

Research Article

Targeted Molecular Magnetic Resonance Imaging Detects Brown Adipose Tissue with Ultrasmall Superparamagnetic Iron Oxide

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The peptide (CKGGRAKDC-NH₂) specifically targets the brown adipose tissue (BAT). Here we applied this peptide coupled with polyethylene glycol (PEG)-coated ultrasmall superparamagnetic iron oxide (USPIO) nanoparticles to detect BAT in vivo by magnetic resonance imaging (MRI). The peptide was conjugated with PEG-coated USPIO nanoparticles to obtain targeted USPIO nanoproboscopes. Then the nanoproboscopes for BAT were evaluated in mice. T₂*-weighted images were performed, precontrast and postcontrast USPIO nanoparticles. Finally, histological analyses proved the specific targeting. The specificity of targeted USPIO nanoproboscopes was observed in mice. The T₂* relaxation time of BAT in the targeted group decreased obviously compared to the controls (P<0.001). Prussian blue staining and transmission electron microscope confirmed the specific presence of iron oxide. This study demonstrated that peptide (CKGGRAKDC-NH₂) coupled with PEG-coated USPIO nanoparticles could identify BAT noninvasively in vivo with MRI.

1. Introduction

Obesity has become globally epidemic. It is associated with an increased risk for many chronic diseases, such as diabetes, hypertension, and heart disease. Obesity will develop when energy intake exceeds expenditure [1].

As known, white adipose tissue (WAT) is the main site of energy storage in the form of triglycerides, while brown adipose tissue (BAT) is the major depot of adaptive thermogenesis in mammals [2]. BAT, which is mainly located in intrascapular and paraspinal region in rodents and humans [3], plays a key role in the balance of energy metabolism. The increase in the amount or activity of BAT can effectively increase the energy consumption, reduce the WAT accumulation, improve metabolism, and resist obesity and hyperlipidemia as well as other metabolic disorders [4].

The functional activity of BAT has been researched a lot in animals; then extrapolation to humans of results of these studies firstly needs noninvasive approaches to assess human BAT [5]. The positron emission tomography/computed

tomography (PET/CT) imaging with 2-deoxy-2-[18F]fluoro-D-glucose ([18F]DG) is the most common technique for BAT imaging [6]. However, the PET-CT imaging depends on high glucose utilization which is associated with metabolic activity. [18F]DG PET-CT imaging only shows activated BAT and is not sensitive to BAT in the nonactivated thermoneutral state. Another limitation of PET imaging is owing to the use of ionizing radiation [7].

Recently, molecular imaging has arisen to visualize and characterize BAT at the molecular as well as cellular level with different imaging modalities that include positron emission tomography, single photon emission computed tomography, and optical imaging. Because of the high spatial resolution and simultaneous anatomic, physiologic, and functional information, magnetic resonance imaging (MRI) is one of the best noninvasive methods in molecular imaging for assessing function of tissues or diagnosing diseases [8].

Various nanoparticulate contrast agents for MR molecular imaging have been developed, like ultrasmall superparamagnetic iron oxide (USPIO), lutetium oxide nanoparticles,

and so on [9–11]. With smaller particle, high biocompatibility, and low toxicity, USPIO has become an attractive contrast agent in molecular imaging. USPIOs possess ultrasmall size as well as superparamagnetic properties and can produce large magnetism in the weak external magnetic field. In vivo, USPIO will result in the nonuniformity of local magnetic field which can lead to rapid dephasing of proton. So USPIOs can shorten transverse relaxation time and induce signal decrease on T2*-weighted MR images [12].

Since the peptide (CKGGRAKDC) that targeted WAT vasculature was found by Kolonin MG et al., we researched this peptide for over ten years [13]. Our previous study identified the peptide-drug conjugate (CKGGRAKDC-BVT.2733) protecting against diet-induced obesity, and amino-modified peptide (CKGGRAKDC-NH₂) specifically homing to BAT [14]. We have achieved two national patents: a targeted peptide for BAT (Patent No. 201010159916.3); a new targeted drug to BAT for treatment of obesity (Patent No. 201010159909.3).

In this study, we innovatively conjugated the targeted peptide (CKGGRAKDC-NH₂) with polyethylene glycol (PEG)-coated USPIO nanoparticles to synthesize BAT-targeted USPIO nanoprobe and investigated its potential application in detecting BAT with MRI in vivo.

2. Materials and Methods

2.1. Animals. C57BL/6J mice were purchased from Nanjing Biomedical Research Institute of Nanjing University. The weight of mice was 27.3±1.1g. The mice were housed five per cage in a room kept at 23 ± 1°C with 12-h light/dark cycle at Animal Core Facility of Nanjing Medical University and were allowed free access to water and food. All animal designs and studies were approved by the Animal Care and Use Committee of Nanjing Medical University.

2.2. Peptide and USPIO Nanoparticles. The targeted peptide CKGGRAKDC-NH₂ was synthesized by GL Biochem (Shanghai) Ltd. USPIO nanoparticles were commercially available and purchased from Nanjing Nanoeast Biotech Co.

The morphological characters of USPIOs were examined by transmission electron microscopy ([TEM] JEM-200CX, Japan). The nanoparticle size and distribution were calculated by measuring the diameters of 300 particles at least. The hydrodynamic diameter and Zeta potential were analyzed by dynamic light scattering ([DLS] Zetasizer Nano-z, UK). The magnetic saturation moment was measured by Vibrating Sample Magnetometer (Lakeshore7407, USA). To determine the relaxivity of nanoparticles, USPIO nanoparticles were diluted to the concentration of 0, 0.01395, 0.0279, 0.0558, 0.1116, 0.2232, and 0.4464mmol/L and performed on Siemens Prisma 3.0 T MR scanner (Erlangen, Germany). The detailed image parameters were repetition time (TR) = 10000ms and echo time (TE) = 12, 24, 48, 96, 192, 384ms. T2 relaxivity (r₂) was plotted against the iron concentration in the nanoparticles dilutions. T2 relaxation rate (r₂) was determined by a linear fit.

2.3. Synthesis of Targeted USPIO Nanoprobos. The USPIO nanoparticles were coated by PEG. First, 5mg PEG-USPIO

nanoparticles (1mg/mL) were taken; then 1mg 1-ethyl-3-(dimethylaminopropyl) carbodiimide hydrochloride (EDC) molecule and 0.5 mg N-Hydroxysuccinimide (NHS) molecule were added. They reacted at a constant temperature shaker for 25 min (25°C, 180 rpm) to activate PEG-USPIO nanoparticles. 2 mg peptide and 5 mg activated PEG-USPIO nanoparticles were mixed and reacted at a shaker for 2 hours. Finally, the unreacted peptide was isolated by using molecular sieves and targeted USPIO nanoprobos were obtained. PEG-USPIO nanoparticles were untargeted and used as control.

2.4. In Vivo MRI Scanning. Studies were performed on a 7T preclinical animal MRI scanner by using a dedicated mouse body coil with a coil diameter of 40 mm (Bruker BioSpin, Germany). The animals were maintained under anesthesia with 2%–3% isoflurane in O₂ gas and the flow rate was 0.8L/min. In addition, respiratory rates of the mice were monitored to maintain 30 breaths per minute.

T2*-weighted images were acquired by using the multi-gradient-echo (MGE) sequence with free-breath. Detailed image parameters were as follows: repetition time (TR) = 400 ms, echo time (TE) range = 2.7–37.0 ms, echo spacing = 3.12ms, flip angle = 25°, echoes = 12, matrix = 192×192, field of view (FOV) = 30 × 30mm², slice thickness = 0.5 mm, and number of excitations (NEX) = 4.

The mice were randomly divided into two groups: targeted group received targeted USPIO nanoprobos and control group injected with untargeted USPIO nanoparticles at the same dose of 80 μmol Fe/kg body weight through the mice tail vein. First, we repeated the T2*-weighted images before and at 30min, 60min, 90min, 120min, and 180min after injection (n = 2 per group) to research when maximum negative enhancement effect was observed and disappeared. Then we continued to study the specificity of nanoprobos by imaging before and at 120min after intravenous administration of contrast agents on the basis of the results (n = 7 per group).

2.5. Image Analysis. The T2*MR relaxometry maps were generated automatically on 7T MRI. Later, T2* value was measured on T2*MR relaxometry maps with FireVoxel software. Region of interest (ROI) was manually drawn surrounding the BAT and WAT or at the center of muscle and ROI on muscle was chosen at the same slice of BAT. ΔR2* was calculated by equation: ΔR2* = R2*_{post} - R2*_{pre} and R2* [s⁻¹] = 1/T2* [milliseconds] × 1000.

2.6. HE Staining and Prussian Blue Staining. For hematoxylin and eosin (HE) and Prussian blue staining, tissue was fixed in 10% formalin and embedded into paraffin blocks. Thin sections were cut off and deparaffined and rehydrated. The staining was performed by using a routine protocol. Images were obtained with light microscopy (LeicaDFC450C, Germany).

2.7. Transmission Electron Microscopy. For TEM analyses, Tissue samples (less than 1mm³) were fixed in 5% glutaraldehyde for 2 hours, then fixed in 1% OsO₄ for 2 hours and stained with 2% uranyl acetate, dehydrated with acetone, and

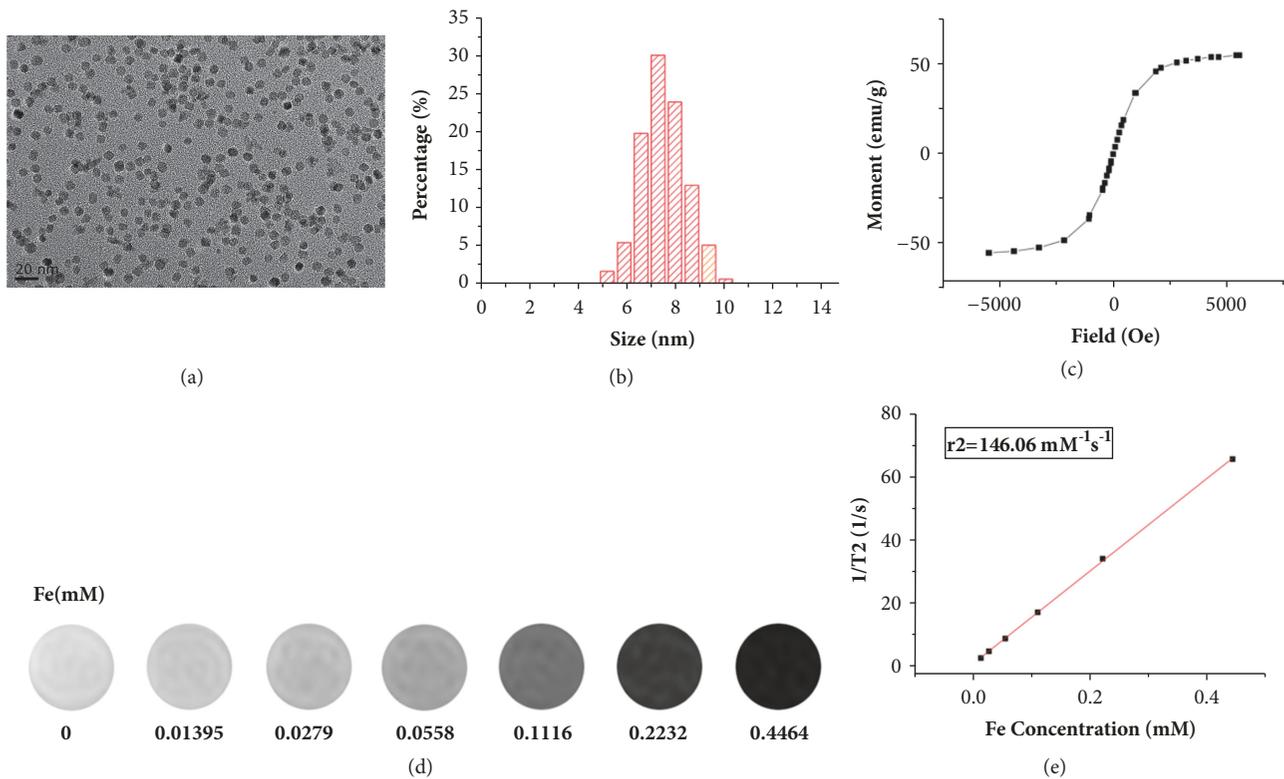


FIGURE 1: Characterization of the USPIOs. (a) TEM image of the USPIO nanoparticles. (b) Distribution of USPIO core size. (c) Magnetization curve of USPIO nanoparticles. (d) Concentration-dependent T₂-mapping of USPIO nanoparticles on 3.0 T MR in vitro. (e) T₂ relaxation time at various iron concentrations. Data are recorded as the mean \pm SD. TEM indicates transmission electron microscope.

embedded in EPON resin. Images of ultrathin sections were analyzed with TEM at 60 kV (Hitachi H-7500, Japan).

2.8. Statistical Analysis. The data were recorded as the mean \pm standard deviation (SD). Statistical comparisons were performed by a one-way analysis of variance (ANOVA) with SPSS19.0. P values of <0.05 were considered statistically significant.

3. Results

3.1. Characterization of Untargeted USPIO Nanoparticles and Targeted USPIO Nanoprobes. TEM images showed that the USPIOs were well dispersed (Figure 1(a)) and the mean core size was 7.56 ± 0.92 nm (Figure 1(b)). Research suggests that the size-dependent magnetism is important for biomedical application [15]. The saturation magnetization value was 55 emu/g (Figure 1(c)), which was high enough to achieve significant superparamagnetism. With the concentration of USPIO increased, T₂-mapping MR signal was gradually attenuated (Figure 1(d)), and the r_2 value of USPIOs was $146.06 \text{ mM}^{-1} \cdot \text{s}^{-1}$ (Figure 1(e)). The hydrodynamic diameters of untargeted USPIO nanoparticles and targeted USPIO nanoprobes were 17.50 ± 5.90 nm and 20.55 ± 5.70 nm, respectively, which were within the range of ultrasmall particles of iron oxide (10-40nm) [16]. Zeta potentials of untargeted

USPIO nanoparticles and targeted USPIO nanoprobes were -33.9 ± 6.72 mV and -22.0 ± 5.86 mV, respectively, demonstrating the stability [17].

The targeting specificity of the peptide (CKGGRAKDC-NH₂) homing to BAT was proved by confocal immunofluorescence analysis (see Figures S1 in the Supplementary Material).

3.2. In Vivo MRI Studies. The ability of targeted USPIO nanoprobe to detect BAT in vivo was assessed by MRI. Figure 2 was the representative color-coded T₂* map before and 30min, 60min, 90min, 120min, and 180min after administering targeted USPIO nanoprobes or untargeted USPIO nanoparticles. The T₂* map of BAT in control group (Figure 2(a)) and WAT in targeted group (Figure 2(b)) showed no obvious shortening of the T₂* time. BAT in targeted group (Figure 2(c)) exhibited the shortening of T₂* time after injection until 120min which is due to the properties of USPIO, and this negative enhancement phenomenon disappeared at 180min. However, no change was observed in muscle. The reduction of T₂* time corresponded to the increase in the estimated R₂*. The calculated ΔR_2^* (Figure 3(a)) curves displayed that the ΔR_2^* of BAT was ascendant after injecting targeted USPIO nanoprobes and kept in steady for some time until 120min and then declined. The ΔR_2^* of WAT and muscle in targeted group and the ΔR_2^* of BAT in control group experienced almost no change.

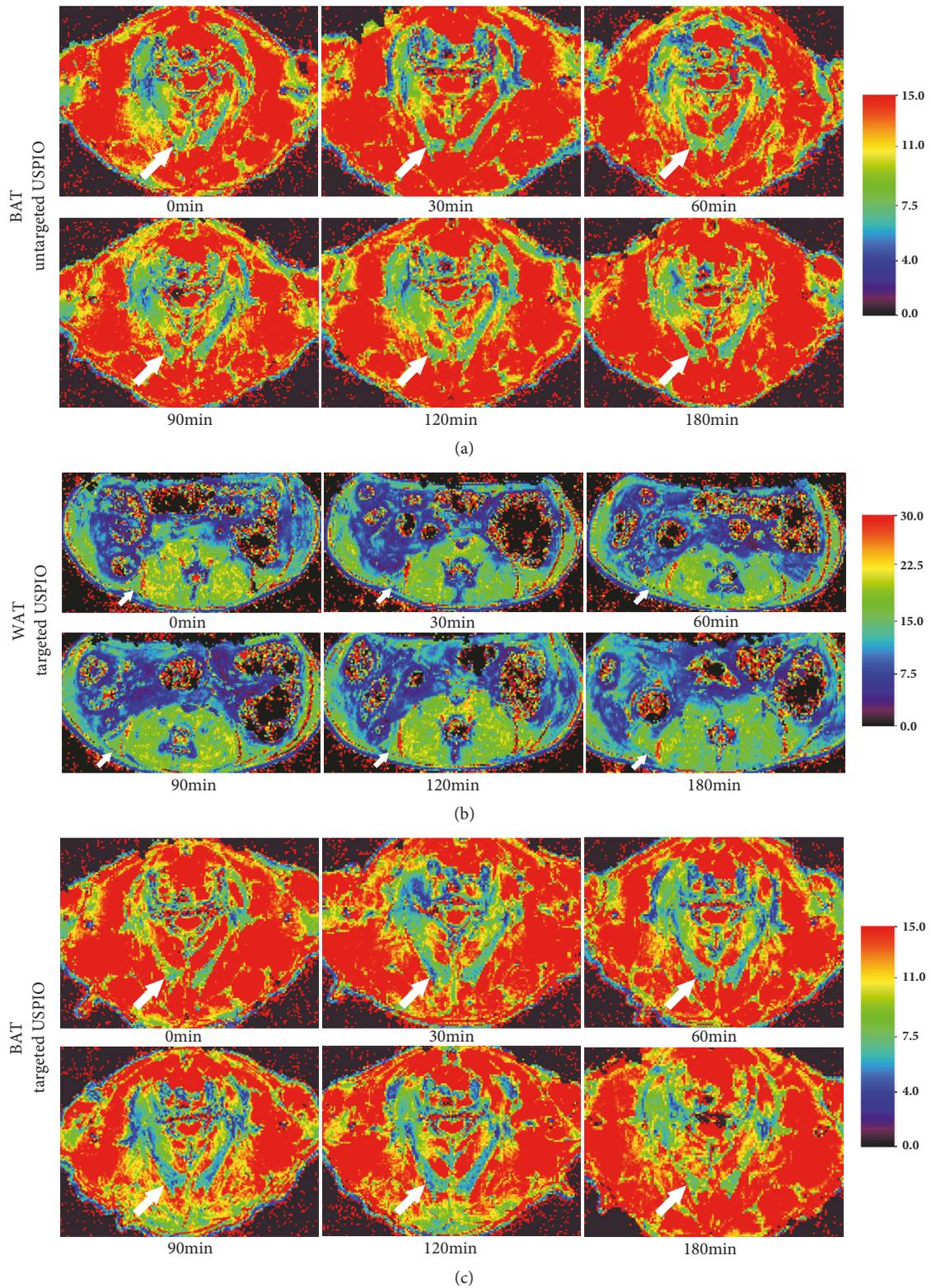


FIGURE 2: Representative color-coded MR T2* map. T2* map of BAT injected with untargeted USPIO (a). T2* map of WAT (b) and BAT (c) received targeted USPIO (T2*WI indicates T2*-weighted image). Arrows point to BAT in (a) and (c) and point to white adipose tissue in (b).

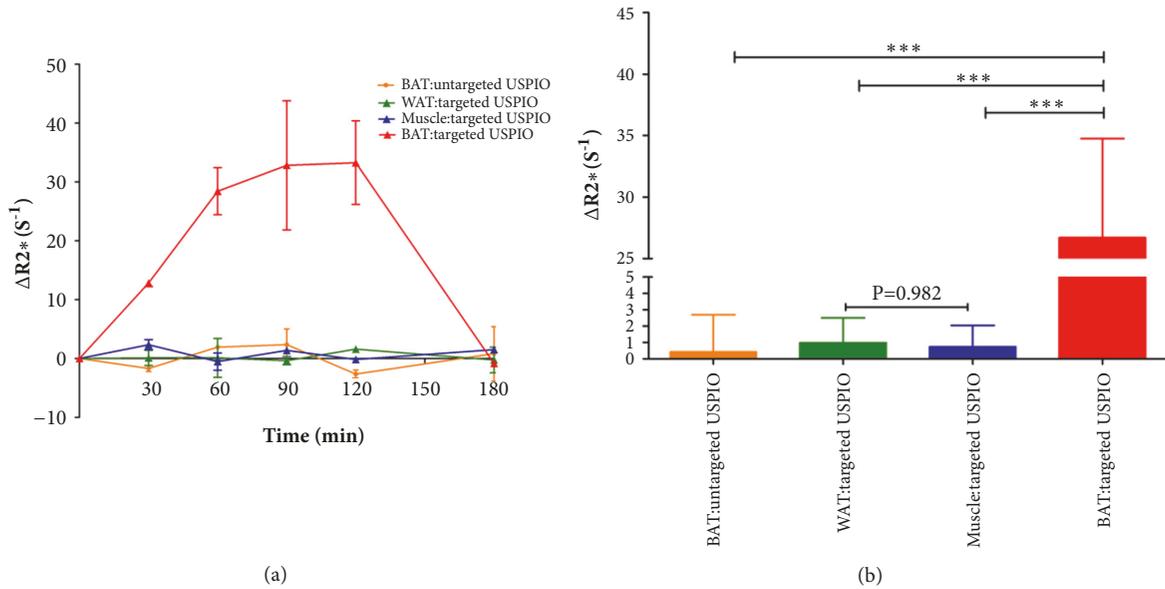


FIGURE 3: Quantitative analysis of $\Delta R2^*$ values. (a) The $\Delta R2^*$ values curve of BAT, WAT, and muscle in targeted group and BAT in control group injected with targeted USPIO or untargeted USPIO, respectively (n = 2). The error bars represent the SD of $\Delta R2^*$ for two mice in different group. (b) Statistic analysis of $\Delta R2^*$ values in the BAT in control group and WAT, muscle, and BAT in targeted group (n = 7). *** P<0.001. The error bars represent the SD of $\Delta R2^*$ for seven mice in different group. Data are recorded as the mean \pm SD.

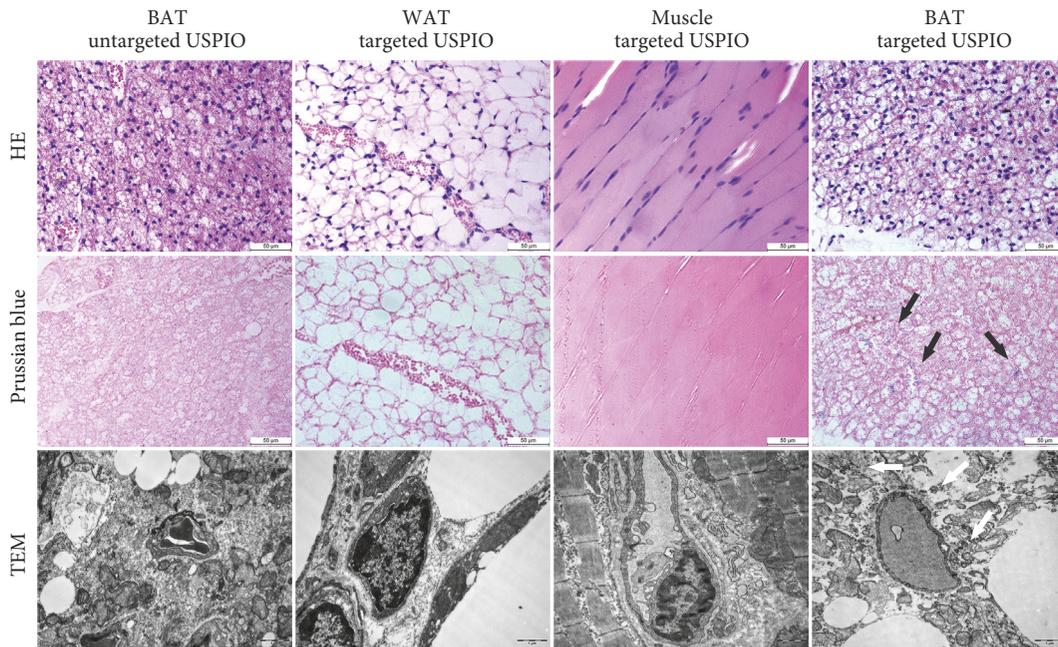


FIGURE 4: Histological analyses. HE staining, Prussian blue staining, and TEM of BAT in control mice and WAT, muscle, and BAT in targeted group. The arrows refer to the iron particles. Scale bar in staining is 50 μ m. Scale bar in TEM is 1 μ m.

Later, we chose to perform T2*MR imaging on mice at 120min after administration. As observed in Figure 3(b), the $\Delta R2^*$ (26.71s⁻¹) of BAT of mice in targeted group was significantly higher (p \leq 0.05) than all the other negative controls.

3.3. *Histological Analyses.* Prussian blue staining and TEM analysis have confirmed the MRI results. In Prussian blue

staining, iron particles appeared as blue stain and only existed in BAT, not in WAT or muscle in targeted group. As expected, the BAT in control group was Prussian blue negative. Transmission electron microscopy demonstrated that iron particles were located in BAT while no obvious particles were found in the controls, which were consistent with the results of Prussian blue staining and MRI results (Figure 4).

4. Discussion

In this study, we innovatively used targeted peptide (CKGGRAKDC-NH₂) coupled with USPIO nanoparticles to specifically identify BAT in vivo on MR. According to our knowledge, it has not been researched before. In previous research, a noninvasive method to image BAT with superparamagnetic iron oxide nanoparticle (SPIO) has been used [18]. However, the applicability is limited without BAT-specific probes.

With intrinsic unique magnetic properties that can lead to rapid dephasing of protons, SPIOs offer sufficient sensitivity for T₂*-weighted MR imaging. However, their hydrodynamic diameter is >50 nm, resulting in a fast clearance rate by phagocytic cells [19]. Then, USPIO is smaller than SPIO and also has high relaxivity. So USPIO nanoparticles have become an attractive targeted contrast agent for T₂*-weighted MR imaging [20]. In our study, the mean hydrodynamic diameter of our USPIOs was about 17.50nm, which helps nanoparticles escape from phagocytes and maintain a higher blood half-life [19]. After measuring, there were about 10⁴ Fe atoms and 230 peptides per nanoparticle. And the calculated amount of targeted ligand per unit area was similar to that of Kazmierczak et al.'s study [21]. The saturation magnetization value (55 emu/g) and the relaxation coefficient r₂ of USPIOs (146.06mM⁻¹·S⁻¹) were high enough to cause obvious T₂ contrast enhancement [22, 23]. Due to their strong relaxation properties, low dose of contrast agent (80 μmol Fe/kg body weight) could be detected and was used in our study [24].

USPIO nanoparticles used in this study were stabilized with PEG. The stabilization can prevent aggregation and cleared by macrophages [25]. The Zeta potential of our USPIOs demonstrated that they were stable [17]. PEG is one of the best hydrophilic and biocompatible polymeric coating materials, which can make nanoparticles become water-soluble and does not change the magnetic abilities [26]. At the same time, using PEG to coat USPIO can increase biocompatibility and blood circulation time. In addition, PEG will provide active functional carboxylic acid groups able to conjugate with targeted peptides, which also has been reported by Roberts [27].

Furthermore, recent studies have verified that specific peptide can be linked with USPIO for molecular MR imaging and the peptide properties and functionality as well as the relaxation abilities of USPIO nanoparticles can also be remained [28]. Our research also demonstrated that the targeted peptide (CKGGRAKDC-NH₂) conjugated with USPIO can also retain its affinity for BAT.

T₂*-weighted imaging was used in our study and acquired with multi-gradient-echo sequence. The signals were acquired at different TE and an appropriate model was fitted to calculate the T₂* value, which reflected tissue characterization [29]. USPIO will result in the nonuniformity of local magnetic field. As it happens, T₂*WI is sensitive to local field variation and iron oxide nanoparticle detection [30].

It has been confirmed that this MR targeted USPIO nanoprobe could identify BAT in vivo noninvasively and efficiently. The results of our study revealed that targeted

USPIO nanoprobe mainly caused T₂* signal decrease in BAT of mice on MR relaxometry T₂* map. In contrast, there was no T₂* signal reduction in BAT injected with untargeted USPIO nanoparticles (Figure 2). The T₂* value of WAT was higher than BAT and similar to muscle; in order to observe the difference between WAT and muscle better, the scale range in Figure 2(b) was larger than others. After exploring, we found that when the center value of scale bars was similar to the T₂* value of objective tissue, the tissue contrast was the best. In view of Kuhlpetter's report, the R₂* value, reciprocal of T₂* value, was shown to be proportional to the iron concentration in tissue [31]. Quantitative R₂* analysis will enable noninvasive estimation of iron in tissue. Therefore, R₂* and ΔR₂* value were calculated to perform quantitative analysis in this research. The obvious positive ΔR₂* value demonstrated the existence of iron, which was only observed in BAT injected with targeted USPIO nanoprobe. Statistical analyses also proved that the ΔR₂* was significantly higher at 120min after administrating targeted USPIO nanoprobe (P<0.05), while the controls showed no substantial deviation from zero. Prussian blue staining and TEM confirmed the specific existence of iron in BAT too.

As shown in Figure 3(a), the maximum negative enhancement effect occurred at 120min and disappeared at 180min postcontrast targeted USPIO nanoprobe. Species of mice, targeted tissue, peptide sequence, and dose of contrast agent were all likely to affect the time when maximum negative enhancement effect occurred and the duration it may continue. Burtea et al. studied USPIO conjugated with targeted peptides to probe pancreatic beta cells [32]. They reported that the lowest signal was attained at 53min and the enhancement still lasted until they stopped to perform MRI at 133min after injection. The dose was similar to ours, but the species of mice, targeted tissue, and sequence of peptide were all different from ours, which might be the cause of different enhancement time. You et al. discovered maximum contrast displayed at 3h and disappeared at 6h after administration of the probe when researching targeted USPIO to detect tumor [33]. In You's study, single-chain oligonucleotide fragment instead of peptide was used. Species of mice and targeted tissue were also different from ours. In addition, the amount of contrast was much higher than ours. These differences might be the reason that the maximum enhancement was generated later and continued longer.

Several limitations may influence the outcome of this study. First, MR T₂* relaxometry for quantitative analysis is affected by large-scale field inhomogeneities, which may lead to signal loss as well as overestimation of relaxation rate R₂*. Second, the T₂* of BAT is relatively short. Theoretically, compared to long T₂* values, short T₂* values may have more defects. With the shortening of the interecho times, R₂* may be highly calculated. If echotimes are very short, the relaxation signal will do not behave exponentially. These defects are based on scanner limitations and the T₂* value of tissue [34]. Third, the signal curves were just acquired from two mice in our study, which needed more mice to further verify maximum enhancement time and contrast disappearance time.

The further objective is to detect BAT function on MRI with this probe, which needs more research. Also, whether this probe can recognize BAT in human will be evaluated in future studies.

5. Conclusions

In summary, we have demonstrated that MRI with peptide (CKGGRAKDC-NH₂) coupled with PEG-coated USPIOs allowed noninvasive assessment of BAT in vivo, which is a specific, sensitive method for imaging BAT and provides new evaluation means for clinical obesity research as well as treatment.

Data Availability

The research article data used to support the findings of this study are included within the article and the supplementary information file.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Qingqiao Hu and Xiangxun Chen contributed equally to this work. All authors were involved in the study and approved its final version.

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Supplementary Materials

In vivo fat homing of the CKGGRAKDC-NH₂ motif in C57BL/6J mice. FITC-CKGGRAKDC-NH₂ peptide was administrated intravenously in C57BL/6J mice; then different tissues were collected and processed for distribution assay. The green immunofluorescence in formalin-fixed paraffin section is the most obvious in BAT, but nearly not detectable in WAT, liver, or muscle. Scale bar, 50 μm. (*Supplementary Materials*)

References

- [1] D. M. Nguyen and H. B. El-Serag, "The epidemiology of obesity," *Gastroenterology Clinics of North America*, vol. 39, no. 1, pp. 1–7, 2010.
- [2] Y.-H. Tseng, E. Kokkotou, T. J. Schulz et al., "New role of bone morphogenetic protein 7 in brown adipogenesis and energy expenditure," *Nature*, vol. 454, no. 7207, pp. 1000–1004, 2008.
- [3] K. Almind, M. Manieri, W. I. Sivitz, S. Cinti, and C. R. Kahn, "Ectopic brown adipose tissue in muscle provides a mechanism for differences in risk of metabolic syndrome in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 7, pp. 2366–2371, 2007.
- [4] F. Villarroya, R. Cereijo, J. Villarroya, and M. Giralt, "Brown adipose tissue as a secretory organ," *Nature Reviews Endocrinology*, vol. 13, no. 1, pp. 26–35, 2017.
- [5] K. A. Virtanen, "The rediscovery of BAT in adult humans using imaging," *Best Practice & Research Clinical Endocrinology & Metabolism*, vol. 30, no. 4, pp. 471–477, 2016.
- [6] K. Y. Chen, A. M. Cypess, M. R. Laughlin et al., "Brown Adipose Reporting Criteria in Imaging Studies (BARCIST 1.0): Recommendations for Standardized FDG-PET/CT Experiments in Humans," *Cell Metabolism*, vol. 24, no. 2, pp. 210–222, 2016.
- [7] V. Stahl, F. Maier, M. T. Freitag et al., "In vivo assessment of cold stimulation effects on the fat fraction of brown adipose tissue using DIXON MRI," *Journal of Magnetic Resonance Imaging*, vol. 45, no. 2, pp. 369–380, 2017.
- [8] M. G. Shapiro, R. M. Ramirez, L. J. Sperling et al., "Genetically encoded reporters for hyperpolarized xenon magnetic resonance imaging," *Nature Chemistry*, vol. 6, no. 7, pp. 629–634, 2014.
- [9] Y. Yuan, Z. Ding, J. Qian et al., "Casp3/7-Instructed Intracellular Aggregation of Fe₃O₄ Nanoparticles Enhances T₂ MR Imaging of Tumor Apoptosis," *Nano Letters*, vol. 16, no. 4, pp. 2686–2691, 2016.
- [10] Z. Liu, K. Dong, J. Liu, X. Han, J. Ren, and X. Qu, "Antibiofouling polymer-decorated lutetium-based nanoparticulate contrast agents for in vivo high-resolution trimodal imaging," *Small*, vol. 10, no. 12, pp. 2429–2438, 2014.
- [11] Z. Liu, F. Pu, S. Huang, Q. Yuan, J. Ren, and X. Qu, "Long-circulating Gd₂O₃:Yb³⁺, Er³⁺ up-conversion nanoprobe as high-performance contrast agents for multi-modality imaging," *Biomaterials*, vol. 34, no. 6, pp. 1712–1721, 2013.
- [12] Y. Wang, C. Xu, Y. Chang et al., "Ultrasmall Superparamagnetic Iron Oxide Nanoparticle for T₂-Weighted Magnetic Resonance Imaging," *ACS Applied Materials & Interfaces*, vol. 9, no. 34, pp. 28959–28966, 2017.
- [13] M. G. Kolonin, P. K. Saha, L. Chan, R. Pasqualini, and W. Arap, "Reversal of obesity by targeted ablation of adipose tissue," *Nature Medicine*, vol. 10, no. 6, pp. 625–632, 2004.
- [14] J. Liu, L. Wang, A. Zhang et al., "Adipose tissue-targeted 11β-hydroxysteroid dehydrogenase type 1 inhibitor protects against diet-induced obesity," *Endocrine Journal*, vol. 58, no. 3, pp. 199–209, 2011.
- [15] J. Xie, Y. Zhang, C. Yan et al., "High-performance PEGylated Mn-Zn ferrite nanocrystals as a passive-targeted agent for magnetically induced cancer theranostics," *Biomaterials*, vol. 35, no. 33, pp. 9126–9136, 2014.
- [16] M. Mahmoudi, V. Serpooshan, and S. Laurent, "Engineered nanoparticles for biomolecular imaging," *Nanoscale*, vol. 3, no. 8, pp. 3007–3029, 2011.
- [17] S. Bhattacharjee, "DLS and zeta potential - What they are and what they are not?" *Journal of Controlled Release*, vol. 235, pp. 337–351, 2016.
- [18] C. S. L. Jung, M. Heine, B. Freund et al., "Quantitative activity measurements of brown adipose tissue at 7 T magnetic resonance imaging after application of triglyceride-rich lipoprotein 59Fe-superparamagnetic iron oxide nanoparticle: Intravenous versus intraperitoneal approach," *Investigative Radiology*, vol. 51, no. 3, pp. 194–202, 2016.
- [19] J. W. M. Bulte and D. L. Kraitchman, "Iron oxide MR contrast agents for molecular and cellular imaging," *NMR in Biomedicine*, vol. 17, no. 7, pp. 484–499, 2004.

- [20] V. Pourcelle, S. Laurent, A. Welle et al., "Functionalization of the PEG corona of nanoparticles by click photochemistry in water: Application to the grafting of RGD Ligands on PEGylated USPIO imaging agent," *Bioconjugate Chemistry*, vol. 26, no. 5, pp. 822–829, 2015.
- [21] P. M. Kazmierczak, M. Schneider, T. Habereeder et al., " $\alpha v\beta 3$ -Integrin-Targeted Magnetic Resonance Imaging for the Assessment of Early Antiangiogenic Therapy Effects in Orthotopic Breast Cancer Xenografts," *Investigative Radiology*, vol. 51, no. 11, pp. 746–755, 2016.
- [22] Y.-W. Jun, J.-H. Lee, and J. Cheon, "Chemical design of nanoparticle probes for high-performance magnetic resonance imaging," *Angewandte Chemie International Edition*, vol. 47, no. 28, pp. 5122–5135, 2008.
- [23] D. Högemann-Savellano, E. Bost, C. Blondet et al., "The transferrin receptor: a potential molecular imaging marker for human cancer," *Neoplasia*, vol. 5, no. 6, pp. 495–506, 2003.
- [24] S. K. Mishra, B. S. H. Kumar, S. Khushu, R. P. Tripathi, and G. Gangenahalli, "Increased transverse relaxivity in ultrasmall superparamagnetic iron oxide nanoparticles used as MRI contrast agent for biomedical imaging," *Contrast Media & Molecular Imaging*, vol. 11, no. 5, pp. 350–361, 2016.
- [25] C. Corot, P. Robert, J.-M. Idée, and M. Port, "Recent advances in iron oxide nanocrystal technology for medical imaging," *Advanced Drug Delivery Reviews*, vol. 58, no. 14, pp. 1471–1504, 2006.
- [26] S. Laurent, D. Forge, M. Port et al., "Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications," *Chemical Reviews*, vol. 108, pp. 2064–2110, 2008, Erratum in *Chemical Reviews*, vol. 110, p. 2574, 2010.
- [27] M. Roberts, M. Bentley, and J. Harris, "Chemistry for peptide and protein PEGylation," *Advanced Drug Delivery Reviews*, vol. 54, no. 4, pp. 459–476, 2002.
- [28] C. Burtea, S. Laurent, T. Sanli et al., "Screening for peptides targeted to IL-7R α for molecular imaging of rheumatoid arthritis synovium," *Arthritis Research & Therapy*, vol. 18, no. 1, 2016.
- [29] V. Positano, B. Salani, A. Pepe et al., "Improved T2* assessment in liver iron overload by magnetic resonance imaging," *Magnetic Resonance Imaging*, vol. 27, no. 2, pp. 188–197, 2009.
- [30] H. E. Daldrup-Link, "Ten things you might not know about iron oxide nanoparticles," *Radiology*, vol. 284, no. 3, pp. 616–629, 2017.
- [31] R. Kuhlperter, H. Dahnke, L. Matuszewski et al., "R2 and R2* mapping for sensing cell-bound superparamagnetic nanoparticles: In vitro and murine in vivo testing," *Radiology*, vol. 245, no. 2, pp. 449–457, 2007.
- [32] C. Burtea, S. Laurent, D. Crombez et al., "Development of a peptide-functionalized imaging nanoprobe for the targeting of (FXVD2) γ a as a highly specific biomarker of pancreatic beta cells," *Contrast Media & Molecular Imaging*, vol. 10, no. 5, pp. 398–412, 2015.
- [33] X. You, R. Tu, and M. Peng, "Molecular magnetic resonance probe targeting VEGF165: preparation and in vitro and in vivo evaluation," *Contrast Media & Molecular Imaging*, vol. 9, pp. 349–354, 2014.
- [34] H. Dahnke and T. Schaeffter, "Limits of detection of SPIO at 3.0 T using T2* relaxometry," *Magnetic Resonance in Medicine*, vol. 53, no. 5, pp. 1202–1206, 2005.



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