Research Article

Characterization of C-S Lyase from *C. diphtheriae*: A Possible Target for New Antimicrobial Drugs

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The emergence of antibiotic resistance in microbial pathogens requires the identification of new antibacterial drugs. The biosynthesis of methionine is an attractive target because of its central importance in cellular metabolism. Moreover, most of the steps in methionine biosynthesis pathway are absent in mammals, lowering the probability of unwanted side effects. Herein, detailed biochemical characterization of one enzyme required for methionine biosynthesis, a pyridoxal-5′-phosphate (PLP-) dependent C-S lyase from *Corynebacterium diphtheriae*, a pathogenic bacterium that causes diphtheria, has been performed. We overexpressed the protein in *E. coli* and analyzed substrate specificity, pH dependence of steady state kinetic parameters, and ligand-induced spectral transitions of the protein. Structural comparison of the enzyme with cystalysin from *Treponema denticola* indicates a similarity in overall folding. We used site-directed mutagenesis to highlight the importance of active site residues Tyr55, Tyr114, and Arg351, analyzing the effects of amino acid replacement on catalytic properties of enzyme. Better understanding of the active site of *C. diphtheriae* C-S lyase and the determinants of substrate and reaction specificity from this work will facilitate the design of novel inhibitors as antibacterial therapeutics.

1. Introduction

The emergence of resistance to antibacterial agents is a pressing concern for human health. New drugs to combat this problem are therefore in great demand. Acquired bacterial resistance has caused several antibiotics to become useless or, at best, compromised in their ability to counteract bacterial infection [1]. The potential of amino acid biosynthesis as an antimicrobial target has been validated both chemically and biochemically [2]. Methionine represents a key amino acid in prokaryotes and it is an attractive antimicrobial target because of its important role in cell metabolism. Methionine, in the form of S-adenosylmethionine, is the methyl donor for a number of essential biochemical reactions. The biosynthesis of methionine is therefore of vital importance to microbial growth. This has been validated by the fact that several natural products, including 2-amino-5-hydroxy-4-oxopentanoic acid [3], azoxybacilin [4], and rhizoctcin [5], target important enzymes for methionine biosynthesis and have antimicrobial properties. Moreover, most of the steps in the methionine biosynthesis pathway are unique to bacteria and plants.

We have studied one of the enzymes required for methionine biosynthesis, namely, NCBI protein NP_940074, which has been annotated as a pyridoxal-5′-phosphate (PLP-) dependent C-S lyase from *Corynebacterium diphtheriae*, a pathogenic bacterium that causes diphtheria. The threat of diphtheria, even in countries with good coverage in childhood immunization programs by vaccination, has not disappeared and outbreaks in many European countries have revealed a phenomenon of decreasing immunity to diphtheria among adults [6]. The treatment of diphtheria patients is based on administration of antibiotics to eliminate the corynebacteria from the site of infection, thus stopping ongoing toxin production with or without additional treatment with specific immunoglobulins [7]. Thus, while diphtheria...
antitoxin (DAT) contributes to reducing the case fatality rates of infections due to this microorganism, it is unlikely, in the foreseeable future, that it will substantially reduce the world’s consumption of antimicrobial agents. This highlights the importance of the development of new antibacterial drugs.

C-S lyase catalyzes the α,β-elimination of sulfur-containing amino acids, such as L-cystathionine (L-Cth), to generate ammonia, pyruvate, and homocysteine, the penultimate step in methionine biosynthesis transsulfuration pathway. The crystal structure of *C. diphtheriae* C-S lyase has been determined by X-ray crystallography at 1.99 Å resolution (PDB code: 3FDB, Joint Center for Structural Genomics, http://www.jcsg.org/). According to its folding pattern, the enzyme belongs to the fold type I family of the PLP-dependent enzymes [8]. The commonly known fold type I PLP-dependent enzymes have an aromatic amino acid residue located at the *re* face of the PLP-Lys internal aldime and stacking with the pyridine ring of PLP. The structure shows that a conserved binding site for PLP which is covalently linked to Lys222 has a stacking interaction with Tyr114 and H-bond interaction with Asn60, Asp188, and His91. There is one molecule in each asymmetric unit (PDB code: 3FDB). Although the crystal structure has been solved, no protein characterization and structure-based design studies have been reported to date.

The availability of the protein in purified form has allowed us to obtain an insight into the biochemical properties of the enzyme by thorough characterization of its kinetic and spectral properties. With increasing resistance of bacteria to common antibiotics, C-S lyase appears to be an interesting novel target for the development of specific inhibitors against the pathogen *C. diphtheriae*.

### 2. Materials and Methods

#### 2.1. Chemicals.

PLP, L-cystathionine, L-cysteine, L-cysteine, L-djenkolic acid, L-serine, aminoethyl-L-cysteine, O-acetyl-L-serine, L-homoserine, L-methionine, phenylhydrazine hydrochloride, and 10mM imidazole), and lysed by sonication. The cell debris was removed by centrifugation (20,000 × g for 30 min) and the supernatant was loaded onto an Ni-affinity column equilibrated with 20 mM sodium phosphate at pH 7.5, 200 mM sodium chloride, and 10 mM imidazole. The imidazole concentration was increased stepwise, first to 30 mM to remove nonspecifically bounded proteins, and then to 500 mM to elute the enzyme. Monomer concentration was determined from the calculated extinction coefficient ($ε_{280\text{nm}} = 65048 \text{ M}^{-1} \text{cm}^{-1}$; http://web.expasy.org/protparam/). PLP content of holoenzyme was determined by releasing the coenzyme in 0.1 M NaOH and by using $ε_{388\text{nm}} = 6600 \text{ M}^{-1} \text{cm}^{-1}$. The yield from a standard purification was approximately 25 mg/L culture. The protein was stable at −80°C for many months and freezing and thawing did not alter the activity of the isolated enzyme.

#### 2.2. Enzyme Activity Assays.

Activity was measured at 30°C on a Jasco-V560 UV-Vis spectrophotometer. The assay buffer was composed of 50 mM MOPS, 50 mM bicine, 50 mM proline pH 9.0, and 20 μM PLP. Reactions were initiated by the addition of 1 μM enzyme. The activity of C-S lyase toward L-Cth was detected via two spectrophotometric assays by using the reaction of 5,5′-dithiobis-(2-nitrobenzoic acid) (DTNB) with the free thiol of the L-homocysteine product ($ε_{412\text{nm}} = 13600 \text{ M}^{-1} \text{cm}^{-1}$) and by monitoring pyruvate formation with the coupling enzyme NADH-dependent lactate dehydrogenase (LDH) which reduces the pyruvate product to lactate, with the concomitant conversion of NADH to NAD+ ($ε_{340\text{nm}} = 6200 \text{ M}^{-1} \text{cm}^{-1}$). The LDH assay was also employed to monitor C-S lyase activity toward other substrates.

Initial velocity data obtained by varying substrate concentrations were fitted to the hyperbolic form of the Michaelis-Menten equation

$$ \frac{v}{E_t} = \frac{k_{cat} S}{K_m + S}. \quad (1) $$

In the case of deviations from hyperbolic kinetics, initial velocity data obtained by varying substrate concentrations were fitted to a nonhyperbolic curve to (2) that takes into account substrate inhibition, that is, binding of a second molecule of substrate to the ES complex to form an inactive ternary complex SES [11]:

$$ \frac{v}{E_t} = \frac{k_{cat}}{1 + (K_m S) + (S/K_i)}, \quad (2) $$

where $E_t$ is the total enzyme concentration, $k_{cat}$ is the maximum velocity, $S$ is the substrate concentration, $K_m$ is the...
apparent Michaelis-Menten constant, and $K_i$ is the dissociation constant for the inhibitory SES ternary complex.

2.5. Evaluation of the pH Dependence. The pH dependence for C-S lyase towards L-Cth and L-cysteine was determined using both DTNB and LDH assays in 50 mM MOPS, 50 mM bicine, 50 mM proline, and 20 μM PLP. Kinetic measurements were carried out between pH 7.8 and 10.3. The values for $\log k_{\text{cat}}/K_m$ as a function of pH were fitted to the appropriate equations:

$$\log Y = \log \frac{C}{1 + (H/K_A)},$$

$$\log Y = \log \frac{C}{1 + (H/K_A) + (K_B/H)},$$

where $K_A$ and $K_B$ represent the ionization constants for enzyme or reactant functional group, $Y$ is the value of the parameter observed as a function of pH, and $C$ is the pH-independent value of $Y$. All data were fitted to the previous equations using the Origin8 program (OriginLab).

2.6. Spectroscopic Measurements. Absorption measurements were carried out on a Jasco V-560 UV-Vis spectrophotometer, using 10 μM enzyme in a buffer solution containing 50 mM MOPS, 50 mM bicine, and 50 mM proline pH 8.5.

Fluorescence spectra were obtained with a Jasco FP8200 spectrofluorometer using 5 nm bandwidths on both sides at a protein concentration varying from 1 to 10 μM in a solution containing 20 mM Bis-Tris propane pH 8.5. Spectra of blanks, that is of samples containing all component except the lyase, were taken immediately prior to the measurements of samples containing protein. Blank spectra were subtracted from spectra of samples containing enzyme.

CD measurements were carried out on a Jasco J-710 spectropolarimeter using 1 mg/mL enzyme in a buffer solution containing 50 mM Tris-HCl pH 8.5. For near-UV and visible wavelengths, three automatically averaged spectra were recorded in a cuvette with a 1 cm path length at a scan speed of 50 nm/min.

2.7. Preparation and Reconstitution of Apoenzyme. The coenzyme PLP was removed as a phenylhydrazone following the protocol in [12]. After the removal of the cofactor, C-S lyase exhibited no residual activity and no absorption peak at 413 nm. Titration of the apoenzyme with PLP was carried out in equilibrium by measuring the fluorescence emission at 500 nm ($\lambda_{\text{exc}} = 413$ nm) after incubation of apoprotein (5 μM) with various concentrations of the cofactor (0–100 μM) at 25 °C for 15 min in 20 mM Bis-Tris propane pH 8.5. The increase in fluorescence emission at 500 nm was dependent on added PLP and the values were used to calculate the $K_d$ according to a tight-binding hypothesis (5):

$$F_{500} = \Delta F_{\text{max}} \left( e_0 + l_0 + K_d - \sqrt{(e_0 + l_0 + K_d)^2 - 4e_0l_0} \right),$$

where $e_0$ and $l_0$ are the total concentrations of C-S lyase subunit and PLP, respectively, $K_d$ is the equilibrium dissociation constant for the encounter complex, and $\Delta F_{\text{max}}$ represents the fluorescence change at 500 nm recorded in the presence of an excess of PLP over apoprotein. The reconstituted enzyme completely recovered its original activity.

2.8. Size Exclusion Chromatography. The molecular weight of recombinant enzyme was monitored by analytical size exclusion chromatography (Superdex 200 HR 10/30) using AKTA FPLC system (GE Healthcare). The chromatography was performed using 20 mM sodium phosphate buffer pH 7.2 and 150 mM NaCl as the mobile phase, at a flow rate of 0.1 mL/min. The calibration curve was determined with gel filtration molecular weight standards containing bovine thyroglobulin (669 kDa), apoferritin (443 kDa), β-amylase (200 kDa), alcohol dehydrogenase (150 kDa), bovine serum albumin (66 kDa), carbonic anhydrase (29 kDa), and cytochrome c (12.4 kDa). The curve was linear between 12.4 and 669 kDa.

2.9. Structural Similarity Analysis. The structure of the C-S lyase in complex with the PLP-aminoethoxyvinylglycine (AVG) was obtained by optimal superposition of the main chain of the C-S lyase (PDB code: 3FD2) and the structure of the cystalysin from Treponema denticola (PDB code: 1C7O) as in [13].

3. Results

Recombinant C-S lyase was purified as a His-tagged protein to greater than 99% purity as confirmed by SDS-PAGE; the size of the protein, calculated with a molecular size marker, was about 44 kDa that corresponds well to the molecular mass elucidated from the sequence information. As determined by gel filtration, an apparent molecular mass of 85 kDa was calculated for the native enzyme. Therefore, these results indicate that, as suggested by interface interaction, recombinant C-S lyase from C. diphtheriae is a homodimer.

3.1. Spectral Properties of Recombinant C-S Lyase. The visible absorption spectrum of the native enzyme is dominated by a band centered around 413 nm (Figure 1). This peak indicates that the predominant tautomer of the internal aldimine is the ketenamine, and the imine nitrogen of the cofactor is protonated allowing the formation of an intramolecular hydrogen bond with the 3’-oxygen atom of PLP and the conjugation of the π system of the imine with the pyridine ring [14, 15].

The intensity of the band did not change in the pH range from 6 to 11, indicating a $pK_a$ of the internal aldimine higher than 11. The observed ratio $A_{380 \text{nm}}/A_{413 \text{nm}}$ was ~8. The stoichiometric ratio of PLP to enzyme was calculated from the $A_{380 \text{nm}}$ of the free cofactor, which was released upon incubation of the native holoenzyme in 0.1 M NaOH. By this method, a PLP content of 2 mol/mol of the native dimer was found.

As shown in Figure 2(a), the CD spectrum of the enzyme displays a positive dichroic band at 415 nm whose intensity does not change with pH. The near-UV spectrum is characterized by pronounced negative dichroic bands in the aromatic region at 260–296 nm, which would indicate the
Figure 1: Absorption spectra of C-S lyase in the presence of substrates and substrate analogues. (a) Absorption spectra of 10 μM C-S lyase in the absence (solid line) and presence of 100 mM L-serine (dashed line) and 100 mM O-acetyl-L-serine (dotted line). The spectra were subtracted from pyruvate absorbance. Pyruvate concentration was evaluated by a coupled LDH assay (see Section 2). (b) Absorption spectra of 10 μM C-S lyase in the absence (solid line) and presence of 450 mM L-homoserine (dotted line) and 150 mM L-methionine (dashed line). All spectra were recorded in a solution containing 50 mM MOPS, 50 mM bicine, and 50 mM proline pH 8.5.

Figure 2: CD and fluorescence emission spectra of C-S lyase in the presence of substrates and substrate analogues. (a) CD spectra of enzyme (solid line) in the presence of 450 mM L-homoserine (dotted line) and 150 mM L-methionine (dashed line) in 50 mM Tris-HCl pH 8.5. (b) Fluorescence emission spectra of 1 μM enzyme (solid line) in the presence of 150 mM L-methionine (dotted line), 100 mM L-serine (dashed line), and 100 mM O-acetyl-L-serine (dash dotted line) upon excitation at 295 nm. Inset: emission spectra of 10 μM enzyme (solid line) in the presence of 100 mM O-acetyl-L-serine (dash dotted line), 100 mM L-serine (dashed line), and 150 mM L-methionine (dotted line) upon excitation at 418 nm. All spectra are recorded in 20 mM Bis-Tris-propane pH 8.5.

asymmetry of certain aromatic amino acids, most likely associated with the active site (Figure 2(a)).

The fluorescence emission spectra of C-S lyase recorded for the direct excitation at 295 nm of Trp residues exhibit a pronounced peak centered around 340 nm and a much lower intensity peak at around 500 nm (Figure 2(b)). The peak at 340 nm can be attributed to direct emission from Trp residues, while the peak centered at 505 nm can be attributed to emission from PLP, resulting from an intramolecular energy-transfer process from the excited Trp residues to the cofactor [16, 17]. Direct excitation of the C-S lyase ketoamine tautomer of PLP at 413 nm gives a single emission peak centered at 502 nm (Figure 2(b), inset), which is typical for emission from Schiff bases of PLP in a neutral, aqueous
environment [18, 19]. None of the fluorescence emission properties of the enzyme is affected by pH in the range from 5.7 to 10.2.

At pH 8.5 in 20 mM Bis-Tris propane, the excitation spectrum for the longer-wavelength emission of C-S lyase measured at 502 nm shows two bands. The peak of the shorter wavelength excitation band is located at 279 nm, and the peak of the longer-wavelength excitation band is located at 413 nm (data not shown).

The emission spectrum obtained for apo C-S lyase, when excited at 295 nm, exhibits a single peak centered at 345 nm, with a 4-fold increase in emission intensity and a 2 nm red shift compared to the holoenzyme (data not shown). The apoenzyme has no residual activity and does not exhibit either absorbance bands in the visible region or PLP emission fluorescence. Titration of apo C-S lyase with PLP resulted in an increase in the fluorescence emission at 500 nm; the values were plotted against PLP concentrations and used to calculate the external aldimine is the only evident intermediate during identification of specific bands and is likely due to the accumulation of reaction products, that is, pyruvate, which absorbs at 318 nm [21].

The emission spectrum obtained for apo C-S lyase, when excited at 295 nm, exhibits a single peak centered at 345 nm, with a 4-fold increase in emission intensity and a 2 nm red shift compared to the holoenzyme (data not shown). The apoenzyme has no residual activity and does not exhibit either absorbance bands in the visible region or PLP emission fluorescence. Titration of apo C-S lyase with PLP resulted in an increase in the fluorescence emission at 500 nm; the values were plotted against PLP concentrations and used to calculate the $K_{d, PLP}$ value for the enzyme by a theoretical curve fit of the experimental data using (5). The obtained $K_{d, PLP}$ value was $0.09 \pm 0.03 \mu M$, indicating a tight binding of PLP to the enzyme.

3.2. Steady-State Kinetic Studies. Several sulfur- and nonsulfur containing amino acids were utilized as potential substrates for the enzyme. The enzyme showed maximum activity at pH 9 and 30°C, as determined in 50 mM MOPS, 50 mM bicine, and 50 mM proline. Table 1 shows that the enzyme has a relatively broad substrate specificity. L-Cth, L-djenkolate, and aminoethyl-L-cysteine were the most effective substrates. Cystine and L-cysteine, to a lesser extent, also acted as substrates. O-acetyl-L-serine and L-serine proved to be very poor substrates, whereas no activity could be detected with L-homocysteine, L-homoserine, or L-methionine. These latter inhibited, although to different extents, C-S lyase activity (data not shown), thus indicating their binding to the enzyme. The recombinant C-S lyase obeyed Michaelis-Menten kinetics with all substrates examined except for L-cysteine, for which substrate inhibition was observed. By using (2) (describing substrate inhibition, [11, 20]), initial velocity data obtained by varying L-cysteine concentrations fit a nonhyperbolic curve to yield $K_m$ and $K_i$ values of 0.95 ± 0.18 and 3.98 ± 0.66 mM, respectively.

3.3. pH Dependence of Kinetic Parameters for C-S Lyase. The pH dependence of the kinetic parameters for C-S lyase toward L-Cth and L-cysteine was determined, and the results are shown in Figures 3(a) and 3(b). $\log k_{cat}$ is pH independent from 7.8 to 10.2 for both substrates. The $\log k_{cat}/K_m$—pH profile for L-Cth is bell shaped, consistent with the involvement of two ionizable groups, one with an apparent $pK_a$ of 8.05 ± 0.07 and the other with an apparent $pK_a$ of 9.90 ± 0.07 (Figure 3(a)). The $\log k_{cat}/K_m$—pH profile for L-cysteine exhibited a single $pK_a$ of 8.26 ± 0.01 for a group that must be deprotonated for activity (Figure 3(b)).

The emission spectrum obtained for apo C-S lyase, when excited at 295 nm, exhibits a single peak centered at 345 nm, with a 4-fold increase in emission intensity and a 2 nm red shift compared to the holoenzyme (data not shown). The apoenzyme has no residual activity and does not exhibit either absorbance bands in the visible region or PLP emission fluorescence. Titration of apo C-S lyase with PLP resulted in an increase in the fluorescence emission at 500 nm; the values were plotted against PLP concentrations and used to calculate the $K_{d, PLP}$ value for the enzyme by a theoretical curve fit of the experimental data using (5). The obtained $K_{d, PLP}$ value was $0.09 \pm 0.03 \mu M$, indicating a tight binding of PLP to the enzyme.

3.4. Absorption, Fluorescence, and CD Spectral Changes of C-S Lyase with Substrates and Substrate Analogues. Except for L-serine and O-acetyl-L-serine, the rate of reaction for the substrates tested (Table 1) was very high, and reaction of the enzyme with these substrates cannot be studied by conventional spectroscopy.

Addition of O-acetyl-L-serine to C-S lyase at pH 8.5 produces an immediate shift from 413 to 420 nm (Figure 1(a)). This shift is consistent with the conversion of internal to external aldimine (Figure 1(a)). A broad increase in the absorbance at wavelengths shorter than 350 nm hampers the accurate identification of specific bands and is likely due to the accumulation of reaction products, that is, pyruvate, which absorbs at 318 nm [21]. After subtraction of the contribution of pyruvate absorbance to the absorbance spectra of enzyme, the external aldimine is the only evident intermediate during
### Table 1: Kinetic parameters for substrates in 50 mM MOPS, 50 mM bicine, and 50 mM proline pH 9.0 at 30°C.

<table>
<thead>
<tr>
<th>Substrate</th>
<th>$K_m$ (mM)</th>
<th>$k_{cat}$ (s$^{-1}$)</th>
<th>$k_{cat}/K_m$ (mM$^{-1}$ s$^{-1}$)</th>
<th>Structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>L-cystathionine</td>
<td>1.07 ± 0.22</td>
<td>92.67 ± 4.44</td>
<td>86.60 ± 21.46</td>
<td></td>
</tr>
<tr>
<td>L-djenkolic acid</td>
<td>0.23 ± 0.087</td>
<td>49.11 ± 3.08</td>
<td>213.50 ± 94.14</td>
<td></td>
</tr>
<tr>
<td>Aminoethyl-L-cysteine</td>
<td>1.29 ± 0.18</td>
<td>22.69 ± 0.75</td>
<td>17.59 ± 3.03</td>
<td></td>
</tr>
<tr>
<td>L-cystine</td>
<td>1.14 ± 0.22</td>
<td>4.67 ± 0.25</td>
<td>4.10 ± 1.01</td>
<td></td>
</tr>
<tr>
<td>L-cysteine</td>
<td>0.95 ± 0.18</td>
<td>4.36 ± 0.47</td>
<td>4.59 ± 1.85</td>
<td></td>
</tr>
<tr>
<td>L-serine</td>
<td>8.28 ± 1.57</td>
<td>2.65 ± 0.13</td>
<td>0.32 ± 0.07</td>
<td></td>
</tr>
<tr>
<td>O-acetyl-L-serine</td>
<td>11.21 ± 3.48</td>
<td>0.17 ± 0.018</td>
<td>0.015 ± 0.006</td>
<td></td>
</tr>
</tbody>
</table>
steady-state conditions. On the other hand, binding of L-serine at pH 8.5 to the enzyme results in mixtures of external aldimine and quinonoid species absorbing at 428 and 495 nm, respectively (Figure 1(a)).

Binding of the substrate analogues L-methionine or L-homoserine at pH 8.5 to the enzyme results in mixtures of external aldimine (λ_{max} = 418 nm) and quinonoid species (λ_{max} = 505 nm). This equilibrium is attained immediately upon mixing (Figure 1(b)).

CD spectra of the PLP cofactor were obtained in the presence of L-methionine and L-homoserine (Figure 2(a)). The spectrum for unliganded enzyme exhibits a positive Cotton effect centered on the visible absorption band of the cofactor, as it has been found for many PLP enzymes including cystalysin [12] and O-acetylserine sulfhydrylase [22] which catalyze reactions very similar to that catalyzed by C-S lyase. In the presence of saturating concentrations of L-methionine and L-homoserine, a positive Cotton effect is still evident, but with only 30% ellipticity relative to enzyme alone. The presence of L-homoserine gives a small negative dichroic signal centered around the visible absorption band of the quinonoid intermediate at 502 nm, and a modest negative dichroic band at 337 nm, mimicking what is observed in the visible spectrum.

Differences were also seen between the unliganded and liganded enzymes in the near-UV CD spectrum. Upon addition of ligands, a decrease of the negative dichroic signals in the Tyr spectral region was observed, which was more pronounced for L-methionine. Both these ligands have a pronounced effect on Trp residue(s) whose fine structure between 293 and 305 nm is clearly affected (Figure 2(a)).

Upon excitation at 295 nm, in the presence of L-serine, the cofactor emission band of C-S lyase exhibits a decrease in intensity compared to the internal aldimine emission spectrum (Figure 2(b)), while the Trp emission band is red shifted to 344 nm, maintaining almost the same intensity. The decrease in the emission band of the ketoenamine tautomer is confirmed by the direct excitation of the ketoenamine tautomer at 425 nm, which shows a decrease and a red shift to 517 nm compared to the emission of the internal aldimine (Figure 2(b), inset). When the enzyme was excited at 418 nm in the presence of O-acetyl-L-serine or L-methionine, the emission spectrum exhibited a quenching of the fluorescence emission at 500 nm, comitant with a modest red shift to 504 or 506 nm, respectively (Figure 2(b), inset).

### Table 2: Kinetic parameters for C-S lyase wt and mutants using L-Cth in 50 mM MOPS, and 50 mM bicine, 50 mM proline pH 9.0 at 30°C.

<table>
<thead>
<tr>
<th>Protein</th>
<th>K_{m} (mM)</th>
<th>k_{cat} (s^{-1})</th>
<th>Relative activity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>wt</td>
<td>1.07 ± 0.22</td>
<td>92.67 ± 4.44</td>
<td>100</td>
</tr>
<tr>
<td>Y114F</td>
<td>0.20 ± 0.01</td>
<td>1.20 ± 0.02</td>
<td>1.3</td>
</tr>
<tr>
<td>Y55F</td>
<td>0.62 ± 0.21</td>
<td>0.45 ± 0.01</td>
<td>0.5</td>
</tr>
</tbody>
</table>

* n.d.: not detected.

3.5. Effects of Arg351, Tyr114, and Tyr55 Mutation on the Enzymatic Activity of C-S Lyase. To identify the residues involved in catalysis and/or substrate binding, we aligned the amino acid sequences of seven bacterial C-S lyases (Figure 4). The C-S lyases share similar sequences and exhibit identities to C-S lyase from C. diphtheriae ranging from 27% (cystalysin from T. denticola) to 54% (C-S lyase from C. glutamicum). The alignment indicates that 40 amino acids are strictly conserved, and structural comparison of C-S lyase with cystalysin indicates similarity in overall fold as well as in active site residues (Figure 5). Of the 20 residues bordering the active site and the cofactor, we focused on three amino acids that are invariant among all the enzymes of the transsulfuration pathway.

3.6. Structural Similarity Analysis. Guided by structural superimposition and structural similarity analysis, interesting differences were found between cystalysin from T. denticola and C-S lyase from C. diphtheriae in the AVG adduct. Although the root mean square deviations (RMSD) between both structures are small, that is, 1.5 Å, there are important differences in the residues present in the binding cavity. Indeed, while in the cystalysin-AVG complex hydrophobic interactions are formed with Phe273 (cyan in Figure 5(b)), this Tyri is located on the dimer interface, and its side chain directly interacts with the PLP of the other subunit. Mutant R351A reduced the activity below detectable levels. On the other hand, the Y114F mutant retained 1.3% of the turnover rate of wild type enzyme. Although Tyr114 and Arg351 are positioned near the dimer interface, there are no direct interactions with any residues of the other subunit.

4. Discussion

The enzymes of the bacterial transsulfuration pathway, cystathionine γ-synthase and cystathionine β-lyase, are attractive targets for the development of novel antimicrobial compounds because this pathway is unique to plants and bacteria.
C. diphtheriae
---HQPSPIDLRANMTKWTRLY---GQQLPLWAESDFSTCPAVLQAITDAVR
C. glutamicum
---HADAY---
C. halotolerans
---HGPPSDLNLKBRRTKVTVY---GDDPLWIAESDFSTPPKVQAIADAV
C. glucuronolyticum
---KCPDDLDLRBRTKVTAF---EPDPLWIAESDFSTNPCTLALQVAIDE
S. mutans
MSKYFNPQAPNRLSHTKYKWET---ETDPQFLPAIAMDFEYMKVQAIHDVAEQ
T. denticola
-MIYDFTTKISRKNGLSWLDMYQPEVLPVEISKLKDKNWKL

C. diphtheriae
EAFQTPQDGLSSLQAQATEAYFRQYPARPEWIPFIPDVRRLGILYAIHNDPTAQPKSVYV
C. glutamicum
EVPPFPPATDALDLALTGFYKRYGFQPMPESVFAIPPPVGRLLAIEHFKPSAI1VP
C. halotolerans
ESFTPATPSDQLSFALSDFAYARWFPDCHVAVPDVPVRGLALLAAYRMTPGSPVYP
C. glucuronolyticum
EQFQTPQAGHVEALASQFCQARYQHVDPSIRHLATUDVMLGAIALLKYVAP---PVYIP
S. mutans
LVQTYTASDELQAVLQDKSEHYSFDKEDVFYVEGPVAIS1IAIAEYKGEAEVLIN
T. denticola
TVLTQGTPTEEYKVTWKMHRGQIDTWDWINTAQVPAVRNFEEVTPGPDGVIII

C. diphtheriae
TPAFFPPFPWLIATRRQ---EIGIFDATGIMHLDEFGA--QARSILCNPYPGLM
C. glutamicum
LPAAFFPFLPKVTRG---QAIYIDAE-DYEKIEKRXAFOAG--GASLFPCHNPPLGT
C. halotolerans
VPAAPPELPLEPETR---EVEVLHAYGLDAEIAEAEFRNG--AGSLICPNPHGLY
C. glucuronolyticum
TTFSPFPFFACQASQR---EVTYLDL---SDWAEFTQDHAPAIFLCPSHNPGV
S. mutans
VRSPFPQASFLKLRLVNLQVSEKEEOFLQIDPQLELD3VYDQWVLQCSSPNPHGR
T. denticola
SVPFYVPARTLRDLHDRLSEHVLNSLOLQGREENFDQLEKXDINNWKLYCPSHNPGR

C. diphtheriae
VFAPEVLNCELHDLARYDAVLRDLHEIAPFLVFDG--QHTVAAGVSDTAASVCITAPS
C. glutamicum
FSIZEEELTLTDIAKYDARIAEIHAPLVYEG--THVAGVSEHAAANTCITATTA
C. halotolerans
TLSFEELFLDLVALARYDLVVDLHEIAPFLVG--NAVAGLHDAEAAKCYVTATTA
C. glucuronolyticum
VHERYDLNLALALDLRTAVDVLSEIAPFLVMHCHPSAASVDASERTITFSTNG
S. mutans
VWEREYLGHCILKQHLVLSDIEHQDLYGSFVRDFKDFAVLSAATK
C. halotolerans
VWODDLKLAELCKWNLVLYSEIQDQALLAPKTVHLHSLQDAKQDFTILLASATK
T. denticola
WVKDELQIKDLOLSDLSAEIHDFLMPYGEHTVQISDDELATKTTPSAKT

C. diphtheriae
WNIAGLCAQ1IFSNEPDHDAWQQLS--PVXEDGASTLGLIAEAAARYGDSDLFVWYAV
C. glutamicum
WNTAGLCAQ1IFSNEPDHDAWQQLS--ITDRCVGDSLGLIAEAYTEYNEEELFDESISE
C. halotolerans
WNIAGLCAQ1IFSNEPDHDAWQQLS--PVXEDGASTLGLIAEAAARYGDSDLFVWYAV
C. glucuronolyticum
WNAGLRSALQ1IFTWNWDAQKLK--PAVPPPSLINVWAAYTCTDISSLDDEAYL
S. mutans
FIAIGTNSYIAIIEPTLCAPQKHQLVNNHESLSGLIYATTAFRGKFLVALKAV
C. halotolerans
FIAGTNSYIAIIEPTLCAPQKHQLVNNHESLSGLIYATTAFRGKFLVALKAV
T. denticola
FIAGTNSYIAIIEPTLCAPQKHQLVNNHESLSGLIYATTAFRGKFLVALKAV

C. diphtheriae
KNNDLFLLEHPKIPGAKTPMQATLYMWDIFRDTTIEG---SPEEFIEKAVWMD
C. glutamicum
KDNDRFAAELEK---LGKYVAPDSTYMWLDFAGTAKIE---APSKLEEQGVMLND
C. halotolerans
RGDNLWELPEKPKVPLQVSNPEATLWMLDFSGTVIDLG---PQAALWRKAVLNGN
C. glucuronolyticum
VENERYDLTHLPEVPLGMSHPATLWMLDFSQICEALLNRPQVLAEKVR
S. mutans
EENIQAEFYAFAKAPKLVPQKPVQTYLULWDSYGLLTD---ALFPTLHDQKAVLNLG
C. halotolerans
FIAGTNSYIAIIEPTLCAPQKHQLVNNHESLSGLIYATTAFRGKFLVALKAV
T. denticola
FIAGTNSYIAIIEPTLCAPQKHQLVNNHESLSGLIYATTAFRGKFLVALKAV

C. diphtheriae
AWFEDGTQDCFNLNFSRELVELEAADMRMACKVHST---376
C. glutamicum
AAIF--GFTCARNLFCASRETELEGRLIASV---325
C. halotolerans
TTFGGYESAOCERLAEELSSMLAGRRAEVAQAVED---381
C. glucuronolyticum
TCOGHAVDGFQANGCDFSSTLAEIRNRTMF---373
S. mutans
SDDYSEQELGHANLIAPKLVLVREIECKRVCCLP---388
S. mutans
AHFKEGYFKFALNYATPKVQALNLSSISIVFGK---387
T. denticola
YIFDGKGIPELIVLAAAAPDIVQEXELKALKDLKRNHKL

Figure 4: Sequence alignment of C-S lyases. The target residues for mutational analysis are highlighted in gray. The arrow above the sequences indicates the PLP-binding lysine. The C-S lyases used in this alignment (UniProt accession numbers) and their sources are Q6NFZ9, Corynebacterium diphtheriae NCTC 13129; Q46061, Corynebacterium glutamicum; M1NUD7, Corynebacterium halotolerans; COVUP4, Corynebacterium glucuronolyticum; A6BMJ3, Streptococcus anginosus IMU102; Q8DST5, Streptococcus mutans; Q56257, Treponema denticola ATCC 3540. All sequence alignment were carried out using the ClustalW program.
Recent studies probing the active sites of these enzymes are providing information that will guide the design of novel therapeutics [2, 23]. As members of the γ-subfamily of fold-type I of PLP-dependent enzymes, they also represent a useful model system for studies exploring the molecular mechanisms that underlie specificity among enzymes dependent on this catalytically versatile cofactor. A thorough understanding of the mechanisms controlling substrate and reaction specificity is a necessary step to enable the engineering of PLP-dependent enzymes for biotechnological applications [24].

The structure of \textit{C. diphtheriae} C-S lyase has been solved at 1.99 Å resolution (PDB code: 3FDB). We isolated the recombinant enzyme as a His-tagged protein at high yield and examined the spectral properties and steady state kinetic parameters of the purified enzyme, in addition to structural similarity analysis with cystalysin.

The internal aldimine absorbance spectra of C-S lyase exhibit a band centered at 413 nm (Figure 1(a)) which indicates that the predominant tautomer of the internal aldimine is the ketoenamine and that the imine nitrogen of the cofactor is protonated. This band is associated with a positive Cotton effect and gives rise to an emission maximum at about 500 nm. The enzyme spectrum is independent of pH from 6 to 11, indicating that the pK\text{a} of the internal aldimine is higher than pH 11. In the structurally related enzyme cystalysin, a band at 320 nm was attributed to a substituted aldimine, a species stabilized with respect to the ketoenamine by alkaline pH [12]. Both cystalysin and C-S lyase are catalytically more competent at alkaline pH, where a remarkable portion of the coenzyme in cystalysin exists as inactive aldime structure with a nucleophilic group. Interestingly, the spectrum of the \textit{C. diphtheriae} is pH independent; thus, this nucleophilic group is either missing or has a different location.

The apoenzyme showed no absorption in the visible range and no emission fluorescence due to PLP. When the intrinsic fluorescence emissions of holo- and apoenzyme were compared, it was evident that a substantial fraction (about 70\%) of the Trp emission in the apoenzyme is quenched in the holoenzyme. Such quenching is concomitant with the appearance of the 500 nm emission indicative of energy transfer from Trp residue(s) to the internal Schiff base. The interaction between the apoenzyme and PLP is characterized by a dissociation constant of 0.09 \(\mu\)M, which is indicative of very tight binding of the cofactor to the enzyme.

Conversion to the external aldime upon addition of substrate or substrate analog is expected to involve a conformational change that causes, among other things, the exclusion of water molecules from the active site. Formation of the external aldime with L-serine is accompanied by a bathochromic shift of the ketoenamine tautomer absorption band (15 nm; Figure 1(a)) compared to the same species with O-acetyl-L-serine, L-methionine, or L-homoserine (about 6 nm). This finding suggests a pronounced increase in the polarity of the microenvironment in the active site, likely due to a repositioning of charged or polar residues around the cofactor. Binding of L-serine to the enzyme results in mixtures of external aldime and quinonoid species (Figure 1(a)). \(\alpha,\beta\)-Elimination mechanisms generally proceed via a quinonoid intermediate, in which the carbanion formed upon abstraction of the \(\alpha\)-proton is stabilized by delocalization into the pyridinium ring of PLP. This transient intermediate with \(\lambda_{\text{max}} \sim 450-500\) nm is detectable when...
the elimination of the leaving group is slow, as might be expected for the hydroxyl group of L-serine [25]. The steady-state spectrum represents an intermediate whose formation is rate limiting or represents the sum of more than one intermediate if multiple steps are partially rate limiting [26]. Therefore, the presence of both the external aldimine and the substrate quinonoid in the steady-state spectrum of C-S lyase with L-serine indicates that formation of these intermediates may be partially rate determining. In contrast, a quinonoid intermediate is not observed for the elimination of acetate from O-acetyl-L-serine as the acetate group of O-acetyl-L-serine does not require protonation and is a better leaving group than the hydroxyl group of the L-serine. Although L-methionine and L-homoserine bind to C-S lyase in an unproductive manner, they nonetheless give equilibrating mixtures of external aldimine and quinonoid species (Figure 1(b)). This implies that L-methionine or L-homoserine stops the step at the reaction involving the quinonoid intermediate. The effects of all amino acids tested on the CD and fluorescence spectra of C-S lyase consist, albeit to different extents, in a modest enhancement of intrinsic Trp fluorescence, quenching and red shift of the long wavelength PLP emission, and a change in the dissymmetry of bound cofactor and aromatic amino acids located near the active site (Figures 2(a) and 2(b)). Taken together, in terms of local perturbations these data can be interpreted as changes in the orientation of the PLP and in the relative orientations of an aromatic residue(s) and of cofactor upon binding of ligand.

The $k_{\text{cat}}$ profile for the $\alpha,\beta$-elimination reaction of L-Cth or L-cysteine is pH independent. The $k_{\text{cat}}/K_m$ versus pH profile for L-Cth is bell-shaped, with $pK_a$ values of $8.05 \pm 0.07$ (p$pK_{a1}$, acidic limb) and $9.90 \pm 0.07$ (p$pK_{a2}$, basic limb). The $k_{\text{cat}}/K_m^{L-\text{Cth}}$ versus pH profile decreased on the acidic and basic sides with limiting slopes of 1 and –1, indicating that the ionization of the two groups is relevant to the enzymatic reaction, with one being protonated and the other unprotonated (Figure 3(a)).

The pH independence of the internal aldimine of C-S lyase indicates that this group cannot be assigned to one of those that titrates in the pH-activity profiles. The $pK_a$ values of the R-NH$_2$ groups of L-Cth were determined by direct titration with NaOH as $8.51 \pm 0.02$ and $9.60 \pm 0.02$ at 25°C, which are within experimental error of those determined for the acidic and basic limb of the $k_{\text{cat}}/K_m^{L-\text{Cth}}$ versus pH plot.

The basic limb (p$pK_{a2}$) of the $k_{\text{cat}}/K_m^{L-\text{Cth}}$ versus pH profile was assigned to the p$pK_a$ of the R-amino group of L-Cth, since p$pK_{a2}$ was not present in the $k_{\text{cat}}/K_m^{L-\text{Cys}}$ profile (Figure 3(b)). The acidic limb (p$pK_{a1}$) can be assigned to the $\alpha$-amino group of L-Cth (p$K_{a1} = 8.54$), indicating that the substrate must bind in the deprotonated state. Alternatively, the p$pK_{a1}$ could reflect ionization of an enzyme group other than the imino moiety of the internal aldimine.

It is generally accepted that PLP-dependent $\beta$-eliminases, including $\beta$C-S lyases, have three key intermediate steps [8] (Scheme 1). Plausible catalytic mechanisms for the formation of intermediates have been reported for several $\beta$C-S lyases on the basis of structural and spectroscopic information [12, 27–30]. Contrary to their common intermediates, the quaternary structures (dimer and tetramer) and the active site residues of the $\beta$C-S lyase enzymes are somewhat different, possibly depending on their substrate specificity. When studying the $\alpha,\beta$-elimination reactions of sulfur containing amino acids in bacteria, determination of enzyme substrate specificity is crucial since it allows distinction between true cystathionine $\beta$-lyase activities involved in the synthesis of methionine and other C-S lyase activities. Thus, cystathionine $\beta$-lyase supports the cleavage of either L-Cth or djenkolate at high levels and shows low activity towards L-cystine. Conversely, C-S lyases are characterized by marginal cleavage of L-Cth and efficient synthesis of cysteine persulfide from L-cysteine [31, 32].

To define the substrate specificity of C-S lyase from C. diphtheriae, we examined the reaction of the enzyme with sulfur- and nonsulfur containing amino acids as well as with disulfidic amino acids (Table 1). Among the substrates tested, only L-cysteine exhibits substrate inhibition, similar to T. denticola cystalysin [12], C-DES from Synechocystis [33], $\beta$-cystathionase [31], and D-cysteine desulphydrase from E. coli [34].

Recombinant C-S lyase from C. diphtheriae showed a relatively broad substrate specificity toward sulfur-containing amino acids, as previously observed for the enzyme from A. thaliana [35], E. coli [31], B. avium [36], and S. typhimurium [37]. For all these enzymes, the highest rates of $\beta$-cleavage activity were obtained with L-Cth and djenkolate, whereas L-cysteine and L-cysteine were poor substrates. In the case of L-cysteine and L-cysteine, compared with L-Cth, substrate selectivity does not appear to derive from substrate binding,
since $K_m$ values are similar, but rather from chemical steps that follow the binding (Table 1). Finally, no activity could be detected for the enzyme when either L-homocysteine or L-methionine was used as substrate (yC-S linkages).

The present observations are in agreement with previous analysis of the substrate specificity of cystathionine $\beta$-lyase from A. thaliana [35] which showed the following: (1) all active substrates contain a C3 L-$\alpha$-amino acid moiety; (2) the nature of the sulfur-containing bridge (thioether, thiaoacetal, or disulfide) influences the rate of the $\alpha,\beta$-elimination process; and (3) the length of the second moiety, and therefore the spatial separation of the two carboxyl groups of the substrate, has a critical role in the efficiency of cleavage. In light of these findings, C. diphtheriae should more properly be considered as a cystathionine $\beta$-lyase, rather than a C-S lyase, as annotated (NCBI protein NP_940074). Interestingly, although both C. diphtheriae C-S lyase and cystathionin belong to fold type I, and share similar sequences (Figure 4) and structural similarity in overall fold and active site residues (Figure 5), they do not display desulphhydrase activity with similar substrate specificity. Cystathionin, in fact, displays a strong substrate preference for L-djenkolic acid and L-cystine over L-cysteine and, among sulfur-containing amino acids, the lowest $\beta$-cleavage activity was obtained with L-Cth [12]; its substrate specificity is similar to that of L-cysteine/cysteine C-S lyase (C-DES) from Synechocystis.

Based on structural and kinetic studies, a postulated catalytic mechanism with the intermediates has been proposed for cystathionin [38, 39] and Streptococcus anginosus $\beta$C-S lyase (Lcd) [40], the enzymes with the highest sequence identity to C. diphtheriae C-S lyase (Figure 5(a)) among structurally and enzymatically characterized $\beta$C-S lyases. The structural information on the intermediate complexes enabled us to select conserved residues for site-directed mutagenesis. Mutational analysis demonstrated the important roles of several active site residues, namely, Tyr55, Tyr114, and Arg351 (Figure 5(a)). In particular, Tyr114 recognizes the $\gamma$-atom (or $\delta$-atom) of the substrate by the phenolic hydroxyl group and fixes PLP at the proper position by a stacking interaction throughout the $\alpha,\beta$-elimination reaction [27, 38].

Mutational analysis further revealed that Y114F retained only 1.3% of wt enzymatic activity (Table 2). On the basis of kinetic studies, Cellini et al. [38] concluded that Tyr64 of cystathionin (Tyr55 of C. diphtheriae C-S lyase) plays a role in the properly orienting Lys238 (Lys222). In agreement with structural observations, substitution of Tyr55 to Phe reduced the catalytic activity to 0.5% (Table 2), suggesting the importance of the phenolic hydroxyl group of this residue.

In both Lcd external aldimine and $\alpha$-aminoacrylate forms, the guanidino group of Arg365 formed a salt bridge with the carboxyl moiety of the amino acid substrate bound to PLP [40]. This Arg is widely conserved among type I PLP-dependent enzymes and is supposed to act as a general $\alpha$-carboxylate docking site [41]. A similar salt bridge has been observed between the corresponding Arg and the carboxyl moiety of the coenzyme bound substrate in other PLP-dependent $\beta$-eliminases [28, 42] suggesting a common role of the Arg for substrate binding. Substitution of Arg351 with Ala notably reduced catalytic activity toward L-Cth to below a detectable level (Table 2).

Although the residues involved in PLP and substrate binding are fully conserved, structural similarity analysis between cystathionin from T. denticola and C-S lyse from C. diphtheriae in the AVG adduct revealed subtle differences in the binding cavity. These involve just three amino acids, that is, two charged residues present in C-S lyse and not in cystathionin and vice versa, a Phe residue present in cystathionin and not in C-S lyse. Indeed, the availability of two charged residues and the lack of bulky hydrophobic one will dramatically affect the physicochemical properties of the binding cavity of C-S lyse. These elements can thus be exploited for rational design of novel and species specific inhibitors to be used in antibacterial therapeutics.

**Abbreviations**

- PLP: Pyridoxal 5'-phosphate
- DTNB: 5,5'-dithiobis-2-nitrobenzoate
- LDH: L-lactic dehydrogenase
- L-Cth: L-cystathionine
- AVG: L-aminoethoxyvinylglycine
- $K_d$: Equilibrium dissociation constant for PLP
- CD: Circular dichroism
- PDB: Protein Data Bank
- $k_{cat}$: Turnover rate
- $K_m$: Michaelis constant.

**References**


