Research Article

Genotypic Diversity of *Mycobacterium tuberculosis* Clinical Isolates in the Multiethnic Area of the Xinjiang Uygur Autonomous Region in China

Jie Liu,1,2,3 Junlian Li,4 Jiao Liu,1,2 Xiuqin Zhao,1,2 Lulu Lian,1,2 Haican Liu,1,2 Bing Lu,1,2 Qin Yu,1,2 Jingrui Zhang,1,2 Yingcheng Qi,4 and Kanglin Wan1,2

1 National Institute for Communicable Disease Control and Prevention, Chinese Center for Disease Control and Prevention, State Key Laboratory for Infectious Disease Prevention and Control, Beijing 102206, China
2 Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, Hangzhou 310003, China
3 Chaoyang Center for Disease Control and Prevention, Beijing, 100021, China
4 The Chest Hospital of Xinjiang Uygur Autonomous Region, Ürümqi 830001, China

Correspondence should be addressed to Kanglin Wan; wankanglin@chinatb.org

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**Objectives.** We studied the genetic diversity of clinical isolates from patients with tuberculosis in the multiethnic area of Xinjiang autonomous region in China. A total of 311 clinical *M. tuberculosis* isolates were collected in 2006 and 2011 and genotyped by two genotyping methods. All isolates were grouped into 68 distinct spoligotypes using the spoligotyping method. The Beijing family was dominant, followed by T1 and CAS. MIRU-VNTR results showed that a total of 195 different VNTR types were identified. Ten of the 15 loci were highly or moderately discriminant according to their HGDI scores, and 13 loci had good discriminatory power in non-Beijing family strains, whereas only two loci had good discriminatory power in Beijing family strains. Chi-square tests demonstrated that there were no correlations between four characteristics (sex, age, type of case, and treatment history) and the Beijing family. In summary, Beijing family strains were predominant in Xinjiang, and the VNTR-15 locus-set was suitable for genotyping all Xinjiang strains, but not for the Beijing family strains. Thus, these data suggested that different genotype distributions may exist in different regions; MLVA locus-sets should be adjusted accordingly, with newly added loci to increase resolution if necessary.

1. Introduction

Tuberculosis (TB) is a severe chronic infectious disease caused by *Mycobacterium tuberculosis* (*M. Tuberculosis*), which remains prevalent despite intense global efforts to control and eliminate this disease. In 2014, 9.6 million people were estimated to have contracted TB, and 1.4 million TB-related deaths occurred [1]. Of the 22 countries accounting for 79% of the world’s burden of TB, China is ranked second and has the highest absolute number of cases annually worldwide [2].

The Beijing family genotype of *M. tuberculosis* was first described in 1995 by Van Soolingen, and 86% of isolates from Beijing, China, were found to have this Beijing family genotype [3]. However, the distribution of *M. tuberculosis* and the proportions of Beijing family isolates in Xinjiang are unclear.

The Xinjiang Uygur autonomous region is located in northwestern China, surrounded by India, Russia, Pakistan, Mongolia, and other countries, and covers one-sixth of the land area of China (a total of 1.66 million km²). Thus, this province is the largest province in China and has the longest land borderline with neighboring countries. In 2010, the population of this region was 22 million, and the region was multiethnic, comprised of Uygur, Kazak, Hui, and other ethnic minorities. The minority population in this region accounts for approximately 60.5% of the population. The Xinjiang autonomous region has a high TB burden and TB
prevention and control measures are needed owing to the unique geographical location and complex ethnic composition of this region.

In this study, we used spacer-oligonucleotide typing (spoligotyping) and multiple locus variable number tandem repeat (VNTR) analysis (MLVA), which both employ polymerase chain reaction-(PCR-) based genotyping technology, to characterize M. tuberculosis genotypes circulating in the Xinjiang Uygur autonomous region and to explore whether there were relationships between the spread of Beijing family strains and patient characteristics, including sex, age, type of case, and treatment history. In addition, we evaluated the discriminatory power of 15-loci-set MLVA (VNTR-VNTR, and spacers from 35 to 43) to characterize the strains from the Xinjiang Uygur autonomous region.

2. Materials and Methods

2.1. M. tuberculosis Strains. A total of 311 M. tuberculosis isolates were obtained from sputum samples collected from patients with pulmonary TB in 2006 and 2011 at the Provincial Tuberculosis Hospital of Xinjiang. All patients with TB were diagnosed based on the national guidelines of China. Demographic, epidemiological, and clinical data were obtained from the medical records of all patients; these data included sex, age, present address, diagnosis results, drug susceptibility test results, previous TB history, symptoms, and associated medical data from local doctors working in the hospital using uniform epidemiological investigation methods. For the 311 M. tuberculosis strains, detailed information is provided in Table 1.

2.2. DNA Sample Preparation. Bacteria were isolated and inoculated on Löwenstein-Jensen (L-J) medium. Culture was performed for all samples and the bacteria were kept at the National Laboratory of TB, ICDC, China CDC, Beijing, China. Mycobacterial genomic DNA was extracted from mycobacterial colonies grown on L-J medium by resuspending one loopful of mycobacterial colonies in 200μL TE buffer (10 mM Tris-HCl, 1 mM ethylenediaminetetraacetic acid [EDTA]) and was incubated at 85°C for 30 min. The supernatant containing the DNA was then collected by centrifugation at 12,000 rpm for 5 min and stored at −20°C for further use.

2.3. Spoligotyping. Spoligotyping involves PCR-based amplification of the whole CRISPR region with the primers DRa and DRb (DRa: 5’-GGT TTT GGG TCT GAC GAC-3’, DRb: 5’-CGG AGA GGG GAC GGA AAC-3’), followed by hybridization of the amplified DNA to a set of 43 spacer-oligonucleotides probes corresponding to each spacer, covalently linked to a membrane. Because clinical isolates vary in the nature of spacer sequences, the spoligotype patterns obtained were strain specific. Detailed procedures were described previously [5]. The results were analyzed using BioNumerics software (version 5.0). The Beijing genotype here was any isolate missing spacers 1 to 34, with at least three spacers from 35 to 43.

2.4. MLVA Typing. In addition to spoligotyping, the MLVA typing method based on VNTR-15\(\text{China}\) was also carried out, described by Wan et al. [7], for most of the isolates collected from more than 14 provinces in China [4, 7, 8]. The discrimination of the locus combination was calculated using the Hunter-Gaston discriminatory index (HDGI), calculated using the following formula [9]:

\[
\text{HDGI} = 1 - \frac{1}{N(N-1)} \sum_{j=1}^{s} n_j (n_j - 1),
\]

where \(N\) is the total number of isolates in the typing method, \(s\) is the number of distinct patterns discriminated by MIRU-VNTR, and \(n_j\) is the number of isolates belonging to the \(j\)th pattern.

2.5. Data Analysis. Genotype results were entered in binary format into a Microsoft Excel spreadsheet, as shown in Table S1. The patterns were established based on clusters generated in BioNumerics software version 5.0 (Applied Maths, Sint-Martens-Latem, Belgium). Spoligotypes were designated according to the updated version of the international spoligotype database SITVIT2 (http://www.pasteur-guadeloupe.fr:8081/SITVITDemo). Statistical data were analyzed using the Chi-square test. All statistical tests were two-sided, and differences with \(P\) values of less than 0.05 were considered significant. Statistical analyses were carried out using SPSS software 19.0 (SPSS Inc., Chicago, IL, USA).

3. Results

We collected 311 isolates from patients clinically diagnosed with pulmonary TB in this study in 2006 and 2011. Patient demographics are shown in Table 1.

3.1. Spoligotyping Analysis. Spoligotyping results showed that the 311 isolates in this study could be grouped into 68 distinct spoligotypes; 54 strains represented a single isolate, whereas the other 257 isolates were grouped into 14 clusters containing from two to 212 isolates, with a cluster rate of 78.14%. According to SITVIT2, 271 (87.13%) strains were classified into 29 shared international types (SITs), and 40 (12.87%) strains were found to not have SIT number. In the seven families (Beijing, T, Haarlem, CAS, LAM9, MANU2, and U), MANU2 and LAM9 had only one isolate, and the other families had two or more strains. The Beijing family was the dominant genotype, with 224 isolates (72.03%), followed by T (20, 6.43%) and CAS (12, 3.86%; Table 2).

A total of 311 strains were collected in the Xinjiang area; 171 strains were collected in 2006, and 140 strains were collected in 2011. The genotype distribution of strains in different years is shown in Table 3. Compared with that in 2006, the proportions of strains belonging to the Beijing and T families increased in 2011 and the proportion of new genotypes decreased.

3.2. MLVA. A total of 195 different VNTR types were identified among the 311 strains. One hundred fifty-four (49.52%)
strains were unique, and 157 (50.48%) strains could be grouped into 41 clusters, containing from two to 31 strains, with a cluster rate of 37.30%.

Next, we evaluated the discriminating ability of MIRU-VNTR loci based on HGDI scores classified as high (>0.6), moderate (0.3 to 0.6), and poor (<0.3) [10]. For the genotyping of all 311 strains, HGDI scores were calculated from 0.778 to 0.069 among IS MIRU-VNTR loci. MIRU26 (0.778) and Mtb21 (0.691) had high discriminating ability. Additionally, four loci (ETRB, ETRC, ETRD, and MIRU27) had poor discrimination ability (HGDI < 0.3), and MIRU23 (0.069) showed almost negligible diversity (HGDI < 0.1; Table 4). Thirteen loci had good discriminatory power in non-Beijing family strains, whereas only two loci had good discriminatory power in Beijing family strains.

3.3. Comparisons between Spoligotyping and 15-Loci MLVA. As shown in Figure S1 (in Supplementary Material available online at https://doi.org/10.1155/2017/3179535), there was good agreement between the two methods; only two non-Beijing family strains were clustered with Beijing family strains when we used 15-loci-set MLVA. This may be due to the presence of two independent strains in some clinical samples. Eight spoligotype variants of the Beijing family were detected in the 224 Beijing family strains, and the HGDI score was 0.103. Moreover, these strains were distributed into 119 genotypes by MLVA, with an HGDI score of 0.986 confirming that VNTR-15China MLVA was more suitable for typing Beijing strains than spoligotyping.

In 224 Beijing family strains, 89 (39.73%) strains were unique, and 135 (60.27%) strains could be grouped into 33 clusters, with a cluster rate of 45.53%. Sixty-five (74.71%) strains were unique in a total of 87 non-Beijing family strains, and 22 (25.29%) strains could be grouped into eight clusters, with a cluster rate of 16.09%.

3.4. Relationship between Beijing Family Genotypes and Strain Characteristics. Four factors associated with TB, that is, sex, age, case type, and treatment history, were included in this study. We found that there were no correlations between Beijing family genotypes and the four factors associated with TB ($P > 0.05$ for all; Table 5).

4. Discussion

Efficient disease control can be achieved using epidemiological surveillance systems to accurately monitor epidemic trends at the regional and global levels [11]. Genotyping of $M.\ tuberculosis$ plays an important role in epidemiological studies [12], and genetic analyses have suggested that $M.\ tuberculosis$ exhibits substantial genetic variations [13], such as large sequence polymorphisms (LSPs) [14], single-nucleotide polymorphisms (SNPs), variable numbers and locations of insertion element (IS) 6110 [15], and VNTRs [16], all of which have been commonly employed in molecular epidemiology. However, because a single genotyping method cannot define all unique isolates, the current studies undertaken require various strategies to increase the power of strain differentiation [17, 18]. In this study, by the comparison of MLVA and spoligotyping methods, we confirmed that the VNTR-15China loci set was suitable for typing strains in China. Spoligotyping results showed that the cluster rate was 78.14%. In the MLVA results, we found that patients infected with a Beijing family strain were more likely to be clustered
than patients who were infected with a non-Beijing family strain. The MLVA-VNTR cluster case, which was produced by the same source of infection spread, had the same genotype in the short term. Moreover, in this study, the cluster rate of the MLVA-VNTR was 37.30%, indicating that 37.30% of patients may have acquired infections from the recent spread. However, this analysis was likely to have overestimated the recent spread. A study showed that, during the course of evolution, some strains at great distances may form the same VNTR genotype [19]. This is a limitation of this method, which we called VNTR homoplasy [20]. The genetic distance of Beijing family strains was relatively close; thus, it may be more likely for these strains to form the same VNTR genotype during evolution. In addition, Beijing family strains are quite abundant, leading to a higher cluster rate.

*Note:* As studies showed, during the course of evolution, some strains at great distances may form the same VNTR genotype [19]. This is a limitation of this method, which we called VNTR homoplasy [20]. The genetic distance of Beijing family strains was relatively close; thus, it may be more likely for these strains to form the same VNTR genotype during evolution. In addition, Beijing family strains are quite abundant, leading to a higher cluster rate.

**M. tuberculosis** Beijing family strains are the most prevalent strains in China [8]. In this study, according to spoligotyping results, the Beijing genotype was also predominant in the Xinjiang region; however, the proportion of the Beijing genotype in Xinjiang, which is located in northwestern China, was lower than that in other provinces in northern China [8, 21–24], but higher than those of the areas in southern China [25–30]. These results could be explained by the particular features of the regions, including geographic, climatic, or ethnic differences [8]. Beijing family strains were also found to be dominant in some East Asian countries,

### Table 2: Spoligotypes of prevalent clades by SITVIT2 in this study.

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<td>56</td>
<td>U</td>
<td>2</td>
</tr>
</tbody>
</table>

*a* Presence of spacer (◼); absence of spacer (◻).

*b* Representing spoligotype families as assigned in SITVIT2.

*c* N: number of strains.
such as South Korea (97.1%) [31], Thailand (44%) [32], and
Vietnam (53%) [33, 34]. Moreover, within the past decade,
the molecular epidemiological data from some areas have
revealed that the Beijing family genotype is widespread
around the world [35].

In our study, in addition to Beijing family strains, we also
detected strains belonging to other families, such as T1, U,
H4, T2, MANU2, and LAM9. One interesting finding in this
study was that 12 (3.86%) strains tested belonged to the CAS
family, which has only been found in Tibet [23] and Xinjiang
in China. All CAS family strains originated from patients
of Tibetan and Uygur ethnicity before 2010. This finding
suggested that the CAS family may have associated with the
patient’s ethnic groups and regional distribution. Moreover,
Tibet and Xinjiang share geographic borders with India,
where the CAS family is dominant [36]. This family of strains
may also be transported by trade, tourism, or migration
from India. When we first found CAS family strains in 2006,
all strains were isolated from Uygur individuals. Now, we
detected one CAS strain from a Han ethnic patient in 2011
and found CAS strains isolated from a Han ethnic patient
in Gansu, 2011 [4]. These findings suggested that CAS family
strains have the potential to spread inland. The LAM family
was also found in Jiangsu province and Taiwan in China
[26, 30], although this family of strains is predominantly
prevalent in South America [37] and West Africa [38].

The spoligotyping typing method has the advantage of
identification of Beijing family strains, but with lower ability
to distinguish among strains in comprehensive analysis. Thus,
we performed spoligotyping in combination with VNTR-
M. tuberculosis Beijing family genotype, particularly the high
infection rate in China, we aimed to determine the cause of
this problem. We attempted to combine Beijing family strains
with demographic data in order to investigate the correla-
tions between the Beijing family genotype and the general
characteristics of patients with TB. The results showed that
there were no significant correlations of sex, age, treatment
history, and case type with the distribution of Beijing family
strains; thus, we can speculate that these four factors were
likely not correlated with the prevalence of Beijing family
strains. These results were similar to those of previous studies
[16, 40, 41]. Therefore, we suggest that there may be other
factors promoting the transmission of the M. tuberculosis
Beijing family. Some researchers speculated that the long-
term M. bovis BCG vaccine may be one of the selective forces
implicated in the successful spread of the Beijing genotype
[42, 43] and that drug resistance (particularly multiple drug
resistance) may be a factor enhancing the spread of this family
[44]. These two hypotheses are still controversial because they
have not been investigated sufficiently. Thus, although other
factors may also promote the spread of the Beijing family
genotype of M. tuberculosis, additional studies are required
to confirm this assertion.

In summary, this is the first report applying spoligo-
typing in combination with VNTR-15_China loci-set MLVA
technology for genotyping of TB strains in the Xinjiang
autonomous region of China. Our results showed that Beijing
family strains were predominant in Xinjiang and that the
VNTR-15_China loci-set was suitable for genotyping all Xin-
jiang strains, but not all Beijing family strains. Thus, these
data suggested that different genotype distributions may exist
in different regions; MLVA loci sets should be adjusted
accordingly with newly added loci to increase resolution if
necessary.

**Disclosure**

The funders had no role in the study design, data collection
and analysis, decision to publish, or preparation of the
manuscript.

**Competing Interests**

The authors declare that there are no competing interests
regarding the publication of this manuscript.
Table 4: HGDI scores of the different MIRU-VNTR loci.

<table>
<thead>
<tr>
<th>MIRU/VNTR</th>
<th>All strains</th>
<th>Beijing family strains</th>
<th>Non-Beijing family strains</th>
</tr>
</thead>
<tbody>
<tr>
<td>MIRU26</td>
<td>0.778</td>
<td>0.717</td>
<td>0.766</td>
</tr>
<tr>
<td>Mtub21</td>
<td>0.691</td>
<td>0.526</td>
<td>0.795</td>
</tr>
<tr>
<td>MIRU10</td>
<td>0.498</td>
<td>0.217</td>
<td>0.746</td>
</tr>
<tr>
<td>ETRE</td>
<td>0.469</td>
<td>0.187</td>
<td>0.500</td>
</tr>
<tr>
<td>MIRU40</td>
<td>0.415</td>
<td>0.217</td>
<td>0.711</td>
</tr>
<tr>
<td>ETRA</td>
<td>0.405</td>
<td>0.205</td>
<td>0.615</td>
</tr>
<tr>
<td>MIRU16</td>
<td>0.402</td>
<td>0.207</td>
<td>0.703</td>
</tr>
<tr>
<td>Mtub30</td>
<td>0.400</td>
<td>0.062</td>
<td>0.408</td>
</tr>
<tr>
<td>MIRU39</td>
<td>0.385</td>
<td>0.096</td>
<td>0.464</td>
</tr>
<tr>
<td>Mtub39</td>
<td>0.306</td>
<td>0.139</td>
<td>0.618</td>
</tr>
<tr>
<td>ETRC</td>
<td>0.243</td>
<td>0.130</td>
<td>0.435</td>
</tr>
<tr>
<td>ETRB</td>
<td>0.187</td>
<td>0.027</td>
<td>0.443</td>
</tr>
<tr>
<td>ETRD</td>
<td>0.129</td>
<td>0.045</td>
<td>0.315</td>
</tr>
<tr>
<td>MIRU27</td>
<td>0.122</td>
<td>0.045</td>
<td>0.288</td>
</tr>
<tr>
<td>MIRU23</td>
<td>0.069</td>
<td>0.045</td>
<td>0.127</td>
</tr>
</tbody>
</table>

Table 5: Statistical analysis of the relationship between M. tuberculosis Beijing family and the factors associated with TB.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Total</th>
<th>Beijing family (%)</th>
<th>Non-Beijing family</th>
<th>$\chi^2$</th>
<th>$P$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>174</td>
<td>123 (70.69)</td>
<td>51</td>
<td>0.070</td>
<td>0.791</td>
</tr>
<tr>
<td>Female</td>
<td>136</td>
<td>98 (72.06)</td>
<td>38</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age group</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$\leq 20$</td>
<td>23</td>
<td>19 (82.61)</td>
<td>4</td>
<td>3.165</td>
<td>0.367</td>
</tr>
<tr>
<td>$\sim 20$</td>
<td>134</td>
<td>97 (72.39)</td>
<td>37</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$\sim 40$</td>
<td>81</td>
<td>58 (71.60)</td>
<td>23</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$\geq 60$</td>
<td>67</td>
<td>43 (64.18)</td>
<td>24</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Type of case</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Relapse</td>
<td>67</td>
<td>51 (76.12)</td>
<td>16</td>
<td>3.412</td>
<td>0.065</td>
</tr>
<tr>
<td>New case</td>
<td>133</td>
<td>84 (63.16)</td>
<td>49</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treatment history</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>93</td>
<td>62 (66.67)</td>
<td>31</td>
<td>0.045</td>
<td>0.832</td>
</tr>
<tr>
<td>No</td>
<td>91</td>
<td>62 (68.13)</td>
<td>29</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Authors’ Contributions

Jie Liu, Junlian Li, and Jiao Liu contributed equally to this paper. Kanglin Wan, and Yingcheng Qi conceived and designed the experiments. Jie Liu, Jiao Liu, Junlian Li, Lulu Lian, Xiuqin Zhao, Qin Yu, and Jingrui Zhang performed the experiments. Kanglin Wan, Jie Liu, Jiao Liu, Haican Liu, and Bing Lu analyzed the data. Kanglin Wan and Yingcheng Qi contributed reagents/materials/analysis tools. Kanglin Wan, Jie Liu, and Jiao Liu wrote the manuscript.

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References


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