Research Article

Anticancer Activity of Marine Sponge Hyrtios sp. Extract in Human Colorectal Carcinoma RKO Cells with Different p53 Status

Hyun Kyung Lim,1 Woori Bae,1 Hyi-Seung Lee,2 and Joohee Jung1

1 College of Pharmacy, Duksum Women's University, No. 33, 144-gil, Samyang-ro, Dobong-gu, Seoul 132-714, Republic of Korea
2 Korea Institute of Ocean Science & Technology, 787 Haean-ro, Ansan 426-744, Republic of Korea

Correspondence should be addressed to Joohee Jung; joohee@duksung.ac.kr

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1. Introduction

The ocean occupies about 70% of the earth and contains a huge number of marine organisms. Collection and identification of marine organisms were difficult for researchers and drug developers, but marine resources are still attractive for the application of pharmaceutical fields. Among marine resources, marine sponges are known to have about 15,000 species worldwide [1]. Marine sponges take in nutritive through body pores and produce secondary metabolites with bioactivity. In our ongoing research, we investigated the bioactivity of marine sponges before classifying and isolating their active compounds. Crude extracts were made from marine sponges collected from the Chuuk islands in Micronesia and investigated for anticancer effect. Screening tests identified some specimens with anticancer effects. One of the specimens was identified as Hyrtios sp. (Figure I). Recently, Hyrtios sp. was reported to have cytotoxic [2, 3] and antioxidant activities [4]. Several Hyrtios metabolites [5, 6] and active compounds [2, 4] have been reported, but the anticancer effects of Hyrtios sp. have not been reported. Herein, Hyrtios sp. extract was investigated for anticancer activity in human colorectal carcinoma RKO cells with different p53 status.

2. Materials and Methods

2.1. Specimen Preparation. Marin sponge specimens were collected by hand with scuba equipment at Chuuk state, Federated States of Micronesia, in October, 2010. Freshly collected specimens were washed by sterilized artificial sea water three times, immediately frozen, and stored at −20°C until use. Lyophilized specimens were extracted with methanol (3 × 3 L) as previous study [7]. All of the samples were provided from KIOST (Korea Institute of Ocean Science & Technology). The extracts of specimens (10 mg) were dissolved in sterile distilled water (the final concentration,
2.2. Cells and Treatment. Human colorectal carcinoma RKO (CRL-2577) and RKO-E6 (CRL-2578) cells (ATCC, Man-
assas, VA) were cultured in Dulbecco’s modified Eagle’s medium (DMEM, GenDEPOT) supplemented with 10% fetal bovine serum (GenDEPOT) and 1% penicillin/streptomycin (GenDEPOT) in a humidified 5% CO2 incubator. Cells used for the assays were in exponential growth phase. The samples were treated to the cell culture for 24 h or 48 h.

2.3. Cell Cytotoxicity. Cell cytotoxicity was examined by Cell Counting Kit-8 (CCK-8, Dojindo, Japan). Briefly, cells were seeded in 96-well plates at a density of 3 × 103 cells/well. After incubation for 24 h, cells were treated with sponge samples for 24 h or 48 h. CCK-8 reagent (10 μL) was added to each well and incubated for 3 h at 37°C. Absorbance at 450 nm was determined through microplate reader (Infinite M200 PRO, TECAN, Austria).

2.4. Cellular Morphology. Cells were seeded in 6-well plate at 4–6 × 104 cells/well and treated with Hyrtios sp. extract (80 μg/mL) for 48 h. Cells were observed under light microscopy (×40 and ×400) (Nikon eclipse TS100, Japan).

2.5. Western Blot Analysis. Cells were seeded in 6-well plate at a density of 4–6 × 104 cells/well. Samples were treated to each well and incubated for 24 h or 48 h. Cells were harvested and lysed in RIPA buffer (GenDEPOT) with protease inhibitors (Xpert protease inhibitor cocktail solution, GenDEPOT) and phosphatase inhibitors (Xpert phosphatase inhibitor cocktail solution, GenDEPOT). Cell lysates were boiled in 5× sample buffer and separated by 10% SDS-PAGE. Proteins were transferred onto PVDF membranes (Millipore) using a semidyrid electroblotter (Peglab, Germany). Membranes were blocked with 5% skim milk in TBST (50 mM Tris-
HCl pH 7.4, 150 mM NaCl, 0.1% Tween 20) and incubated sequentially with primary antibodies at 4°C, overnight. After washing membranes with TBST, the membranes reincubated with secondary antibody at room temperature for 3 hr. Immunoreactive protein was visualized using ECL reagents and developed with X-ray film. Antibodies and ratios at which they were used were p53, p21 (1: 2000, Millipore), c-Jun N-terminal kinase (JNK, 1: 500, Santa Cruz Biotechnology), β-actin (1: 5000, Sigma-aldrich), and anti-mouse IgG (H+L) horseradish peroxidase conjugate and anti-rabbit IgG (H+L) horseradish peroxidase conjugate (1: 3000, Bio-Rad).

2.6. Apoptosis. Cells were seeded in 6-well plate and treated as described for western blots. After incubation, cells were stained with Annexin V-FITC (Nexelom Bioscience LLC) and propidium iodide (PI) solution. Apoptosis was detected using a Cellometer (Nexelom Bioscience LLC).

3. Results and Discussion

3.1. Cytotoxicity of Hyrtios sp. Extract in RKO and RKO-E6. To evaluate cytotoxicity to cells with different p53 status, serially diluted samples of Hyrtios sp. extracts were treated to RKO and RKO-E6 cells for 24 h and 48 h. As shown in Figure 2(a), cytotoxicity slightly increased in both RKO and RKO-E6 cells. Hyrtios sp. extract (100 μg/mL) inhibited 52.7 ± 0.9% viability of RKO cells and 66 ± 0.9% viability of RKO-E6 cells. Hyrtios sp. extract (100 μg/mL) for 48 h inhibited 26.5 ± 0.7% and 44.5 ± 1% of cell viability in RKO and RKO-E6 cells, respectively (Figure 2(b)). Hyrtios sp. extract increased cytotoxicity time-dependently for RKO cells and RKO-E6 cells. The result showed that RKO cells were more sensitive than RKO-E6 cells to Hyrtios sp. extracts and indicated that the anticancer effects of Hyrtios sp. were different for RKO and RKO-E6 cells depending on their p53 status.

3.2. Hyrtios sp. Extract-Induced Mitotic Catastrophe. To investigate cell death induced by Hyrtios sp. extracts, cellular morphology was observed. RKO cells had fewer cells than RKO-E6 cells after treatment with Hyrtios sp. extracts (Figure 3(a)) consistent with cytotoxicity results. In particular, RKO cells treated with Hyrtios sp. extracts exhibited multinucleation and increased cell size, indications of mitotic catastrophe (Figure 3(b), upper right arrow). Hyrtios sp. might trigger the cell death induced by DNA damage.

3.3. Changes in p53, p21, and JNK Levels after Treatment with Hyrtios sp. Extract. To examine differences in cellular mechanisms between RKO and RKO-E6 cells, protein levels were determined by western blots. Expression of p53 protein was detected in RKO cells but not RKO-E6 cells (Figure 4). RKO cells treated with Hyrtios sp. extracts showed an increase of p53 protein levels and also increased expression of p21 protein which is encoded by a p53 target gene. In addition, p21 protein also increased in RKO-E6 cells treated with Hyrtios sp. extracts. JNK protein expression was slightly reduced in RKO and RKO-E6 cells (Figure 4). These results indicated that RKO cells exposed to Hyrtios sp. extracts induced cell death by increase of p53 protein, which activated the expression of p21 protein for 24 h (not shown) and

![Figure 1: Morphology of Hyrtios sp. specimen before methanol extraction.](image)
Figure 2: Inhibition of cell viability by *Hyrtios* sp. in RKO and RKO-E6 cells. Cells were treated with *Hyrtios* sp. extract and incubated for 24 h (a) and 48 h (b). Cytotoxicity was determined by CCK-8 assay. Data represent mean ± standard deviations (n = 7), t-test (*P < 0.001).

Figure 3: Change in cellular morphology after treatment with *Hyrtios* sp. extracts. Cells were treated with *Hyrtios* sp. extract (80 μg/mL) and incubated for 48 h. (a) Scale bar = 500 μm. (b) Scale bar = 50 μm.

48 h, consistent with previous studies [8–10]. Surprisingly, RKO-E6 cells, which lack functional p53, showed slightly induced p53 protein expression after treatment with *Hyrtios* sp. extracts. However, RKO-E6 cells treated with *Hyrtios* sp. extracts inhibited cell growth through induction of p21 protein via a p53-independent pathway. These results suggested that suppression of JNK protein might be involved in p21 protein expression as previous study [11]. The JNK pathway is involved in phosphorylation of c-Jun [12] and p53 [13] and regulation of cell growth [14]. As reported by Potapova [15], inhibition of JNK reduced the growth of p53-deficient cells and induced p21 protein expression. Our results were consistent with this study [15].

3.4. Induction of Apoptosis by *Hyrtios* sp. Extract. To verify cellular apoptosis, we used Annexin V and PI staining to investigate RKO and RKO-E6 cells treated with *Hyrtios* sp. extracts. As shown in Figure 5, apoptotic cells were detected similarly in RKO and RKO-E6 cells without treatment of
3.73% 84.49%
Control Hyrtios sp.

100 101 102 103 104 105
FL2 (intensity)
102 103 104 105
FL1 (intensity)

RKO
RKO-E6

Figure 4: Expression of p53, p21, and JNK. Cells were treated with Hyrtios sp. extract (80 µg/mL) and incubated for 48 h. Protein expression was determined by western blots according to “Section 2”.

Figure 5: Hyrtios sp. extracts induced apoptosis. Cells were treated with Hyrtios sp. extract (80 µg/mL) and incubated for 24 h (a) or 48 h (b). Apoptotic cells were detected by Annexin V and PI staining.

Hyrtios sp. extracts. Hyrtios sp. extracts time-dependently decreased live cells and increased apoptotic cell death. RKO cells treated with Hyrtios sp. extract showed more apoptotic cells than RKO-E6 cells treated with Hyrtios sp. Extract. Therefore, Hyrtios sp. extracts induced more apoptosis in RKO cells than in RKO-E6 cells.

4. Conclusions
In this study, we investigated anticancer effects of Hyrtios sp. extract on p53 wild-type RKO cells and p53-deficient RKO-E6 cells. Our results indicated that Hyrtios sp. extract suppressed cell growth in both cell lines, but RKO cells,
which have functional p53, were more sensitive to the cytotoxic effects of Hyrtios sp. extract than RKO-E6 cells. The anticancer effect of Hyrtios sp. extract was induced via activation of the p53 pathway in RKO cells and suppression of the JNK pathway in RKO-E6 cells. Hyrtios sp. is a natural marine sponge that could be a good candidate for cancer treatments. In a further study, the bioactive components of Hyrtios sp. will be investigated.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

