Research Article

Cytoplasmic Localization of WT1 and Decrease of miRNA-16-1 in Nephrotic Syndrome

Pablo Zapata-Benavides,¹ Mariela Arellano-Rodríguez,¹,² Juan José Bollain-y-Goytia,² Moisés Armides Franco-Molina,¹ Gloria Azucena Rangel-Ochoa,³ Esperanza Avalos-Díaz,² Rafael Herrera-Esparza,² and Cristina Rodríguez-Padilla¹

¹Departamento de Microbiología e Inmunología, Facultad de Ciencias Biológicas, Universidad Autónoma de Nuevo León (UANL), 66450 San Nicolás de los Garza, NL, Mexico
²Laboratorios de Inmunología y Biología Molecular, Unidad Académica de Ciencias Biológicas, Universidad Autónoma de Zacatecas (UAZ), 98040 Zacatecas, ZAC, Mexico
³Hospital General Zacatecas “Luz González Cosío”, Ciudad Gobierno, 98160 Zacatecas, ZAC, Mexico

Correspondence should be addressed to Pablo Zapata-Benavides; pablozapata@hotmail.com

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Nephrotic syndrome (NS) is a glomerular disease that is defined by the leakage of protein into the urine and is associated with hypoalbuminemia, hyperlipidemia, and edema. Steroid-resistant NS (SRNS) patients do not respond to treatment with corticosteroids and show decreased Wilms tumor 1 (WT1) expression in podocytes. Downregulation of WT1 has been shown to be affected by certain microRNAs (miRNAs). Twenty-one patients with idiopathic NS (68.75% were SSNS and 31.25% SRNS) and 10 healthy controls were enrolled in the study. Podocyte number and WT1 location were determined in the study. Podocyte number and WT1 location were determined by immunofluorescence, and the serum levels of miR-15a, miR-16-1, and miR-193a were quantified by RT-qPCR. Low expression and delocalization of WT1 protein from the nucleus to the cytoplasm were found in kidney biopsies of patients with SRNS and both nuclear and cytoplasmic localization were found in steroid-sensitive NS (SSNS) patients. In sera from NS patients, low expression levels of miR-15a and miR-16-1 were found compared with healthy controls, but only the miR-16-1 expression levels showed statistically significant decrease (p = 0.019). The miR-193a expression levels only slightly increased in NS patients. We concluded that low expression and delocalization from the WT1 protein in NS patients contribute to loss of podocytes while modulation from WT1 protein is not associated with the miRNAs analyzed in sera from the patients.

1. Introduction

Nephrotic syndrome (NS) is one of the most common diseases in children and is characterized by the leakage of large amounts of protein into the urine, lipidemia, hypoalbuminemia, and dysfunction of the glomerular filtration barrier [1]; NS is also a major cause of podocyte injury [2]. Podocytes, highly specialized cells of the visceral epithelium, are found in the glomerular membrane of the kidney [3]; they constitute the terminal part of the ultrafiltration barrier that prevents protein loss, which can result in damage and detachment of podocytes. The reduction of glomerular podocytes in lupus nephritis (LN) patients is correlated with the cumulative loss of urinary podocytes and proteinuria during the progression of kidney disease [2, 4].

Some urinary biomarkers have been found to be associated with injured podocytes in urine sediment, such as podocalyxin (PODXL), nephrin (NPHSI), podocin, and Wilms tumor protein (WT1) [5]. WT1 itself has been found in exosomes in urine sediment, indicating podocyte injury [6].

WT1 is essential for urogenital development, and its expression is limited to mature podocytes [7]. The WT1 gene encodes for a transcription factor that is critical for the development of podocytes and viability; the regulation of podocyte homeostasis occurs via PODXL and NPHSI [7].
Recent studies have demonstrated that mutations in WT1 can lead to syndromic forms of steroid-resistant NS (SRNS), such as Denys-Drash or Frasier syndrome, and can cause isolated SRNS [8, 9]. Between 10% and 20% of patients fail to respond to steroid therapy; thus, the prognosis for SRNS is usually poor, due to the increased risk of developing end-stage renal disease [10, 11]. Beltcheva et al. reported a case of a pediatric patient with SRNS that was caused by a novel dominant WT1 mutation, C1184A, identified in exon 9 [10]. Mutations within WT1 are frequent causes of sporadic, isolated cases of SRNS in girls [8]. WT1 expression is modulated by microRNAs (miRNAs) [12], which are small (20–25 nucleotides), noncoding RNA molecules that act as posttranscriptional regulators of gene expression in animals and plants, participating in many key biological processes [13]. In the kidney, miRNAs play critical roles in renal development, homeostasis, and physiological function. Several studies have shown that miRNAs are also key mediators of the pathogenesis of various renal diseases [14–16]. Furthermore, miRNAs can be secreted into the extracellular environment and detected in fluids, including urine and serum. They have been proposed as biomarkers for a wide range of diseases [17, 18].

Gao et al. discovered that miRNAs 15a and 16-1 may function as tumor suppressors to regulate leukemic cell proliferation, potentially by downregulating the WT1 oncogene [12]; pure curcumin was also recently shown to modulate the expression of WT1, partly by upregulating the expression of miR-15a and miR-16-1 in leukemic cells [19].

In the podocytes of patients with focal segmental glomerulosclerosis (FSGS), the expression of miR-193a was induced, which in turn inhibited the expression of WT1, modulating the expression of genes critical for podocyte architecture [20]. The presence of miRNAs in plasma, serum, and urine was also shown to be a possible biomarker of diabetic kidney disease [21]. Another mechanism of modulation of WT1 that has been observed in experimental mice during the sepsis is the decrease of nuclear WT1 in podocytes and was associated with transcriptional suppression of nephrin and cause albuminuria [22].

The aim of this study was to determine whether there are different expression levels and protein localization of WT1 in SRNS and SSNS patients and its relationship with the miR-15a, miR-16-1, and miR-193a, which modulate the WT1 expression in other models.

2. Methods

Twenty-one biopsies, serum and urine samples from 11 males and 10 females, from patients diagnosed with NS who met the criteria of International Study of Kidney Disease in Children with proteinuria levels greater than 3.5 g/24 h were included, in the control group serum and urine samples were obtained from 10 subjects without kidney disease, renal biopsies were obtained of autopsy subjects without renal disease who died in accidents as a result of a head injury. Patients who agreed to participate in the study signed a letter of informed consent. This study was performed with patients living in the north-central region from Mexico, according to the principles of the Declaration of Helsinki, and was approved by the ethics committees of our institutions.

2.1. Sample Processing. Freshly urine samples were processed in less than 30 min, and 10 mL of urine was centrifuged for 5 min at 1,500 rpm. After decanting the supernatant, the pellet was resuspended in 1 mL PBS-TS (0.1% Tween-20 and 0.02% SDS in phosphate-buffered saline [PBS]); 25 μL of which was fixed at 55°C for 10 min for continued permeabilization as described below.

2.2. Immunofluorescence. As the localization of WT1 is essential for its biological action, WT1 localization was assessed for all samples. For the analysis of the number of podocytes per glomerulus in both NS patients and healthy subjects, WT1 and nephrin were used as biomarkers for immunofluorescence of 4 μm thick sections of renal tissue mounted on microscope slides. The specimens were dewaxed, permeabilized with Triton X-100 (Triton 0.1% with 1% sodium citrate in PBS), and washed three times with PBS. The tissues were blocked with 20% fetal bovine serum (FBS) in PBS for 30 min and incubated for 1 h with monoclonal anti-WT1 antibody (WT1 [F-6] sc-7585, monoclonal mouse IgG1, Santa Cruz Biotechnology, Santa Cruz, CA, USA) or anti-nephrin monoclonal antibody (H-300 sc-28192, polyclonal rabbit IgG, Santa Cruz Biotechnology) diluted 1:100 in 10% FBS in PBS. After washes with PBS, the presence of bound antibody was identified by goat anti-mouse IgG1 Texas Red (sc-2979, Santa Cruz Biotechnology) and goat anti-rabbit IgG-FITC (sc-2012, Santa Cruz Biotechnology) staining. Some slides were counterstained with 4’,6-diamidino-2-phenylindole (DAPI). Finally, the slides were mounted and examined by confocal scanning microscopy. The intensity of the signal was expressed in pixels and analyzed using Image-Pro Plus software, version 7.0 (Media Cybernetics, Rockville, MD, USA). The WT1 localization was determined by Pearson correlation coefficient (PCC) [23].

2.3. Blood Collection. Five mL whole blood samples from NS patients and healthy controls were collected in tubes (BD Vacutainer; Becton Dickinson Vacutainer Systems, Franklin Lakes, New Jersey, USA) and then processed for obtain the serum by centrifugation at 1,500 rpm for 10 min. The sera were stored at −70°C until analysis.

2.4. RNA Isolation and Quantification. Total RNA was extracted from patient sera using the miRNeasy Serum/Plasma Kit (QIAGEN, Hilden, Germany), according to the manufacturer’s instructions. Two aliquots of 200 μL each were used to obtain sufficient RNA to perform all quality tests. RNA was dissolved in diethylpyrocarbonate-treated water, and RNA quality and quantity were determined by spectrophotometry using the NanoDrop ND-1000 (NanoDrop Technologies, Wilmington, DE, USA).

2.5. cDNA Synthesis and Real Time RT-PCR (qPCR) Assay for miRNAs Expression. To determine whether miR-15a, miR-16-1, and miR-193a are involved in regulating the expression of WT1, their expression was quantified in sera from NS patients
and healthy controls. Total RNA was polyadenylated and subjected to reverse transcription using an NCodermiRNAFirst-Strand cDNA Synthesis Kit (Invitrogen Carlsbad, CA) according to the manufacturer’s instructions.

Real-time qPCR analysis was carried out using SYBR® Green PCR Master Mix (Applied Biosystems; Foster City, CA, USA) on an ABI 7500 Fast Real-Time PCR System (Applied Biosystems). The primers reported by Gao et al. [12] were used to quantify the relative expression levels of miR-15a, miR-16-1, and miR-193a in the samples. Experiments were performed in triplicate. U6 was used as an endogenous control. The expression of each miRNA relative to U6 RNA was calculated using the equation $2^{-\Delta\Delta CT}$ [24–26].

2.6. RNA Isolation and qPCR for WT1 mRNA Expression. Total RNA was isolated from renal biopsies using the High Pure RNA Isolation Kit and High Pure RNA Paraffin Kit (Roche Life Science, Mexico) according to the manufacturer’s instructions. Single-stranded cDNA was synthesized by reverse transcription using the miScript Reverse Transcription Kit (Qiagen, United States) according to the manufacturer’s instructions. Real-time PCR was performed using the ABI 7500 Fast Real-Time PCR System (Applied Biosystems, Foster City, CA, USA) with TaqMan gene expression assays. Comparative real-time PCR assays were performed for each sample in triplicate. The primers for WT1-F: 5′-TCTGGCAGACCTATACAG-3′, WT1-R: 5′-CACATCCTGAACTGCTGAGA-3′; and WT1-P FAM: 5′-CACCCTGGGTGTAGATT-3′ were used. The comparative quantification cycle threshold ($C_T$) method was used to determine the relative expression levels of the WT1 gene. The 18S rRNA (Applied Biosystems, Foster City, CA, USA) gene was used for normalization.

2.7. Statistical Analyses. Statistical analyses were performed using SPSS 17.0 software. The results are presented as the mean ± SD. Comparisons were made using an analysis of variance and Student’s t-test, and the association between miRNA levels and expression of WT1 was assessed by Pearson’s correlation statistical program and considered significant at $p$ value less than or equal to 0.05.

3. Results

3.1. Sample Description. Twenty-one samples from patients with NS and 10 healthy subjects without renal disease as control group (Table 1) were analyzed for levels of creatinine (0.39 ± 0.25 mg/dL), proteinuria (4.01 ± 2.7 g/24 h), cholesterol (207.06 ± 91.72 mg/dL), triglycerides (141.56 ± 93.55 mg/dL), and albumin (3.75 ± 0.83 g/dL). Control renal biopsies were obtained from autopsy subjects without renal disease who died in accidents as a result of a head injury.

3.2. WT1 Localization in Podocytes in NS Patients and Healthy Subjects. Immunofluorescence of renal biopsy tissue using WT1 as a biomarker showed a decrease of 27% in NS patients compared to healthy subjects (Figure 1(a)). The localization of WT1 in SRNS patients was primarily cytoplasmic, whereas WT1 in SSNS patients showed both a nuclear and a cytoplasmic localization while in healthy controls it was mainly nuclear. The localization assay was performed by the colocalization of DAPI (nuclear localization) and Texas Red (WT1), showing a statistically significant difference between SSNS (PPC = 0.51 ± 0.03) and SRNS (PPC = 0.29 ± 0.01) patients compared with healthy subjects (PPC = 0.73 ± 0.02) ($p = 0.00001$) (Figure 1(c)). In urine sediments of SRNS patients, detached podocytes showed a cytoplasmic localization of WT1 (PPC = 0.35 ± 0.08) (Figure 1(b)). These results were significantly different from those in healthy subjects (PPC = 0.64 ± 0.03) ($p \leq 0.0001$; Figure 1(d)).

3.3. Analysis of WT1 Expression in Podocytes of NS Patients and Healthy Controls. WT1 expression analysis in podocytes was performed by two methods: immunofluorescence and RT-qPCR. WT1 expression by intensity pixels in renal biopsies of patients with NS was statistically different than healthy controls ($p = 0.0001$) and between patients with SSNS and SRNS ($p = 0.05$; Figure 2(a)), with highest levels in healthy controls and lowest levels in SRNS patients. WT1 RNA expression levels were analyzed by RT-qPCR in renal biopsies of NS patients and healthy controls, showing significantly lower (66%) WT1 mRNA levels in NS patients ($p = 0.017$), indicating a decrease in basal WT1 levels in the podocytes of NS patients (Figure 2(b)).

3.4. Expression of miR-15a, miR-16-1, and miR-193a Levels in Sera from NS Patients. The expression levels of miR-15a ($2^{-\Delta\Delta CT}$) did not differ between the different groups (SRNS 0.33 ± 0.12, SSNS 0.91 ± 0.44, and HS 1.54 ± 0.54 of relative expression) ($p = 0.138$; Figure 3(a)) or between patients with SRNS and SSNS compared to the control group ($p = 0.31$; Figure 3(b)). While the expression of miR-16-1 was significantly lower (0.54 ± 0.144 relative expression) in the sera of NS patients compared with healthy controls (1.86 ± 0.82 relative expression) ($p = 0.019$; Figure 3(c)), there was no difference between SSNS and SRNS patients ($p = 0.14$; Figure 3(d)). The relative expression of miR-193 ($2^{-\Delta\Delta CT}$) did not differ between NS patients (2.14 ± 0.36 relative

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<th>Table 1: Description of NS patients and controls.</th>
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SSNS: steroid-sensitive nephrotic syndrome; SRNS: steroid-resistant nephrotic syndrome.
expression) and controls (2.30 ± 0.75 relative expression, \( p = 0.59 \); Figure 3(e)) or between patients with SRNS and SSNS \( (p = 0.91 \); Figure 3(f)).

3.5. Relationship between WT1 and miRNAs. We did not find a significant correlation between WT1 expression and serum concentrations of miR-15a, miR-16-1, and miR-193a in NS patients (miR-15a: \( r = 0.118, p = 0.882 \); miR-16-1: \( r = 0.301, p = 0.969 \); and miR-193a: \( r = 0.213, p = 0.787 \)). However, a positive correlation was found between miR-15a and miR-16-1 \( (r = 0.994, p = 0.006) \). There was no significant relationship between the particular miRNA levels.

4. Discussion

Epidemiological studies indicate that NS remains the most common manifestation of glomerular disease in childhood, often causing death from untreated infections [27]. The present study reviewed 21 children with NS in the Mexican mestizo population, with an average age of 5.8 years.

Idiopathic NS is one of the most common glomerular diseases in pediatrics. The response to steroids is the best prognostic factor in this disease, and most children with NS respond to corticosteroids. However, 70% of children experience recurrence, with episodes of edema and proteinuria [28]; fewer than 3% of patients with SSNS progress to chronic renal failure compared to 50% patients with SRNS [29].

The WT1 protein has been used as a biomarker of podocyte loss from glomeruli in kidney disease [30, 31] and of the presence of damaged podocytes in urinary sediment [32, 33]. In this work, we clearly observed a reduction of podocytes in renal biopsies of children with NS and the presence of podocytes in the urine, but the reason for such podocyte
detachment is not clear. The cause may be an imbalance between different isoforms of WT1 or decreased basal protein expression. Different research groups have observed a decrease in WT1 protein expression in children with SRNS, which was often accompanied by WT1 mutations in exons 8 and 9 [8,34,35]. However, another report suggested that reduced expression levels of WT1 cause glomerulonephritis and mesangial sclerosis, depending on the level of the protein [36].

We found normal WT1 expression in the nuclei of podocytes from biopsies of healthy controls without kidney disease; however, in SSNS patients, WT1 expression was localized to both the nucleus and cytoplasm, whereas WT1 expression is primarily cytoplasmic in SRNS patients, suggesting that delocalization of WT1 from the nucleus can lead to a loss of function as a transcription factor. Decreased expression levels of WT1 lead to downregulation of its target genes PODXL (podocalyxin) and NPHSI (nephrin), as well as several other genes crucial for the architecture of podocytes, initiating a catastrophic collapse of the entire podocyte-stabilizing system [20]. Additionally, Kato et al. [22] observed that in Lipopolysaccharide (LPS) treated mice, a loss of albumin decreased nephrin levels and the nuclear localization of WT1 in podocytes. The nuclear localization of WT1, as well as nephrin mRNA and protein levels, returned to near basal levels 72 hours after LPS with recovery of albumin levels [22].

The loss of WT1 gene expression could be a fatal consequence as described above [9]; the regulation of WT1 is provided by certain genes, such as PAX2 and PAX8, or epigenetic modulation mechanisms, such as miRNAs [12, 19]. In addition, WT1 has the ability to self-regulate [37]. In this study, we assessed whether low WT1 expression in podocytes correlates with the circulating levels of miR-15a, miR-16-1, and miR-193a in the sera of NS patients. Our results indicate that the expression of miR-15a was higher in healthy controls than in NS patients but was not statistically significant. Similar results were observed for miR-16-1, but the difference between the expression of miR-16-1 in NS patients and in healthy controls was significant, indicating an inverse relationship between miRNAs and WT1 expression. Gao et al. reported that miR-15a and miR-16-1 inhibit the proliferation of leukemic cells by decreasing WT1 levels via joining WT1-3'UTR. The expression of these miRNAs inhibits cell proliferation, promotes apoptosis of cancer cells, and suppresses tumorigenicity both in vitro and in vivo [12]. Both miR-15a and miR-16-1 have been shown to negatively affect several oncogenes, including BCL2, MCL1, CCND1, and WNT3A. Downregulation of these miRNAs has been reported in chronic lymphocytic lymphoma, pituitary adenoma, and prostatic carcinoma [37, 38], while miR-193a was not found to elicit any significant effects on these cancers. However, the data reported by Gebeshuber et al. [20] indicated that miR-193a is involved in FSGS by decreasing WT1 levels. In conclusion, we found low WT1 expression and delocalization in NS patients. We also observed higher expression levels of miR-15a and miR-16-1 in healthy controls compared with NS patients, but only miR-16-1 levels were statistically different. However, we did not find a correlation between miRNA expression and WT1 levels. It is important to extend the search for miRNAs as biomarkers of renal diseases as well as to understand the basic mechanisms of gene regulation of podocytes and the glomerular filtration system.

Disclosure

The authors alone are responsible for the content and writing of the article.

Competing Interests

The authors report no conflict of interests.
Figure 3: Expression of miR-15a, miR-16-1, and miR-193a in sera from patients with NS. Relative expression levels of miR-15a (a) in patients with NS (nephrotic syndrome) and healthy subjects (HS) and (b) in patients with steroid-sensitive NS (SSNS) and steroid-resistant NS (SRNS). miR-16-1 (c) in patients with NS and HS and (d) in patients with SSNS and SRNS and miR-193a (e) in patients with NS and HS and (f) in patients with SSNS and SRNS. *p < 0.05, **p < 0.01, and ***p < 0.001.

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