Review Article

Carum copticum L.: A Herbal Medicine with Various Pharmacological Effects

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Carum copticum L. commonly known as “Ajwain” is cultivated in many regions of the world including Iran and India, states of Gujarat and Rajasthan. Traditionally, C. copticum has been used in the past for various therapeutic effects including bloating, fatigue, diarrhea, abdominal tumors, abdominal pain, respiratory distress, and loss of appetite. It has other health benefits such as antifungal, antioxidant, antibacterial, antiparasitic, and hypolipidemic effects. This plant contains different important components such as carbohydrates, glucosides, saponins and phenolic compounds (carvacrol), volatile oils (thymol), terpiene, paracymene and beta-pinene, protein, fat, fiber, and minerals including calcium, phosphorus, iron, and nicotinic acid (niacin). In the previous studies, several pharmacological effects were shown for C. copticum. Therefore, in this paper, the pharmacological effects of the plant were reviewed.

1. Introduction

C. copticum or Ajwain belongs to the Apiaceae plants family and its seeds are used extensively as a food additive in India and mainly therapeutically effective, with hot nature. C. copticum is an Egyptian aborigine plant. This plant grows in arid and semiarid fields in different regions of central Europe, Asia, India (most crops are in the states of Rajasthan, Gujarat, and West Bengal), Iran (especially eastern regions of Baluchistan), Iraq, Afghanistan, and Pakistan [1, 2].

In traditional medicine, different therapeutic applications for C. copticum have been described and in Persian traditional medicine it is used for thousands of years [3]. The bronchodilatory, antitussive, and antidyspnea effects were demonstrated for C. copticum [3]. The therapeutic effects of this plant in gastrointestinal disorders, such as reflux, cramps, abdominal tumors, abdominal pain, and Helicobacter pylori, as well as in eye infection disorders, have been demonstrated [3].

Therapeutic uses of C. copticum seeds also include carminative, antiseptic, amoebiasis expectorant, antimicrobial, antiparasitic, antiplatelet-aggregatory, and antilithiasis as well as treating common cold and acute pharyngitis [3]. Abortifacient, galactagogic, and diuretic activities have been observed for this plant [4, 5]. There is also anticarcinogenic potential evidence for C. copticum [6]. It has been shown that this plant has also foetotoxicity, abortion potential, and galactogogue properties [7].

In previous studies, different pharmacological effects were shown for C. copticum. In addition, the plant has been used widely in traditional medicine. Therefore, different pharmacological effects of C. copticum and its constituents were reviewed in the present paper.

2. Methods

The following databases and electronic journals were searched from September, 2012, to December, 2013, including Google Scholar, Pubmed, Wiley, Science Direct,
3. Phytochemistry

C. copticum is identified in different regions of the world by different names as follows.

Scientific name: Trachyspermum ammi and Sprague, it is synonym of Carum copticum Benth and in some documents Aromaticum has been named by different herbalists.

Different names of the plant in various languages (vernacular name) are Sanskrit: Yamini, Assamese language: Jain, English: Bishop’s weed, Hindi, Baluchi: Ajwain and Spica, Gujarati Language: Ajmo, Canada: Oma, Malaysia: Oman, Arabic: Khella or khellin, Persian: nankhah, zenian, khordaneh, and South Khorasan: aghio [8].

C. copticum is a perennial plant; its height is a little more than black cumin and about a meter, but the leaf shape and color of the flowers of the plant are similar to black cumin. Its stem is ramose; its leaves are slurred and filiform with small white flowers. The plant’s fruit which is called C. copticum is small, oval, and dark yellow and the fruit surface has five long thin lines of light yellow. Fruits and roots are highly regarded in traditional medicine.

4. Chemical Components

The constituents of the seed of C. copticum included carbohydrates (38.6%), fat (18.1%), protein (15.4%), fiber (11.9%), tannins, glycosides, moisture (8.9%), saponins, flavone, and mineral matter (71%) containing calcium, phosphorous, iron, cobalt, copper, iodine, manganese, thiamine, riboflavin, and nicotinic acid [3, 9]. C. copticum grows in different areas of the world containing different compounds. Main components of the oil of Iranian and African C. copticum oil are carvacrol, γ-terpinene, and p-cymene while thymol (97.9%) is the main component of south Indian plant oil. It was also reported that thymol (45.9%), γ-terpinene (20.6%), and o-cymene (19%) are the major components of the oil of C. copticum but ethylene methacrylate (6.9%), β-pinene (1.9%), and hexadecane (1.1%) were the other constituents of the plant [10]. Thymol (72.3%), terpinolene (13.12%), and o-cymene (11.97%) were also identified as constituents of C. copticum [11]. Chemical composition of C. copticum in two areas in Iran was assessed and results showed that the plant in Kamfiruz contains γ-terpinene (48.07%), p-cymene (33.73%), and thymol (17.41%) compared to the composition of plant in Eghlid area which included γ-terpinene (50.22%), p-cymene (31.90%), and nerolidol (4.26%) as main components [12].

Chemical constituents of the essential oil of C. copticum and its acetone extract were also examined by GC and GC-MS analysis. Results showed that 96.3% of the total amount of the essential oil contains 26 components including thymol (39.1%), p-cymene (30.8%), γ-terpinene (23.2%), β-pinene (1.7%), and terpinene-4-ol (0.8%) while 68.8% of the total amount of its acetone extract has thymol (39.1%), oleic acid (10.4%), linoleic acid (9.6%), γ-terpinene (2.6%), p-cymene (1.6%), palmitic acid (1.6%), and xylene (0.1%) [13]. Hydrodistillation and supercritical fluid (CO2) extraction (SFE) methods of the plant were also performed. In hydrodistilled oil, there were 8 components including thymol (49.0%), γ-terpinene (30.8%), p-cymene (15.7%), b-pinene (2.1%), myrcene (0.8%), and limonene (0.7%), but in SFE method with the best condition of temperature, pressure, and dynamic extraction time there were 3 components including γ-terpinene (14.2%), p-cymene (23.1%), and thymol (62.0%) [14].

According to the results of study of Srivastava et al., the main constituents of fruit oil of C. copticum were p-cymene (41.98%), carvacrol (45.20%), and thymol (0.48%) [15]. The content of chromone, an isomer of the coumarin which is a drug with anticoagulant performance, in various stages of growth of C. copticum was determined by high performance liquid chromatography (HPLC) and the results showed that the amount of chromone was higher in unripe than dried [16].

Chemical compounds of C. copticum seeds, cultivated in different studies using gas chromatography (GC) and gas chromatography mass spectrometry (GC-MS), are listed in Table I.

5. Pharmacological Effects

C. copticum has aromatic odor and spicy taste and is widely used as a spice in the curry powder (curry). The odor of the plant is due to thymol and its aromatic compounds are mainly obtained from methanol extract [19]. Several therapeutic effects were shown for C. copticum and its main constituents which were reviewed in the rest of this paper.

5.1. Respiratory Effects. One of the therapeutic effects of C. copticum is its effect on respiratory system. This plant is used as antiasthma and antidyspnea in traditional medicine. In this context, multiple studies have been carried out including relaxant and inhibitory effects on histamine receptors, stimulatory effect on adrenoreceptors of guinea pigs’ tracheal smooth muscles, antitussive effect in guinea pigs, and its bronchodilatory effect on airways of asthmatic patients.

C. copticum showed potent relaxant effect on tracheal smooth muscles which was not due to its content of thymol or competitive antagonistic effect on cholinergic receptors. The existence of α-pinene in essential oil of this plant showed anticholinergic activity (functional antagonism) [20]. Relaxant effects of different fractions from C. copticum including fractions 1, 2, 3, and 4 in guinea pigs’ tracheal smooth muscle were shown. For preparation of four fractions, the essential oil was freeze at 0°C overnight. The white crystals were collected by filtration, air dried, and subjected to NMR analysis. The filtrate (1 mL) was chromatographed on a silica gel (70–230 mesh). The column was eluted with solvent mixtures comprising petroleum ether (40–60°C) and chloroform with varying concentrations. Fractions (25 mL) were collected and
Table 1: Chemical composition of *C. copticum* based on geolocation or type of extraction.

<table>
<thead>
<tr>
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</thead>
<tbody>
<tr>
<td>Thymol</td>
<td>45.9</td>
<td>72.3%</td>
<td>17.41%, 50.22%</td>
<td>39.1%, 39.1%</td>
<td>54.50%</td>
<td>57.18</td>
</tr>
<tr>
<td>p-Cymene</td>
<td>—</td>
<td>—</td>
<td>33.73%, 31.90%</td>
<td>30.8%, 1.6%</td>
<td>19.38%</td>
<td>22.55</td>
</tr>
<tr>
<td>γ-Terpinene</td>
<td>20.6</td>
<td>—</td>
<td>48.07%</td>
<td>23.2%, 2.6%</td>
<td>22.96%</td>
<td>13.07</td>
</tr>
<tr>
<td>α-Cymene</td>
<td>19.0</td>
<td>11.97%</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Ethylene methacrylate</td>
<td>6.9</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>Hexadecane</td>
<td>1.1</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>β-Piynene</td>
<td>1.9</td>
<td>—</td>
<td>1.7%</td>
<td>0.78</td>
<td>0.43</td>
<td></td>
</tr>
<tr>
<td><em>cis</em>-Limonene oxide</td>
<td>0.7</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>β-Myrcene</td>
<td>0.7</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.34</td>
</tr>
<tr>
<td>Myrcene</td>
<td>0.7</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.48</td>
<td>—</td>
</tr>
<tr>
<td>α-Terpinene</td>
<td>0.4</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.29</td>
<td>0.31</td>
</tr>
<tr>
<td>α-Thujene</td>
<td>0.4</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.30</td>
<td></td>
</tr>
<tr>
<td>Carvacrol</td>
<td>0.2</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.46%</td>
<td>0.524</td>
</tr>
<tr>
<td>β-Phellandrene</td>
<td>0.4</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.541</td>
<td>—</td>
</tr>
<tr>
<td>α-Pinene</td>
<td>0.3</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.10</td>
<td>0.29</td>
</tr>
<tr>
<td>Sabinene</td>
<td>0.3</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.2</td>
<td></td>
</tr>
<tr>
<td>Limonene</td>
<td>0.2</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.2</td>
<td></td>
</tr>
<tr>
<td>Dodecane</td>
<td>0.2</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.2</td>
<td></td>
</tr>
<tr>
<td>4-Terpineol</td>
<td>0.1</td>
<td>—</td>
<td>0.8%</td>
<td>0.11</td>
<td>0.155</td>
<td></td>
</tr>
<tr>
<td>β-Fenchyl alcohol</td>
<td>0.1</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>0.11</td>
<td>0.155</td>
</tr>
<tr>
<td>Terpinolene</td>
<td>0.1</td>
<td>13.12%</td>
<td>—</td>
<td>—</td>
<td>0.095</td>
<td></td>
</tr>
<tr>
<td>Nerolidol</td>
<td>—</td>
<td>4.26%</td>
<td>—</td>
<td>—</td>
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</tr>
</tbody>
</table>

analogous fractions according to their TLC profile were mixed (solvent system comprising petroleum ether 40–60°C; chloroform (4:1) and the spots were visualized using sulfuric acid (50%, v/v). The relaxant effect of fraction 2 of the plant (suggested to be carvacrol) was comparable to the effect of theophylline and more potent than other fractions. Fraction 3 also showed a relaxant effect on tracheal smooth muscle to lesser extent. In addition the results showed that the relaxant effect of fractions 2 and 3 was not due to their inhibitory effect on muscarinic or stimulatory property on beta-adrenergic receptors [21].

Inhibitory effect of *C. copticum* on histamine (H1) receptors of isolated guinea pig tracheal smooth muscle showed a competitive antagonistic effect of the plant on H1 receptors; however, its effect was lower than chlorpheniramine [22].

Stimulatory effect of essential oil, aqueous, and ethanolic extract of *C. copticum* on beta 2 adrenoceptors was examined in isolated guinea pigs tracheal chain. The results showed a stimulatory effect only for ethanolic extract of *C. copticum* on beta 2 adrenoceptors [23]. A xanthine-like activity was also shown for the extract of *C. copticum* [24].

In the study of Gilani et al. bronchodilator effect of *C. copticum* seed extract in presence of high K+ (50 mM) and carbachol on guinea pig tracheal preparation was evaluated. Results demonstrated that *C. copticum* made dose-dependent relaxation (dose 0.1–1 mg/mL) with a possible mechanism of calcium channel blocking effect [25].

The antitussive effects of aerosols of two different concentrations of aqueous and macerated extracts, carvacrol, codeine, and saline were examined by enumerating the number of coughs due to citric acid aerosol 10 min after exposing animals to aerosols of different solutions. Results showed that antitussive effects of aqueous and macerated extracts were similar to codeine which is possibly due to its bronchodilatory properties. Nevertheless, carvacrol, one constituent of *C. copticum* with potent bronchodilatory effect, did not show any antitussive effect which suggested different afferent neural route between cough and bronchoconstriction [26].

Bronchodilatory effect of oral administration of boiled extract from *C. copticum* and theophylline in asthmatic patients was also examined. Different pulmonary function tests (FEV1, PEF, MMEF, MEF75, MEF50, MEF25, and sGaw) were measured 15 min after administration of different drugs and continued until 180 min after drug administration. The results showed that *C. copticum* has a relatively bronchodilatory effect on asthmatic airways which was comparable with the effect of theophylline at concentrations used [27]. The results of this study suggest that this plant could be of therapeutic value as a bronchodilatory drug in patients with obstructive airway diseases.

One of the main components of *C. copticum* is thymol. The effect of thymol on tracheal and ileum smooth muscles and ciliary motion of respiratory system in rat showed that thymol has a dose-dependent antispasmodic property and increases mucosa transfer due to ciliary motion [28]. Additionally, the antispasmodic effect of thyme extract was
Table 2: Respiratory effects of C. copticum and its constituents thymol and carvacrol.

<table>
<thead>
<tr>
<th>Effect</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Relaxant effect on tracheal smooth muscles</td>
<td>[20, 40]</td>
</tr>
<tr>
<td>Inhibitory effect of C. copticum on histamine (H1) receptors</td>
<td>[22]</td>
</tr>
<tr>
<td>Stimulatory effect on beta 2 adrenoceptors</td>
<td>[23]</td>
</tr>
<tr>
<td>Antitussive effects</td>
<td>[26]</td>
</tr>
<tr>
<td>Bronchodilatory effect</td>
<td>[20, 27]</td>
</tr>
<tr>
<td>Increase of mucosa transfer due to ciliary motion</td>
<td>[28]</td>
</tr>
<tr>
<td>Antispasmodic effect</td>
<td>[29]</td>
</tr>
<tr>
<td>Relaxant effect</td>
<td>[30, 31, 40]</td>
</tr>
<tr>
<td>Competitive antagonistic effect at histamine H1 receptors</td>
<td>[32]</td>
</tr>
<tr>
<td>Stimulatory effect on beta-2 adrenoceptor</td>
<td>[33]</td>
</tr>
<tr>
<td>Blocking effect at muscarinic receptors</td>
<td>[34]</td>
</tr>
<tr>
<td>Inhibitory effect on secretion of TNF-α and IL-1β in porcine alveolar macrophage</td>
<td>[35, 37]</td>
</tr>
<tr>
<td>Inhibitory effect on COX-1 and COX-2 and 5-lipoxygenase (anti-inflammatory effect)</td>
<td>[37]</td>
</tr>
</tbody>
</table>

Demonstrated which is suggested to be due to phenolic volatile oil compounds such as thymol [29].

The relaxant effect of carvacrol, one of the main constituents of C. copticum, on tracheal smooth muscle of guinea pigs has been shown which was greater than the effect of theophylline [30].

Other plants containing carvacrol such as Carum carvi [31] also showed relaxant effects on tracheal smooth muscle. Fraction 2 of C. copticum, which is suggested to be carvacrol, also revealed relaxant effect on tracheal smooth muscle [21]. Therefore, the main constituent of C. copticum, carvacrol, may have relaxant effects on the tracheal smooth muscle.

To examine the possible mechanism(s) responsible for the relaxant effect of carvacrol on tracheal smooth muscle, its effect on histamine receptors was evaluated in tracheal smooth muscle of guinea pigs by measuring EC$_{50}$ histamine (effective concentration of histamine causing 50% of maximum response) in the presence of carvacrol and chlorpheniramine. The results of this study showed a competitive antagonistic effect of carvacrol at histamine H1 receptors. In addition, the results suggested its stimulatory effect on β-adrenergic receptors and also a blocking effect at muscarinic receptors [32] for carvacrol. In fact, stimulatory effect of carvacrol on β2-adrenoceptors was proved by performing isoprenaline concentration response curve and measurement of EC$_{50}$ in the presence of the carvacrol, propranolol, and saline on tracheal smooth muscle of guinea pigs in nonincubated and incubated with chlorpheniramine (to block histamine H1 receptors) conditions. The results showed parallel leftward shift of isoprenaline concentration response curve and lower EC$_{50}$ in the presence of carvacrol and higher EC$_{50}$ in the presence of propranolol compared to the results of saline [33]. These results showed a clear β2-adrenoceptors stimulatory effect for carvacrol. In addition, the inhibitory effect of carvacrol on muscarinic receptors which is the other possible mechanism for its relaxant effect on the tracheal smooth muscle was also studied. The rightward shift in methacholine-response curves and the increased EC$_{50}$ in the presence of different concentrations of carvacrol compared with saline were seen which showed possible competitive antagonistic effects of carvacrol at muscarinic receptors [34]. These results suggest that the mechanism of relaxant effect of carvacrol similar to plant extract could have inhibitory effects on muscarinic and histamine receptors and stimulatory effect on β2-adrenoceptors or combinations of the three mechanisms.

However, carvacrol with a potent relaxant effect on tracheal smooth muscle shows no antitussive effect [26].

With regard to the lung inflammation in different respiratory diseases, mainly asthma, the anti-inflammatory and immunomodulatory effects of carvacrol were also examined in several studies. The effect of carvacrol on cell culture supernatants of macrophages in porcine induced alveolar inflammatory showed inhibitory effect of carvacrol on TNF-α, IL-1β, and TGF-β [35]. Carvacrol also inhibited secretion of TNF-α and IL-1β in porcine alveolar macrophage [36]. Anti-inflammatory effect of carvacrol was also evaluated by measurement of exudates volume and leukocyte migration in plural cavity due to carrageenan injection to this cavity which showed a preventive effect of carvacrol on exudates volume and leukocyte migration (in vivo and in vitro) and suggested an inhibitory effect on COX-1 and COX-2 and 5-lipoxygenase [37]. In addition carvacrol also depicted a preventive effect on serum levels of endothelin, total protein, histamine, NO, and total white blood cells, differential white blood cells (WBC) count and tracheal responsiveness in ovalbumin sensitized guinea pigs [38, 39]. Table 2 summarizes respiratory effects of C. copticum and its constituents thymol and carvacrol.

5.2. Cardiovascular Effect. Due to calcium channel blocking effect, C. copticum has remarkable role in heart rate and blood pressure. Thymol also made fall in blood pressure and heart rate [41]. Several cardiovascular effects of C. copticum and its constituents were shown. Negative inotropic and chronotrophic effects due to administration of 1–10 mg/kg thymol in mice were shown which lead to decrease in blood pressure. It was suggested that this effect of thymol could be due to calcium channel blocking property [25].

Kumar et al. examined the effect of juice of C. copticum leaves on isolated frog heart. It had positive ionotropic effect
and negative chronotropic effect on cardiac muscle perfused heart [42].

The cholinomimetic effects of aqueous extracts from C. coticum seeds on guinea pigs illume were shown [43], which could cause bradycardia. However, this effect of the plant is not supported by the results of more recent studies. In addition, in a pilot clinical trial, the impact of C. coticum on syndrome of cardiovascular disease (angina) was reported which showed that this plant can cause vasodilation of coronary arteries and decreased systemic blood pressure [44].

Lipid-lowering effect of C. coticum seeds has been studied in rabbit. In these studies, methanolic extract of the plant (2 g/kg) significantly decreased total cholesterol, triglycerides, and LDL-cholesterol (71%, 53%, and 63%, resp.) and increased HDL up to 60% which was comparable to the effect of simvastatin (0.6 mg/kg). It was also suggested that antilipidemic effect of the plant is possibly due to enhanced removal or catabolism of lipoproteins and inhibition of HMG COA reductase [45, 46]. In addition, it was shown that C. coticum seed powder was also effective in increasing secretion of lipase and amylase from pancreas gland in rat [47].

Rajput et al. administered extract of Ajwain with dose of 50 mg/kg and warfarin (0.54 mg/kg) orally to rats and measured coagulation parameters (PT and aPTT). On the 14th day, extract significantly increased PT time compared with warfarin but did not have effect on aPTT. They demonstrated its possible effects on the extrinsic pathway [48].

Administration of thymol orally twice daily (14 mg/kg) to high fat diet rats caused decremented effect on body weight gain and serum lipid peroxidation and increased antioxidant levels [49].

5.3. Urogenital Effects. In an in vivo study, the effect of the extract of C. coticum seeds on urinary stone of 350 patients was investigated. According to data of this study, Ca oxalate, Ca oxalate/uric acid, and Ca-oxalate/hydroxyapatite stones were treated by 100%, 53%, and 31.25%, respectively, with the extract [50]. Recently in India an anticalcifying protein from the seeds of C. coticum has been extracted and was administered in urolithic rat model. This protein inhibited calcium oxalate deposition by adhesion to calcium oxalate and prevented growth of stones in vitro and also in vivo [51]. However, other observations did not show any effect of this plant on the production of urea in 24 hours. The results showed that traditional use of C. coticum in the treatment of kidney stones was not statistically significant in laboratory setting [52].

C. coticum was tested for abortion in some states of India in 1987. The result of the study showed that C. coticum leads to abortion in 50 cases of 75 pregnant women and possibly has fetotoxicity feature. However, the possibility of congenital defect in this region of India increased during the study period. C. coticum dry seed has phytoestrogen content with 473 ppm value that can increase milk production [9].

5.4. Gastrointestinal Effects. Traditional use of the C. coticum seeds in many gastrointestinal diseases, including intestinal disorders, abdominal pain (colic), or diarrhea, is reported [56]. The alcoholic extract of the plant fruit showed significant reduction effect in ulcer index in an animal model of gastric ulcer [57]. In addition, the extract of crushed fruit from C. coticum was effective in relieving stomach pain but increased stomach acid secretion.

Aqueous extract of C. coticum (125, 250, and 500 mg/kg) treatment for two weeks improved peptic ulcer induced by ibuprofen in rats which was comparable with the effect of omeprazole. It was also suggested that antulcer effect of this plant is possibly due to its antioxidant effect [58].

C. coticum is able to increase the gastric acid secretion time and the amount of gastric acid. In addition, it was shown that the plant can reduce the transit time of food in the digestive system of mice [59]. Inhibitory effect of C. coticum on the contractions of the digestive tract smooth muscle, especially the intestines, increased activities of digestive enzymes and bile secretion was reported [60], which support its effect on gastrointestinal tract.
Table 4: The effects of *C. copticum* on gastrointestinal tract.

<table>
<thead>
<tr>
<th>Effect</th>
<th>Reference</th>
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<tbody>
<tr>
<td>Antiluerc effect</td>
<td>[58]</td>
</tr>
<tr>
<td>Increase of liver enzymes</td>
<td>[58]</td>
</tr>
<tr>
<td>Increasing of time and amount of gastric acid secretion</td>
<td>[64]</td>
</tr>
<tr>
<td>Inhibitory effect on the gastrointestinal contractions</td>
<td>[59, 60]</td>
</tr>
<tr>
<td>Hepatoprotective effects</td>
<td>[25]</td>
</tr>
<tr>
<td>Antispasmodic effects</td>
<td>[63]</td>
</tr>
<tr>
<td>Carvacrol</td>
<td>Apoptosis and antiproliferation effect on HepG2 cells of human hepatocellular carcinoma</td>
</tr>
</tbody>
</table>

In several studies, hepatic effects of *C. copticum* have been observed. The effect of 125, 250, and 500 mg/kg from *C. copticum* was assessed on peptic ulcer induced by ibuprofen in rat. In addition, the effect of the extract on liver enzymes including aspartate transferase (AST) and alanine transferase [7] in the serum was examined. Both high and low doses of the extract increase liver enzymes. Thus low dose of this plant is recommended for treatment of peptic ulcer and liver disorders [58].

In a study, the hepatoprotective effects of polyherbal formulations (containing several plants such as *C. copticum*) administered twice daily for one week after paracetamol (500 mg/kg) administration were evaluated on day 8. Paracetamol increases liver enzymes but treatment with polyherbal formulations improved the liver enzyme which was suggested to be due to cell membrane stabilization and recovery of hepatic tissue [61].

The effect of *C. copticum* on liver injury induced by CCL4 and lethal dose of paracetamol (1 g/kg) in mice was also examined. Oral administration of *C. copticum* reduced liver enzymes (ALT, ALP, and AST) and improved paracetamol- and CCL4-induced hepatic injuries [25]. On the other hand, carvacrol caused apoptosis and antiproliferation on HepG2 cells of human hepatocellular carcinoma. Carvacrol selectively decreases phosphorylation of ERK1/2 and activated phosphorylation of p38 but did not affect JNK MAPK phosphorylation. A significant reduction effect on Bcl-2 gene expression was also shown 24 h after carvacrol treatment. In addition, carvacrol inhibited DNA synthesis and decreased the number of cancer cells and total protein content [62].

The effect of *C. copticum* on isolated guinea pig ileum showed antispasmodic activity of extract of the plant and suggested that this effect may be due to cholinergic receptors inhibition by *C. copticum* [63]. Table 4 summarizes the effects of *C. copticum* on gastrointestinal tract.

5.5. Antiparasitic Effects. Infection with filarial nematodes makes lymphatic filariasis and synthetic drug not adequately effective in killing these parasites. Therefore, antifilarial effects of medicinal plant, namely, fruit extract of *C. copticum*, were shown in vitro and in vivo. *C. copticum*, thymol, and carvacrol have macrofilaricidal properties against adult bovine filarial worm *S. digitata in vitro*. In addition, the plant increased mortality and infertility of female worm of human filarial worm *Brugia malayi in vivo* [65]. The effect of *C. copticum* seeds on treatment of leishmaniasis parasitic was also reported. Hydroalcoholic extract of *C. copticum* showed antileishmanial activity with IC\(_{50}\) 15.625 \(\mu\)M which was less than IC\(_{50}\) for macrophage cell line (43.76 \(\mu\)M) [66].

Anthelmintic effect of *C. copticum* in comparison with levamisole (an anthelmintic and immunomodulator drug) on sheep infected with mixed nematode was also evaluated. *C. copticum* powder dose dependently caused reduction in eggs per gram of feces which was more potent compared with levamisole [67].

*Plasmodium falciparum* is genus of parasitic protozoa. Infection with this genus is known as malaria. Ethyl acetate extract of *C. copticum* seed with values of 25 \(\mu\)g/mL also showed *in vitro* antimalarial activity [68].

Pinewood nematode (PWN) makes pine wilt disease. Nematicidal activity of *C. copticum* oil against *B. xylophilus* was evaluated *in vitro* and mortality of nematodes after 24 h was studied. *C. copticum* and its components killed nematodes and likely are suitable as natural nematicides. It was also shown that thymol and carvacrol have a significant effect on nematodes [69, 70]. Considering that one of the most important worldwide parasitic diseases (especially in dirty and unsanitary areas) is hydatid cysts, it was shown that *C. copticum* play a significant role in the removal of hydatid cysts *in vitro*. In a study, Protoscoleces were exposed to essential oil of *C. copticum* (3, 5, and 10 mg/mL) for 10, 20, 30, and 60 min. The results showed that the higher concentration in the least time period of the study killed 100% of hydatid cyst protoscoleces which was suggested to be due to its phenol compounds [71]. *Coccidian protozoa* such as *Eimeria tenella* live in intestinal tract of animal and cause coccidiosis which in severe cases lead to death. Herbal complex (containing *C. copticum*) with three concentrations (2, 4, and 6 g) was added to water of broiler chickens infected with *Eimeria tenella* and symptoms were compared with amprolium group. This herbal complex in a concentration-dependently manner improved broiler chickens with *Eimeria tenella* [72].

In addition, there are several studies regarding the disinfesting and insecticide effects of *C. copticum* extracts, such as its effects on adult male and female German cockroaches by inhibition of acetylcholine esterase (AChE). In addition, *C. copticum* oil, 0.1 mg/mL, caused 100% larval mortality against *A. aegypti* mosquito larvae. Thus *C. copticum* can be used as botanical insecticides [73, 74]. The effect of thymol vapor on eggs laying of malaria mosquito (*Anopheles stephensi*) was more effective with LD\(_{50}\) 1.6-fold than *C. copticum* oil (80.77
versus 48.88 µg/mL [75]. Table 5 summarizes antiparasitic effects of C. copticum.

<table>
<thead>
<tr>
<th>Effect</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Macrofilaricidal properties (e.g., S. digitata), increase of mortality, and infertility of female worm (Brugia malayi)</td>
<td>[65]</td>
</tr>
<tr>
<td>Antileishmanial activity</td>
<td>[66]</td>
</tr>
<tr>
<td>Dysbosis treatment</td>
<td>[81]</td>
</tr>
<tr>
<td>Anthelmintic effect</td>
<td>[61]</td>
</tr>
<tr>
<td>Antimalarial activity</td>
<td>[68]</td>
</tr>
<tr>
<td>Nematicidal activity</td>
<td>[69, 70]</td>
</tr>
<tr>
<td>Killing of hydatid cyst protoscolices</td>
<td>[71]</td>
</tr>
<tr>
<td>Insecticidal activity</td>
<td>[73, 74]</td>
</tr>
</tbody>
</table>

5.6. The Antimicrobial Effects. Essential oil from Iranian C. copticum including 72.3% thymol inhibited gram-positive and gram-negative bacteria and viruses in which inhibition rate is associated with thymol content. High dose of thymol inhibits gram-positive more than gram-negative bacteria. It was shown that phenolic compounds interfere with cell membrane, change pH and ions homeostasis, and perhaps in this way act as antimicrobial agents. At all these studies the antimicrobial activity was examined by broth microdilution method [10, 12, 76, 77].

The effect of aqueous extract of C. copticum on several strains of bacteria showed antibacterial effect on Enterococcus faecalis, Staphylococcus aureus, Escherichia coli, P. aeruginosa, S. typhimurium, and Shigella flexneri [78]. The effect of C. copticum on fifty-five bacterial strains showed antimicrobial activity with minimum inhibitory concentration <2% (v/v) except Pseudomonas aeruginosa [79]. It was also shown that ether fraction of C. copticum had better antibacterial and antifungal activity against multidrug resistant (MDR) strains of Candida albicans, Candida krusei, Candida tropicalis, Candida glabrata, Escherichia coli, and reference strains of Streptococcus mutans and Streptococcus bovis than other fractions [80].

Dysbiosis disease occurs due to microbial imbalance in intestinal flora as lactobacilli, bifidobacteria, and coliform bacteria which are lower in fecal counts. In this disease, useful bacteria decreased and harmful bacteria increased in intestinal flora which leads to reduction in energy and body weight. It was shown that C. copticum can lead to reduction in pathogenic microorganisms such as Candida albicans, Clostridium spp., and Bacteroides fragilis while having little effect on microflora and therefore could be effective in dysbiosis treatment [81].

The effect of C. copticum with thymol chemotype (when main component is thymol in contrast carvacrol chemotype) on bacterial strains (S. aureus, B. cereus, L. monocytogenes, E. coli O157:H7, and S. enteritidis) was also evaluated. Bacteria were cultured overnight at 37°C, and the essential oil of the plant and antimicrobial standards (chloramphenicol and ascorbic acid) were added. After incubation at 37°C for 22–24h, the MIC (mg/mL) was calculated and the microorganism growth inhibition was assayed using an ELISA reader. The results of this study showed that the antimicrobial of C. copticum was more potent than B. persicum and C. cyminum [82].

The antimicrobial effects of C. copticum as MIC and MBC were shown in Table 6.

The overall results of these studies showed that C. copticum essential oil is rich in monoterpenes compounds and could be used as a natural antimicrobial agent in the food and pharmaceutical industries.

Regarding ophthalmic disorders and cataract, it was claimed that the herbal ophthalmic drops (Ophthacare), which is a C. copticum extract product, treat infection, inflammation, and cataract in an experimental study [83]. C. copticum is also able to protect food against microbial invasion in vitro. These antimicrobial properties of C. copticum are due to its two ingredients, thymol and carvacrol [84]. Thymol has microbial killing property against common resistant microbial pathogens to multiple antibiotics drugs from the third generation. Therefore, it can be named as the fourth generation plant antibiotic [85].

Gilani et al. studied antibacterial effect of C. copticum by applying cream containing 5% essential oil of C. copticum to healing wound in rabbits in comparison with iodine tincture. Wound contraction on the 15th day in C. copticum group was 99.68%, compared with the healing effect of iodine tincture group, 100%, and nontreatment group, 96.57%, which indicates a wound healing effect of C. copticum [86].

In a study, bactericidal properties of C. copticum were shown on gram-negative Erwinia carotovora in vitro which is suggested to be due to its phenolic compounds such as thymol and carvacrol [87].

5.7. Antifungal Effects. Antifungal activity of essential oil of C. copticum seeds is also documented against toxigenic Aspergillus species. The oil of this plant also is able to inhibit the growth of this parasite [88]. In another study, C. copticum (900 ppm concentration) showed fungitoxicity activity
Table 6: The antibacterial activity of C. copticum [82].

<table>
<thead>
<tr>
<th>Species of bacteria</th>
<th>G MIC (mg/mL)</th>
<th>MBC (mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. aureus</td>
<td>+ 0.06</td>
<td>0.06</td>
</tr>
<tr>
<td>B. cereus</td>
<td>+ 0.03</td>
<td>0.03</td>
</tr>
<tr>
<td>L. monocytogenes</td>
<td>+ 0.25</td>
<td>0.25</td>
</tr>
<tr>
<td>E. coli O157:H7</td>
<td>– 0.25</td>
<td>0.5</td>
</tr>
<tr>
<td>S. enteritidis</td>
<td>– 0.5</td>
<td>1.0</td>
</tr>
</tbody>
</table>

G (+): gram-positive, G (−): gram-negative. Minimum bactericidal concentration = MBC.

Minimum inhibitory concentration = MIC.

against Epidermophyton floccosum, Microsporum canis, and Trichophyton mentagrophytes [89].

Anti-Candida activity of 10–55 μL concentrations of C. copticum was assessed by agar disc diffusion assay. The concentrations 50–55 μL of C. copticum were more effective in inhibition of the growth of Candida [90]. The effect of methanolic extract of C. copticum on Saccharomyces cerevisiae, Candida albicans, and C. utilis was studied in vitro which has more effect on Candida albicans and C. utilis [91].

5.8. Antitoxic Effects. Aflatoxins are mycotoxins that are produced by Aspergillus flavus and Aspergillus parasiticus, species of fungi, which infect crops such as corn and rice; C. copticum seed extract is shown to have destructive effect on aflatoxin G1 (AFG1) and significantly reduced aflatoxin activities down to 65%. In addition, C. copticum seed extract makes significant damage in other types of aflatoxins (AFB1, AFB2, and AFG2). Damage of aflatoxin G1 caused by C. copticum extract was more than 98% in 24 hours, and during 6 hours their destruction has been reported to be over than 78% [92]. The effect of C. copticum extract as inhibitors of chromium toxicity has also been shown, as C. copticum extract can increase cell viability and decrease DNA damage by reduction of caspase-3 and apoptosis, increase of the mitochondrial membrane potential, and reduction in reactive oxygen species [93].

5.9. Neural Effects. C. copticum has been used in traditional medicine for relieving rheumatic, joint, headache, and neuralgic pain. Dashti-Rahmatabadi et al. demonstrated that analgesic effect of ethanolic extract of C. copticum is comparable with morphine and this effect is suggested to be due to its parasympathomimetic through descending pain modulating pathways [94]. Analgesic effect of C. copticum essential oil in formalin test was also assessed and pain scores were recorded during one hour (every 5 minutes). Results showed that essential oil affected the late phase of pain by formalin compared to morphine. The mechanism of this effect of the plant was not due to opioid receptors because it was not reversed by naloxone [95]. Study of Ghannadi et al. on morphine withdrawal syndrome in mice showed that C. copticum leads to suppression of morphine withdrawal. It was suggested that this effect was modulated via potentiation of GABA neurotransmission and suppression of glutamate receptors and nitric oxide pathway [96].

Antiepileptic and sedative effects of C. copticum in PTZ and amygdala kindling models and its depressant effect in open field test in male rats were also demonstrated and suggested to be due to increase in GABAergic neurotransmission in the brain which reduces neural activity [97].

5.10. Dose and Administration Rote. Three to six grams of the seed powder with food or by means of other ways can be consumed daily. Although the seeds are small, they should be powdered for more effectiveness. In addition, it may be extracted or boiled and used. Dried extract of C. copticum seeds can be consumed up to 125 mg daily. The liquid extract (tincture) can be also consumed up to 6 mL daily.

5.11. Conclusion. C. copticum or Ajwain belongs to the Apiaceae plants family and its most important constituents are thymol and carvacrol. C. copticum seeds have various important medicinal properties such as antipyretic, antitussive, antispasmodic and cardiovasodilator, respiratory, liver protection, urogenital, gastrointestinal, antiparasitic, antimicrobial, and lipid lowering effects. Therefore this plant could be of therapeutic value in treating of various disorders. Therefore, further clinical studies regarding various effects of C. copticum and its main constituents are recommended. If significant clinical results were found, proper industrial drug products need to be prepared for clinical use.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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