Research Article

Antigenicity and Immunogenicity of Rotavirus VP6 Protein Expressed on the Surface of Lactococcus lactis

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Received 29 April 2013; Revised 25 June 2013; Accepted 9 July 2013

A cademic Edi to r: Jose R. Botella

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Group A rotaviruses are the major etiologic agents of acute gastroenteritis worldwide in children and young animals. Among its structural proteins, VP6 is the most immunogenic and is highly conserved within this group. Lactococcus lactis is a food-grade, Gram-positive, and nonpathogenic lactic acid bacterium that has already been explored as a mucosal delivery system of heterologous antigens. In this work, the nisin-controlled expression system was used to display the VP6 protein at the cell surface of L. lactis.

Conditions for optimal gene expression were established by testing different nisin concentrations, cell density at induction, and incubation times after induction. Cytoplasmic and cell wall protein extracts were analyzed by Western blot and surface expression was confirmed by flow cytometry. Both analysis provided evidence that VP6 was efficiently expressed and displayed on the cell surface of L. lactis. Furthermore, the humoral response of mice immunized with recombinant L. lactis was evaluated and the displayed recombinant VP6 protein proved to be immunogenic. In conclusion, this is the first report of displaying VP6 protein on the surface of L. lactis to induce a specific immune response against rotavirus. These results provide the basis for further evaluation of this VP6-displaying L. lactis as a mucosal delivery vector in a mouse model of rotavirus infection.

1. Introduction

L. lactis is a food-grade, Gram-positive, and nonpathogenic lactic acid bacteria that has already been explored as a mucosal delivery system of heterologous antigens [1]. The most commonly used system for protein expression in L. lactis is the nisin-controlled expression system (NICE) based in the combination of a vector containing a nisin-inducible promoter (PnisA) and the regulatory genes (nisK and nisR) inserted in the bacterial genome of L. lactis strain NZ9000 [2]. Diverse genetic constructs can be used to target the heterologous antigen to different cell compartments (cytoplasm, cell wall, or extracellular medium). In particular, for cell-wall anchoring, the antigen can be fused to a fragment of Streptococcus pyogenes M6 protein which allows peptidoglycan binding of the heterologous protein [3, 4].

Rotavirus particles are nonenveloped with a triple-layered protein capsid, belonging to the Reoviridae family. Among them, group A rotaviruses are the major etiologic agents of acute gastroenteritis worldwide in children and young animals. The rotavirus diarrhea is associated with a high mortality rate, particularly in developing countries, as well as to a considerable economic burden. These facts have led to an extensive research in rotavirus vaccinology to prevent its morbidity and mortality [5–7].

Rotavirus protein expression in L. lactis was previously reported by Perez et al. [8] (VP7 protein), Marelli et al. [9], Rodriguez-Diaz et al. [10] (VP8∗ protein), Li et al. [11] (VP4 protein), and Enouf et al. (NSP4 protein) [12]. However, among rotavirus structural proteins, the intermediate layer protein VP6 is the most immunogenic and determines group specificity since it is highly conserved among strains belonging to the same group [13]. When coadministered with an adjuvant and tested in the murine infection model, VP6 (as the only viral antigen) induced a protective immune response. This protection did not depend on the murine rotavirus strains used for the challenge, revealing that VP6 contains at least some of the epitopes shared between strains. Moreover, the fact that crossed protection was induced by two divergent VP6 proteins indicates that a vaccine including VP6 from any group A rotavirus would protect against infection with any other group A rotavirus strain [14, 15].
In the present paper, *L. lactis* NZ9000 was evaluated as a cell-wall display vector of rotavirus VP6 protein and used as an antigenic carrier for mice immunization.

2. Materials and Methods

2.1. Bacterial Strains and DNA Manipulation. *L. lactis* strain NZ9000, kindly provided by Dr. Christian Magni (Instituto de Biología Molecular y Celular de Rosario, Argentina), was grown in M17 broth (Biokar Diagnostics, Beauvais, France) supplemented with 0.5% glucose at 30°C without shaking. *Escherichia coli* strain TOP10 was grown in the Luria-Bertani medium at 37°C with shaking. Clones were selected by the addition of antibiotics as follows: for *L. lactis*, chloramphenicol 10 μg/mL and for *E. coli*, ampicillin 100 μg/mL.

Plasmid DNA isolation and general procedures for DNA manipulations were essentially performed as described previously [16]. Reverse transcriptase reaction was performed using AMV RT (Promega, Madison, WI, USA) polymerase chain reactions (PTC-200 Thermo Cycler, MJ Research, Waltham, MA, USA) were performed using Pfu DNA polymerase or Taq DNA polymerase (Promega, Madison, WI, USA). Plasmids were sent to Macrogen Sequencing Service (Seoul, Republic of Korea) for DNA sequencing.

2.2. Viruses and Cell Culture. EC rotavirus strain was used to generate VP6-encoding cDNA, while RRV rotavirus strain was used to produce a virus stock providing antigens for ELISA and Western blot assays. The EC strain of rotavirus was kindly provided by Dr. Harry Greenberg (Department of Medicine and Microbiology and Immunology, Stanford University School of Medicine, Stanford, CA, USA) and RRV strain was kindly provided by Dr. Viviana Parreño (Instituto de Virología, CICVyA, INTA Castelar, Argentina). Both strains were grown in confluent MA104 cells maintained in Dulbecco's modified Eagle's medium without serum and 2% fetal bovine serum. To be used as antigen for ELISA and Western blot analysis, cell-culture-propagated RRV strain was concentrated by ultracentrifugation through a sucrose cushion (20% sucrose) and then, the bacterial cell walls were digested once with TES buffer (10 mM Tris-HCl pH 5.8, 1 mM EDTA, 25% sucrose) and then, the bacterial cell walls were digested with 200 μL of TES-LLP (TES buffer containing 10 mg/mL lysozyme, 100 μg/mL lysostaphin, and 1 mM phenylmethylsulfonyl fluoride). After 1 h of incubation at 37°C, the protoplasts were recovered by a 10 min centrifugation at 2,000 g. The pellet was washed with TES buffer and resuspended in 100 μL of TES buffer: PAGE loading buffer (1:1). The supernatant (cell wall fraction) was directly mixed with 50 μL of loading buffer for PAGE analysis.

Bacterial protein extracts were subjected to SDS-PAGE and Western blot analysis. For this, SDS-PAGE (10%) gels containing 1% casein at 4°C overnight, the membranes were washed and incubated with a 1/1000 dilution of a mouse polyclonal anti-VP6 mouse antibody diluted with PAGE loading buffer (1:1). The supernatant (cell wall fraction) was directly mixed with 50 μL of loading buffer for PAGE analysis. To obtain total protein extracts, 50 μL of SDS (20%) was added after cell wall digestion and a 1:1 dilution was made with PAGE loading buffer.

Bacterial protein extracts were subjected to SDS-PAGE and Western blot analysis. For this, SDS-PAGE (10%) gels were Coomassie blue-stained or blotted onto PVDF membranes. After blocking with PBS (PBS, 0.2% Tween-20) containing 1% casein at 4°C overnight, the membranes were incubated with a 1/3000 dilution of a mouse polyclonal antibody anti-RRV rotavirus at 37°C for 1 h. Then, the membranes were washed and incubated with a 1/1000 dilution of HRP-conjugated goat anti-mouse IgG (Pierce Biotechnology, Rockford, IL, USA) at 37°C for 1 h followed by detection with a chemiluminescent substrate (PBL, Bernal, Argentina) according to the manufacturer's instructions. Purified and concentrated RRV rotavirus proteins were included as a positive control.

To further confirm the display of the VP6-CWA fusion protein on the bacterial surface, induced *L. lactis* NZ9000/pCWA:VP6 cultures were analyzed by flow cytometry. For this, cultures were centrifuged and washed with PBS and incubated with an anti-VP6 mouse monoclonal antibody diluted in PBS containing 1% casein for 30 min at 37°C. This antibody was produced according to standard procedures [21] by fusing myeloma cells with splenocytes obtained from
mice immunized with purified rotavirus. After washing, cells were incubated with a FITC-conjugated rabbit anti-mouse IgG (Caltag Laboratories, Burlingame, CA, USA). For each sample, 100,000 events were acquired on a FACS calibur flow cytometer (Becton Dickinson, Immunocytometry Systems, San Jose, CA, USA) by gating L. lactis on forward scatter and side scatter dot plots. A band-pass filter of 530 nm (515 to 545 nm) was used to register the cells emitting green fluorescence (FLI-H). Uninduced L. lactis NZ9000/pCWA:VP6 cultures were processed and stained in the same way and used as negative control.

2.6. Preparation of Bacterial Cells for Immunization. Bacterial cultures were optimally induced and cell pellets were washed three times with PBS. Induced (NZ9000/pCWA:VP6) or plasmid-free bacteria (NZ9000) were resuspended in PBS to obtain $10^9$ colony-forming units/mL (CFU/mL).

2.7. Mice Immunization and Sample Collection. Groups of ten male BALB/c mice (6 to 8 weeks old) were inoculated subcutaneously on days 1, 14, and 28 with $1 \times 10^9$ CFU of induced L. lactis NZ9000/pCWA:VP6. Control groups of mice received identical quantities of plasmid-free bacteria NZ9000 or PBS. The final dose volume per mouse was 100 µL. Mice were bled on days 0, 14, 28, and 43 and sera were stored individually at −20°C until use. All animal procedures were conducted in accordance with the regulations of the Ethics Committee of the Universidad Nacional de Quilmes.

2.8. ELISA Analysis of Mice Sera. ELISA 96-well plates were coated overnight at 4°C with concentrated RRV rotavirus strain in carbonate buffer. Sera were tested in twofold dilution series (in PBST containing 1% casein) and plates were incubated for 1 h at 37°C. Mouse polyclonal antibodies anti-RRV rotavirus were used 1/1000 as positive control. After washing, membranes were incubated with HRP-conjugated goat anti-mouse IgG (Pierce Biotechnology, Rockford, IL, USA) followed by detection using a chemiluminescent substrate (Kalium Technologies, Bernal, Argentina).

2.9. Western Blot Analysis of Mice Sera. Purified and concentrated RRV rotavirus proteins were separated by SDS-PAGE (10%) and transferred onto PVDF membranes. After blocking with PBST containing 1% casein, the membranes were incubated with mice serum samples (1/100 dilution). Mouse polyclonal antibodies anti-RRV rotavirus were used 1/1000 as positive control. After washing, membranes were incubated with HRP-conjugated goat anti-mouse IgG (Pierce Biotechnology, Rockford, IL, USA) followed by detection with a chemiluminescent substrate (Kalium Technologies, Bernal, Argentina).

3. Results and Discussion

3.1. Expression of VP6-CWA in L. lactis. The VP6 sequence from the murine rotavirus EC strain was PCR amplified, cloned, and transferred into L. lactis NZ9000 obtaining NZ9000/pCWA:VP6. The nucleotide sequence analysis confirmed that there were no variations in the VP6 sequence and that it was in frame with both SP and CWA fragments. To determine if pCWA:VP6 allowed the expression of VP6-CWA, total protein extracts of induced cultures were analyzed by SDS-PAGE. As can be seen in Figure I(a) after only one hour of induction, one band corresponding to the expected size of the VP6-CWA fusion (62.9 kDa) was detected. It is important to note that this protein was absent in the extract from uninduced cultures (Figure I(a), lane 0 h).
3.2. VP6 Expression Optimization and Localization. To confirm that VP6-CWA fusion protein was properly attached to the cell wall, cultures of *L. lactis* NZ9000/pCWA:VP6 were analyzed by cell fractioning and Western blot of protein extracts using an antirotavirus polyclonal serum. Analysis of the protein content of the cell wall fraction (CW) revealed the presence of the band corresponding to VP6-CWA, which as expected was also detected in the protoplast fraction (PP) [19] (Figure 2(a)). A multiple-banding pattern was observed in both fractions (data not shown) as seen in total cell extracts (Figure 1(b)). VP6 was not detected in the supernatant.
fraction of induced cultures, even after concentrating with trichloroacetic acid (data not shown) [20].

To determine optimal expression conditions, cultures were fractioned and analyzed by Western blot at different time points between 0 and 6 h after induction at different starting OD<sub>600</sub> (0.2, 0.5, and 0.8) and increasing nisin concentrations (0, 1, 10, 50, 100, 200, and 500 ng/mL). Figures 2(a) and 2(b) show example blots obtained during expression optimization and Figure 2(c) shows growth curves plotted for every nisin concentration. To summarize, VP6-CWA expression was found to be optimal when L. lactis NZ9000/pCWA:VP6 cultures were induced at 0.5 OD<sub>600</sub> for two hours at 30°C with 100 ng/mL of nisin, which showed the highest targeting efficiency with minimal bacterial growth impact. These conditions were used for further experiments.

The targeting efficiency of VP6-CWA (the ratio of VP6-CWA protein detected in the cell wall as a fraction of total VP6-CWA protein detected) could be estimated to be around 40% under optimal conditions as determined by Western blot densitometry. The band considered for this estimate was that corresponding to undegraded VP6-CWA. This means that VP6-CWA is efficiently exported to the cell wall, in accordance with previous results obtained with L. lactis NZ9000/pCWA:Nuc [19].

In order to confirm VP6 localization, optimally induced L. lactis NZ9000/pCWA:VP6 cultures were analyzed by flow cytometry (Figure 3). The right shift to higher fluorescence values observed for induced L. lactis (Figure 3, black line) not only confirmed VP6 was present on the cell wall, but it also reflected that the protein was properly exposed on the outer side of the L. lactis NZ9000/pCWA:VP6 cell wall.

3.3. Immunogenicity of VP6-CWA in Mice. To evaluate whether VP6-CWA produced by L. lactis could induce a VP6 specific humoral response, 1 × 10<sup>9</sup> CFU of L. lactis NZ9000/pCWA:VP6 were used to immunize mice on a three-dose schedule via the subcutaneous route. Immune sera were analyzed by testing their reactivity against VP6 by ELISA (Figure 4(a)) and Western blot (Figure 4(b)). Mice immunized with L. lactis NZ9000/pCWA:VP6 exhibited a marked increase in specific serum IgG compared to mice immunized with plasmid-free NZ9000 or PBS. Rotavirus-specific antibodies could be detected in sera at day 28 following first immunization while sera from control mice remained negative after completing the immunization protocol (Figure 4(a)). The highest anti-VP6 IgG titer was obtained after the third booster immunization reaching an average endpoint titer of 2280 (95% confidence interval: 1000–3560) for mice immunized with L. lactis NZ9000/pCWA:VP6 and was found to be significantly different when compared to sera from control mice immunized with L. lactis NZ9000 (P < 0.05) or PBS (P < 0.01). Representative membrane strips for sera from each immunized group analyzed by Western blot are shown in Figure 4(b) and indicate that sera from mice immunized with L. lactis NZ9000/pCWA:VP6 were directed against VP6, while sera from control mice did not recognize rotavirus proteins.

Although antigen production at the L. lactis cell wall is in general less efficient compared to intracellular production [22], the obtained results indicated that the amount of VP6, produced by genetically engineered L. lactis NZ9000/pCWA:VP6, was sufficient for eliciting a specific immune response against rotavirus.

Moreover, it has been previously described that, for some antigens expressed in L. lactis, only cell-wall associated but not secreted or intracytoplasmic expression strategies were able to induce specific IgG in serum [23]. On the contrary, other antigens like tetanus toxin fragment C (TTFC) [24–26] did induce a specific immune response regardless of the subcellular localization. This demonstrates that not only the subcellular location of antigen expression affects the immune response generated, but the antigen’s characteristics are relevant as well.

In this particular case, VP6 was chosen as the rotavirus antigen for expression in L. lactis, considering that although antibodies against the VP6 protein are not associated with classical extracellular neutralizing activity, they have been associated with protection in some studies. For example, it has been demonstrated that monoclonal antibodies directed against VP6 protect mice against rotavirus infection by intracellular interference of the viral cycle, when hybridoma cells are injected into the backs of immunodeficient mice [27, 28]. In addition, a DNA vaccine encoding VP6 induced protective active immunity in mice [29, 30], and an E. coli-expressed fusion VP6-maltose-binding protein or a 14-amino-acid VP6 peptide induced protection from viral challenge [31]. Furthermore and importantly, this protection is heterotypic since VP6, which represents 51% of the virion mass, is antigenically conserved among most circulating group A rotavirus strains [14, 32, 33].

The results obtained so far with L. lactis NZ9000/pCWA: VP6 indicate that the VP6 protein is efficiently expressed and correctly displayed on the cell wall and that specific antibodies can be elicited, demonstrating correct folding of the epitopes and good immunogenicity. These recombinant bacteria can now be further explored as a mucosal delivery vehicle to be administered via oral or intranasal routes. In this
sense, mucosal immunization is mandatory to determine if a protective immune response against rotavirus can be elicited, by testing this VP6-expressing vector on the mouse model of rotavirus infection [34–36]. Furthermore, research on inoculation routes as well as immunization protocols will bring insight into this bacterial display platform strategy, suggesting a safer alternative to attenuated viral pathogens, which is the current strategy for human immunization against rotavirus [5, 7]. Additionally, these bacterial vectors expressing heterologous proteins are economical to produce and have a great potential for large-scale use of the NICE system [37].

4. Conclusions

This is the first time *L. lactis* surface display system was exploited as a means of expressing the rotavirus VP6 protein in the form of an immunogenic cell-wall anchored fusion protein. VP6 expression levels were optimized in order to improve cell-wall anchoring and surface exposure. Furthermore, *L. lactis* NZ9000/pCWA:VP6 proved to induce rotavirus-specific serum antibodies in a mouse model in the absence of any exogenous adjuvant. These results provide the basis for this bacterial vector to be further evaluated as a strategy for mucosal immunization against rotavirus in a mouse model of rotavirus infection.

Acknowledgments

The authors would like to thank Dr. Christian Magni (Instituto de Biologia Molecular y Celular de Rosario, Universidad Nacional de Rosario, Argentina) and Dr. Luis Bermúdez-Humarán (INRA-Institut National de la Recherche Agronomique, Jouy-en-Josas, France) for their assistance in the *L. Lactis* expression system. L. E. Estebarán and C. F. Tempran are recipients of doctoral and postdoctoral fellowships, respectively, granted by CONICET (Comisión Nacional de Investigaciones Científicas y Técnicas, Argentina). This work was supported by a research grant from the Universidad Nacional de Quilmes.

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