Clinical Study

Impact of Genetic Variants of Apolipoprotein E on Lipid Profile in Patients with Parkinson’s Disease


1 Department of Molecular Biology, Sao Jose do Rio Preto Medical School (FAMERP), 15090000 Sao Jose do Rio Preto, SP, Brazil
2 Sao Jose do Rio Preto Medical School (FAMERP), Center of Research in Biochemistry and Molecular Biology, NPBIM, Avenida Brigadeiro Faria Lima 5416, Vila Sao Pedro, 15090000 Sao Jose do Rio Preto, SP, Brazil
3 Federal University of São Paulo (UNIFESP), 04021001 São Paulo, SP, Brazil
4 Hospital de Base, 15090000 Sao Jose do Rio Preto, SP, Brazil
5 Sao Jose do Rio Preto Medical School (FAMERP), 15090000 Sao Jose do Rio Preto, SP, Brazil
6 Department of Neuroscience/Molecular Biology, Sao Jose do Rio Preto Medical School (FAMERP), 15090000 Sao Jose do Rio Preto, SP, Brazil
7 Department of Molecular Biology, Sao Jose do Rio Preto Medical School—FAMERP, 15090000 Sao Jose do Rio Preto, SP, Brazil

Correspondence should be addressed to Marcela A. S. Pinhel; marcelapinhel@yahoo.com.br

Received 12 April 2013; Revised 7 August 2013; Accepted 19 August 2013

Academic Editor: Patrick Kehoe

Copyright © 2013 Michele L. Gregório et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The pathogenesis of Parkinson’s disease (PD) seems to involve genetic susceptibility to neurodegeneration. APOE gene has been considered a risk factor for PD. This study aimed to evaluate the association of APOE polymorphism with PD and its influence on lipid profile. We studied 232 PD patients (PD) and 169 individuals without the disease. The studied polymorphism was analyzed by PCR/RFLP. The Fisher’s exact test, chi-square, ANOVA, and t-test (P < 0.05) were applied. The APOE3/3 genotype was prevalent in PD patients and Controls (P = 0.713) followed by APOE3/4 (P = 0.772). Both groups showed recommended values for lipid profile, with increase in the values of total cholesterol and LDLc, as well as decreased values of triglycerides in PD patients compared with Controls (P < 0.05 for all of them). Increased levels of HDLc, in PD patients, were associated with the APOE3 genotype versus APOE-/4 genotypes (P = 0.012). The APOE polymorphism does not distinguish PD patients from Controls, as opposed to the lipid profile alone or in association with APOE. Furthermore, a relationship between increase of HDLc levels and APOE3 in homozygous was found in PD patients only.

1. Introduction

Parkinson’s disease (PD) is a complex neurodegenerative disorder, chronic and progressive, affecting 2% of the population older than 65 years [1]. The identification of specific biomarkers for PD is one of the main goals of this clinical research. Today the diagnosis of this second most common neurological disease is possible by clinical evaluation of extrapyramidal signs, such as tremor, rigidity, and bradykinesia. These symptoms occur when the degeneration of nigral dopaminergic neurons of substantia nigra (SN), which project to the striatum [1], disrupting the motor circuit of the basal ganglia, has risen over 70% [2–4]. It has been suggested that cognitive deficit is a common feature of PD [1].

In Brazil, the incidence of PD is equivalent to 150/200 cases per 100,000 inhabitants, with the emergence of 20/100,000 new cases per year [5]. The etiology of PD seems to involve genetic susceptibility and environmental factors [6]. Some evidence shows that mitochondrial dysfunction, oxidative stress, and genetic factors play an important role in
the pathogenesis of this disease [7]. Approximately 85% of PD cases are sporadic, 10–15% of patients have family history, and less than 5% of them present monogenic inheritance [8].

The apolipoprotein E (ApoE) plays an important role in the lipoprotein metabolism [9] and transport of lipids to tissues [10]. It serves as a ligand for at least two specific receptors of lipoproteins, the low density lipoprotein receptor (LDLR) or ApoB/E and the liver receptor to apoE, the lipoprotein receptor-related proteins (LRP = LDL receptor related protein), allowing the removal of these particles by the liver [11]. The apolipoprotein E (ApoE) is also the main apolipoprotein in the central nervous system, with evidence of its association with cerebrovascular diseases [12], and neurodegenerative diseases as late onset of Alzheimer’s Disease (AD) [13,14] and PD [15,16]. Thus, APOE gene has become a significant target for investigation in neurodegenerative diseases.

The APOE gene, containing four exons, is mapped on the human chromosome 19q13.2 [17]. The APOE polymorphism, located on exon 4, is identified in the form of three major alleles APOE2 (prevalence of 0.01 to 0.15), APOE3 (0.49 to 0.91), and APOE4 (0.06 to 0.37) [18], which determines three protein isoforms (E2, E3, and E4, resp.) and six possible genotypes (e2/e2, e2/e3, e2/e4, e3/e3, e3/e4, and e4/e4). E2 and E3 clear plaques 20 times more efficiently than E4 [19]. E3 seems to be the normal isoform in all known functions, while E4 and E2 can each be dysfunctional [19]. Some researchers and a meta-analysis study have shown that the e2 allele is associated with higher risk of PD development [18, 20, 21], whereas other studies have shown that the e4 allele is responsible for PD development [15, 21–23]. Therefore, both the e2 and the e4 alleles may play a role in PD development. APOE is responsible for clearance of the b-amyloid plaques which impair the nervous system [24]. Therefore, the APOE4 allele is associated with high concentrations of LDLc and APOE2 at low plasma levels of LDLc [25]. The apoE is synthesized by astrocytes within the brain, and among its polymorphisms, the APOE4 allele, in particular, seems to have a risk effect on PD and a possible relationship with the existing neurodegeneration among Parkinson’s disease patients. Some studies have considered e4 as a risk factor for the age of onset for PD, decline in cognitive impairment, and/or development of dementia in PD. APOE2 has also been identified as an important risk factor for PD, however, with weak and inconsistent effect across studies [1, 26–34].

Therefore, the change in lipid metabolism mediated by different isoforms of apoE could influence the neuronal degenerative processes. Several mechanisms have been proposed associating increased risk of PD development with higher plasma levels of total cholesterol (TC), low density lipoprotein cholesterol (LDLc), high density lipoprotein cholesterol (HDLc) and very low density (VLDLc), and triglycerides (TG). Lipid profile tests were partially done for the PD patients (N = 86) as it was difficult to collect enough biological material (peripheral blood) for all analyses (genetic and biochemical). The study of APOE polymorphism (rs429358 and rs7412) was performed in the Center for Research in Biochemistry and Molecular Biology of Sao Jose do Rio Preto Medical School and consisted of genomic DNA extraction from whole blood samples (5 mL) [41] and DNA amplification by conventional PCR (polymerase chain reaction) and enzymatic restriction (Hha I).

Peripheral blood was collected in order to obtain analysis of genetic polymorphisms for APOE and lipid profile, including plasma levels of total cholesterol (TC), low density lipoprotein cholesterol (LDLc), high density lipoprotein cholesterol (HDLc) and very low density (VLDLc), and triglycerides (TG). Lipid profile tests were partially done for the PD patients (N = 86) as it was difficult to collect enough biological material (peripheral blood) for all analyses (genetic and biochemical). The study of APOE polymorphism (rs429358 and rs7412) was performed in the Center for Research in Biochemistry and Molecular Biology of Sao Jose do Rio Preto Medical School and consisted of genomic DNA extraction from whole blood samples (5 mL) [41] and DNA amplification by conventional PCR (polymerase chain reaction) and enzymatic restriction (Hha I).

Each reaction was performed in Eppendorf-Mastercycler thermocycler; each tube contained 0.5 μL of nucleotides (0.8 mM), 2.5 μL of buffer PCR 10X, 2.5 μL of dimethyl sulfoxide 10%, 2.5 μL of each primer (2.5 μM), 0.2 μL of Taq polymerase (5 U/μL), 11 μL of Milli Q water, and 2 μL of diluted genomic DNA (0.2 μg). Complementary primers were used in regions next to the polymorphic codons I12 and I158, located in exon 4 of APOE (rs429358 and rs7412) PI: 5‘-ACA GAA TTC GCC CGG CCT GGT ACA C-3’ and P2: 5‘-TAA GCT TGG CAC GGC TGT CCA AGC A-3’ [42]. Initial DNA denaturation was obtained at 94°C during 5 minutes, and the reaction mix submitted to 40 cycles of 94°C during
Table 1: Distribution of absolute and relative frequencies of alleles and genotypes of APOE polymorphism in individuals with Parkinson's disease (PD) and without (Controls).

<table>
<thead>
<tr>
<th>Genotype</th>
<th>PD (N = 232)</th>
<th>Controls (N = 137)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>%</td>
<td>N</td>
</tr>
<tr>
<td>APOE2/4</td>
<td>1</td>
<td>0.4</td>
<td>0</td>
</tr>
<tr>
<td>APOE2/3</td>
<td>19</td>
<td>8.1</td>
<td>8</td>
</tr>
<tr>
<td>APOE3/3</td>
<td>169</td>
<td>73.0</td>
<td>103</td>
</tr>
<tr>
<td>APOE3/4</td>
<td>37</td>
<td>15.9</td>
<td>24</td>
</tr>
<tr>
<td>APOE4/4</td>
<td>6</td>
<td>2.6</td>
<td>2</td>
</tr>
<tr>
<td>Total</td>
<td>232</td>
<td>100</td>
<td>137</td>
</tr>
<tr>
<td>APOE-/-4</td>
<td>44</td>
<td>18.9</td>
<td>26</td>
</tr>
<tr>
<td>APOE-/-2</td>
<td>20</td>
<td>8.6</td>
<td>8</td>
</tr>
</tbody>
</table>

Allele

<table>
<thead>
<tr>
<th>Allele</th>
<th>PD (N = 232)</th>
<th>Controls (N = 137)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td>APOE2</td>
<td>20</td>
<td>0.04</td>
<td>8</td>
</tr>
<tr>
<td>APOE3</td>
<td>394</td>
<td>0.85</td>
<td>238</td>
</tr>
<tr>
<td>APOE4</td>
<td>50</td>
<td>0.11</td>
<td>28</td>
</tr>
<tr>
<td>Total</td>
<td>464</td>
<td>1.0</td>
<td>274</td>
</tr>
</tbody>
</table>

*p Fisher or Chi-squared tests; N: total number of individuals; Abs. Freq.: absolute frequency.

Table 2: Distribution of means and standard deviation for lipid profile in patients with Parkinson's disease (PD) and without (Controls).

<table>
<thead>
<tr>
<th>Biochemical profile</th>
<th>PD (N = 86)</th>
<th>Controls (N = 169)</th>
<th>p*</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC</td>
<td>Mean</td>
<td>202.5</td>
<td>186.8</td>
</tr>
<tr>
<td>SD</td>
<td></td>
<td>45.7</td>
<td>51.1</td>
</tr>
<tr>
<td>HDLc</td>
<td>Mean</td>
<td>58.0</td>
<td>56.8</td>
</tr>
<tr>
<td>SD</td>
<td></td>
<td>13.2</td>
<td>20.9</td>
</tr>
<tr>
<td>LDLc</td>
<td>Mean</td>
<td>122.0</td>
<td>102.8</td>
</tr>
<tr>
<td>SD</td>
<td></td>
<td>38.7</td>
<td>44.9</td>
</tr>
<tr>
<td>VLDLc</td>
<td>Mean</td>
<td>22.8</td>
<td>31.3</td>
</tr>
<tr>
<td>SD</td>
<td></td>
<td>13.3</td>
<td>29.4</td>
</tr>
<tr>
<td>TG</td>
<td>Mean</td>
<td>115.5</td>
<td>136.9</td>
</tr>
<tr>
<td>SD</td>
<td></td>
<td>66.7</td>
<td>73.5</td>
</tr>
</tbody>
</table>

*p-t-test; TC: total cholesterol; HDLc: high density lipoprotein cholesterol; LDLc: low density lipoprotein cholesterol; VLDLc: very low density lipoprotein cholesterol; TG: triglycerides; SD: standard deviation; N: number of individuals.

30 seconds and 65°C during 2 minutes, extension at 72°C during 1 minute, and ending cycle at 72°C during 7 minutes [42].

The analysis of ApoE genetic variants was performed by restriction fragment length polymorphism (RFLP) analysis, with the restriction enzyme Hha I fast (5 U per reaction tube) in double boiler at 37°C during 45 minutes, for cleavage of amplified sequences in specific regions (GCGC), identifying the alleles APOE2, APOE3, and APOE4. The DNA fragments were separated by electrophoresis on 6% polyacrylamide gel nondenatured, under constant electric current of 180 V during 2 hours. A standard DNA sample (100 base pairs, Invitrogen) was used as comparison with the electrophoretic bands of patients. The gel was colored by GelRed (Uniscience of Brazil) during 10 minutes, and DNA fragments were visualized under ultraviolet light (UV).

3. Statistical Analysis

Categorical variables (including demographic data and genetic variants) were analyzed applying Fisher’s exact test and the Chi-square test. Continuous variables (age, values for lipid profile) were analyzed by ANOVA, Tukey, t-test, or Mann-Whitney test. P < 0.05 was considered statistically significant. Statistical analysis also included Hardy-Weinberg equilibrium (Fisher’s exact test and the Chi-square test).

4. Results

Table 1 shows the genotype distribution and allele frequencies of the APOE polymorphism. We observed the prevalence of APOE3/3 genotype in PD patients (73.0%) and Controls (75.2%, P = 0.713), followed by APOE3/4 (15.9%, 17.5%, resp.; P = 0.772). There was a similar distribution of alleles of the ApoE in both groups (P > 0.05).

Concerning the lipid profile (Table 2), both groups showed values within the reference limits of normality, except for patients with slightly increased TC mean value (202.5 ± 45.7 mg/dL). This group showed higher TC and LDLc levels (122.0 ± 38.7 mg/dL) and lower VLDLc and TG levels (22.8 ± 13.3 mg/dL), compared with Controls (186.8 ± 51.1 mg/dL; P = 0.017; 102.8 ± 44.9 mg/dL; P = 0.001; 31.3 ± 29.4 mg/dL; P = 0.001; 136.9 ± 73.5 mg/dL; P = 0.024, resp.).

Table 3 shows the relationship between lipid profile and APOE polymorphism. PD patients present a relationship between APOE-/-4 and a decrease in HDLc levels (51.8 ± 10.5 mg/dL), compared with APOE3/3 (60.3 ± 13.3 mg/dL; P = 0.025). Furthermore, patients with APOE3/3 genotype showed significantly higher levels of HDLc (60.3 ± 13.3 mg/dL) and reduced levels of VLDLc and TG (22.7 ± 13.1 mg/dL; 113 ± 65.7 mg/dL, resp.), compared with Controls with the same genotype (52.0 ± 15.5 mg/dL; P = 0.001; 36.6 ± 37.4 mg/dL; P = 0.001; 143.5 ± 79.8 mg/dL; P = 0.017, resp.). In Controls, individuals with APOE3/3 genotype presented higher values of VLDLc, compared with those with APOE-/-4 genotypes (P = 0.002).

5. Discussion

This study shows similar genotype distribution and allele frequency for the APOE polymorphism in PD patients and Controls, as described by other authors [1, 25–28]. On
Table 3: Means and standard deviations for lipid profile in patients with Parkinson's disease (PD) and without (Controls) considering APOE polymorphism.

<table>
<thead>
<tr>
<th>Biochemical profile (mg/dL)</th>
<th>PD (N = 232)</th>
<th>Controls (N = 137)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>APOE-/-4 (a) (N = 15)</td>
<td>APOE3/3 (b) (N = 58)</td>
<td></td>
</tr>
<tr>
<td>TC</td>
<td>206.5</td>
<td>203.7</td>
<td>200.7</td>
</tr>
<tr>
<td>LDLc</td>
<td>128.7</td>
<td>121.7</td>
<td>123.1</td>
</tr>
<tr>
<td>HDLc</td>
<td>51.8</td>
<td>60.3</td>
<td>56.5</td>
</tr>
<tr>
<td>VLDLc</td>
<td>24.3</td>
<td>22.7</td>
<td>22.6</td>
</tr>
<tr>
<td>TG</td>
<td>126.0</td>
<td>113.3</td>
<td>129.7</td>
</tr>
<tr>
<td>SD</td>
<td>48.7</td>
<td>47.4</td>
<td>59.4</td>
</tr>
<tr>
<td></td>
<td>43.3</td>
<td>38.8</td>
<td>55.0</td>
</tr>
<tr>
<td></td>
<td>10.5</td>
<td>13.3</td>
<td>15.9</td>
</tr>
<tr>
<td></td>
<td>17.1</td>
<td>13.1</td>
<td>9.0</td>
</tr>
<tr>
<td></td>
<td>82.5</td>
<td>65.7</td>
<td>83.5</td>
</tr>
</tbody>
</table>

*One-Way ANOVA Test; †t-test; TC: total cholesterol; HDLc: high density lipoprotein cholesterol; LDLc: low density lipoprotein cholesterol; VLDLc: very low density lipoprotein cholesterol; TG: triglycerides; SD: standard deviation; N: number of individuals.

The other hand, a meta-analysis study highlights the increased susceptibility to the disease associated with presence of the APOE2 allele (odds ratio: 1.16; confidence interval 95%: 1.03–1.31) [43], however, not observed in this study. Differences among studies may be explained by changes in methodology, sample size, age of manifestation of the disease, ethnicity, and other environmental and geographic factors which can be considered in the analysis of genetic polymorphisms.

ApoE genotype distribution differs among populations. Eichner et al. [44] reviewed the frequencies of E2, E3, and E4 alleles in different populations and reported that these range from 0.02 to 0.13 for E2, 0.06 to 0.85 for E3, and 0.11 to 0.31 for E4. The prevalence of the E4 allele in Brazil is similar to that observed in other South American countries with frequencies between 0.23 and 0.26 [14], except for Chileans with a frequency of 0.40. However, in Controls, the prevalence ranges between 0.08 and 0.19 [45]. These frequencies for the E4 allele are lower than in populations from the northern hemisphere, whose frequencies vary between 0.38 and 0.48 [46, 47].

The possible association of apoE and risk of PD has been widely investigated in different populations. However, it remains underrepresented in the Brazilian population. Thus, this is a pioneer study in terms of the distribution of ApoE genetic polymorphisms and its relationship with lipid profiles in patients with PD, helping understand possible genetic markers for this disease. Considering the relationship of apoE and lipid metabolism [48], we also evaluated the lipid profile. The mean values were within the reference limits except for TC, with slight increase in PD patients [49], who also showed increased levels of LDLc and reduction in VLDLc and TG levels, compared with Controls.

Clinical and subclinical conditions [50, 51] can be associated with reduced concentrations of cholesterol (TC, LDLc and HDLc), as observed in inflammatory diseases [50, 52]. Studies also show changes in lipid metabolism associated with neurodegenerative diseases, including Alzheimer’s disease and PD [53, 54]. In this case, there is reference to reduction in the synthesis of cholesterol in skin fibroblasts from patients with PD [55], as well as low values of TC compared with Controls [35].

In an analysis stratified by gender, Pena et al. [40] observed a reduction in the risk of PD with increased levels of TC only in women. In men, only higher values of TC accounted for reduced risk for PD, compared with men with lower values of TC. However, de Lau et al. [38] found no relationship among LDLc concentration, duration of PD, and use of dopaminergic agents. Therefore, both authors suggest that the relationship between low levels of LDLc and PD is a consequence of the disease or its treatment.

Additionally, there is speculation that high levels of TC raise the risk of PD [36], partly due to the excess of body weight, as some studies observed a relationship between excess weight and a higher risk for PD [36, 56]. Also, there is an inverse association between intake of total fat and unsaturated fatty acids with risk of PD [57, 58]. Thus, a number of factors strongly influence the variation of lipid profile in PD patients, which requires strict criteria to be applied on the selection of the studied groups.

This study also highlights the clear influence of apoE polymorphisms and lipid profile between the groups. In this case, PD patients showed increased HDLc levels, compared with Controls, only in those patients with APOE3/3 genotype,
who also showed reduced levels of TG and VLDLc when compared with Controls. Additionally, only PD patients showed the relationship between the presence of APOE4 and reduced serum levels of HDLc, compared with APOE3/3 genotype. Thus, PD patients and Controls show differences in the combination of lipid profile and the apoE-Hha I polymorphism. On the other hand, Huang et al. [18] detected increased risk of PD, associated with reduced plasma levels of TC related to the allele APOE2. Thus, these controversies may reflect changes in lifestyle habits, drug treatment, and medical advice to control the disease, which are aspects to be investigated in further studies.

Therefore, this study allows us to conclude that APOE polymorphism does not distinguish PD patients from Controls, as opposed to the lipid profile alone or in association with APOE. In this case, and increase in TC and LDLc levels can be observed in PD patients, whereas higher VLDLc and TG levels are prevalent in Controls. Moreover, only PD patients show a relationship between increase of HDLc and TG levels are prevalent in Controls. Additionally, only PD patients showed the relationship between increases of HDLc and APOE3 in homozygous. Further studies including subgroups of patients with and without family history of PD need to clarify the influence of genetic polymorphisms and their respective mechanisms in PD.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This paper was supported by Grants no. 2009/17222-0, no. 2008/53950-8, and no. 2009/18476-6, from São Paulo Research Foundation (FAPESP) and Sao Jose do Rio Preto Medical School (FAMERP).

References


[52] W. H. Ettinger Jr., T. Harris, "Low serum cholesterol and mortality: which is the cause and which is the effect?" *Circulation*, vol. 92, no. 9, pp. 2396–2403, 1995.


