Some provocative thoughts on damage and repair of DNA

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The instability of nucleic acid bases, responsible for hereditary information storage is amazing. Is there a system in this madness? All the nitrogen bases may undergo different kinds of rapid, spontaneous, or induced modification. They may form adducts and photoproducts, may be deaminated, methylated, and oxidized, or may be lost spontaneously or in the course of enzymatic processes. The list of potentially mutagenic lesions in DNA is long (some of them are given in parentheses) and includes products of deaminations (hypoxanthine, uracil, 5-methyluracil (thymine)); of methylation (3-methyladenine, O-6-methylguanine); of oxidation (8-oxoguanine, thymine glycol, 5-hydroxycytosine, 5-hydroxyuracil, 5-formyluracil), and apurinic/aprimidinic (AP) sites. The frequency of base oxidation much outweighs the remaining modifications. The most susceptible base to oxidation damage (and to depurination), both in nucleotide pools as well as in DNA, is guanine, with formation of the highly mutagenic base, 8-oxoguanine. It has been calculated that in a single cell per day as much as $\sim 100\,000$ modification events occur [1, 2]. Modified bases may be restored by (1) direct reversal of the modifying reaction (e.g., by action of photolyases that reverse pyrimidine dimers and 6-4 pyrimidine photoproducts to native pyrimidines [3–5] or by action of O-6-methylguanine DNA methyltransferase, a suicide enzyme that removes methyl (or alkyl) from O-6-alkylguanine residues in DNA and accepts it on its own Cys321 residue [6] and (2) by processes of base excision- or nucleotide excision-repair (BER or NER) [7–10]. About six distinct DNA-glycosylases, at least, are found in bacteria and their homologues have been detected in mammalian cells. There is a common mechanism for BER which includes: (i) attachment of the enzyme to the damaged site, (ii) flipping out a modified base (or AP-sites) of the double-stranded structure, excising a base, (iii) interruption of the phosphodeoxyribose chain to form a 3'OH end, (iv) filling the gap with matching nucleotides, and (v) sealing to restore the continuity of the DNA. BER begins with removal of a modified base, and then 1 to 10 nucleotides are undergoing exchange. In NER, a fragment of 12–13 nucleotides or 24–32 nucleotides is excised in bacteria and mammalian cells, respectively [11]. The NER system excises a broad spectrum of structurally unrelated DNA lesions, pyrimidine dimers, and 6-4 photoproducts,

but also excises 8-oxoguanine, and thymine glycols from DNA [9]. Yet another, mismatch repair system, acts on native (or modified) mismatched bases shortly after replication, when distinction between the new synthesized and the old strand of DNA is possible [12, 13]. In the course of the repair of a single mismatch by mismatch repair system as many as 1000–2000 nucleotides may be excised [14]. When all kinds of modifications and repair are considered, it can be calculated that in a single cell, in the course of DNA repair per day as many as 2×10^6 base pairs are exchanged ($\sim 10^6$ arise from base modification, and 10^6 from mismatch repair). This covers a large fraction of genomic DNA. Therefore, the question may be posed of whether the huge number of enzymes for DNA repair, arose in a long process of evolution as a defense system against different modifications, or if they are part of normal DNA metabolism to keep molecules in a continuous dynamic state? Apparently, nature prefers modification and repair to firm stability. Therefore, is modification of the bases a method for quick response to situations in the environment and thus a source of evolution? A natural bases in DNA are necessary before it is replicated, the cell cycle apparatus and checkpoint proteins control DNA damage and halt cell division until repair of DNA is complete.

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