aerocellulose may have a more porous structure. SEM images of cross section...
Figure 1: Photographs of aerocelluloses prepared from 5%, 9%, and 11% cellulose concentrations.

Figure 2: SEM images of cross section of aerocelluloses prepared from different cellulose concentrations: (a-b) 5%, (c-d) 9%, and (e-f) 11%. Images of (b), (d), and (f) are images at a higher magnification of (a), (c), and (e), respectively.

of aerocelluloses are shown in Figure 2. At low magnification, aerocelluloses prepared from 5% and 9% cellulose concentrations have similar porous morphologies, whereas aerocellulose from 5% cellulose concentration exhibits more uniform pores aligned in an ordered arrangement that can be observed at high magnification (Figure 2(b)). Comparably, the aerocellulose prepared from 11% cellulose concentration exhibits a more compact structure with much more irregular pores shown at low magnification and more open pores shown at high magnifications. It is noticed that some visible
chunks and small white aggregates are present in the aerocellulose from 11% cellulose concentration, suggesting that cellulose in this case was not well dissolved, which led to inhomogeneous cellulose solution to prepare aerocellulose at this high concentration. More porous structure generally enables aerocelluloses to obtain ultra-light weight and high specific surface area, and thereby 5% cellulose concentration may be the best option to prepare aerocelluloses with satisfactory porosity in this study.

Figure 3 shows $N_2$ adsorption/desorption isotherms for aerocelluloses prepared from 5%, 9%, and 11% cellulose concentrations. The isotherms for three aerocellulose samples all exhibit type IV characteristics. The H1 hysteresis loops present in these isotherms reveal that both capillary condensation and evaporation occurred in a cylindrical channel, suggesting that many cylindrically shaped pores open at both ends [23–25]. Accordingly, the aerocelluloses prepared in this study are considered mesoporous materials with a pore diameter of 2–50 nm. Table 2 summarizes the variance analysis of the surface area of aerocelluloses. These aerocelluloses exhibit high BET surface areas varying from 80 to 125 m$^2$/g. The average BET surface area of aerocelluloses prepared from 5% cellulose concentration is 124 m$^2$/g, which is significantly higher than aerocelluloses prepared from 9% and 11% cellulose concentrations ($p < 0.05$). The pore size distributions of aerocelluloses prepared from different cellulose concentrations are shown in Figure 4 and Table 3. The aerocelluloses prepared from 5% cellulose concentration exhibit higher pore volume corresponding to a higher surface area than aerocelluloses prepared from 9% and 11% cellulose concentrations. The statistical analysis shows that the amount of cellulose dissolved in the solvent system plays a key role in the surface area of the aerocellulose. The low surface area of aerocelluloses prepared from 9% and 11% cellulose concentrations may be attributed to the incomplete dissolution of cellulose at high cellulose concentrations.

The FTIR spectra of cellulose and aerocelluloses exhibit different structural characteristics (Figure 5). The vibration at 3292 cm$^{-1}$ is assigned to intermolecular hydrogen bonding (C3-(OH)-C6 (O)) while the vibration at 3335 cm$^{-1}$ is due to intramolecular hydrogen bonding (O (3) H-O (5)), which are only present in the FTIR spectrum of cellulose [26]. The conversion from MCC to aerocelluloses is accompanied by a shift of these vibrations to 3443 and 3481 cm$^{-1}$. The vibration at 3443 cm$^{-1}$ is assigned to intramolecular hydrogen bonding (O (2) H-O (6)) and the vibration at 3481 cm$^{-1}$ is assigned to the intramolecular hydrogen bonding in cellulose II [26, 27]. The disappearance of the vibrations at 3292 and 3335 cm$^{-1}$
Table 2: The average and multicomparisons of BET and Langmuir surfaces.

<table>
<thead>
<tr>
<th>Variable</th>
<th>5% aerocelluloses</th>
<th>9% aerocelluloses</th>
<th>11% aerocelluloses</th>
</tr>
</thead>
<tbody>
<tr>
<td>$S_{BET}$ single (m$^2$/g)</td>
<td>120.61 (A)</td>
<td>77.61 (B)</td>
<td>97.09 (AB)</td>
</tr>
<tr>
<td>$S_{BET}$ (m$^2$/g)</td>
<td>124.42 (A)</td>
<td>80.01 (B)</td>
<td>100.17 (AB)</td>
</tr>
<tr>
<td>$S_{Langmuir}$ (m$^2$/g)</td>
<td>160.42 (A)</td>
<td>103.13 (B)</td>
<td>129.17 (AB)</td>
</tr>
</tbody>
</table>

Note: $S_{BET}$ single: surface area determined by BET method at single point $P/P_0 = 0.199$; $S_{BET}$: surface area determined by BET method; $S_{Langmuir}$: surface area determined by Langmuir method; A, B, and C: groups in multicomparisons.

Table 3: The average and multicomparisons of the pore volumes and pore diameters.

<table>
<thead>
<tr>
<th>Variable</th>
<th>5% aerocellulose</th>
<th>9% aerocellulose</th>
<th>11% aerocellulose</th>
</tr>
</thead>
<tbody>
<tr>
<td>$S_{pore}$ BJH (m$^2$/g)</td>
<td>113.93 (A)</td>
<td>75.73 (B)</td>
<td>97.65 (AB)</td>
</tr>
<tr>
<td>Cumulative $V_{BJH}$ (cm$^3$/g)</td>
<td>0.4546 (A)</td>
<td>0.2798 (B)</td>
<td>0.3249 (B)</td>
</tr>
<tr>
<td>Av. pore diameter (Å)</td>
<td>138.08 (A)</td>
<td>123.63 (A)</td>
<td>118.92 (A)</td>
</tr>
</tbody>
</table>

Note: $S_{pore}$ BJH: surface area of pores determined by the BJH method. Cumulative $V_{BJH}$: BJH adsorption cumulative volume of pores; Av. pore diameter: BJH desorption average pore diameter; A, B, and C: groups in multicomparisons.

Figure 4: Pore size distributions of aerocelluloses prepared from 5%, 9%, and 11% cellulose concentrations.

Figure 5: FTIR spectra of cellulose and aerocelluloses prepared from 5%, 9%, and 11% cellulose concentrations.

and the appearance of the vibrations at 3443 and 3481 cm$^{-1}$ indicate the transformation of the crystalline structure from cellulose to aerocellulose. The peak at 1429 cm$^{-1}$ is assigned to CH$_2$ of the crystalline cellulose [28], and its intensity significantly decreased from cellulose to aerocelluloses, suggesting that the conversion of cellulose to aerocelluloses may result in a decrease of crystallinity. The vibration at 1315 cm$^{-1}$ is due to CH$_2$ wagging while the vibration at 1337 cm$^{-1}$ is assigned to C-OH in plane bending [29], which are characteristics of cellulose I. The decrease of the intensity of the peaks at 1315 and 1337 cm$^{-1}$ for aerocelluloses further demonstrates the transition from cellulose I for cellulose to cellulose II for aerocelluloses. The intensity of the peak at 1161 cm$^{-1}$ that slightly decreased in the FTIR spectra of aerocelluloses, associated with the antisymmetric bridge C$_1$-O-C$_4$ stretching, is characteristic of cellulose I [30]. The intensity of the peak at 1104 cm$^{-1}$ associated with the antisymmetric in-plane ring stretching mode decreased in the FTIR spectra of aerocelluloses, suggesting the transformation of cellulose I to cellulose II [30]. The increase in the intensity of the peak at 898 cm$^{-1}$, which is assigned to C-O-C stretching ($\beta$-(1-4)-glycosidic linkage) for aerocelluloses, also suggests the reduction of crystallinity during the conversion from cellulose to aerocelluloses [31]. Due to the transformation of the crystalline structure from cellulose I to cellulose II, the CH$_2$ rocking vibration located at 710 cm$^{-1}$ related to crystalline cellulose I$_\beta$ is less intense in aerocelluloses [30], which suggests a reduction in the crystallinity of aerocelluloses as compared to cellulose.

Principal Components Analysis (PCA) is applied to reduce the dimensionality of the original spectral data from
thousands of variables (3,350 wavenumbers) to fewer dimensions. The variability in each spectrum regarding the mean of the population presumably represents a smaller set of values (axes), termed as principal components (PCs) (Figure 6); the main sources of variability in the data are concentrated into the first two PCs (PC1 and PC2) that are plotted against each other. PC1 accounts for 80% of the variance and clearly summarizes variables of the FTIR spectra into two groups: Group 1 for aerocelluloses and Group 2 for cellulose; these two distinct categories further demonstrate the significant transformation of the crystalline structure from cellulose I for microcrystalline cellulose to cellulose II for aerocelluloses.

The thermograms in Figure 7(a) show two regions of thermal decomposition. The weight loss in region I from 37°C to 150°C is attributed to the loss of adsorbed water while the weight loss in region II from 150°C to 425°C is attributed to the decomposition of cellulose. Statistical analysis shows that the amount of adsorbed water for aerocelluloses is higher than cellulose, which suggests a higher surface area of aerocelluloses (80–125 m²/g) than cellulose with a low surface area (~1 m²/g). The derivative weight loss curves show two major peaks of thermal decomposition (Figure 7(b)). The peak around 47°C is associated with maximum desorption rate of water (weight%/min) and the peak around 380°C is associated with the maximum decomposition rate of cellulose [32, 33]. The water evaporation temperature at the maximum rate for cellulose is significantly lower than aerocelluloses (p < 0.05), suggesting that water in cellulose is easier to desorb from cellulose while porous aerocelluloses exhibited a better capability of holding water. Furthermore, the cellulose decomposition temperature at the maximum rate for cellulose is similar to aerocellulose prepared from 5% cellulose concentration while aerocelluloses prepared from 9% and 11% cellulose concentrations exhibit higher decomposition temperatures. This suggests that aerocelluloses are thermostable structure compared to cellulose.

The XRD pattern of cellulose in Figure 8 shows 4 major characteristic peaks of cellulose I at 14.9°, 15.8°, 22.5°, and 34.6°, which are assigned, respectively, to 1–10, 110, 200, and 004 [34]. However, the peaks at 20.0° and 21.6° in XRD patterns of aerocelluloses show a typical cellulose II pattern. The crystallite index (CI) is calculated using the following
equation: \( CI = \frac{(I_{200} - I_{am})}{I_{200}} \times 100 \), where \( I_{200} \) is the peak intensity at \( 2\theta = 22.5^\circ \) and \( I_{am} \) is the intensity of the peak at \( 2\theta = 18^\circ \) [35]. It is noticed that there is a significant difference of CIs between cellulose and aerocelluloses \( (p < 0.05) \). These results are consistent with FTIR analysis above.

3.2. Cationic Functionalization of Aerocelluloses. Cationic functionalization of aerocelluloses was successfully achieved using cationic reagent (CHPTAC) as shown in Scheme I. The resulting cationized aerocelluloses are expected to remove the anionic dye (CI RB19) from wastewater due to the presence of a large number of positively charged sites (quaternary ammonium cations) on the surface of the aerocelluloses. As shown in Figure 9, the peak around 1500 cm\(^{-1}\), associated with quaternary ammonium cation in the FTIR spectrum of the cationized aerocellulose, indicates the presence of CHPTAC on aerocellulose [28, 36]. Figure 10 shows the variation of the dye concentration as a function of time in DI water containing CI RB19 dye using cationized aerocelluloses prepared at 5% and 11% cellulose concentrations. Regardless of the cellulose concentration used, the cationized aerocelluloses exhibit strong ability to remove dye molecules. This is illustrated by the drastic decrease of the dye concentration in solution until the complete removal on day 6 (Figure 11(e)). The aerocellulose without cationic functionalization only absorbs a limited amount of dye as shown in Figure 11(b). The cationic functionalization of aerocellulose immobilizes negatively charged dye on positively charged sites, while aerocellulose with porous morphology may absorb limited amount of dyes only. As shown in Figures 11(b) and 11(d), no difference in the appearance of aerocellulose and cationized aerocellulose is present, suggesting that the cationic functionalization of aerocellulose does not impact the surface area and pore volume of aerocelluloses due to the steric effect resulting from positively charged quaternary ammonium molecules grafted onto the aerocellulose [37]. Therefore, cationic functionalization and good porosity enable aerocelluloses to exhibit excellent removal of negatively charged dye molecules from wastewater.

4. Conclusions

In this study, microcrystalline cellulose was dissolved in NaOH/water solvent system followed by gelation and supercritical drying operations to prepare aerocelluloses. The aerocelluloses prepared from different cellulose concentrations exhibited a porous structure and a specific surface area of 80–125 m\(^2\)/g. FTIR and XRD analyses indicated typical transformation of crystalline structure from cellulose I (cellulose) to cellulose II (aerocelluloses). Aerocelluloses were cationically functionalized and the resulting material exhibited a strong ability to completely remove negatively charged dyes from
Figure 11: Comparison of the removal efficiency of reactive dye (CI RB19) from the water for aerocellulose and cationized aerocellulose prepared from 5% cellulose concentration: (a) aerocellulose and (c) cationized aerocellulose before dye absorption; (b) aerocellulose and (d) cationic aerocellulose after dye absorption; (e) photographs of the dye fixation over time for cationized aerocellulose.

water. In the light of sustainability, biodegradability, and low cost of cellulose, highly porous aerocelluloses prepared in this study could be a promising material as a fast absorbent to clean environmental pollutants.

Competing Interests

The authors declare that there are no competing interests regarding the publication of this paper.

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