

Research Article

Antimicrobial Properties of Copper Nanoparticles and Amino Acid Chelated Copper Nanoparticles Produced by Using a Soya Extract

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This paper reports a comparison of the antibacterial properties of copper-amino acids chelates and copper nanoparticles against *Escherichia coli*, *Staphylococcus aureus*, and *Enterococcus faecalis*. These copper-amino acids chelates were synthesized by using a soybean aqueous extract and copper nanoparticles were produced using as a starting material the copper-amino acids chelates species. The antibacterial activity of the samples was evaluated by using the standard microdilution method (CLSI M100-S25 January 2015). In the antibacterial activity assays copper ions and copper-EDTA chelates were included as references, so that copper-amino acids chelates can be particularly suitable for acting as an antibacterial agent, so they are excellent candidates for specific applications. Additionally, to confirm the antimicrobial mechanism on bacterial cells, MTT assay (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) was carried out. A significant enhanced antimicrobial activity and a specific strain were found for copper chelates over *E. faecalis*. Its results would eventually lead to better utilization of copper-amino acids chelate for specific application where copper nanoparticles can be not used.

1. Introduction

It is well known that the copper has a wide activity against bacteria and fungi [1–4]. Due to the development of antibiotic resistance and the surging of infectious diseases, a lot of researchers are searching for new antibacterial agents [5–11].

Although copper is one of the most widely used materials in various applications, its nanosynthesis requires some special care because of their high holding to rust. Compared to gold and silver, copper is extremely sensitive to air, and copper oxides phases are thermodynamically more stable.

Therefore, the formation of an oxide layer on the surface of the copper nanoparticles is inevitably causing a marked decrease of its antibacterial properties [12–14].

In the last few years, several organics copper complexes of Ni, Cu, and Co have been tested for their antibacterial and antitumor properties. Their mode of action is probably the binding of amino acids and metals ions, which is likely to leave some potential donors of free atoms and enhance the biological activity [13, 15–17].

Chelates are highly stable products capable of maintaining the surrounded metal ions from an organic molecule

(chelating agent); so its precipitation as insoluble hydroxides should be avoided with the care of the storage [15, 18].

We have previously reported about a soybean aqueous extract mediated synthesis of very stable copper nanoparticles; in this work we report the antibacterial activity of copper nanoparticles and copper chelated species that are an intermediate step between ion copper and copper nanoparticles [19]. The main objective was to compare the antibacterial properties of different species of copper, such as copper nanoparticles and copper chelates. Such comparative study would reveal strain specificities and would eventually lead to better utilization of copper-amino acids chelate for specific application where copper nanoparticles could be not used.

2. Materials and Methods

2.1. Reagents. Textured soy was purchased in a local natural products supermarket; sodium hydroxide (NaOH) and copper sulfate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) of Sigma-Aldrich were used without any further purification. Milli-Q water (18.2Ω) was used throughout the experiment; and the synthesis reagents are phosphate buffer Na_2HPO_4 and KH_2PO_4 (Fermont), Mueller-Hinton broth (BD Difco), sodium chloride (NaCl, CTR Scientific), and Kit MTT 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide (Sigma-Aldrich).

2.2. Characterization. Optical absorption spectra were obtained with an Ocean Optics S2000-UV-Vis system, Transmission Electron Microscopy (TEM) images at 100 kV were carried out using a JEOL-1230, Thermogravimetric Analysis (TGA) was made, and the average particle size was measured with a Dynamic Light Scattering Nanosizer (DLS). Infrared spectroscopy (FTIR) was made with an IR Affinity-1.

2.3. Experimental Method. Six samples were prepared to compare the bactericidal activity; the samples with copper were synthesized with a concentration of 40 mM. The syntheses are described as follows:

- (i) Soybeans extract: the solution was prepared by heating to boiling point 100 mL of deionized water with 3 g of textured soya for 10 min; the fiber was then separated by filtration.
- (ii) Ionic solution of Cu^{2+} species: this solution was prepared adding CuSO_4 in deionized water.
- (iii) Copper-soybean extract chelates: they were synthesized according to the method previously reported by Guajardo-Pacheco et al. [19]. A soybean extract solution was prepared, as described above; after that, 1 gr of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ was added and mixed for 15 min in magnetic stirring; the pH was adjusted to 7 using a 1 M solution of sodium hydroxide.
- (iv) Colloidal dispersion of copper nanoparticles: the particles were prepared in the aqueous phase by a chemical reduction of copper chelate complex solution using sodium borohydride as reducer. 100 mL solutions of copper chelate complex were purged with N_2 gas for 10 min to remove the dissolved oxygen;

1 mL of aqueous solution of sodium borohydride (0.1 M) was then added to it drop-wise under constant stirring. The solution turned into a black color on the complete addition of the reducing agent, denoting the formation of copper nanoparticles. The nanoparticles were washed repeatedly with ethylic alcohol [19].

- (v) EDTA-copper chelate: for EDTA chelated copper solution, 1 gr of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ and 1 gr of EDTA were mixed in 100 mL of deionized water. The Cu/EDTA ratio was calculated previously by 11.68.

2.4. Antimicrobial Test. The antimicrobial activity of the chelates was tested using the standard microdilution method, which determines the minimum inhibitory concentration (MIC) and the minimum bactericide concentration (MBC), leading to the inhibition of bacterial growth (CLSI M100-S25 January 2015). The strains tested were *Escherichia coli* (ATCC 25922), *Staphylococcus aureus* (ATCC 29213), and *Enterococcus faecalis* (ATCC 29212). The bacterial concentration was standardized to an optical density of 0.2 at 568 nm (approximately 1×10^8 CFU/mL) using the McFarland scale. The chelates in dispersion were diluted with 50 μL of Mueller-Hinton broth and 50 μL of phosphate buffer inoculated with the tested strains at a concentration of 1×10^5 CFU/mL was grown for 24 h at $35 \pm 1^\circ\text{C}$ [20].

2.5. MTT Assay. The MTT assay was used for measuring the proliferation of bacterial cells. Bacterial cells (10^5 CFU/mL) were incubated into Mueller-Hinton broth at 50 μL per well in 96-well microtiter plates. Threefold serial dilutions of copper-amino acids chelate were added to wells containing bacterial cells. After 24 h of incubation at 37°C , each concentration was tested in triplicate. 24 hours later, 10 μL of the MTT (5 $\mu\text{L}/\text{mL}$) reagent was added to each well and the plates were incubated for 4 h at 37°C . Then, DMSO (100 μL) was added to finish the reaction; the plate was shaken gently to redissolve the crystals formed. The absorbance was read at 490 nm wavelength by a Microplate Reader (Bio-Rad Laboratories) spectrophotometer. The results were expressed as the inhibition ratio of cell increase calculated as $[(A - B)/A] * 100\%$, in which *A* and *B* represent the average of live bacteria of the control and samples, respectively [21].

3. Results and Discussion

Figure 1(a) shows UV-Vis absorption spectra of the copper ions solution and Figure 1(b) copper-soybeans extract solution. It is possible to observe the typical wide band associated to copper free ions in the range between 650 and 1000 nm with a maximum at approximately 800 nm [22]. In Figure 1(b) this band disappears indicating the depletion of the copper free ions and formation of a copper-soya extract complex. The band in the range between 400 and 550 nm has been ascribed to soybeans extract.

Figure 2 shows the absorption infrared spectrum of the copper-soybean extract solution. There can be seen bands in the range from 400 to 500 cm^{-1} corresponding to the formation of the copper chelate, the band at 457 cm^{-1} is

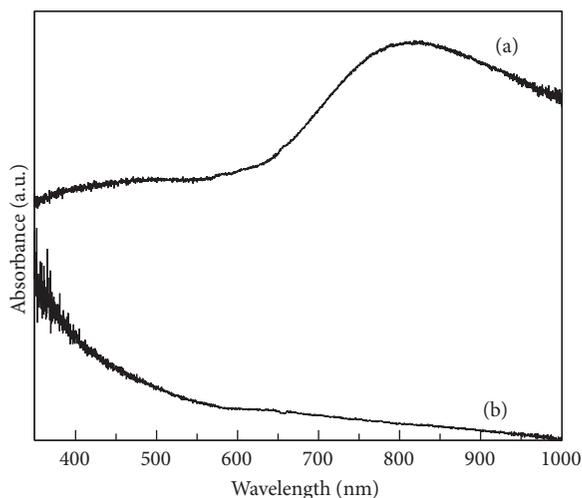


FIGURE 1: UV-Vis spectrum of (a) ionic solution of copper and (b) copper-amino acids chelate.

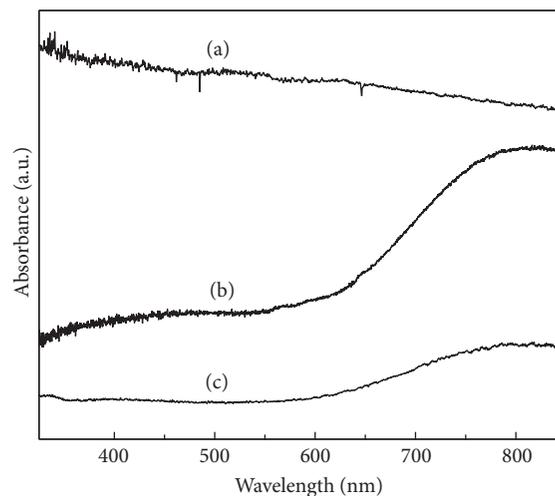


FIGURE 3: UV-Vis spectrum of (a) EDTA solution, (b) ionic solution of copper, and (c) EDTA-copper chelate.

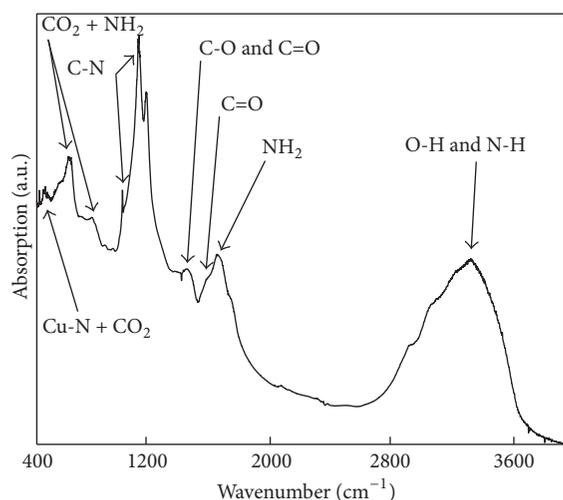


FIGURE 2: Infrared absorption spectra of copper-amino acids chelate.

assigned to the contribution of the Cu-N symmetric vibration and CO_2 wagging, and the bands at 614 cm^{-1} and 773 cm^{-1} can be ascribed to the contribution of CO_2 rocking and NH_2 rocking due to complex with amino acids; from 1000 to 1100 cm^{-1} are the C-N vibrations. The peaks of ν (C-O) and ν (C=O) at 1421 cm^{-1} , 1580 cm^{-1} , and 1626 cm^{-1} are the contribution of ν (C=O) and NH_2 scissoring, respectively. The range from 3290 to 3390 cm^{-1} corresponds to O-H and N-H vibrations. All of these vibrations modes confirm the copper-amino acid complex formation [23]; it is well known that the soybean is rich in proteins content, of which the 95% of the total protein is called soluble protein, and approximately 16% is attributed to the Nitrogen presence [24, 25], favoring the copper-amino acids chelates formation as can be seen in the infrared absorption spectra.

Figure 3 shows the UV-Vis absorption spectrum of (a) EDTA starting solution, where a wide band in the range of

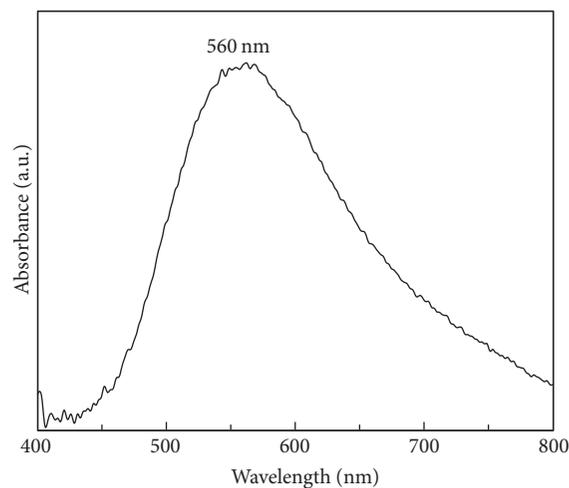


FIGURE 4: UV-Vis of copper nanoparticles sample.

400 to 900 nm can be observed, and (b) ionic copper solution with its typical band wide range between 400 and 900 nm and the maximum being approximately at 800 nm. And (c) corresponds to the EDTA-copper chelate sample; as it is possible to see, the maximum of ionic copper band decreases due to the copper free ions depletion and formation of copper chelate with the EDTA; this might represent a proof of chelate formation.

Figure 4 shows the UV-Vis absorption spectrum of the copper nanoparticles sample with a typical absorption plasmon band at approximately 560 nm.

Figure 5 shows a TEM image of copper nanoparticles sample, in which there can be observed that the nanoparticles have quasi-spherical shapes with an average diameter of $5.67 \pm 0.5337\text{ nm}$ as it can be seen in the histogram, where 169 nanoparticles were measured. The TEM image and the histogram also show a narrow size distribution of the

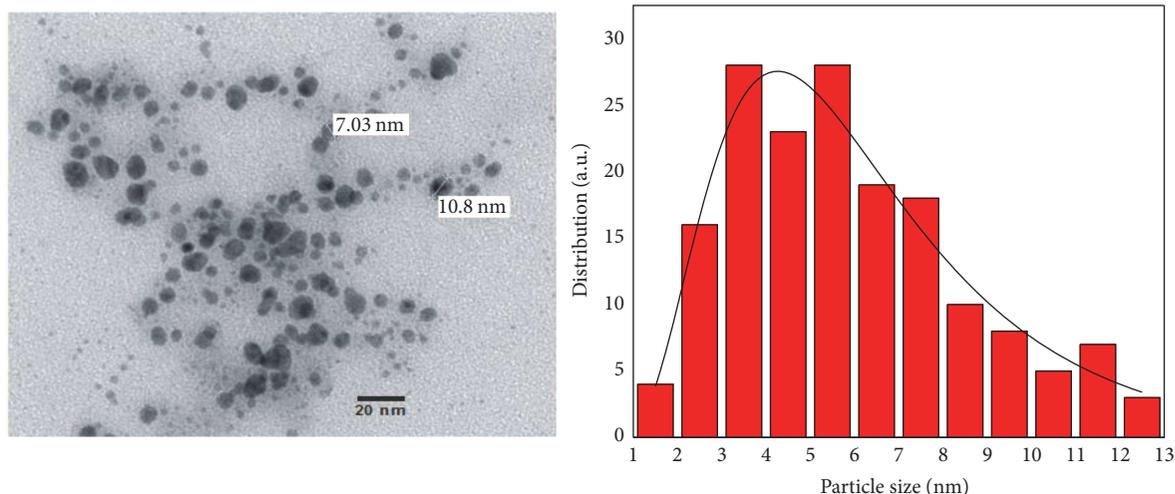


FIGURE 5: TEM image of copper nanoparticles sample and the particle size distribution histogram.

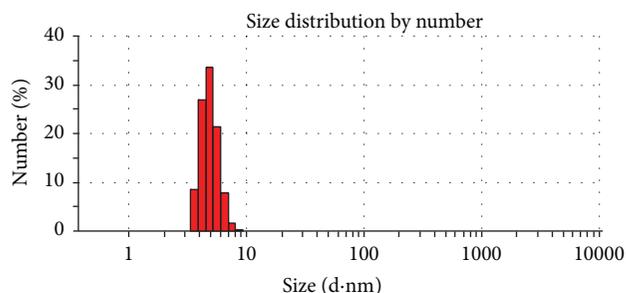


FIGURE 6: Dynamic Light Scattering measurement of copper nanoparticles sample.

nanoparticles; this could be corroborated by Dynamic Light Scattering measurements (Figure 6).

Figure 6 shows the Dynamic Light Scattering measurement of copper nanoparticles; the average size obtained was of 4.9 nm with a narrow size distribution.

3.1. Antibacterial Results. MIC values were obtained for the chelates tested against *E. coli* (ATCC 25922), *S. aureus* (ATCC 29213), and *E. faecalis* (ATCC 29212). The results are presented in Table 1, where it can be seen that the copper-amino acids chelate sample presents the best antibacterial activity against the three strains tested in comparison with copper ions, copper-EDTA chelate, and copper nanoparticles.

The MIC of the copper nanoparticles and the copper-amino acids chelates is lower when it is tested against *E. faecalis* than when tested against *E. coli* and *S. aureus*. These results may refer to differences in the cell wall of each strain; the cell wall of Gram-positive strains is wider than the cell wall of Gram-negative strains [26]. In addition, the reported mechanism for the toxicity of copper ions on *E. faecalis* includes a rapid DNA degradation, followed by a reduction of bacterial respiration; it is also known that copper ions inhibit certain cytochromes in the membrane of *E. faecalis* by altering the conformation and electron transference of

TABLE 1: Minimum inhibitory concentrations (MIC).

Sample	MIC copper (mM)		
	Bacterial strains		
	<i>E. coli</i> (ATCC 25922)	<i>S. aureus</i> (ATCC 29213)	<i>E. faecalis</i> (ATCC 29212)
Soybeans extract	— ^a	— ^a	— ^a
Ionic solution of Cu	20 ± 0	20 ± 0	20 ± 0
Copper-amino acids chelate	10 ± 0	10 ± 0	5 ± 0
Copper nanoparticles	40 ± 0	40 ± 0	20 ± 0
EDTA-copper chelate	20 ± 0	20 ± 0	20 ± 0
EDTA solution	40 ± 0	40 ± 0	40 ± 0

^aThe antibacterial activity was not found with the mM concentrations used in this work.

associated reductases [27]. These affectations of copper over *E. faecalis* could be potentiated with the presence of amino acids in the copper chelate and this could be the reason why *E. faecalis* is more sensitive to copper-amino acids chelate than *E. coli* and *S. aureus*.

As for the copper nanoparticles and the EDTA reference there is no significant difference between the MIC results of *E. coli* and *S. aureus*.

Figure 7 shows the MICs of copper nanoparticles and copper-amino acids chelate against bacterial strains *E. faecalis*. The serial dilutions of nanoparticles were exposed to bacterial strains, and we can see that the copper-amino acids chelate antimicrobial effect is better than that of the copper nanoparticles; in general, the copper-amino acids chelate showed the highest activity among all the six samples in the three bacteria tested (Table 1).

In Figure 8, we can see MTT assay results for the copper-amino acids chelate sample; the Inhibition Rate value for *E. faecalis* is higher compared to *E. coli* and *S. aureus* at a

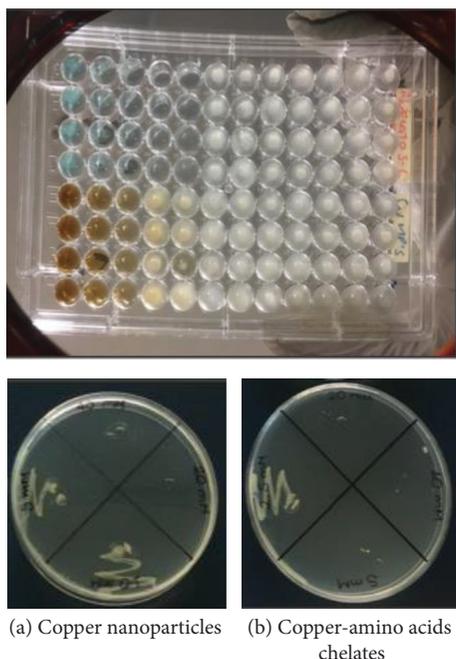


FIGURE 7: MICs of nanoparticles against bacterial strain *Enterococcus faecalis*. (a) Copper nanoparticles; (b) copper-amino acids chelate.

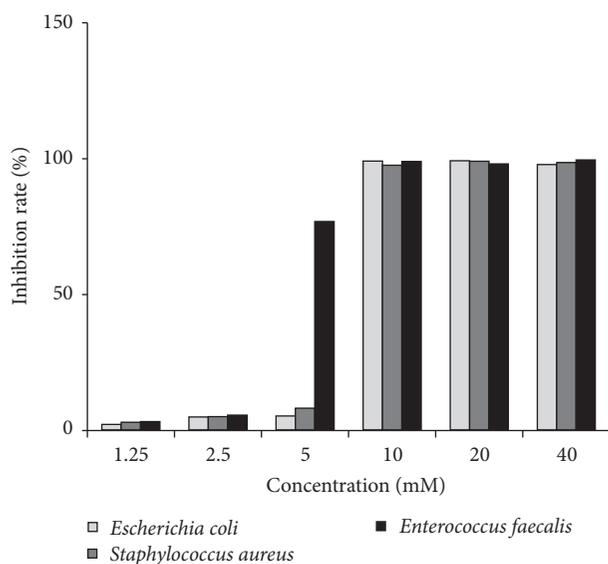


FIGURE 8: Effect of copper-amino acids chelate sample by MTT assay.

concentration of 10 mM, representing one order of magnitude smaller compared to the concentrations obtained for the other tested solutions.

4. Conclusions

Copper-amino acids chelate was synthesized in aqueous solution and antimicrobial assays, which demonstrates that its strain specificities and enhanced activity are about ten

times more effective comparing to the copper nanoparticles and the copper-EDTA chelate, obtaining a better antimicrobial effect against *E. faecalis* rather than both *E. coli* and *S. aureus*. These results show that copper-soy chelates can be particularly suitable as antibacterial agent candidates for specific application instead of copper nanoparticles.

Competing Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

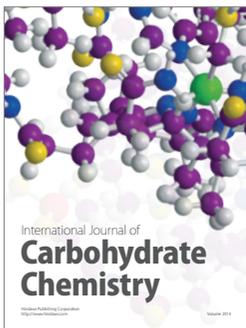
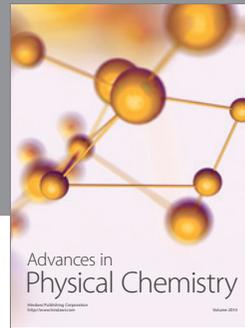
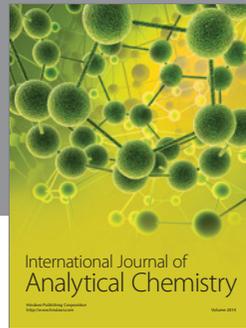
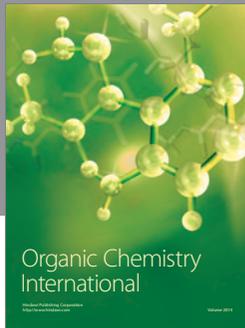
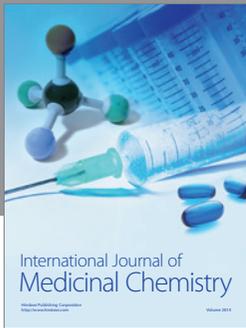
Acknowledgments

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References

- [1] J. P. Ruparelia, A. K. Chatterjee, S. P. Duttagupta, and S. Mukherji, "Strain specificity in antimicrobial activity of silver and copper nanoparticles," *Acta Biomaterialia*, vol. 4, no. 3, pp. 707–716, 2008.
- [2] C. Cervantes and F. Gutierrez-Corona, "Copper resistance mechanisms in bacteria and fungi," *FEMS Microbiology Reviews*, vol. 14, no. 2, pp. 121–137, 1994.
- [3] S. Silver, "Bacterial silver resistance: molecular biology and uses and misuses of silver compounds," *FEMS Microbiology Reviews*, vol. 27, no. 2-3, pp. 341–353, 2003.
- [4] G. Ren, D. Hu, E. W. C. Cheng, M. A. Vargas-Reus, P. Reip, and R. P. Allaker, "Characterisation of copper oxide nanoparticles for antimicrobial applications," *International Journal of Antimicrobial Agents*, vol. 33, no. 6, pp. 587–590, 2009.
- [5] M. J. Hajipour, K. M. Fromm, A. Akbar Ashkarran et al., "Antibacterial properties of nanoparticles," *Trends in Biotechnology*, vol. 30, no. 10, pp. 499–511, 2012.
- [6] M. Agarwala, B. Choudhury, and R. N. S. Yadav, "Comparative study of antibiofilm activity of copper oxide and iron oxide nanoparticles against multidrug resistant biofilm forming uropathogens," *Indian Journal of Microbiology*, vol. 54, no. 3, pp. 365–368, 2014.
- [7] R. Y. Pelgrift and A. J. Friedman, "Nanotechnology as a therapeutic tool to combat microbial resistance," *Advanced Drug Delivery Reviews*, vol. 65, no. 13-14, pp. 1803–1815, 2013.
- [8] J. R. Morones, J. L. Elechiguerra, A. Camacho et al., "The bactericidal effect of silver nanoparticles," *Nanotechnology*, vol. 16, no. 10, pp. 2346–2353, 2005.
- [9] R. Chakraborty, R. K. Sarkar, A. K. Chatterjee, U. Manju, A. P. Chattopadhyay, and T. Basu, "A simple, fast and cost-effective method of synthesis of cupric oxide nanoparticle with promising antibacterial potency: unraveling the biological and chemical modes of action," *Biochimica et Biophysica Acta*, vol. 1850, no. 4, pp. 845–856, 2015.

- [10] C. Sámano-Valencia, G. A. Martínez-Castanon, F. Martínez-Gutiérrez et al., "Characterization and biocompatibility of chitosan gels with silver and gold nanoparticles," *Journal of Nanomaterials*, vol. 2014, Article ID 543419, 11 pages, 2014.
- [11] C. Larimer, M. S. Islam, A. Ojha, and I. Nettleship, "Mutation of environmental mycobacteria to resist silver nanoparticles also confers resistance to a common antibiotic," *BioMetals*, vol. 27, no. 4, pp. 695–702, 2014.
- [12] S. Jeong, K. Woo, D. Kim et al., "Controlling the thickness of the surface oxide layer on Cu nanoparticles for the fabrication of conductive structures by ink-jet printing," *Advanced Functional Materials*, vol. 18, no. 5, pp. 679–686, 2008.
- [13] S. Chen and J. M. Sommers, "Alkanethiolate-protected copper nanoparticles: spectroscopy, electrochemistry, and solid-state morphological evolution," *Journal of Physical Chemistry B*, vol. 105, no. 37, pp. 8816–8820, 2001.
- [14] C. Salzemann, I. Lisiecki, A. Brioude, J. Urban, and M.-P. Pileni, "Collections of copper nanocrystals characterized by different sizes and shapes: optical response of these nanoobjects," *Journal of Physical Chemistry B*, vol. 108, no. 35, pp. 13242–13248, 2004.
- [15] J. J. Lucena, "El empleo de complejantes y quelatos en la fertilización de micronutrientes," *Revista Ceres*, vol. 56, no. 4, pp. 527–535, 2009.
- [16] T. Ganz, "Defensins and other antimicrobial peptides: a historical perspective and an update," *Combinatorial Chemistry & High Throughput Screening*, vol. 8, no. 3, pp. 209–217, 2005.
- [17] F. D. Silva, C. A. Rezende, D. C. P. Rossi et al., "Structure and mode of action of microplusin, a copper II-chelating antimicrobial peptide from the cattle tick *Rhipicephalus (Boophilus) microplus*," *The Journal of Biological Chemistry*, vol. 284, no. 50, pp. 34735–34746, 2009.
- [18] F. Basolo and R. Johnson, *Química de los Compuestos de Coordinación*, Reverté, Bellvei, España, 1980.
- [19] M. J. Guajardo-Pacheco, J. E. Morales-Sánchez, J. González-Hernández, and F. Ruiz, "Synthesis of copper nanoparticles using soybeans as a chelant agent," *Materials Letters*, vol. 64, no. 12, pp. 1361–1364, 2010.
- [20] P. A. Wayne, "CLSI. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Fifth Informational Supplement," CLSI Document M100-S25, Clinical and Laboratory Standards Institute, 2015.
- [21] X. Wang, Y. C. Wu, S. P. Xia, Y. Gao, and C. X. Li, "Inquiring into the method of counting live germ with MTT," *Journal of Luzhou Medical College*, vol. 25, pp. 291–293, 2002.
- [22] P. De Vreese, N. R. Brooks, K. Van Hecke et al., "Speciation of copper(II) complexes in an ionic liquid based on choline chloride and in choline chloride/water mixtures," *Inorganic Chemistry*, vol. 51, no. 9, pp. 4972–4981, 2012.
- [23] A. Cuevas, I. Viera, M. H. Torre, E. Kremer, S. B. Etcheverry, and E. J. Baran, "Infrared spectra of the Copper(II) complexes of amino acids with hydrophobic residues," *Acta Farmaceutica Bonaerense*, vol. 17, no. 3, pp. 213–218, 1998.
- [24] K. A. Kuiken and C. M. Lyman, "Essential amino acid composition of soy bean meals prepared from twenty strains of soy beans," *The Journal of Biological Chemistry*, vol. 177, pp. 29–36, 1948.
- [25] C. H. Van Etten, J. E. Hubbard, J. M. Mallan, A. K. Smith, and C. W. Blessin, "Amino acid composition of soybean protein fractions," *Agricultural and Food Chemistry*, vol. 7, no. 2, pp. 129–131, 1959.
- [26] J. Thiel, L. Pakstis, S. Buzby et al., "Antibacterial properties of silver-doped titania," *Small*, vol. 3, no. 5, pp. 799–803, 2007.
- [27] S. L. Warnes and C. W. Keevil, "Mechanism of copper surface toxicity in vancomycin-resistant enterococci following wet or dry surface contact," *Applied and Environmental Microbiology*, vol. 77, no. 17, pp. 6049–6059, 2011.



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