Review Article

Anti-Inflammatory and Antiapoptotic Responses to Infection: A Common Denominator of Human and Bovine Macrophages Infected with Mycobacterium avium Subsp. paratuberculosis

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Mycobacterium avium subsp. paratuberculosis (Map) is the causative agent of a chronic intestinal inflammation in ruminants named Johne's disease or paratuberculosis and a possible etiopathological agent of human Crohn's disease (CD). Analysis of macrophage transcriptomes in response to Map infection is expected to provide key missing information in the understanding of the role of this pathogen in establishing an inappropriate and persistent infection in a susceptible host and of the molecular mechanisms that might underlie the early phases of CD. In this paper we summarize transcriptomic studies of human and bovine peripheral blood mononuclear cells (PBMC), monocyte-derived macrophages (MDMs), and macrophages-like cell lines in vitro infected with Map. Most studies included in this paper consistently reported common gene expression signatures of bovine and human macrophages in response to Map such as enhanced expression of the anti-inflammatory cytokines IL-10 and IL-6, which promote bacterial survival. Overexpression of IL-10 could be responsible for the Map-associated reduction in the expression of the proapoptotic TNF-α gene observed in bovine and human macrophages.

1. Association of Mycobacterium avium Subsp. paratuberculosis (Map) with Chronic Inflammatory Bowel Diseases of Cattle and Humans

Mycobacterium avium subsp. paratuberculosis (Map) is the causal agent of Johne's disease or paratuberculosis, a chronic inflammatory bowel disease of domesticated ruminants and wildlife species worldwide. Johne's disease causes major economic losses to the global dairy industry due to reduced milk production, lower weight gains, infertility, premature culling, and increased cow replacement costs [1]. Map has a worldwide distribution and is of considerable concern in cattle, sheep, goats, and farmed red deer. Although it is still controversial, Map has been implicated as a causal or exacerbating agent in human Crohn's disease (CD), a chronic inflammatory bowel disease characterized by transmural inflammation and granuloma formation [2–4]. Evidences that Map may be associated to CD in humans include similarity between the clinical signs of CD in humans and those found in animals with paratuberculosis; detection of Map in feces, intestinal tissues, breast milk, macrophages, and peripheral blood of patients with CD; association between Map DNA in blood and cellular and humoral immune responses in CD; and anti-Map antibiotic therapy resulting in reduction of bacteremia and remission or substantial improvement in disease condition in many patients [5–10]. In addition, meta-analysis and epidemiological studies have confirmed an association of Map with CD [11, 12].

Map may enter the food chain from a variety of sources. The organism, shed from infected animals, may contaminate pastures and potable water, where it is resistant to standard purification with chlorine [13]. Because Map can survive pasteurization conditions, dairy products such as milk and cheese have been proposed as possible sources of exposure of humans to Map [14]. Recently, we have
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demonstrated that Map can be detected and cultured from
diaphragm muscle of Map-infected cattle destined for human
carcass consumption and suggested a possible risk of exposure of
to Map via contaminated meat [15]. After oral ingestion, Map invades the intestinal wall preferably through
epithelial cells or M cells present in the follicle-associated epithelium covering the continuous Peyer’s patches in the
distal ileum [16–18]. Although the mechanism of entry in the mucosa is important in establishing Map infection, many
of the bacterial components involved in the interaction with the intestinal epithelium are still unknown. It has
been recently demonstrated that Map3464 gene encodes an
NADH-flavin oxidoreductase involved in invasion of bovine
epithelial cells through the activation of host cell Cdc42 [19].
After translocation of Map across the intestinal epithelium, Map is subsequently phagocytised by macrophages in the
intestinal lamina propria and submucosa. Upon phagocytosis of Map by naive macrophages, there is both intracellular
replication of Map and bacterial killing by the host, which
results as an initial T-helper 1 (Th-1) or proinflammatory
immune response [20–24]. Bacterial killing is due to a rapid
phagosome acidification response by the host that enables
phagosome-lysosome fusion and presentation of antigens to
CD8+ T cells via MHC to occur in some infected cells. However,
since many phagosomes containing Map fail to acquire sig-
ificant amounts of lysosomal-associated membrane protein
(LAMP-1) and to fuse with lysosomes, this can allow Map to
survive and proliferate inside macrophages. An active role for
Map in preventing phagosome-lysosome fusion is supported by the observation that live bacteria are able to persist within
phagosomes, while phagosome maturation is not interrupted
following the uptake of killed Map [25, 26]. In addition, recent
studies have suggested that Map alters the ability of infected
macrophages to react to extracellular signals from T cells, particularly through the CD154-CD40 system [27]. This leads
to an enhanced IL-10 and TGF-β expression in Map-infected
macrophages, which favor bacterial survival by suppression of
Th-1 responses and IFN-γ in T cells [28–31]. In cattle, clinical signs of infection and bacterial shedding are usually
not evident until 2–5 years post-infection (p.i.) [32]. During
the subclinical phase of the infection, Map persists and slowly
proliferates within macrophages of the gut without the innate
system being able to clear the infection.

2. Anti-Inflammatory, Antiapoptotic, and
Anti-Invasive Responses Induced in Bovine
Macrophages Infected with Mycobacterium
avium Subsp. paratuberculosis

Several in vitro studies have investigated gene expression
profiles induced by Map on bovine macrophages obtained
from uninfected cattle and on a bovine macrophage cell line
(Table 1). Compared to uninfected cultures, in vitro chal-
enges of monocytes-derived macrophages (MDMs) from
healthy cows with live Map resulted in enhanced production of
the anti-inflammatory cytokine interleukin-10 (IL-10) at
6, 24, and 72 h p.i. that antagonizes the proinflammatory
immune response by downregulating the production of
interleukin-12 (IL-12), tumor necrosis factor-α (TNF-α),
and interferon-γ (IFN-γ) as estimated by qRT-PCR [33].
Similarly, other authors also observed a downregulatory
trend in TNF-α mRNA expression from 16 h to 96 h p.i.
and upregulation of IL-10 mRNA levels that peak from
48 h to 96 h p.i. [34]. Using microarray technology, three
cytokines including transforming growth factor-β (TGF-
β), interleukin-6 (IL-6), and macophage inflammatory
protein-1β (MIP-1β) had greater expression in Map-infected
MDM at 16 h p.i. when compared with inactivated control
macrophages [35]. The matrix metalloproteinase (MMP-12)
and the thrombopsondin-1, both involved in cell migration
and tissue destruction, were also significantly upregulated. In
contrast, the TNF-β receptor and the major histocompatibil-
ity complex (MHC) class II DQ-β had lower expression in
Map-infected macrophages. Decreased expression of the cell
surface MHC class I and class II molecules was previously
documented in macrophages phagocytising Map organisms
indicating a reduced capacity to present antigens to T lym-
phocytes [44]. Consistently with these results, Murphy et al.
also detected high levels of the anti-inflammatory cytokines
tGFJ-β and IL-6 in MDM infected with Map at 24 h p.i. [36].
In another study, significantly downregulation in expression
of the proinflammatory cytokine IL-1β and of the metallopro-
teinases MMP-1, MMP-23, and MMP-9 involved in tissue
destruction was observed in Map-stimulated PBMC when compared with control cells [37]. Using cDNA microarrays
focused on expressed sequences from a bovine total leukocyte
library (BOLTS) and 10 distinct Map strains to measure total
transcriptomic alterations in Map-infected MDM, a total of
78 annotated bovine genes were found to be differentially
expressed at 6 h p.i., relative to uninfected cells [38]. Within
the group of differentially expressed genes significant down-
regulation of two proapoptotic genes, BCL2 antagonist
of cell death (BAD) and TNF receptor (TNFR), was observed
in Map-infected MDM cells relative to uninfected cells.
Uregulation of the apoptotic inhibitor BCL2A1 and of the
proinflammatory cytokines IL-1α, IL-1β, and IL-8 relative
to uninfected control cells was also observed. By using a
pan-genomic analysis of bovine MDM gene expression in
response to in vitro infection with Map, Machugh et al.
revealed that many of the highly upregulated genes at 2 h p.i.
had proinflammatory related functions, particularly IL-1α,
IL-1β, TNF, IL-6, chemokine ligand 2 gene (C-X-C motif;
CXCL2), and the chemokine ligand 20 gene (C-C motif;
CCL20) [39]. At 6 h p.i. immune-related genes were among
the differentially expressed genes showing the highest relative
increase in expression; however, the fold-change induction
of these genes was not as high as those detected as 2 h p.i.
Uregulated genes at 6 h p.i. included IL-1β, TNF, CXCL2,
CCL4, CCL5, CCL20, CD40, and the complement factor B
gene (CFB). Of the differentially expressed gene identified
24 h p.i. that had a known immune function were the serum
amyloid A3 genes (SAA3), C-type lectin domain family 4
member E (CLEC4E), C-type lectin domain family 2 mem-
ber D (CLEC2D), CD40, and CFB. Overall, several pro-
and anti-apoptotic genes were upregulated at 2 h and 6 h p.i. suggesting
that this process is highly regulated. Pro- and anti-apoptotic
genes upregulated included TNF (proapoptotic), caspase 1,
Table 1: Immune related, apoptosis-related and tissue destruction genes differentially expressed relative to uninfected cells in monocytes-derived macrophages (MDM), peripheral blood mononuclear cells (PBMC), and in a bovine macrophage cell line after stimulation with live bovine isolates of *Map*. Genes upregulated or downregulated in at least three of the studies are shown in bold.

<table>
<thead>
<tr>
<th>Cell model</th>
<th>Map strain</th>
<th>Map/cells ratio</th>
<th>Assay</th>
<th>Time p.i. (h)</th>
<th>Upregulated genes</th>
<th>Downregulated genes</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>MDM</td>
<td>ATCC 19698</td>
<td>10 : 1</td>
<td>qRT-PCR</td>
<td>6h</td>
<td>IL-10, GM-CSF</td>
<td>IFN-γ, TNF-</td>
<td>[33]</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>24h</td>
<td>IL-10, GM-CSF</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>72h</td>
<td>IL-10, GM-CSF, and IL-8</td>
<td>IL-12</td>
<td></td>
</tr>
<tr>
<td>MDM</td>
<td>Field strain B1018</td>
<td>5 : 1</td>
<td>qRT-PCR</td>
<td>16h</td>
<td>IL-10, TNF-</td>
<td>IL-10, TNF-</td>
<td>[34]</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>24h</td>
<td>IL-10</td>
<td>IL-10, TNF-</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>48h</td>
<td>IL-10</td>
<td>TNF-</td>
<td>TNF-</td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>96h</td>
<td>IL-10</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MDM</td>
<td>ATCC 19698</td>
<td>10 : 1</td>
<td>Microarray</td>
<td>16h</td>
<td>IL-6, TGF-, Thrombospondin-1, MIP-1β, and MMP-12</td>
<td>TNFR, MHC class II DQ-β</td>
<td>[35]</td>
</tr>
<tr>
<td>MDM</td>
<td>ATCC 19698</td>
<td>10 : 1</td>
<td>Microarray</td>
<td>24h</td>
<td>IL-6, TGF-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PBMC²</td>
<td>ATCC 19698</td>
<td>10 : 1</td>
<td>Microarray</td>
<td>16h</td>
<td>IL-1β, IL-8, IL-1, and BCL2A1</td>
<td>IL-1, MMP1, MMP23, and MMP9</td>
<td>[36]</td>
</tr>
<tr>
<td>MDM</td>
<td>ATCC 19698</td>
<td>5 : 1</td>
<td>Microarray</td>
<td>6h</td>
<td>IL-1β, IL-6, IL-1, and IL-8, and BCL2A1</td>
<td>TNFR, BAD</td>
<td>[38]</td>
</tr>
<tr>
<td></td>
<td>Field strain</td>
<td>2.1</td>
<td>Microarray</td>
<td>2h</td>
<td>IL-1β, TNF, CXCL2, CCL20, CFB, CASP-1, CASP-4, CASP-8, CASP-6, BIRC-3, and CD40</td>
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</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>6h</td>
<td>IL-1β, TNF, CXCL2, CCL4, CCL5, CCL20, CCL40, CFB, CASP-1, CASP4, CASP8, CASP6, BIRC3, and CFLAR</td>
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<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>24h</td>
<td>SAA3, CLEC4E, CLEC2D, CD40, and CFB</td>
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<tr>
<td>BoMac³</td>
<td>K10</td>
<td>10 : 1</td>
<td>qRT-PCR</td>
<td>4h</td>
<td>IFN-γ, IL-1, BCL2-1</td>
<td>IL-6</td>
<td>[40]</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>14h</td>
<td>IL-6, BCL2-1</td>
<td></td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>24h</td>
<td>IL-6, TGF-1, and TNFγ-2</td>
<td>MMP3-1, IL-1α, and BCL2-1</td>
<td></td>
</tr>
</tbody>
</table>

¹MDM: monocytes-derived macrophages; ²PBMC: peripheral blood mononuclear cells; ³BoMac: bovine macrophage cell line.
4, 6, and 8 genes (CASP-1, CASP-4, CASP-6, and CASP-8; all proapoptotic), the baculoviral IAP repeat containing 3 gene (BIRC-3; antiapoptotic), and the FADD-like apoptosis regulator gene (CFLAR; pro- and antiapoptotic).

We recently examined whether Map isolates with differential abilities to grow within a bovine macrophage cell line (BoMac) induced a characteristic early immune-inflammatory response [40]. Our results showed significant differences in the expression of several cytokines (IL-6, TGF-β, TNF-α2, IFN-γ, and IL-1α), proteins related to apoptosis (BCL2-1), or tissue destruction (MMP3-1) after the infection of BoMac cells with a bovine or an ovine isolate of Map. The bovine isolate that grew within BoMac cells was a good inducer of the apoptotic inhibitor BCL2-1 at 4 or 14 h p.i. which might cause lower levels of apoptosis than in BoMac cells infected with the ovine isolate. In addition, infection of BoMac cells with the bovine isolate resulted in a significant upregulation of the anti-inflammatory cytokines IL-6 and TGF-β at 24 h p.i., when compared with cells infected with the ovine isolate. Although we did not observe significant differences in IL-10 or TNF-α2 gene expression in BoMac cells infected with the bovine or the ovine isolates at any of the time points, the bovine isolate did induce more IL-10 at 14 and 24 h p.i. and less TNF-α2 at 4 and 14 h p.i. than did the ovine isolate. These results suggest that lower TNF-α2 production and an induction of IL-10 are associated with the growth within bovine macrophages of virulent isolates of Map. We also observed that cells stimulated with the bovine isolate exhibited lower levels of the metalloproteinase MMP3-1 involved in tissue destruction at 4 h and 24 h p.i. relative to cells stimulated with the ovine isolate. Differences in induction of the metalloproteinase inhibitor TIMP-1 were not statistically significant at any of the time points studied but the bovine isolate did induce more TIMP-1 at 4 h p.i. than did the ovine isolate. The ovine isolate was significantly attenuated in growth in BoMac cells and this decrease in survival within the infected cells correlated with a reduced anti-inflammatory response in the infected cells and with a significantly upregulated proinflammatory immune response generally associated with elimination of Map and protection. In particular, the expression of the proinflammatory cytokine IL-1α was highly upregulated in cells infected with the ovine isolate at 14 and 24 h p.i. and downregulated in BoMac cells infected with the bovine isolate at 24 h p.i. Because a strong correlation between the intracellular multiplication of the tested isolates and patterns of production of IL-6, TGF-β, MMP-3, BCL2-1, and IL-1α was observed, the levels of expression of these specific proteins might be used to discriminate between isolates with differential virulence in the BoMac cellular model.

All together, the results of the transcriptomic studies in bovine macrophages included in this paper suggested that Map might stimulate an initial proinflammatory immune response mediated by IL-1α that is followed by an enhanced anti-inflammatory response mediated by IL-6, IL-10, and TGF-β. In addition, downregulation of the proapoptotic gene TNF-α was consistently observed in different studies. Stimulation of anti-inflammatory and antiapoptotic responses might allow Map to successfully persist in bovine macrophages during the persistent, subclinical phase of the infection.

### 3. Immune-Inflammatory Responses Induced in Human Macrophages Infected with Map

Despite the possible role of Map in CD, there is not much known about the interaction of Map with the human innate immune system (Table 2). A transformed human monocytic cell line (THP1) stimulated with Map has shown to differentially respond to infections with well-characterized clinical isolates of Map when compared with unstimulated cells. When human THP1 cells were stimulated with bovine or human isolates of Map several genes associated with apoptosis and cytokine signalling (LRDD, PDCD-8, IL-12, IL-18, IL-23, and TNFα) were significantly downregulated [41]. Data from this study suggested that the human macrophage responses to Map isolates from cattle or human sources, regardless of genotype, follow a common theme of antiapoptotic and anti-inflammatory responses within the host cells, an attribute likely associated with successful infection and persistence.

<table>
<thead>
<tr>
<th>Cell Model</th>
<th>Map strain</th>
<th>Map/cells ratio</th>
<th>Assay</th>
<th>Time p.i. (h)</th>
<th>Upregulated cytokines</th>
<th>Downregulated cytokines</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>THP-1 1,2</td>
<td>Bovine 1018</td>
<td>5:1</td>
<td>Microarray</td>
<td>2 h</td>
<td>IL-18, IL-12B, IL-23A,</td>
<td>TNF-α, LRDD, and</td>
<td>[41]</td>
</tr>
<tr>
<td>PBMC from</td>
<td>Human</td>
<td>ATCC 43019</td>
<td>1:1</td>
<td>Flow</td>
<td>IL-10, IL-6, and</td>
<td>IFN-γ</td>
<td>[42]</td>
</tr>
<tr>
<td>CD patients</td>
<td>ATCC 43015</td>
<td>10:1</td>
<td>Flow</td>
<td>3 h</td>
<td>IL-23</td>
<td>TNF-α</td>
<td>[43]</td>
</tr>
</tbody>
</table>

1 THP-1, human monocytic cell line.
2 Comparisons were made between uninfected and infected cells.
3 Comparisons were made between Map-infected macrophages from CD patients and controls.
The response of macrophages from CD patients to live Map has been recently addressed. Following in vitro exposure to Map, PBMC from CD patients secreted significant more amounts of TNF-α, IL-6, and IL-10 when compared to the levels released by PBMC from healthy volunteers [42]. In contrast, the IFN-γ response to Map was significantly elevated in PBMC isolated from healthy volunteers compared to PBMC derived from CD patients. In another study, human MDMs obtained from CD patients and controls were infected with Map, Mycobacterium avium subsp. avium (Mav), and other live intestinal bacteria such as Escherichia coli or Enterococcus faecalis, and cytokine levels were evaluated at different time points [43]. The results of this study indicated that macrophages from CD patients showed impaired TNF-α secretion in response to bacterial challenge but augmented IL-23 secretion as compared to macrophages from healthy individuals. It is plausible that the MAP-dependent IL-23 secretion enhancement occurred as consequence of phagocytosis, because the high IL-23 concentrations obtained at 3 h p.i. were not observed at 3 d and 7 d p.i. Differences in cytokine expression after bacterial challenge where not Map specific, as other bacteria (E. coli and Mav) showed similar effects.

Although most of the studies presented in this paper used PBMC or MDM cells in vitro stimulated with Map, we should indicate that recently Olsen et al. [45] isolated intestinal T cells from intestinal biopsies of CD patients to investigate cellular immune responses to Map. Interestingly, they observed that CD patients had a high frequency of Map reactive T cells and also a higher frequency of response to Map compared to other bacterial antigens. After stimulation with Map, intestinal T cells secreted the proinflammatory cytokines IFN-γ and IL-17, and, therefore, a role for Map in the excessive inflammation seen in CD cannot be excluded.

4. Conclusions

Despite development of cell-mediated immune responses shortly after infection, Map has the capacity to survive and grow in macrophages from human and cattle hosts. Gene expression studies included in this paper allow us to conclude that the inhibition of apoptosis and enhanced expression of inhibitors of macrophage activation could contribute to the early survival and immune escape of Map. Common gene expression signatures of bovine and human macrophages in response to Map such as enhanced expression of the anti-inflammatory cytokines IL-10 and IL-6, which promote bacterial survival, have been consistently observed. Overexpression of IL-10 could be responsible for the Map-associated reduction in the expression of the proapoptotic TNF-α gene observed in many studies. Differential effects on macrophage gene regulation between studies might be caused by the different in vitro models, multiplicities of infection, time p.i., and/or microarray platforms used. Although this paper does not suggest a casual effect for Map, it does compile the findings that Map is able to alter the normal host immune response against a pathogen in susceptible humans and cattle and contribute to the pathogenesis of Crohn’s and John’s diseases.

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References


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