

Research Article

Dynamics of Alloplastic Bone Grafts on an Early Stage of Corticotomy-Facilitated Orthodontic Tooth Movement in Beagle Dogs

Hyung-Joo Choi,^{1,2} Dong-Yeol Lee,³ and Tae-Woo Kim¹

¹ Seoul National University School of Dentistry & Dental Research Institute, Seoul 110-768, Republic of Korea

² Seoul Barun Dental Clinic, Anyang, Gyeonggi-do 431-050, Republic of Korea

³ Department of Oral and Maxillofacial Pathology and Research Center for Tooth & Periodontal Regeneration (MRC), School of Dentistry, Kyung Hee University, Seoul 130-701, Republic of Korea

Correspondence should be addressed to Tae-Woo Kim; taewoo@snu.ac.kr

Received 22 July 2014; Accepted 5 August 2014; Published 3 September 2014

Academic Editor: Seong-Hun Kim

Copyright © 2014 Hyung-Joo Choi et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Alveolar augmented corticotomy is effective in accelerating orthodontic tooth movement, but the effect only lasts for a relatively short time. Therefore, the purpose of this study was to investigate the underlying biology of the immediate periodontal response to orthodontic tooth movement after a corticotomy with alloplastic bone grafts. The results demonstrated that measurable tooth movement began as early as 3 days after the intervention in beagle dogs. Based on the results and histological findings, augmented corticotomy-facilitated orthodontic tooth movement might enhance the condition of the periodontal tissue and the stability of the outcomes of orthodontic treatment.

1. Introduction

New appliances, materials, and mechanics of orthodontic treatment are being developed every day, but much of the biology of orthodontic tooth movement still needs to be clarified. Many adjunctive modalities are available to accelerate orthodontic tooth movement in humans, such as corticotomy [1–9], distraction osteogenesis [10], mechanical vibration [11], medication with local prostaglandins, and low-level laser treatment [12, 13]. Among these interventions, corticotomy is known to be the most effective means to accelerate orthodontic tooth movement [7, 14]. In experiments that used a split-mouth design, with corticotomy performed on one side and the other side serving as the control, the velocity of the tooth movement was accelerated on the corticotomy side [1, 15–17], and the amount of movement doubled over the duration of the experiments [15–18].

The initial microscopic changes and early application of orthodontic force have been emphasized in corticotomy-facilitated orthodontic tooth movement [14], and it has been hypothesized that a corticotomy or an osteotomy can lead to

intensified osteoclastic activity resulting in local osteopenia and increased bone remodeling [2, 15, 19, 20]. To date, however, the immediate periodontal response has not been fully elucidated. Most researchers have either studied the small animals such as rats, cats, or rabbits [2, 6, 18–20], for relatively long periods of 6–12 weeks [4, 5], or carried out gross observational studies with no histologic measurements [1, 2, 9, 17, 21].

The purpose of this study was to investigate the immediate periodontal response to a corticotomy with alloplastic bone grafts in beagle dogs.

2. Materials and Methods

2.1. Animal Subjects. Five adult male beagle dogs, weighing 10–13 kg, were used in the experiment, and their selection, care, and preparation, together with the surgical protocol, were carried out according to the guidelines for animal experiments (IRB no. KHMC-IACUC2012-024). They were caged separately under regulated conditions and fed a normal diet

and water *ad libitum* to secure the experimental orthodontic appliances.

For the preparation processes and surgical procedures, the animals were anesthetized with a mixture of tiletamine-zolazepam and xylazine, via intramuscular and intravenous injections using a catheter in the vessel of the ear.

2.2. Study Preparation. Alginate impressions of each beagle were taken to make study models, and orthodontic appliances were custom-made for each model. The canine and fourth premolar teeth were banded to form anchor teeth, and a \varnothing 0.9 mm stainless steel wire was welded onto the buccal surface of the bands. The second and third premolars were banded with a lingual button. After 2 weeks, the animals were anesthetized to fit the orthodontic appliances to the teeth (Figure 1).

2.3. Surgical Procedures for the Corticotomy and Alloplastic Bone Graft. Under general anesthesia, 2% lidocaine with 1:100,000 epinephrine was also infiltrated to the surgical sites. An intrasulcus incision was performed with a no. 12 blade from the canine tooth to the first molar, and a full-thickness flap was lifted. The circumscribing corticotomy (Figure 2(a)) was performed with a round bur (\varnothing 1.5 mm) under sterile saline irrigation. Alloplastic bone material (MBCP+, Biomatlante, Vigneux de Bretagne, France), composed of 20% hydroxyl apatite and 80% β -tri-calcium phosphate, was used for the graft. The graft bone was soaked with blood, and 1 g of the MBCP+ was grafted onto the surgical surface (Figure 2(b)). The mucoperiosteal flaps were repositioned and sutured with 5-0 nylon and primary closure was obtained (Figure 2(c)). A closed coil spring made of nickel-titanium shape memory wire of 200 g force was applied to the second and third premolars in a buccolingual direction (Figure 2(d)).

All of the surgical procedures were performed under sterile conditions to prevent infection. After surgery, antibiotics and anti-inflammatory analgesics were administered by intramuscular injection twice a day for 6 days. A 1% chlorhexidine-gluconate solution dressing was applied simultaneously for infection control. A soft diet was supplied for 1 or 2 weeks and then a normal diet. Mechanical plaque control was performed once a week. The animals were euthanized with an overdose of thiopental sodium after 1 day, 3 days, 1 week, 2 weeks, and 4 weeks following the surgery (Figure 3).

2.4. Histological Processing and Analysis. Following a predetermined time schedule, after the animals were killed, their maxillae and mandibles were dissected, and the sections containing the canine to the fourth premolar teeth were retrieved. The block specimens were rinsed in sterile saline and immediately immersed in 10% neutral-buffered formalin fixatives for 14 days. The block specimens were large, and rapid decalcification was performed for 6 days using 5% nitric acid because it is sufficiently strong [22]. Had this study been designed for immunohistology, the use of ethylenediamine-tetra-acetic acid or 10% aqueous or formic acid would have been suitable [23], but this was not the case [5, 24].

The specimens were then dehydrated through a series of ethanol solutions of increasing concentrations and embedded in paraffin. Buccolingual sections were sliced with a microtome set at 5 μ m and stained with the Masson's trichrome solution. One slide was processed per experimental tooth.

Histological examinations were conducted using a light microscope (Olympus BX 51, Olympus, Tokyo, Japan) equipped with a DP controller 3.2.276.2 and DP manager 3.1.1.208 (Olympus, Tokyo, Japan). After microscopic examination, a photograph of each slide was taken with a digital camera (Olympus DP 71, Olympus, Tokyo, Japan). With imaging software (cellSens version 1.6, Olympus), we measured the buccal tipping angle ($^{\circ}$) and distance (μ m). The buccal tipping angle was measured from the reversal line of the lingual/palatal bone wall to the lingual/palatal root surface. The buccal tipping distance was measured from the lingual/palatal alveolar crest to the shortest lingual/palatal root surface. A total of 40 slides were fabricated and examined. After taking photographs, the amount of tooth movement was measured three times in each slide for angular changes and linear displacement.

2.5. Statistical Data Analysis. Statistical data analysis was performed using the R programming language [25]. The data on tooth movement did not fulfill the parametric conditions of normality and equality of variance after the D'Agostino normality test was performed. We therefore conducted the Kruskal-Wallis rank sum test to determine whether there existed a significant between-group difference in general and the Wilcoxon test to find a significant pair between two groups. The Bonferroni correction and the Type I error were applied to counteract the problem of multiple comparisons. The data were analyzed with a confidence level of 95%.

3. Results

3.1. Clinical Findings. Figure 4 shows the amount of tooth movement for each beagle dog. Due to the minimal tooth movement, the buccal tipping angle at 1 day could not be measured. No significant difference was observed between the teeth in the maxilla and those in the mandible or in the right or left part of the dentition. When the tipping movement was measured from the angular changes, no statistically significant difference was found among the experimental groups (Figure 4(a)). However, the linear measurements demonstrated statistically significant differences between 1 day and 3 days and between 2 weeks and 4 weeks after the start of the experiment (Figure 4(b)).

3.2. Histological Observations. After 1 day of orthodontic movement, a microphotograph of the buccopalatal/lingual section (Figure 5) showed compression of the periodontal ligament (PDL) (Figure 5(b)), extravasations of red blood cells (RBC) (Figure 5(c)), and reduced capillaries (Figure 5(e)) on the pressure side. No significant finding was observed on the tension side.

At 3 days, the PDL was more severely compressed and fewer cells were found in the PDL space (Figure 6(c)) on the

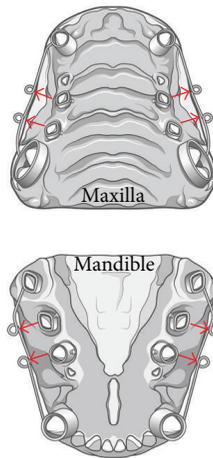


FIGURE 1: Orthodontic appliances were custom-made for each study model. The canine and fourth premolar teeth were banded to form an anchor tooth, and a Ø 0.9 mm stainless steel wire was welded on the buccal surface of the bands. The second and third premolars were banded with a lingual button. After 4 weeks, the animals were anesthetized to fit the orthodontic appliances to the teeth. The arrows indicate the direction of force.

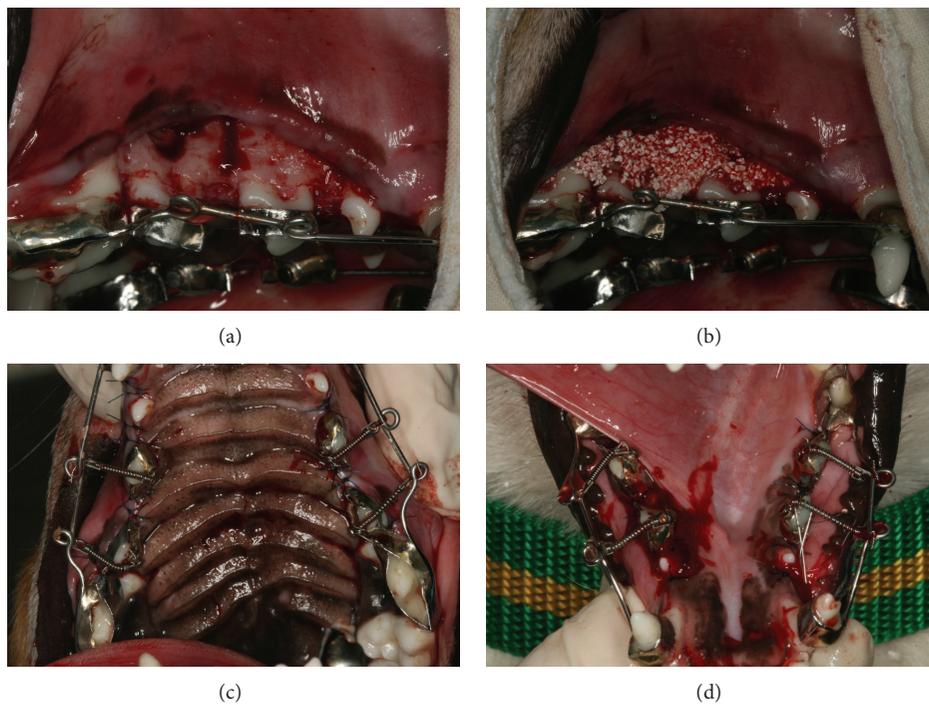


FIGURE 2: Surgical procedure. (a) Corticotomy: the circumscribing corticotomy was performed with a round bur (Ø 1.5 mm) under sterile saline irrigation. (b) MBCP⁺ (Biomatlante, Vigneux de Bretagne, France), composed of 20% hydroxyl apatite and 80% β -tri-calcium phosphate, was used as the alloplastic bone graft material. The graft bone was applied soaked in blood; 1 g of the MBCP⁺ was grafted on the surgical surface. (c) A closed coil spring made of nickel-titanium shaped memory wire of 200 g force was applied to the second and third premolars in a buccolingual direction in the maxilla and (d) in the mandible.

buccal pressure side. The tension side at the lingual alveolar bone crest contained more cells than the pressure side and active osteoblasts forming a new bone (Figure 6(h)).

At 1 week after the start of the experiment, most of grafted MBCP⁺ particles were well maintained (Figure 7). On the pressure side, the PDL space was slightly widened compared

with that at 3 days (Figure 7(b)), and the tension side contained abundant PDL fibroblasts and active osteoblasts (Figures 7(d), 7(e), and 7(f)).

At 2 weeks, undermining resorption and a resorption bay were observed on the buccal pressure side (Figures 8(b), 8(c), 8(q), and 8(l)). In contrast, on the buccal tension

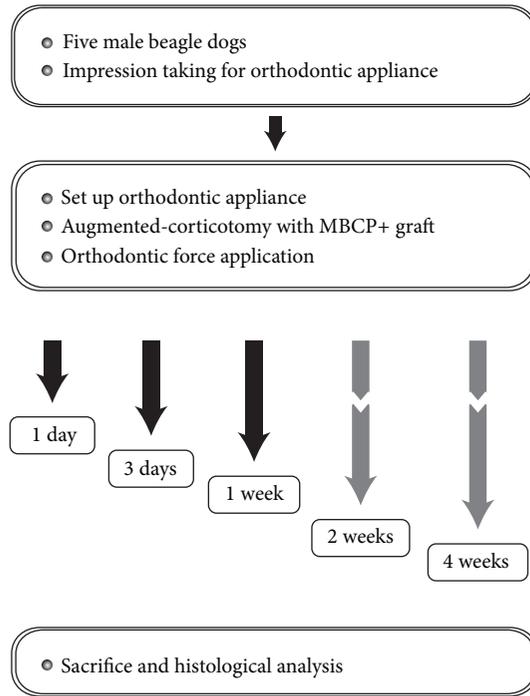


FIGURE 3: Schematic diagram describing the experiment design.

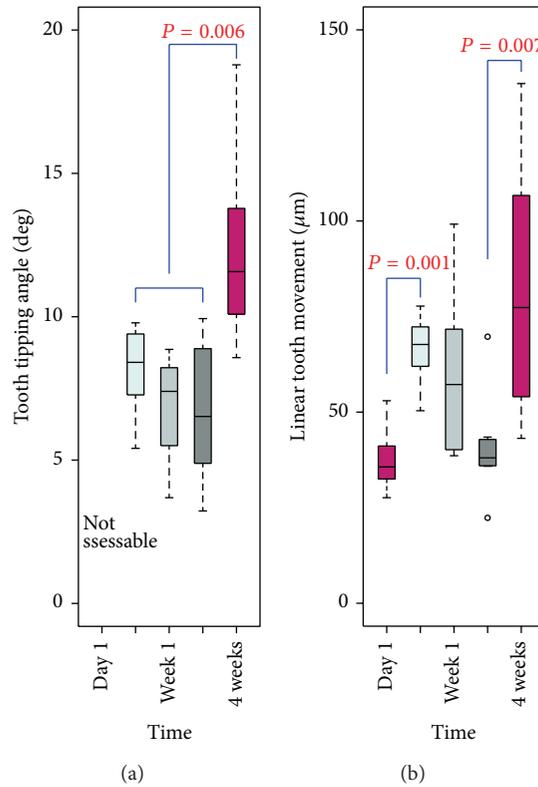


FIGURE 4: Box plots for the tooth movement measurements. (a) No statistically significant difference in the angular tooth movement was observed over time. (b) However, the linear measurements demonstrated statistically significant differences between 1 day and 3 days and between 2 weeks and 4 weeks after the start of the experiment.

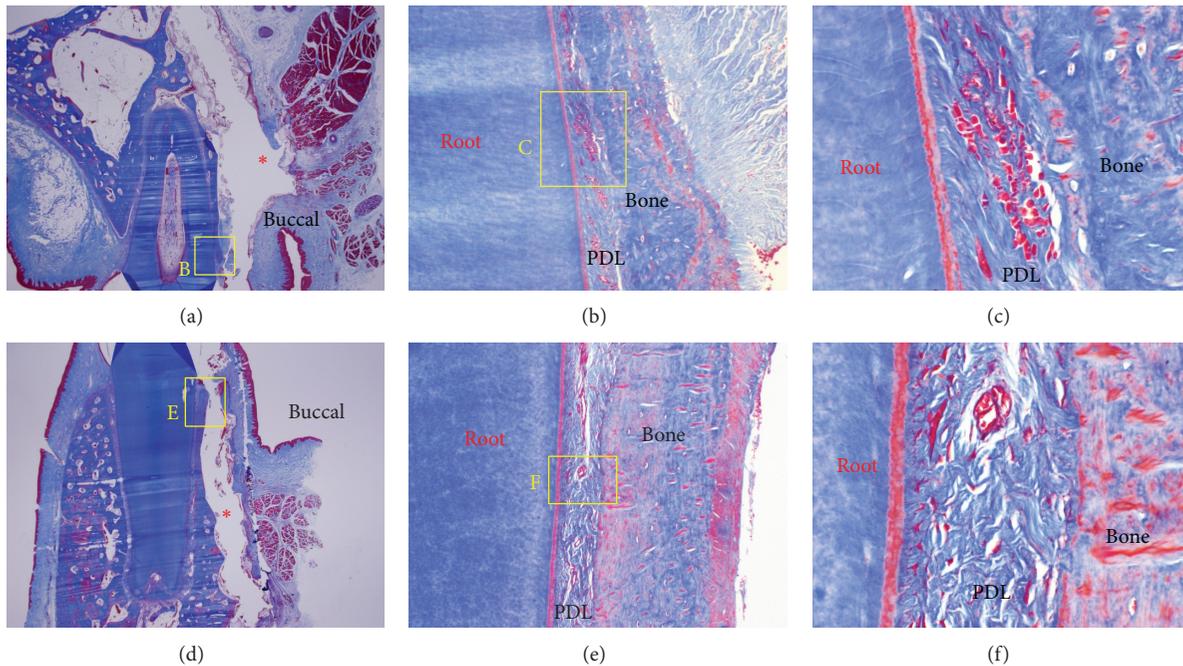


FIGURE 5: Microphotograph of a buccopalatal/lingual section of the 1-day experiment. (a) Maxilla. (b) Higher magnification of (a). On the pressure side, the PDL was compressed. (c) Higher magnification of (b). Extravasation of RBC was observed. (d) Mandible. (e) Higher magnification of (d). Compression of the PDL was shown, and reduced capillaries were identified. (f) Higher magnification of (e). The number of cells was reduced, and grafted MBCP+ particles were lost in the process of making the histological section; the red * in (a) and (d) indicates empty spaces that were occupied by MBCP+ graft particles. Masson's trichrome stain. Original magnification was $\times 12.5$ for (a) and (d), $\times 100$ for (b) and (e), and $\times 400$ for (c) and (f).

side, new bone formation surrounding and bridging the MBCP+ particles was seen (Figures 8(d), 8(j), and 8(f)) due to abundant osteoblasts (Figures 8(e) and 8(k)).

At 4 weeks after the start of the experiment, new bone formation along the PDL formed a new buccal bone wall on the pressure side (Figure 9(a)). Also, a new bone island was formed in the center of the bone-derived mesenchymal matrix (Figure 9(d)), and osteoblasts and osteocytes were observed (Figure 9(e)). On the buccal side, grafted MBCP+ particles were bridged with newly formed bone in the bone-derived mesenchymal matrix. Entrapped osteocytes and aggregated osteoblasts were observed (Figure 9(g)). The palatal crestal (Figure 9(j)) and apical (Figure 9(l)) tension sides showed aggregated osteoblasts and active forms of osteoblasts, and a new bone-forming buccal bone wall and crest were observed (Figure 9(o): native bone (red star) and new bone (yellow star)). New bone was formed on the outer and inner surfaces of the native bone (Figures 9(n) and 9(q)), and the outer portion of the bone-derived mesenchymal matrix could be seen (Figure 9(t)).

4. Discussion

The biological mechanism by which the tooth movement is facilitated after a corticotomy has been suggested to be mediated by a regional acceleratory phenomenon [26, 27],

which might boost the appearance of the macrophages that eliminate the hyaline as early as 1 week after the application of orthodontic force [15, 19]. For this reason, we designed an experiment to determine immediate periodontal responses, which represents the first study to observe the immediate effect of corticotomy-facilitated orthodontic tooth movement. Two studies previously reported observations made 3 days after a corticotomy in rats [2, 20]. However, the results obtained in small animals may differ from those in larger animals [14]. We believe that the current study is unique because we observed the histological responses of periodontal tissue as early as 1 day, 3 days, 1 week, 2 weeks, and 4 weeks after a corticotomy and force application for orthodontic tooth movement in larger animals.

In most experiments in dogs or rats, corticotomy-facilitated tooth movement was observed at a rate of about 1 mm per month, which was almost double that observed on the control side [15–18]. In this study, we observed significant tooth movement within 3 days (Figure 4). We measured the width of the PDL at the crest of the lingual/palatal sides, which does not represent the direct distance of clinical tooth movement. The significant tooth movement observed 4 weeks after the start of the experiment was to be expected, but this obviously occurred earlier in the experiment. The tooth movement measurements demonstrated a similar pattern in angular changes and linear displacement, as shown in Figure 4. We conjectured that this might imply that the

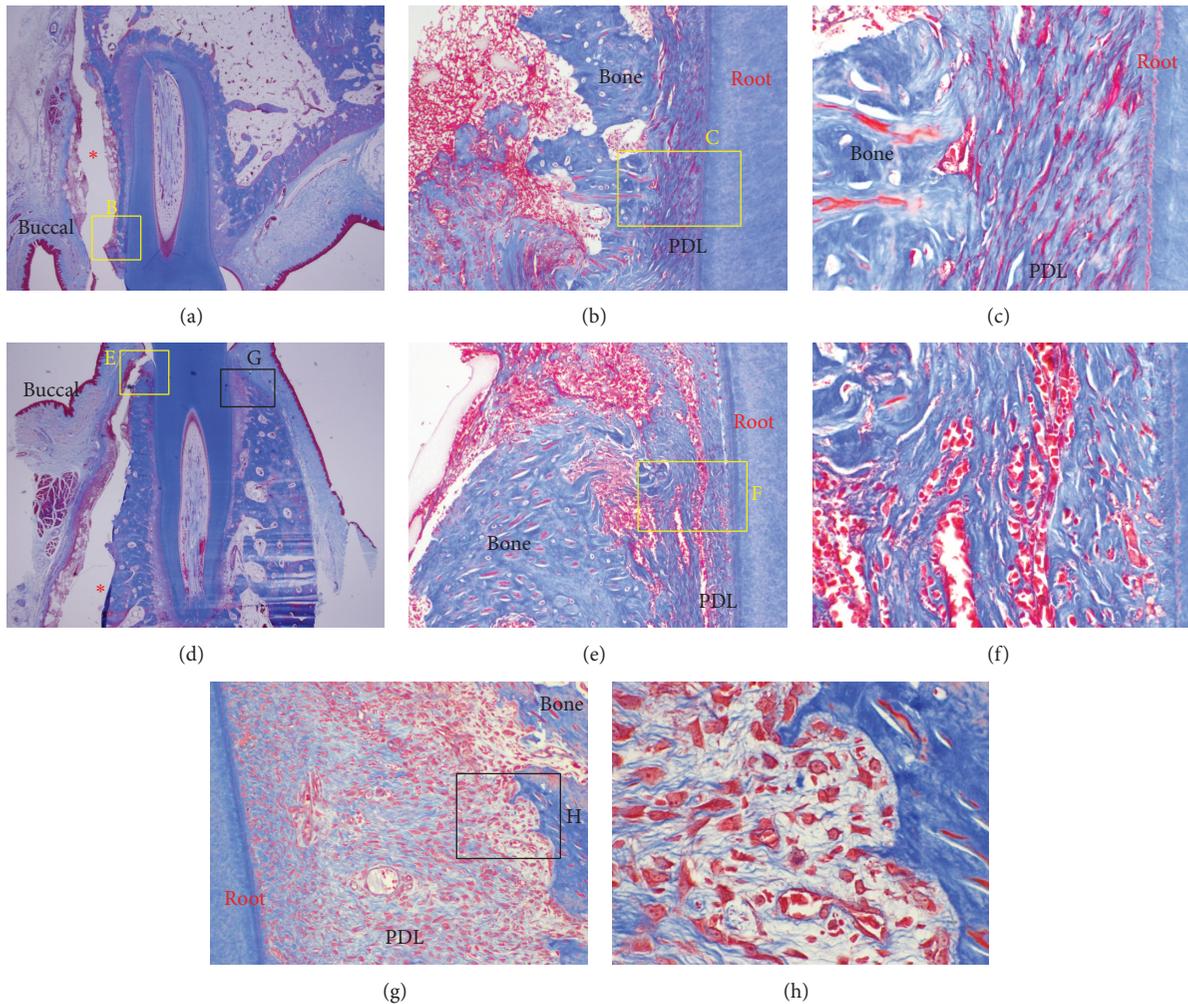


FIGURE 6: Microphotograph of a buccopalatal/lingual section of the 3-day experiment. (a) Maxilla. (b) Higher magnification of (a). On the pressure side, the PDL was more severely compressed. (c) Higher magnification of (b). Few cells were observed in the PDL space. (d) Mandible. (e) Higher magnification of (d). In common with the maxilla, the PDL was severely more compressed on the pressure side. (f) Higher magnification of (d). Extravasation of RBC was observed in the PDL space. (g) Tension side at the lingual bone crest. The tension side showed abundant cells compared with the pressure side. (h) Higher magnification of (g). Active osteoblasts forming new bone were observed. Grafted MBCP+ particles were lost in the process of making the histological section; the red * in (a) and (d) indicates empty spaces which were occupied by MBCP+ graft particles. Masson's trichrome stain. Original magnification was $\times 12.5$ for (a) and (d), $\times 100$ for (b), (e), and (f), and $\times 400$ for (c), (f), and (h).

pattern of rapid tooth movement was not a bodily translation in general but mostly occurred through tipping of the tooth. Therefore, during clinical orthodontic treatment, methods to control for unwanted tipping should also be considered.

Despite the similar pattern between the angular and linear changes, the linear measurements exhibited a statistically significant difference while the angular changes did not. Linear measurements are assessed between two points while degrees of angles are measured between three points, from which a variation in angular measurements can be produced. This may be the cause of the larger variation in angular measurements than in linear measurements.

A typical cell-free zone on the pressure side and an inflammatory reaction were seen on the 1- and 3-day slides

(Figures 5 and 6). A force of 200 g may be strong for tipping movement, but it was difficult to determine the hyalinization layer in the 1- and 3-day slides. The corticotomy was probably responsible for this, but the underlying mechanism was not clear. The progenitor cells regarded as osteoblasts in the 3-day slides showed strong cellular activity.

On the 1-week slide, we observed that the PDL space was widened at the pressure side compared with that of the 3-day slide (Figure 7(b)). This could be explained by undermining resorptions. On the tension side, active osteoblasts forming a new bone were observed. This new bone formation is a common phenomenon in orthodontic tooth movement.

Interestingly, new bone surrounding the graft materials was observed on the compression side on the 2-week slides

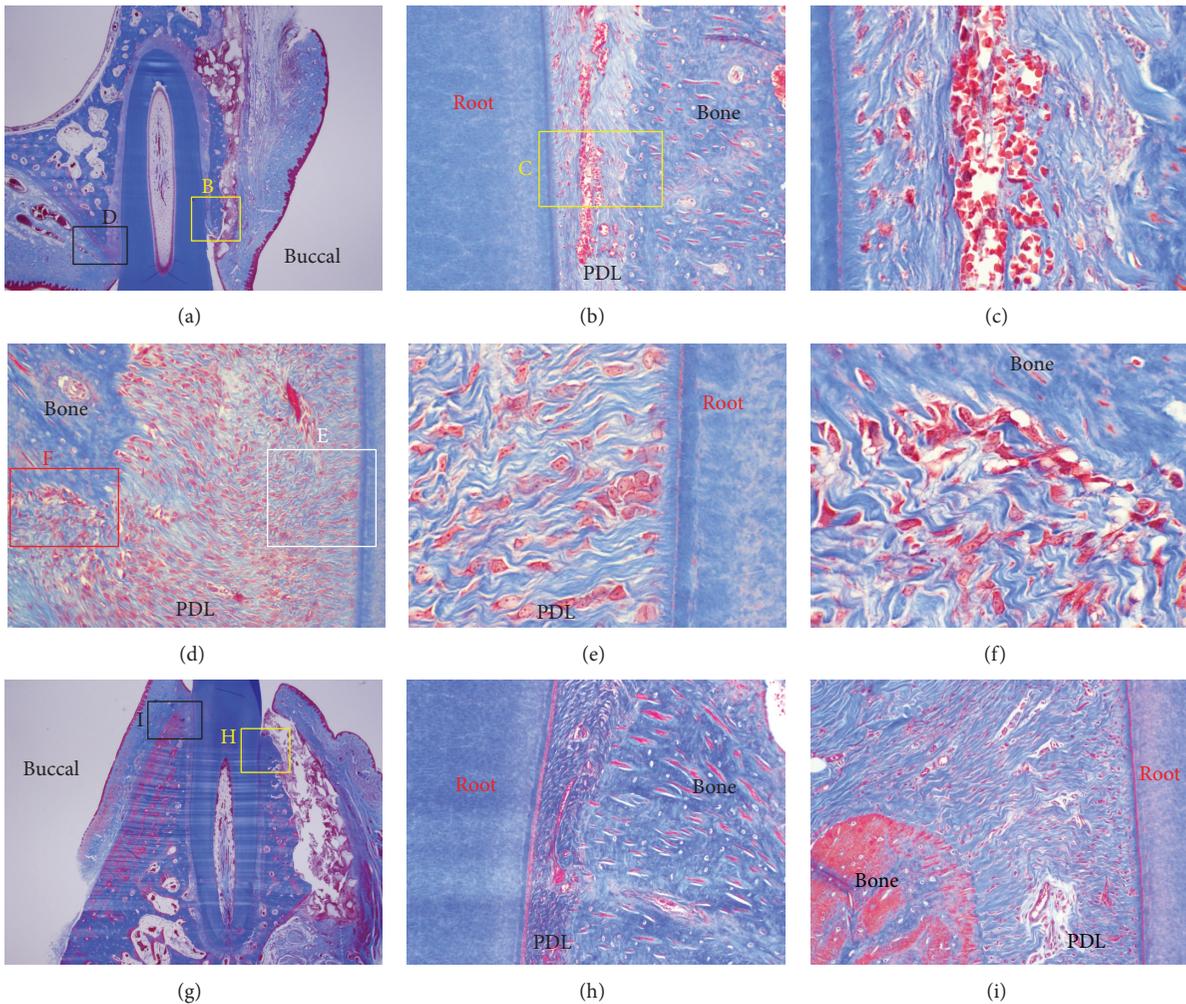


FIGURE 7: Microphotograph of a buccopalatal/lingual section of the 1-week experiment. Most of the grafted MBCP+ particles were well maintained. (a) Maxilla. (b) Higher magnification of (a). On the pressure side, the PDL space was wider than at 3 days. (c) Higher magnification of (b). Extravasation of RBC was observed in the PDL space. (d) Tension side at the palatal bone crest. The tension side showed abundant cells compared with the pressure side and a widened PDL space. (e) Higher magnification of (d). Abundant PDL fibroblasts were seen. (f) Higher magnification of (d). Active osteoblasts forming a new bone were observed. (g) Mandible. Most of the grafted MBCP+ particles were well maintained. (h) Higher magnification of (g). Pressure side. The PDL was compressed. (i) Higher magnification of (g). The tension side showed a widened PDL space. Masson's trichrome stain. Original magnification was $\times 12.5$ for (a) and (g), $\times 100$ for (b), (d), (h), and (i), and $\times 400$ for (c) and (f).

(Figure 8(d)) and at the buccal sides distant from PDL (Figures 8(s), 8(t), and 8(u)). Graft particles were bridged by newly formed bone and osteoclastic and osteoblastic activities were both seen (Figure 8(u)).

From the findings of the 4-week sections, the new bone formation in the center of the MBCP+ graft material on the buccal side was very distinctive (Figures 9(h) and 9(i)), and many entrapped osteocytes and aggregated osteoblasts were observed (Figure 9(g)). However, the cause of this new bone formation on the buccal sides around the graft materials was not certain. Further studies could provide the answer.

The slides taken at 2 and 4 weeks did not show a distinctive loss of periodontal attachment, and the small areas of root resorption that were seen were not significant.

Orthodontic patients have complained about the length of their treatment, and it has become necessary to develop adjunctive methods to tackle this problem [1, 3, 7–13]. Alveolar corticotomy is effective in accelerating orthodontic tooth movement [2, 7, 14]. However, according to pertinent studies [1, 14, 17], the regional acceleratory phenomenon only persists for about 4 months, after which the tooth movement rate returns to normal. To make use of this “window” period in an effective, efficient, and efficacious way, it is imperative to understand the underlying biology of early periodontal responses to tooth movement after augmented corticotomy and to develop appropriate clinical procedures. Also, from the clinical point of view, it is important to determine how to reduce the length of total treatment time. When should

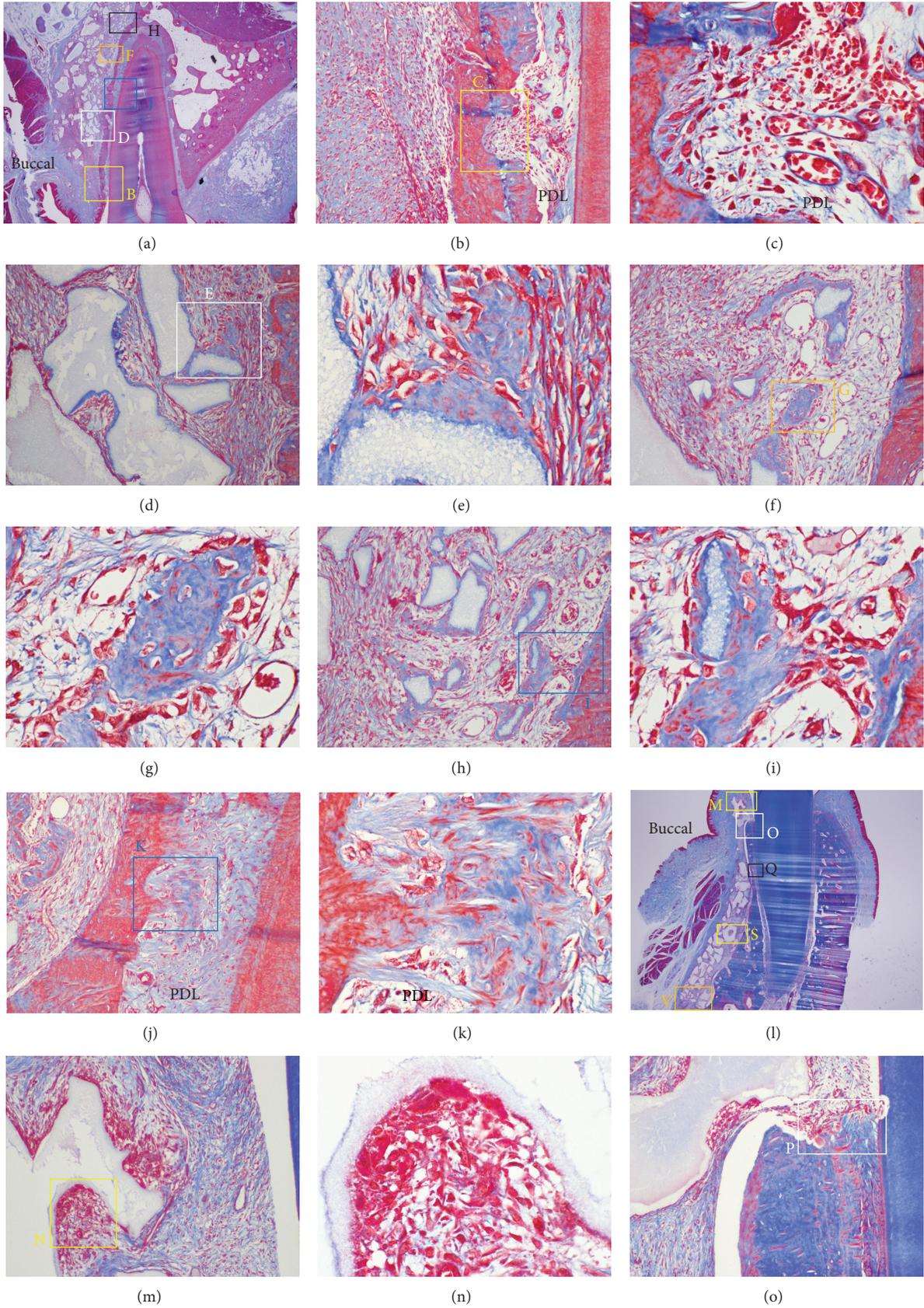


FIGURE 8: Continued.

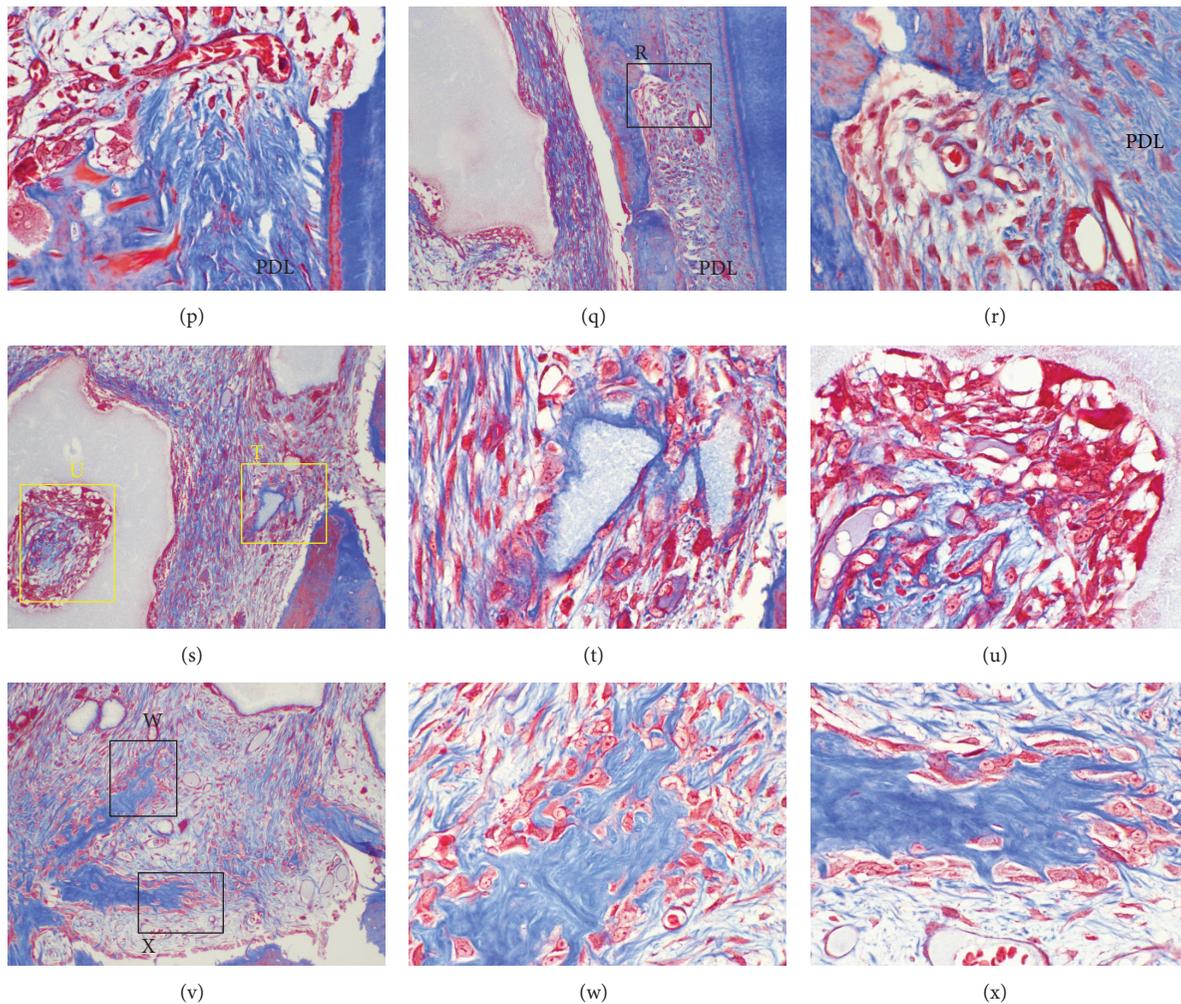


FIGURE 8: Microphotograph of a buccopalatal/lingual section of the 2-week experiment. (a) Maxilla. Most of the grafted MBCP+ particles were well maintained. (b) Higher magnification of (a) at the crest area. Undermining resorption on the pressure side was observed. (c) Higher magnification of (b). Extravasation of RBC was observed in the PDL space. Resorption bays, which indicate undermining resorption on the pressure side, were also observed. (d) Higher magnification of (a) at the buccal bone surface. New bone formation surrounding the grafted MBCP+ particles was observed. Grafted MBCP+ particles were bridged by newly formed bone. (e) Higher magnification of (d). Abundant osteoblasts were forming new bone. (f) Higher magnification of (a). Osteoblasts were forming a new bone island. (h) Higher magnification of (a). Grafted particles encircled by new bone were bridged with the buccal bone surface. (j) Higher magnification of (a). New bone was formed in the PDL space at the buccal tension area. New bone was formed from the bone. (l) Mandible. Most of the grafted MBCP+ particles were well maintained. (m) Higher magnification of (l) at the crest area. Grafted particles were resorbed by osteoclasts. (n) Higher magnification of (m). (o) Higher magnification of (l). (p) Higher magnification of (l). Bone and root surface resorption by osteoclasts were observed. (q) Higher magnification of (l). Undermining resorption at the buccal bone in the PDL area was observed. (r) Higher magnification of (q). Many osteoblasts filled the resorption bay. (s) Higher magnification of (l). Active new bone formation was found at the buccal bone surface in the apical area. (t) Higher magnification of (s). Many osteoblasts were forming a new bone encircling the grafted MBCP+ particles. (u) Higher magnification of (s). Osteoclastic and osteoblastic activities were both observed. (v) Higher magnification of (l). New bone islands were formed. (w) Higher magnification of (v). (x) Higher magnification of (v). Abundant osteoblasts were actively forming new bone islands. Masson's trichrome stain. Original magnification was $\times 12.5$ for (a) and (l); $\times 100$ for (b), (d), (f), (h), (j), (m), (o), (q), (s), and (v), and $\times 400$ for (c), (e), (g), (i), (k), (n), (p), (r), (t), (u), (w), and (x).

the corticotomy be performed? How often should orthodontic force be applied? What magnitude of force would be optimal? These are still widely open questions that are worthy of further investigation.

Although there is no doubt that this procedure could align teeth within a shorter period of time [1, 3, 8, 14, 28–31], at the present time, we cannot expect augmented

corticotomy-facilitated orthodontic tooth movement to reduce the entire length of orthodontic treatment noticeably in adult patients. In addition, there is a paucity of information in the available research to assert that grafting enhances the stability of orthodontic treatment [14]. To prove this, well-designed randomized controlled clinical trials should be performed.

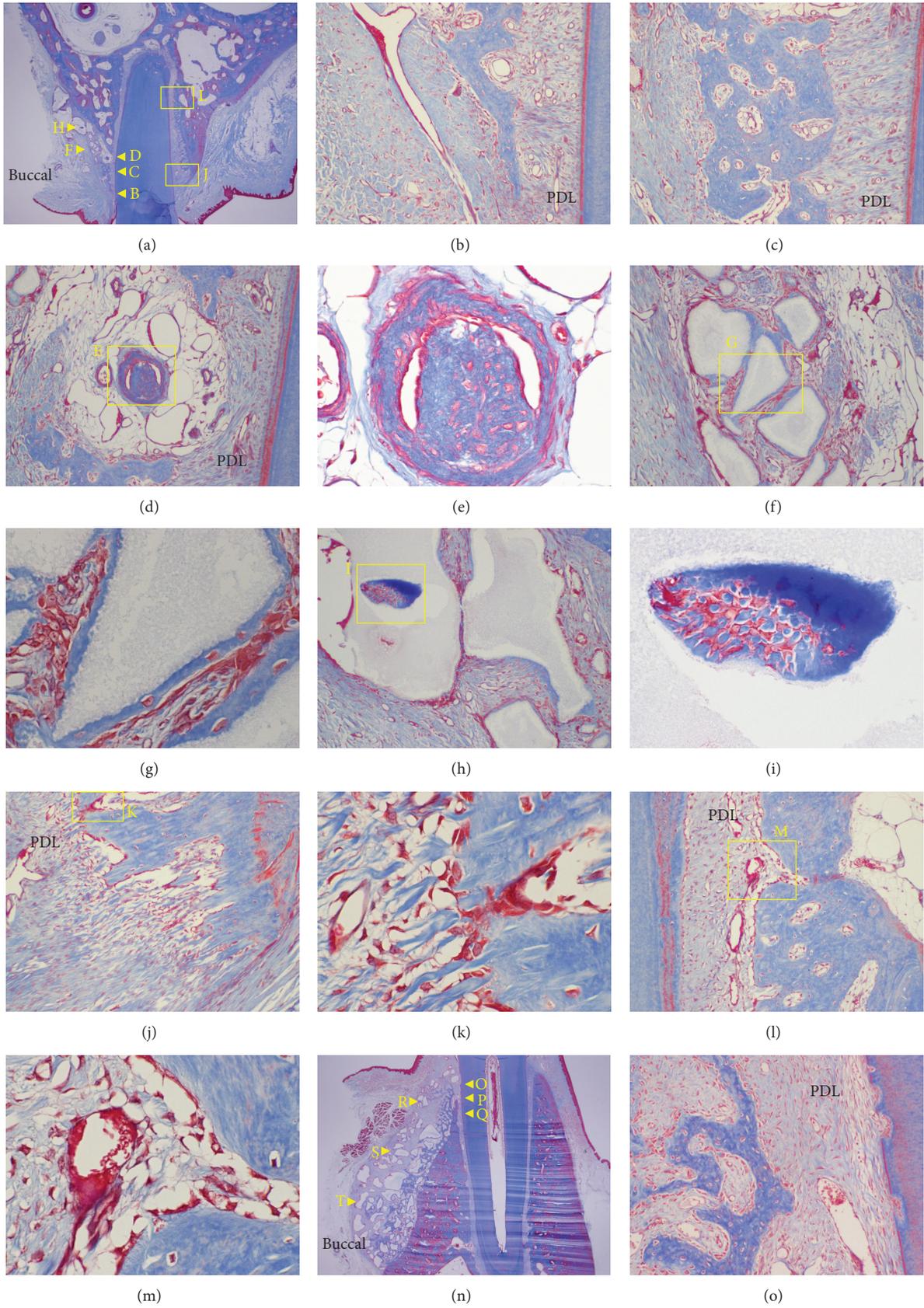


FIGURE 9: Continued.

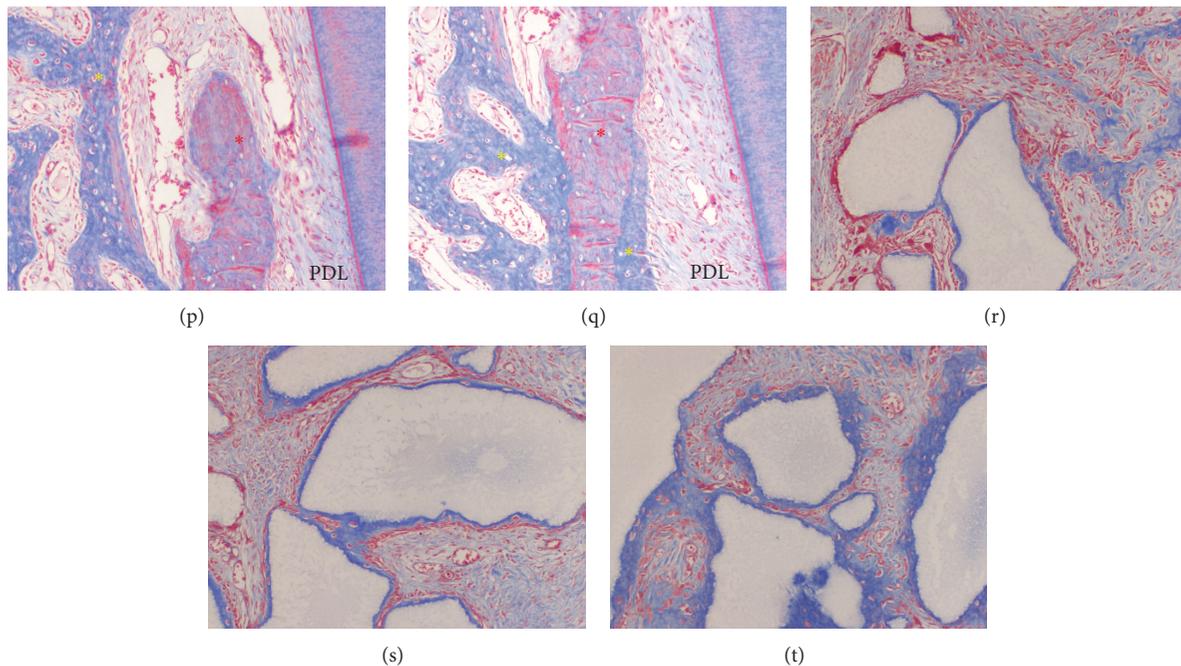


FIGURE 9: Microphotograph of a buccopalatal/lingual section of the 4-week experiment. (a) Maxilla. Most of the grafted MBCP+ particles were well maintained. New bone formation along the PDL formed a new buccal bone wall. ((b) and (c)) Higher magnification of (a) at the crest area. (d) Higher magnification of (a). The bone-derived mesenchymal matrix bordered the PDL-derived mesenchymal matrix. A new bone island was formed in the center of the bone-derived mesenchymal matrix. (e) Higher magnification of (d). Osteoblasts and osteocytes were observed. (f) Higher magnification of (a). The grafted MBCP+ particles were bridged with newly formed bone in the bone-derived mesenchymal matrix on the buccal side. (g) Higher magnification of (f). Entrapped osteocytes and aggregated osteoblasts were observed. (h) Higher magnification of (a). (i) Higher magnification of (h). New bone was formed in the center of the grafted MBCP+ particles at the buccal side. ((j) and (l)) Higher magnification of (a). The palatal tension side at the crestal (j) and apical (l) areas. ((k) and (m)) Higher magnification of (j) and (l), respectively. Aggregated osteoblasts and active form of osteoblasts were seen. (n) Mandible. (o) Higher magnification of (n) at the crestal area. New bone forming a buccal bone wall and crest was observed. (p) Higher magnification of (n). Native bone (red star) and new bone (yellow star). (q) Higher magnification of (n). New bone (yellow star) was formed on the outer and inner surface of the native bone (red star). ((r), (s), and (t)) Higher magnification of (n). The outer portion of the bone-derived mesenchymal matrix. Grafted MBCP+ particles were bridged by the newly formed bone. Masson's trichrome stain. Original magnification was $\times 12.5$ for (a) and (n), $\times 100$ for (b), (c), (d), (f), (h), (j), (l), (o), (p), (q), (r), (s), and (t), and $\times 400$ for (e), (g), (i), (k), and (m).

The combination of corticotomy with an alveolar graft was introduced by Wilcko et al. and is referred to as accelerated osteogenic orthodontics or periodontally accelerated osteogenic orthodontics [8, 9, 32]. They asserted that bone grafting of the labial and lingual cortical bones would increase the stability of orthodontic treatment, enhance the range of possible tooth movements, increase alveolar bone volume, and provide a more structurally stable periodontium. However, no convincing scientific or histological evidence was available other than clinical reports. In some case reports, increased volume of bone around the alveolus was observed after bone grafting [3, 5, 9]. The present study supported the hypothesis that tooth movement with an augmented corticotomy might enhance orthodontic tooth movement, because we found new bone formation at the buccal surface. However, it focused on initial responses, and further investigation is needed to clarify its findings.

Tooth movement that was thought to be difficult or impossible to produce in the past has become possible due to the development of orthodontic mechanics and new appliances and materials. Therefore, orthodontic treatment

is mainly limited by the scope of the alveolar bone, and orthognathic surgery is required when this limit is exceeded. If an augmented corticotomy could increase the alveolar bone volume, this would help patients who have a limited amount of supporting alveolar bone.

Augmented corticotomy surgery is not free from some morbidity. It also requires a skilled clinician, and there may be some discomfort to patients and additional costs.

5. Conclusions

The findings from this study suggest that measurable tooth movement starts as early as 3 days after augmented corticotomy-facilitated orthodontic treatment and that this procedure might enhance the condition of periodontal tissue and the stability of orthodontic treatment outcomes.

Conflict of Interests

The authors declare that they have no conflict of interests. The authors also certify that no financial support was received

from commercial sponsors to conduct this study or the preparation of this paper.

Authors' Contribution

Hyung-Joo Choi and Dong-Yeol Lee contributed equally to this study.

Acknowledgment

This work was supported by the National Research Foundation of Korea, funded by the Korean government (Grant no. 2012RIA5A2051388).

References

- [1] S. M. B. E. Aboul-Ela, A. R. El-Beialy, K. M. F. El-Sayed, E. M. N. Selim, N. H. El-Mangoury, and Y. A. Mostafa, "Miniscrew implant-supported maxillary canine retraction with and without corticotomy-facilitated orthodontics," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 139, no. 2, pp. 252–259, 2011.
- [2] S. S. Baloul, L. C. Gerstenfeld, E. F. Morgan, R. S. Carvalho, T. E. van Dyke, and A. Kantarci, "Mechanism of action and morphologic changes in the alveolar bone in response to selective alveolar decortication-facilitated tooth movement," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 139, no. 4, supplement, pp. S83–S101, 2011.
- [3] S. Kim, I. Kim, D. Jeong, K. Chung, and H. Zadeh, "Corticotomy-assisted decompensation for augmentation of the mandibular anterior ridge," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 140, no. 5, pp. 720–731, 2011.
- [4] D. Y. Lee, H. W. Ahn, Y. Herr et al., "Periodontal responses to augmented corticotomy with collagen membrane application during orthodontic buccal tipping," *BioMed Research International*. In press.
- [5] K. B. Lee, D. Y. Lee, S. H. Ahn et al., "Tooth movement out of the bony wall using augmented corticotomy with non-autogenous graft materials for bone regeneration," *BioMed Research International*. In press.
- [6] W. Lee, G. Karapetyan, R. Moats et al., "Corticotomy-/osteotomy-assisted tooth movement microCTs differ," *Journal of Dental Research*, vol. 87, no. 9, pp. 861–867, 2008.
- [7] H. Long, U. Pyakurel, Y. Wang, L. Liao, Y. Zhou, and W. Lai, "Interventions for accelerating orthodontic tooth movement: a systematic review," *Angle Orthodontist*, vol. 83, no. 1, pp. 164–171, 2013.
- [8] K. G. Murphy, M. T. Wilcko, W. M. Wilcko, and D. J. Ferguson, "Periodontal accelerated osteogenic orthodontics: a description of the surgical technique," *Journal of Oral and Maxillofacial Surgery*, vol. 67, no. 10, pp. 2160–2166, 2009.
- [9] M. T. Wilcko, W. M. Wilcko, J. J. Pulver, N. F. Bissada, and J. E. Bouquot, "Accelerated osteogenic orthodontics technique: a 1-stage surgically facilitated rapid orthodontic technique with alveolar augmentation," *Journal of Oral and Maxillofacial Surgery*, vol. 67, no. 10, pp. 2149–2159, 2009.
- [10] S. Sayin, A. O. Bengi, A. U. Gürton, and K. Ortakoğlu, "Rapid canine distalization using distraction of the periodontal ligament: a preliminary clinical validation of the original technique," *Angle Orthodontist*, vol. 74, no. 3, pp. 304–315, 2004.
- [11] M. Nishimura, M. Chiba, T. Ohashi et al., "Periodontal tissue activation by vibration: intermittent stimulation by resonance vibration accelerates experimental tooth movement in rats," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 133, no. 4, pp. 572–583, 2008.
- [12] D. R. Cruz, E. K. Kohara, M. S. Ribeiro, and N. U. Wetter, "Effects of low-intensity laser therapy on the orthodontic movement velocity of human teeth: a preliminary study," *Lasers in Surgery and Medicine*, vol. 35, no. 2, pp. 117–120, 2004.
- [13] M. Yamaguchi, M. Hayashi, S. Fujita et al., "Low-energy laser irradiation facilitates the velocity of tooth movement and the expressions of matrix metalloproteinase-9, cathepsin K, and alpha(v) beta(3) integrin in rats," *European Journal of Orthodontics*, vol. 32, no. 2, pp. 131–139, 2010.
- [14] D. P. Mathews and V. G. Kokich, "Accelerating tooth movement: the case against corticotomy-induced orthodontics," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 144, no. 1, pp. 5–13, 2013.
- [15] S. Iino, S. Sakoda, G. Ito, T. Nishimori, T. Ikeda, and S. Miyawaki, "Acceleration of orthodontic tooth movement by alveolar corticotomy in the dog," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 131, no. 4, pp. 448–e1, 2007.
- [16] Y. A. Mostafa, M. M. S. Fayed, S. Mehanni, N. N. ElBokle, and A. M. Heider, "Comparison of corticotomy-facilitated vs standard tooth-movement techniques in dogs with miniscrews as anchor units," *American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 136, no. 4, pp. 570–577, 2009.
- [17] P. A. Sanjideh, P. E. Rossouw, P. M. Campbell, L. A. Opperman, and P. H. Buschang, "Tooth movements in foxhounds after one or two alveolar corticotomies," *European Journal of Orthodontics*, vol. 32, no. 1, pp. 106–113, 2010.
- [18] J. Sebouan, A. Kantarci, J. W. Turner, R. S. Carvalho, T. E. Van Dyke, and D. J. Ferguson, "Modeling of trabecular bone and lamina dura following selective alveolar decortication in rats," *Journal of Periodontology*, vol. 79, no. 9, pp. 1679–1688, 2008.
- [19] S. Kim, Y. Park, and S. Kang, "Effects of corticision on paradental remodeling in orthodontic tooth movement," *Angle Orthodontist*, vol. 79, no. 2, pp. 284–291, 2009.
- [20] L. Wang, W. Lee, D. Lei, Y. Liu, D. Yamashita, and S. L. Yen, "Tissue responses in corticotomy- and osteotomy-assisted tooth movements in rats: histology and immunostaining," *The American Journal of Orthodontics and Dentofacial Orthopedics*, vol. 136, no. 6, pp. 770.e1–770.e11, 2009.
- [21] S. Kim, S. Moon, S. Kang, and Y. Park, "Effects of low-level laser therapy after corticision on tooth movement and paradental remodeling," *Lasers in Surgery and Medicine*, vol. 41, no. 7, pp. 524–533, 2009.
- [22] W. H. B. Mawhinney, E. Richardson, and A. J. Malcolm, "Technical methods. Control of rapid nitric acid decalcification," *Journal of Clinical Pathology*, vol. 37, no. 12, pp. 1409–1413, 1984.
- [23] H. H. Verdenius and L. Alma, "A quantitative study of decalcification methods in histology," *Journal of Clinical Pathology*, vol. 11, no. 3, pp. 229–236, 1958.
- [24] J. B. Matthews and G. I. Mason, "Influence of decalcifying agents on immunoreactivity of formalin-fixed paraffin-embedded tissue," *Histochemical Journal*, vol. 16, no. 7, pp. 771–787, 1984.
- [25] R Development Core Team, *R: A Language and Environment for Statistical Computing*, R Foundation for Statistical Computing, Vienna, Austria, 2014.

- [26] H. M. Frost, "The biology of fracture healing: an overview for clinicians. Part II," *Clinical Orthopaedics and Related Research*, no. 248, pp. 294–309, 1989.
- [27] H. M. Frost, "The biology of fracture healing. An overview for clinicians. Part I," *Clinical Orthopaedics and Related Research*, no. 248, pp. 283–293, 1989.
- [28] T. J. Fischer, "Orthodontic treatment acceleration with corticotomy-assisted exposure of palatally impacted canines: a preliminary study," *Angle Orthodontist*, vol. 77, no. 3, pp. 417–420, 2007.
- [29] A. H. Hassan, A. A. Al-Fraidi, and S. H. Al-Saeed, "Corticotomy-assisted orthodontic treatment: review," *The Open Dentistry Journal*, vol. 4, pp. 159–164, 2010.
- [30] H. Nowzari, F. K. Yorita, and H. Chang, "Periodontally accelerated osteogenic orthodontics combined with autogenous bone grafting," *Compendium of Continuing Education in Dentistry*, vol. 29, no. 4, pp. 200–218, 2008.
- [31] W. M. Wilcko, "Rapid orthodontics with alveolar reshaping: Two case reports of decrowding," *International Journal of Periodontics and Restorative Dentistry*, vol. 21, no. 1, pp. 9–19, 2001.
- [32] G. Amit, K. Jps, B. Pankaj et al., "Periodontally accelerated osteogenic orthodontics (PAOO)—a review," *Journal of Clinical and Experimental Dentistry*, vol. 4, no. 5, pp. e292–e296, 2012.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

