Research Article

Localization of Magic-F1 Transgene, Involved in Muscular Hypertrophy, during Early Myogenesis

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1. Introduction

Muscular hypertrophy is controlled by both muscle growth and muscle atrophy. These apparently contrasting biological processes share key molecules triggering active transcriptional programs to induce skeletal muscles atrophy or hypertrophy.

The IGF-1 (insulin-like growth factor-1) pathway affects both hypertrophy and atrophy, increasing protein synthesis and inhibiting the expression of transcription factor FOXO (forkhead box O), respectively. A key molecular player downstream of IGF-1 receptor is Akt (Abelson leukemia kinase) that when phosphorylated activates protein synthesis via mTOR (mammalian target of rapamycin). The two main actors to negatively control the muscle growth are myostatin and atrogin-1. Myostatin belongs to the TGF-β (transforming growth factor beta) family, and it is expressed and secreted predominantly in skeletal muscle, functioning as negative regulator of muscle growth. Mice, sheep, cattle, and humans that present mutations in myostatin gene show a double-muscling phenotype, characterized by extensive muscular hypertrophy [1–4]. In vitro myostatin is able to positively affect the expression of ubiquitin ligases, involved in muscle atrophy [5]. Interestingly, myostatin treatment blocks the IGF1-AKT pathway, allowing the increased expression of atrogin-1, directly involved in muscle atrophy [6]. Among the muscle growth regulators, hepatocyte growth factor (HGF/SF) [7–9], initially unveiled as the major inducers of hepatogenesis, is involved in muscle stem-cell activation through its tyrosine kinase receptor Met [10–14], containing a Pax3-binding site. Pax3 is an early transcription factor involved in embryonic and adult myogenesis, and it is expressed in the lateral dermomyotome of all somites, where also is present Met [15]. Transgenic animals generated to interfere in HGF-Met signaling show abnormality in several muscles during embryogenesis [16–18]. Moreover, HGF-Met...
pathway is important in muscle regeneration, since, sustains the proliferation of muscle stem cells after their activation [19–22]. However, HGF expression is downregulated during myogenesis in order to allow the satellite cells to exit the cell cycle, avoiding a delay in the regeneration process [23, 24].

The production of recombinant proteins allows the great advantage to select a specific biological effect of a given protein able to trigger different biological processes. We recently generated transgenic mice expressing an HGF-related recombinant protein, named Magic factor-1 (Met-activating genetically improved chimeric factor-1 or Magic-F1) and expressed exclusively in skeletal muscles [25]. This animal model develops muscular hypertrophy with no evident side effects or hyperplasia. However, the expression pattern of the transgene in early embryogenesis is unknown.

The effect of Magic-F1 recombinant protein in skeletal muscle tissue offers biological advantages over HGF. Magic-F1 is able to promote myocytes survival and enhance muscle regeneration and a lack of any mitogenic activity could allow a potential safe use of the recombinant protein as therapeutic cytokine for muscle degenerative disorders, promoting muscle regeneration without the potential risk of stimulating uncontrolled proliferation. Because of its selective stimulation of hypertrophy, Magic-F1 is a novel molecule with potential applicative perspective to counteract muscle wasting in muscle diseases such as cachexia or muscular dystrophy.

Here, we evaluate the localization of transgene Magic-F1 in comparison with Pax3, the earliest myogenic transcription factor, in early and late embryogenesis using in situ hybridization on whole-mount and cryosections of transgenic and wt mouse embryos.

2. Materials and Methods

2.1. Mouse Embryos and Staging. Embryos for in situ hybridization were dissected from CD1 wt (wild-type) female mice mated with homozygous Magic-F1 transgenic male mice (Stem Cell Research Institute, H. S. Raffaele, Milan, Italy). Embryos were generated using timed mating, with the morning vaginal plug designated as E0.5.

2.2. Genotyping of Magic-F1 Embryos. Magic-F1 is a recombinant protein containing two HGF NK2 domains joint by a linker. The exact amino acidic sequence of Magic-F1 corresponds to residues 1–285 of human HGF (Gene Bank no. M73239), a linker with the sequence (GGGGS)3; residues 30–285 of human HGF, and a poly-histidine tag with the sequence DDDKHHHHHHH. We generated transgenic mice expressing Magic-F1 construct into a plasmid containing the proper linearization with EcoRI and BamH1, respectively. Magic-F1 was digested with BstXI and Magic-F1 antisense probes were synthesized using T7 RNA (ribonucleic acid) polymerase linearization at a suitable site: pCMV-sport6.1 (Promega). Nonradioactive antisense and sense riboprobes were synthesized by in vitro transcription using digoxigenin-UTP following the manufactures instructions (Boehringer Mannheim). Magic-F1 antisense and sense probes were synthesized using SP6/ T7 RNA (ribonucleic acid) polymerase linearization at a suitable site: pCMV-T Easy Vector System (Promega). Magic-F1 fragments were subcloned into the pGEM-T Easy Vector System (Promega). Nonradioactive antisense and sense riboprobes were synthesized by in vitro transcription using digoxigenin-UTP following the manufactures instructions (Boehringer Mannheim). Magic-F1 antisense and sense probes were synthesized using SP6/ T7 RNA (ribonucleic acid) polymerase linearization at a suitable site: pCMV-sport6.1 (Invitrogen) carrying Pax3 cDNA after proper linearization with EcoRI and BamH1, respectively.

2.3. Plasmid and Riboprobe Synthesis. As probes for hybridization experiments, human Magic-F1 cDNA (complementary deoxyribonucleic acid) (1759 bp) was amplified using oligonucleotide primers FW (5’-ATGTGGTGATCACAAGTGTAT-3’) and REV (5’-CTATGGTGATCAGGTTGTCGGGT-3’). The amplified products were analyzed by agarose gel electrophoresis in TBE (Tris Borate EDTA) 1X.

2.4. In Situ Hybridization. Whole-mount in situ hybridization on mouse embryos was performed as described [26]. Briefly, embryos were dissected free of all membranes in phosphate-buffered saline (PBS) and fixed in 4% paraformaldehyde in PBS for 1-2 h at RT (Room Temperature) with slow rocking speed. After washing twice in PTW (PBS, 0.1% Tween-20), embryos were washed with 50% MeOH/PTW and 100% MeOH then stored in 100% MeOH at −20°C for less than a month. For hybridization,
**Figure 1:** Magic-F1 transgenic mice. (a) Schematic representation of transgene. (b) Genotyping of 13.5 dpc mouse embryos derived by hemizygous-matched mice; +, − positive and negative control, respectively.

**Figure 2:** Expression of Magic-F1 in whole mount embryos. Lateral view of whole-mount in situ hybridization with digoxigenin labeled antisense Magic-F1 (a, b, d, e, f, g, h, and i) and sense Magic-F1 (c) cRNA probes as negative control (ctr-). Specific hybrids are visualized as a purple precipitate in all panels except in (a), 9.5 dpc and (b) 10.5 dpc. (e) Specific Magic-F1 transcripts are detected (f) in the mesodermal cells of the jaw and hyoid bone; in f, tv: telencephalic vesicles, cmt: mesenchymal cephalic tissue, hb: hindbrain, and ba: branchial arches where muscles are localized as well as around the eyes; a faint expression is also revealed in tail (g) and dorsal (h) somites, while a strong signal was selected in somites of fore (f′) and hind (h′) limb buds (i). In embryos hybridized with sense probe (c) background staining has not been detected as well as on 11.5 dpc wild type embryo (wt) hybridized with antisense probes (d).
embryos were incubated horizontally at 65°C overnight in hybridization mix (50% deionized formamide, 1.3xSSC pH 5, 5 mM EDTA pH 8, 50 μg/mL Yeast RNA, 0.2% Tween-20, 0.5% CHAPS, 100 μg/mL Heparin, ddH₂O) with 1 μg/mL digoxigenin-labeled RNA probes. After in situ hybridization, the embryos were washed in MABT buffer (100 mM maleic acid (Sigma), 150 mM NaCl, 0.1% Tween-20, pH 7.5) and NTMT buffer (100 mM NaCl, 100 mM Tris-HCl pH 9.5, 50 mM MgCl₂, 0.1% Tween-20) and incubate with staining solution (4.5 μg/mL NBT and 3.5 μg/mL BCIP). When the developed color reached the desired extent, embryos were washed twice with PTW and refixed in 4% HCHO/0.1% glutaraldehyde/PTW for 2 h at RT or 4°C overnight.

In situ hybridization on mouse cryostat sections was performed according to Sally Dunwoodie, NIMR. Briefly, the sections were fixed 4% paraformaldehyde in PBS, digested with proteinase K and postfixed. Sections were prehybridized in hybridization solution without probe (50% deionized formamide, 10x salt, 250 μg/mL Yeast RNA, 5x Denhardt’s, 50 μg/mL Heparin, 0.1% Tween, ddH₂O) in a humid environment at 67°C for 2 h. After incubation, the prehybridization buffer was removed and replaced with hybridization buffer containing everything in the prehybridization buffer plus 0.1–1 μg/mL labeled cRNA (complementary ribonucleic acid) riboprobe, denatured for 5 min at 80°C. A coverslip was placed over the hybridized area and the slides were then placed in a moist, closed plastic box and hybridized overnight at 67°C h. Following hybridization, the coverslips were washed off by dipping into 50% formamide, 1xSSC prewarmed to 67°C. The tissue was subjected to stringent washing at 67°C in 50% formamide, 1xSSC twice for 30 min, followed by two washes in MABT 1X at RT, for 30 min each. Slides were exposed to BMpurple (Roche) for 2 h. Slides were fixed in PFA 4% and washed in PBS, three times for 5 min. Hybridization signals were observed and photographed using a Nikon microscope. As a negative
control, specimens were incubated with hybridization buffer containing digoxigenin labeled riboprobes corresponding to the sense sequence of Magic-F1.

2.5. Immunocytochemistry. Cryosections placed in a humid, closed plastic box were fixed for 10 min in 4% paraformaldehyde in PBS, permeabilized in 0.2% Triton and incubated, first, with a blocking buffer (2% donkey serum containing 1% blocking solution in PBS) for 30 min at RT and, then, a second blocking buffer (0.5% mouse serum, 0.5% goat serum) containing 1% Blocking Solution (Candorbioscience GmbH) in PBS for 30 min at RT, followed by incubation with primary antibodies (goat polyclonal antibody anti-HGF, 1:100 and Mouse monoclonal antibody anti-Pax3) diluted in 2% BSA (bovine serum albumine) in PBS overnight at 4°C. Sections were rinsed three times in PBS followed by a 2 h incubation with fluorescein-conjugated anti-mouse and anti-goat antibodies diluted 1/1500. Slides were stained with Hoechst dye and mounted in FluorSave. Stained cryosections were observed on an upright microscope Nikon Vico TE-2000 (Nikon, Japan).

3. Results and Discussion

3.1. PCR Analysis. PCR analysis of Magic-F1 showed transgene expression in all the embryonic developmental stages analyzed, and an example at stage 13.5 dpc is reported in Figure 1(b). As expected, no signal was detected in the wild-type embryos due to the high specificity of the primers able to discriminate between HGF and the recombinant protein Magic-F1.

3.2. Whole-Mount In Situ Hybridization. Whole-mount in situ hybridization was performed to detect Magic-F1 and Pax3 mRNA (messenger ribonucleic acid) in embryos at early stages of development (9.5, 10.5 and 11.5 dpc).

Magic-F1 expression was detected in bone cartilage primordia, dorsal somites, in tail and limb buds, where Pax3 signal appears to be downregulated respect to the wt embryos. At 9.5 dpc (30 somite stage) and 10.5 dpc (40 somite stage) (Figures 2(a) and 2(b)), in situ hybridization analysis with digoxigenin- (DIG-) labeled antisense probes showed no expression of Magic-F1 transcript. Magic-F1 transcripts (Figure 2(e)) have been detected from stage of 11.5 dpc (58 somite stage) in developing organs and tissues of mesenchymal origin. Specifically, positive signal was observed in mesodermal cells of the jaw, hyoid bone and in pharyngeal (or brachial) arches from which branchiomeric muscles are formed. This expression profile confirmed the muscle specificity of the MLC1F (myosin light chain type 1F) promoter, regulating Magic-F1 transcription. Furthermore, specific hybrids were also detected around the eyes at the level of the choroid coat and the sclera, both of mesenchymal origin, on the ventral and dorsal side of limb buds (Figure 2(d)), in the dorsal side of some rostral somites (Figure 2(h)), and in the tail bud (Figure 2(g)). No signal was observed in the embryos hybridized with the sense riboprobe (Figure 2(i)). The somites begin to form at the 7.75–8.0 dpc, and, after condensation, they begin to differentiate into three compartments: the sclerotome, the dermatome, and myotome. At 11.5 dpc (58 somite stage), Magic-F1 transcripts have been detected in all three compartments (sclerotome, dermatome, and myotome).
Figure 5: Expression of Pax3 and Magic-F1 at 15.5 dpc. Sagittal section of 15.5 dpc mouse hybridized with digoxigenin-labeled antisense Pax3 riboprobes (a); specific Pax3 transcripts are localized in cartilage primordium of tail vertebral body, cutaneous muscles (arrow) of thorax and trunk regions and in mid-cervical region of spinal cord (double arrow) as shown in (b); expression was also localized in cartilage primordium (arrowhead) of dorsal part of shaft region (c); (d) sagittal section of 15.5 dpc mouse hybridized with Magic-F1 cRNA probes and signal is observed in cartilage primordium of anterior arch of cervical vertebra (arrowheads) and right lobe of thymus gland (arrow); specific Magic-F1 transcripts are also localized in cutaneous muscles (arrow) of thorax and trunk regions (e); (f) higher magnification cartilage primordium of dorsal part of shaft region of ribs.

and myotome) originating from somites. No signal was detected in wt embryos at the same stage.

3.3. In Situ Hybridization on Cryostat Sections. In situ hybridization on 10 μm-cryostat sections was performed to localize Magic-F1 and Pax3 transcripts in embryos at later stages of development (11.5–13.5–15.5–17.5 dpc), since the large size of the samples did not allow a full penetration of the probes.

At 11.5 dpc, transgenic mouse embryos showed Pax3 transcripts expression at the level of hindbrain (Figure 3(a)), neuroepithelium of neural tube (Figure 3(b)), and dorsal somites (Figure 3(c)). Magic-F1 riboprobes showed a similar expression pattern (Figure 3(d)) reported for Pax3 although the signal was no detected at the level of neuroepithelium of neural tube in embryos at the same stage (Figure 3(e)). Specific transcripts were also observed in the cervical dorsal (posterior) root ganglion (Figures 3(c) and 3(f)) and somites (Figure 3(f)). In addition, at this stage, Pax3 signal can be observed in wt embryos mainly in the cerebellar primordium and somites in lumbar region (Figure 3(f)), whereas no signal for Magic-F1 has been detected (Figure 3(g)).

3.4. Localization of Pax3 and Magic-F1 Proteins and Transcripts. Immunofluorescence analysis was performed to localize the presence of recombinant protein. In transgenic embryos at 11.5 dpc, MAGIC-F1 was mainly detected at the level of thoracic (Figure 3(i)) and lumbar (Figure 3(j)) somites, whereas a faint signal was detected in wt embryos (Figure 3(k)), probably due to the interference with HGF localization in the same area. At 13.5 dpc, transgenic embryos showed a weak signal of Pax3 at the level of somites in thoracic (Figure 4(a)) and lumbosacral region (Figure 4(b)), whereas in the wild-type embryo, the signal appeared slightly
Figure 6: Expression of Pax3 and Magic-F1 at 17.5 dpc. Specific Pax3 transcripts are localized on sagittal sections of 17.5 dpc at the level of the head (a), in the nucleus pulposus (b) of lumbar intervertebral disc (arrows) of the trunk, and in cartilage primordium (arrowheads) and wall of spinal cord in midlumbar region (double arrow) as shown in (b) and in cartilage primordium of dorsal part of ribs (c). (d) Magic-F1 signal was observed at the level of cartilage primordium (arrows) of vertebrae.

stronger (Figure 4(c)). A weak signal have been detected in transgenic embryos hybridized by Magic-F1 antisense riboprobes (Figure 4(d)), where Magic-F1 was mainly expressed at the level of somites (Figure 4(e)), whereas no signal was detected in wild-type embryos (Figure 4(f)). Specific Pax3 transcripts can be detected at later stage (15.5 dpc) (Figure 5(a)) in cartilage primordium of tail vertebral body, cutaneous muscles of thorax and trunk regions, and in midcervical region of spinal cord (Figure 5(b)). In addition, at this stage, Pax3 expression demarcated also cartilage primordium of dorsal part of shaft region (Figure 5(c)). In sagittal section of 15.5 dpc embryo hybridized with Magic-F1 cRNA probes a strong signal was detected in cartilage primordium of anterior arch of cervical vertebra and right lobe of thymus gland (Figure 5(d)). Consisted with this, Magic-F1 resulted to be also expressed in cutaneous muscles of thorax and trunk regions (Figure 5(e)) and in primordium of dorsal part of shaft region of ribs (Figure 5(f)). From 17.5 dpc sagittal sections at the level of the head (Figure 6(a)), Pax3 signal was detected in the trunk nucleus pulposus in central part of lumbar intervertebral disc (Figure 6(b)) and in cartilage primordium of dorsal part of ribs (Figure 6(c)). At this stage, Magic-F1 expression was observed at the level of cartilage primordium of vertebræ (Figure 6(d)). These data suggest a role of this factor in muscle development possibly triggering the premature downregulation of Pax3 signal pathway in skeletal muscle precursor cells.

4. Conclusion

Taken together, our data show that Magic-F1 is localized in specific expression pattern in the developing muscular tissues. This could be relevant with the muscle phenotype previously observed in the transgenic animal model. The recombinant protein can be considered as a novel member of Met signaling, since it has been shown to interact with, and thus interfering with, cell migration and myogenesis. It is noteworthy that Magic-F1 expression pattern is overlapping with Pax3, however, limited only in developing muscular tissue, where exerts its myogenic potential. In fact, Pax3 is able to activate Myf5 (Myogenic factor 5) myogenic master gene [25], leading to the formation of skeletal muscle. In addition, Pax3 orchestrates FGF (fibroblast growth factor) cascade [27], targeting directly Fgfr4 (fibroblast growth factor receptor 4) and Sprouty1 known to inhibit FGF pathway. Magic-F1 may interfere with the balance between stem cell self-renewal and cell differentiation in developing muscles, mainly operated by Pax3 and Pax7 (paired box gene 7) [28]. It is unlikely that it is directly involved in stem-cell renewal; however, competing with HGF, the natural Met ligand could be move the balance towards muscle differentiation.

Further studies are necessary to elucidate Magic-F1 specific role in cell specification and differentiation during embryogenesis.

Abbreviations

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<tr>
<td>Akt</td>
<td>Abelson leukemia kinase</td>
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<tr>
<td>ba</td>
<td>Branchial arches</td>
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<td>bp</td>
<td>Base pair</td>
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<tr>
<td>BSA</td>
<td>Bovine Serum Albumine</td>
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<tr>
<td>cmt</td>
<td>Mesenchymal cephalic tissue</td>
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<tr>
<td>dpc</td>
<td>Days post coitum</td>
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<tr>
<td>DNA</td>
<td>Deoxyribonucleic acid</td>
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<tr>
<td>cDNA</td>
<td>Complementary deoxyribonucleic acid</td>
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<tr>
<td>FGF</td>
<td>Basic fibroblast growth factor</td>
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<td>Fgfr4</td>
<td>Fibroblast growth factor receptor 4</td>
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<tr>
<td>FOXO</td>
<td>Forkhead box O protein</td>
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<td>FW</td>
<td>Forward</td>
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<td>GAPDH</td>
<td>Glyceraldehyde-3-phosphate dehydrogenase</td>
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<td>Hindbrain</td>
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<td>HGF</td>
<td>Hepatocyte growth factor</td>
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<td>MLC1F</td>
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<td>mTOR</td>
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REV: Reverse
RNA: Ribonucleic acid
cRNA: Complementary ribonucleic acid
mRNA: Messenger ribonucleic acid
RT: Room temperature
TBE: Tris borate EDTA
TGF-β: Transforming growth factor beta
tv: Telencephalic vesicles
wt: Wild type
WB: Western blot.

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