

Review Article

The Signaling Pathways Involved in the Antiatherosclerotic Effects Produced by Chinese Herbal Medicines

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Cardiovascular diseases (CVDs) are considered to be the predominant cause of death in the world. Chinese herb medicines (CHMs) have been widely used for the treatment of CVDs in Asian countries for thousands of years. One reason of high efficacy of CHMs in treating CVDs is attributed to their inhibition in atherosclerosis (AS) development, a critical contributor to CVDs occurrence. Cumulative studies have demonstrated that CHMs alleviate atherogenesis via mediating pathophysiologic events involved in AS. However, there is deficiency in the summaries regarding antiatherogenic signal pathways regulated by CHMs. In this review, we focus on the signal cascades by which herb medicines and relevant extractives, derivatives, and patents improve proatherogenic processes including endothelium dysfunction, lipid accumulation, and inflammation. We mainly elaborate the CHMs-mediated signaling pathways in endothelial cells, macrophages, and vascular smooth muscle cells of each pathogenic event. Moreover, we briefly describe the other AS-related factors such as thrombosis, autophagy, immune response, and noncoding RNAs and effects of CHMs on them in the way of cascade regulation, which is helpful to further illustrate the molecular mechanisms of AS initiation and progression and discover newly effective agents for AS management.

1. Introduction

Cardiovascular diseases (CVDs) are the most common cause of health loss at home and abroad, by the fact that more than 13 million patients die from CVDs annually [1]. It is demonstrated that atherosclerosis (AS) is the pivotal pathological basis of CVDs. AS, characterized by formation of atherosclerotic plaques in the artery intima, could induce lumen stenosis or occlusion, finally leading to the occurrence of CVDs [2]. Thus, in order to reduce the prevalence of life-threatening CVDs, especially ischemic heart disease and stroke, the prevention and treatment of AS are of vital importance.

Over the past years, several drugs have been developed as therapeutic agents for AS and the representative one is the statin. However, there is evidence indicating that statin therapy is unable to decrease CVD risks in the majority of patients [3]. Moreover, liver dysfunction and

myopathy, which are potentially adverse effects of statin application, make several patients stop receiving statin therapy, especially for those suffering hepatitis [4, 5]. It is urgent to explore alternative and complementary options with high efficiency and less side effects for AS management.

With a holistic and synergistic way, Chinese herbal medicines (CHMs) keep the balance of homeostasis in vivo. It is reported that a variety of herbal drugs and their extractives such as flavonoid, alkaloid, and terpenoid and patent products possess superior pharmacological properties in the prophylaxis and treatment of AS. Considering the effective clinical application of CHMs (Table 1), a plenty of studies have concentrated on the mechanisms of action underlying therapeutic effects for AS [6–8]. In this review, we will focus on the relevant signaling pathways modified by which CHMs exert beneficial effects in AS prevention and therapy.

TABLE 1: The classification of compounds from CHMs with anti-AS roles.

Category	Compound
Terpenoid	Saikosaponin-a, tanshinone IIA, tanshindiol C, ginkgolide B, andrographolide, paeoniflorin, cryptotanshinone, danshenol A, dihydrotanshinone I, celastrol, 1, 6-di-O-caffeoyl- β -D-glucopyranoside, atractylenolide, β -Elemene
Saponin	Xinxuekang, compound K, ginsenoside Rb1, gypenoside XVII, Ginsenoside F1, glycyrrhizic acid, Diosgenin, Elatoside C, Celosin
Alkaloid	Berberine, trichosanatine, ligustrazine, coptisine,
Flavonoid	Kuwanon G, myricitrin, dihydromyricetin, isoquercitrin, icariin, apigenin, isohamnetin, baicalin, hydroxysafflor yellow A, hyperoside, quercetin, wogonin, procyanidin, dracocephalum moldavica, rumex acetose L, delphinidin-3-glucoside, gossypetin
Isoflavonoid	Puerarin, Biochanin-A
Phenolic	Danshensu, paeonol, salvianolic acid B, luteolin
Stilbenoid	Pterostilbene, resveratrol
Iridoid	Geniposide, Genipin
Diarylheptanoid	Curcumin

2. Mechanisms of Action of CHMs

During the development of atheroma, multiple cells in the lesion environment including endothelial cells (ECs), macrophages, vascular smooth muscle cells (VSMCs), platelets, and lymphocytes are involved [9, 10] (Figure 1). CHMs have been shown to target specific signaling cascades in these cells to generate antiatherogenic effects; the detailed information will be discussed below.

2.1. Amelioration of Lipid Metabolism Disorder

2.1.1. Triterpenoid. Reverse cholesterol transport (RCT) is a cholesterol metabolism process through regulating the efflux of cholesterol from lipid-laden foam cells back to liver for recycling or excretion, which is mediated by signal molecules such as ATP-binding cassette transporters A1 and G1 (ABCA1 and ABCG1), peroxisome proliferator-activated receptor γ (PPAR- γ), and liver X receptor α (LXR- α) [11]. One pivotal antiatherogenic mechanism of Saikosaponin-a (Ssa) is attributable to the promotion of signal transduction of PPAR- γ /LXR- α /ABCA1 cascade, stimulating the outflow of cholesterol in macrophages [12] (Figure 2), while, with a LXR- α independent way, Tanshinone IIA (Tan IIA) increases the level of ABCA1 and ABCG1 by facilitating extracellular signal-regulated kinase (ERK)/nuclear factor-erythroid 2-related factor 2 (Nrf2)/heme oxygenase-1 (HO-1) axis [13]. In the presence of Tanshindiol C (Tan C), the content of lipids in macrophages stimulated by oxidized low density lipoprotein (ox-LDL) is markedly reduced, which is attributed to the drug-triggered activation of Nrf2 and Sirtuin 1 (SIRT1) and downstream peroxiredoxin 1/ABCA1 pathway [14].

Several studies report that PPAR- γ is response for cluster of differentiation (CD) 36 expression regulated by ox-LDL and Tan IIA inhibited cholesterol ingestion via suppressing PPAR- γ which transcriptional activates CD36 expression [15]. Moreover, ox-LDL uptake by lectin-like ox-LDL receptor-1 (LOX-1) induces production of reactive oxygen species (ROS) followed by nuclear factor κ B (NF- κ B) activation and subsequent LOX-1 expression, resulting in the

positive feedback of cholesterol inflow in macrophages [16]. It is demonstrated that another mechanism underlying Tan IIA ameliorates atherogenesis to inhibit ox-LDL-triggered ROS/NF- κ B/LOX-1 loop [17]. Besides, ERK/Nrf2/HO-1 pathway activated by Tan IIA enables blocking the activity of activator protein-1 (AP-1) which mediates scavenger receptor-A (SR-A) expression, reducing SR-A-regulated cholesterol influx [13] (Figure 2).

It is documented that Tan IIA induces LDL receptor (LDL-R) production and increase LDL uptake in hepatic cells through stimulating sterol regulatory element-binding protein (SREBP) 2 pathway which mediates LDL-R expression and raising the nuclear Forkhead box O3a (FoxO3a) cascade which inhibits the expression of proprotein convertase subtilisin/kexin type 9 promoting LDL-R degradation [18].

2.1.2. Phenolic Compound. Danshensu (DSS) could afford a cholesterol-lowering role in macrophages by virtue of stimulating the PPAR- γ /LXR- α /ABCA1 pathway [19]. Similarly, DSS derivative (DBZ) and paeonol ameliorate foam cell formation and enhance cholesterol efflux via activation of LXR- α and upregulation of ABCA1 [20, 21]. Moreover, it is proved that DBZ reduces foam cell formation via inhibiting macrophage lipid accumulation by suppressing Toll-like receptor 4 (TLR4)/NF- κ B/adipose differentiation-related protein (ADRP) cascade [22].

2.1.3. Diarylheptanoid. By impeding the activation of ox-LDL evoked p38 MAPK (p38)/PPAR- γ /CD36 cascade, curcumin reveals similar effects in diminishing ox-LDL-upregulated CD36 level [23]. Liu et al. had shown that curcumin was a cholesterol efflux promoter through activating LXR- α and then ABCA1 upregulation [24].

2.1.4. Alkaloid. Berberine (BBR), a kind of cholesterol-lowering herb extractive, activates ERK1/2 to stabilize LDL-R mRNA, leading to upregulation of LDL-R protein and decrease of serum LDL [25]. Additionally, various CHMs attenuate atheroma formation depending on blockade of triglyceride

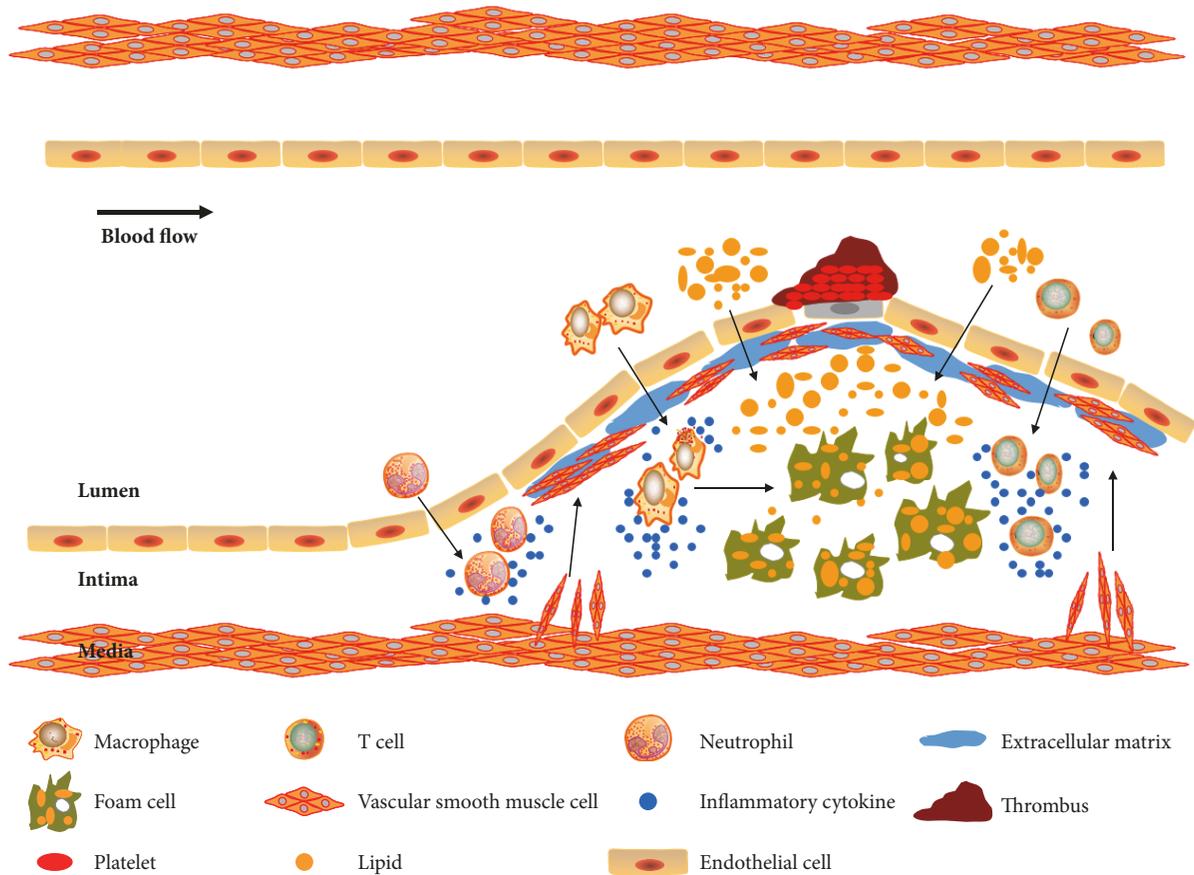


FIGURE 1: The pathogenesis of atherosclerosis. The endothelial dysfunction, inflammation, and lipid dysbolism induce excessive cholesterol disposition and leukocytes accumulation into the vascular intima. Then macrophages and neutrophil secrete cytokines and T cells release immune regulatory factors, which promote AS progression. Moreover, macrophages uptake cholesterols and transform into foam cells, facilitating the formation of lipid core. VSMCs migrate to the subendothelium and release extracellular matrix, leading to the formation of fibrous cap and vascular remodeling. In addition, platelets are activated and aggregate to the injured vascular endothelium, contributing to the thrombosis.

synthesis in hepatocytes. BBR and ginsenosides metabolite compound K (CK) have been proved to stimulate liver kinase BI/AMP-activated protein kinase (AMPK) signaling flow to phosphorylate acetyl-CoA carboxylase (ACC) and inhibit SREBP-1c/fatty acid synthase (FAS) axis, which followed by reduction of lipogenesis [26–28].

2.1.5. Saponin. The liver exerts critical functions in the process of cholesterol synthesis and triglyceride generation and is the primary target organ of RCT. High-density lipoprotein (HDL), reversely associated with AS development, is response for transport of effluent cholesterol from peripheral tissues to the liver for removing. Di'ao Xinxuekang (XXK), saponin extractives of *Dioscorea panthaica* Prain et Burkill, is reported to enhance HDL generation by promoting PPAR- γ /LXR- α /ABCA1 pathway by which XXK improves the RCT process and alleviates atherosclerotic lesions [11] (Figure 3).

2.1.6. Flavonoid. It has been confirmed that Kuwanon G is indicated to accelerate cholesterol elimination in

macrophages by enhancing the signal transduction of LXR- α /ABCA1 cascade [29]. Dong et al. reported that the hawthorn leave flavonoids (HLF) triggered AMPK/PPAR- α /carnitine palmitoyl transferase 1 axis to increase the oxygenolysis of fatty acids, then reducing the generation of triglyceride.

Furthermore, cumulative data indicate that other CHMs improve dyslipidemia via diverse pathways. For instance, Qishen Yiqi pill (QSYQ) accelerates the excretion of bile acids to facilitate serum cholesterol uptake by liver via activating LDL-R/LXR- α /ABCG5 pathway [30].

2.2. Improvement of Cell Apoptosis

2.2.1. Saponin. Biological molecules including ox-LDL, TNF- α , and Ang II are the major driving forces in endothelial dysfunction linked to atherogenesis, through stimulating NADPH oxidase (NOX) and disrupting mitochondria respiration to generate excessive ROS [31]. Cellular analysis illuminates that ginsenoside Rb1 (Rb1) prevents ECs from

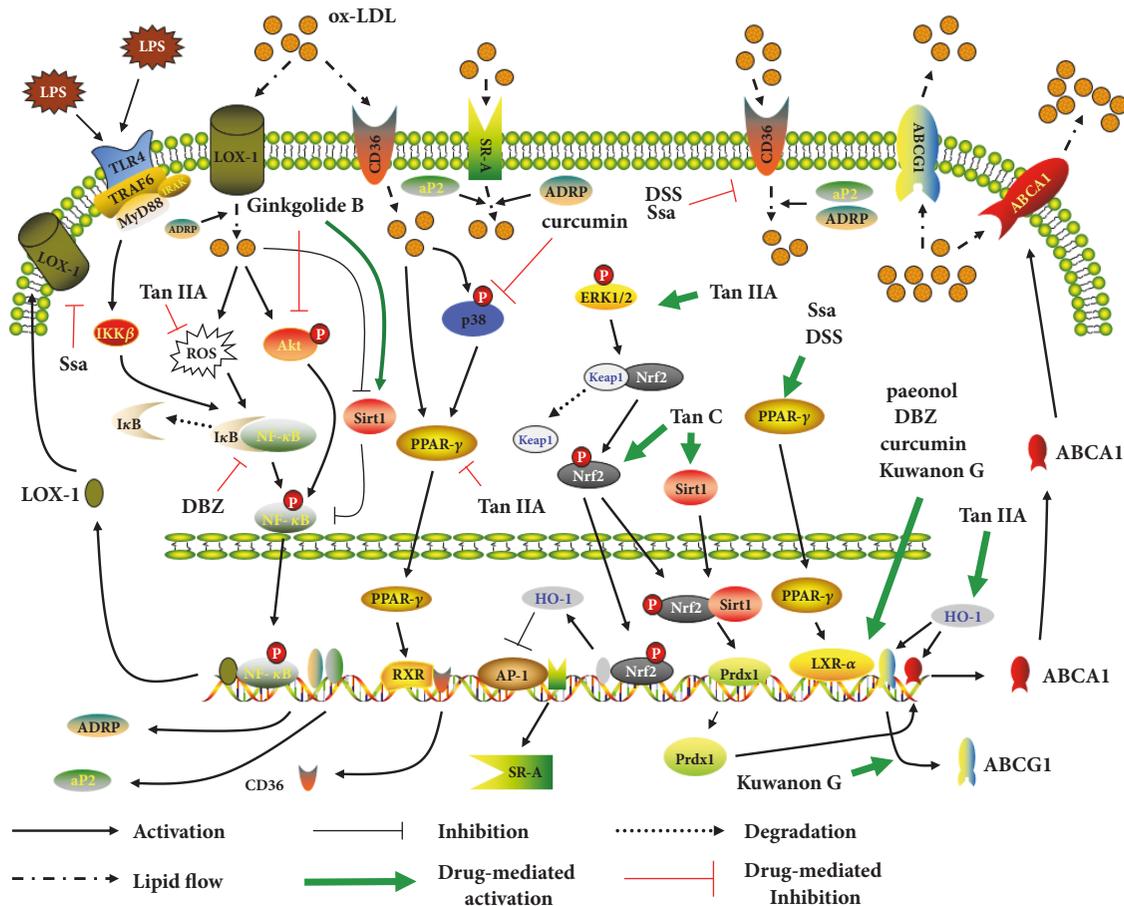


FIGURE 2: The signaling pathways by which CHMs alleviate lipid accumulation in macrophages.

TNF- α -induced insult via disturbing mitochondrial pathway of apoptosis, given that increment of Bcl-2/Bax ratio and caspase-3 evoked by TNF- α /c-Jun N-terminal kinase (JNK)/NF- κ B axis is inhibited by Rb1 [32] (Figure 4).

Considerable evidence has illustrated pivotal actions of PI3K/Akt cascade in cell survival. Gypenoside XVII (GP-17) is thought to directly suppress the apoptotic pathway by activating PI3K/Akt accompanied by Bad dysfunction [33].

2.2.2. Flavonoid. It is demonstrated that p53 upregulated by ROS could aggravate mitochondrial apoptosis, because that p53 elevation facilitates Bax increase and Bcl-2 reduction. The protective ability of dihydromyricetin (DMY) against ECs apoptosis induced by H₂O₂ is partly attributed to inhibition of ROS-activated p53 and then improvement of imbalance of Bcl-2/Bax ratio, cyt-c release, and caspase-3 activation [34]. In addition, on stimulation of PI3K/Akt, isoquercitrin stimulates, resulting in GSK3 β phosphorylation accompanied by Mcl-1 activation which blocks apoptosis of ECs [35]. Laboratory studies suggest that DMY, myricitrin, and GP-17 are able to alleviate ox-LDL-induced ECs apoptosis by activating PI3K/Akt/Nrf2/HO-1 pathway, which enhance intracellular antioxidative abilities to eliminate ROS [33, 36, 37]. Owing that endothelial NO synthase- (eNOS-) synthesized nitric oxide (NO) plays vital roles in maintaining

the integrity of vascular endothelium, CHMs like icariin and Wenxin decoction (WXD) are found to phosphorylate eNOS and release NO by inducing PI3K/Akt signaling for mitigating atherogenic endothelial injury [38, 39].

Zeng et al. supported that ox-LDL maintained the survival of macrophages by upregulating antiapoptotic protein plasminogen activator inhibitor 2 (PAI-2) and apigenin exhibited proapoptotic effects on macrophages via inhibiting Akt/PAI-2 cascade [40]. However, there are clues showing that isohamnetin and Danshen granule are able to alleviate atheroma progression by inhibiting macrophages apoptosis via activating the PI3K/Akt/Nrf2/HO-1 axis [41, 42].

2.2.3. Diterpenoid. Chen et al. showed that andrographolide (Andro) was capable of triggering PI3K/Akt and subsequent Bad inhibition, which impeded the activation of mitochondrial apoptotic pathway and then maintained the survival of ECs [43]. Additionally, in vitro studies report that Tan IIA and flavonoid myricitrin administration maintain the survival of ECs via retarding p53 expression, which hinder the pathway of H₂O₂/ROS/p53/caspase-3 [44, 45].

2.2.4. Phenolic and Alkaloid. Paeonol and trichosanthin protect against ox-LDL-triggered ECs injury through abating

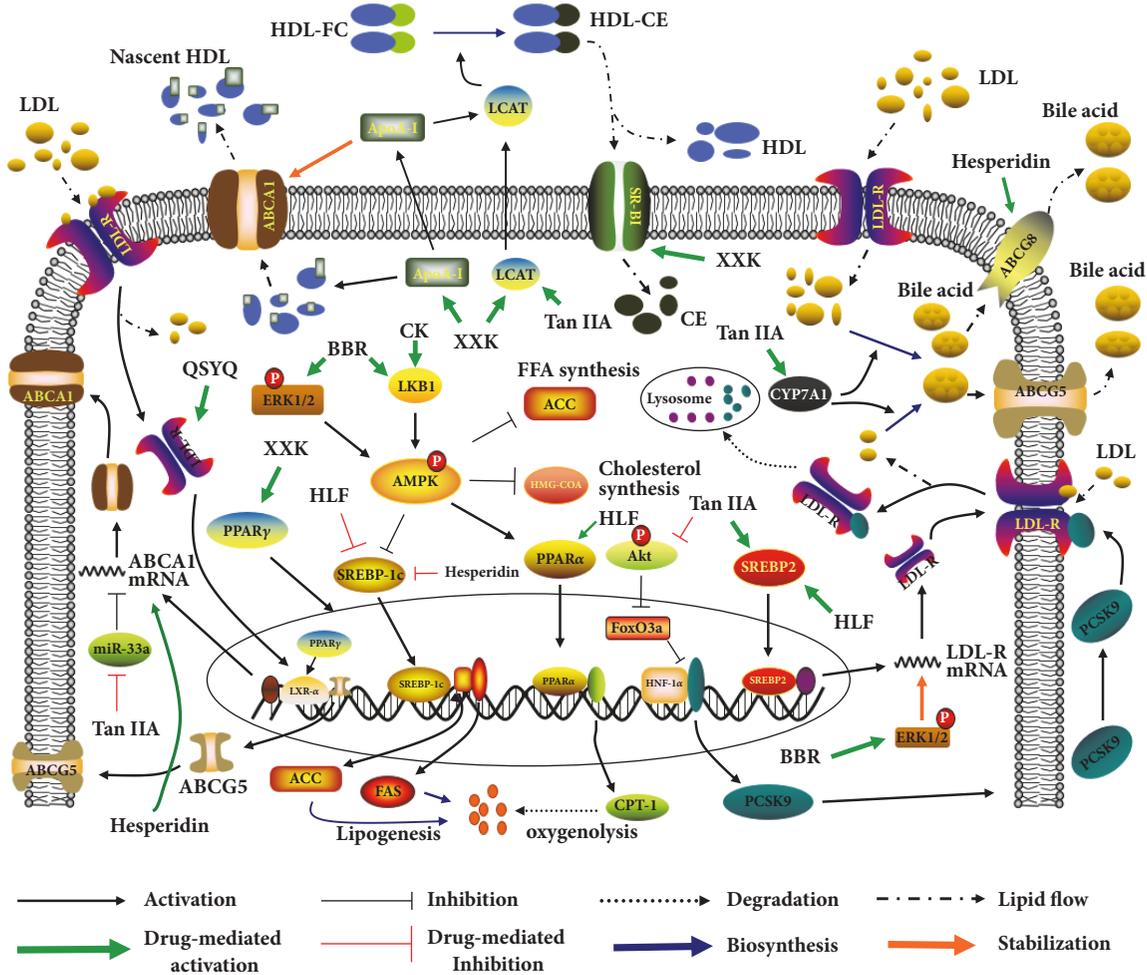


FIGURE 3: The pathway mechanisms underlying CHMs improve lipid metabolism in liver cells.

LOX-1 expression followed by suppression of ox-LDL/LOX-1/ROS/p38 signal axis [46, 47]. One principle of paeonol and Liuwei Dihuang (LWDH) attenuated endothelial dysfunction is inhibiting ER stress, followed by decrease of unfolded protein response and restoration of C/EBP homologous protein and NADPH level, resulting in reduction of ROS production [48–50]. Moreover, MEK/ERK1/2/eNOS and AMPK/PPAR- δ /eNOS cascades are also likely to be applied by herb drugs to produce NO, normalize ROS, and thus abolish oxidative stress-induced ECs apoptosis [38, 48].

2.3. Mediation of Cell Proliferation and Migration

2.3.1. Saponin. Mounting data have confirmed the involvement of ECs in angiogenic process which is critical for accelerating AS development and exacerbating plaque vulnerability [51]. Antiangiogenic functions of CK are associated with activation suppression of p38 and Akt, which probably lead to decrease of proliferation proteins cyclin D1 and VEGF in ECs [52]. Yun et al. demonstrated that Panax notoginseng saponins (PNS) ameliorated plaque angiogenesis via

reducing level of NOX4, resulting in decrement of ROS generation and subsequent VEGF expression [53].

2.3.2. Diterpenoid. It is indicated that aberrant proliferation and migration of VSMCs aggravate atheroma progression and restenosis after balloon angioplasty [54]. Tan IIA has been reported to improve the activation of AMPK/p53/p21 axis to inhibit the expression of cyclin D1 stimulated by high glucose, finally alleviating the proliferation of VSMCs [55]. Wu et al. report that the antimigratory action of Tan IIA on VSMCs occurs by the increase of AMPK activity and subsequent block of NF- κ B cascade, which lower matrix metalloproteinase- (MMP-) 2 expression [55]. Additionally, suppressing TNF- α -activated MEK1/2/ERK1/2/AP-1 and Akt/IKK/NF- κ B cascade is another mechanism underlying Tan IIA reducing MMP-9 induction to decrease the movement of VSMCs [56]. Suh et al. offered evidence that cryptotanshinone (CTS) reduced MMP-9 level through inhibiting TNF- α -induced signal pathway involving in ERK1/2, p38, and JNK and then inactivation of AP-1 and NF- κ B [57].

Furthermore, there is evidence showing that Tan IIA is able to provide protective roles against atherogenesis by

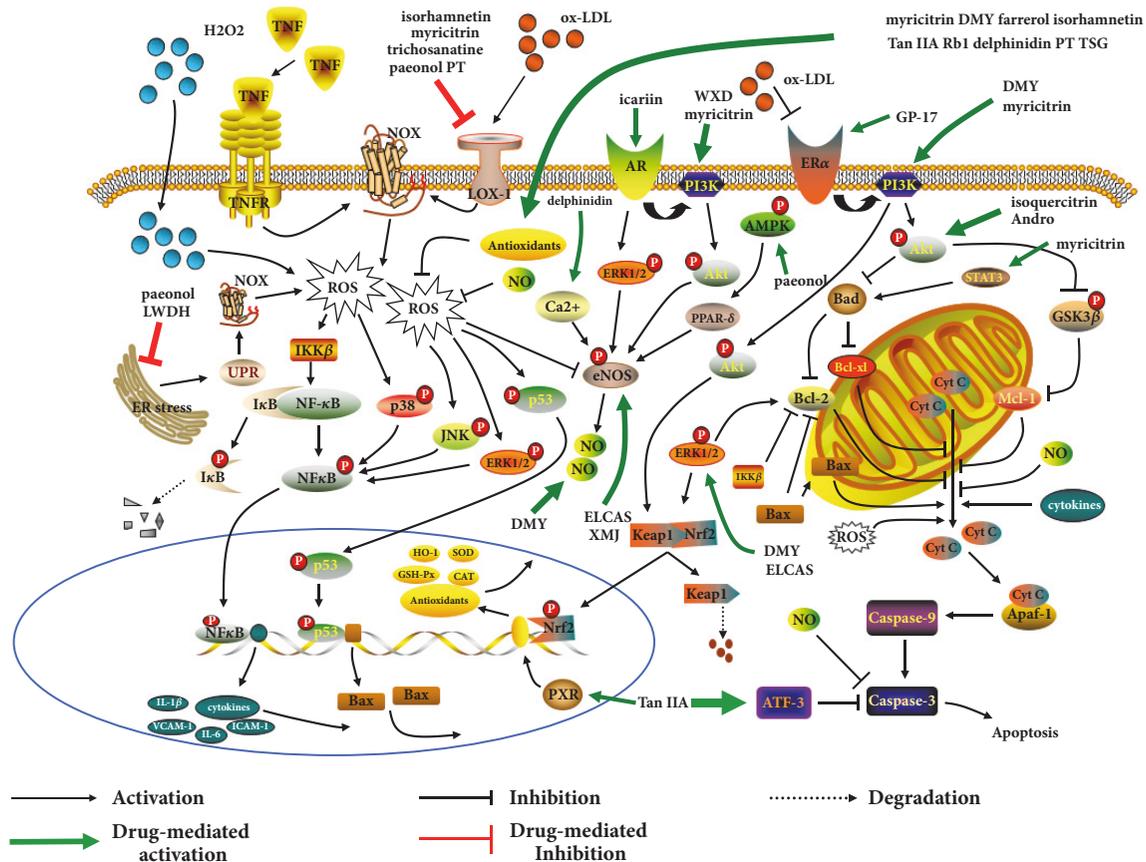


FIGURE 4: The effects at molecular level produced by CHMs in the attenuation of ECs apoptosis. ELCAS: Ligusticum chuanxiong and Angelica sinensis; XMJ: Xinmaiija; WXD: Wenxin decoction; and TSG: 2,3,5,4'-Tetrahydroxystilbene-2-o-β-D-glucoside.

blocking ECs growth via disrupting the vascular endothelial growth factor (VEGF)/VEGF receptor 2 (VEGFR2) axis [58].

2.3.3. Flavonoid. Dong et al. clarified that baicalin obviously inhibited platelet-derived growth factor- (PDGF-) stimulated proliferation of VSMCs, the mechanism of which was blocking signal pathway of PDGF receptor β (PDGFR β)/MEK/ERK1/2, followed by decrease of cyclin E and cyclin-dependent kinase 2 (CDK2) activity and increase of p27 level [59]. Icariin, hyperoside, and monoterpene compound paeoniflorin prevent VSMCs from ox-LDL-stimulated proliferation probably by abrogating LOX-1 expression, ROS generation, and ERK1/2 activation essential for mitogen-related cascade flow [60–62].

2.3.4. Alkaloid. With the modulation of cascades linked to cell growth including Ras/Rac1, AMPK/p53/p21, and MEK/ERK1/2/Egr-1(c-fos), BBR lowers the expression of cell cycle-related proteins crucial for VSMCs proliferation [63, 64]. There are observations suggesting that the molecular basis for the antiproliferative effects of CHMs including alkaloid ligustrazine and flavonoid hydroxysafflor yellow A is that these herbs inhibit PDGF-induced activation of signal factors involved in p38, ERK1/2, and Akt pathway, leading

to the expression depression of downstream cell cycle-associated molecules including cyclinD1, cyclinE, CDK2, and CDK4. In addition, ligustrazine and hydroxysafflor yellow A also enhance NO generation to elevate cGMP for decaying VSMCs growth, given the evidence of cGMP identified as a mitogenic suppressor [65, 66]

2.3.5. Phenolic. Salvanolic acid B (Sal B) obviously attenuates LPS-modulated upregulation of MMP-2 and MMP-9, which might be ascribed to suppression of LPS-induced signaling of TLR4/MyD88, resulting in disorders of downstream pathways including ERK1/2, JNK, and COX-2 responsible for MMPs expression [67]. Moreover, blocking the proliferative signaling of protein kinase C (PKC)/Rac1/ROS and Ras/Raf/ERK1/2 is the potential mechanism for herb drugs like paeonol and isoflavonoid puerarin to extenuate diabetes-induced intimal hyperplasia [68, 69].

2.3.6. Other Groups. Sparstolonin B, an isocoumarin compound, induces cell cycle arrest at G1 phase in ECs and significantly inhibits cell growth and vasculogenesis, and the mechanism might result from inactivation of NF- κ B and thereby decrease of cell cycle-promoting proteins [70]. Stimulating Nrf2-related pathway, antrodia salmonea enhances HO-1 and glutathione expression, favoring the scavenging

of TNF- α -induced ROS, causing blockade of ROS-mediated I κ B kinase (IKK)/NF- κ B/MMP-9 signal transduction and thereby decrease of MMP-9 level [71]. In addition, curcumin, a kind of diarylheptanoid, has antimigratory roles in VSMCs by blocking MMP-9 and MMP-13 production in macrophages via suppressing AMPK/MAPKs and PKC cascade [72]. Moreover, Tongxinluo (TXL), widely used for treating CVDs, has been shown to reduce plaque burden and angiogenesis by decreasing angiogenic factor such as MMP-2 and VEGF through inactivating TNF- α /bone marrow kinase in chromosome X/NF- κ B/MAPKs pathway [73].

2.4. Inflammation Amelioration

2.4.1. Diterpenoid. Upon induction of proinflammatory substances, the expressions of adhesion molecules and chemokines of ECs are elevated, favoring macrophages attachment and transmigration into subendothelium, then promoting AS lesions [9]. CTS encumbers expression of vascular cell adhesion molecule-1 (ICAM-1), intercellular adhesion molecule-1 (VCAM-1), and E-selection forced by ox-LDL on ECs via suppressing NOX4 to abrogate ox-LDL-induced ROS generation and sequential IKK β phosphorylation, I κ B degradation, and NF- κ B activation, and by restoring eNOS activity to produce NO [74]. Danshenol A and dihydrotanshinone I, other components isolated from *Salvia miltiorrhiza* Bunge, also lower macrophage adhesion to ECs triggered by LPS and TNF- α via hindering NOX4/IKK β /NF- κ B pathway [75, 76]. Moreover, Tan IIA perform antiatherogenic properties to lessen expression of adhesion molecules and chemokines in ECs, depending on modulating key signaling cascades containing Rho/Rho kinase, PI3K/Akt, Jak/STAT-3, and Wnt pathways [77–81].

In terms of inflammation attenuation of macrophages, CTS and Tan IIA are able to lessen synthesis and release of proinflammatory factors by mediating multitargets in pathways including TLR4/IKK/NF- κ B and TLR4/MAPKs [82–85]. It is illustrated that compounds from Danshen could activate the cascade of PI3K/Akt/Nrf2/HO-1 in macrophages to enhance the generation of CO that weaken NF- κ B activity and AS development [86–88].

In VSMCs, Andro counteracts LPS/IFN- γ -elicited upregulation of iNOS and MMP-9; the mechanism underlying is the enhancement of nSMase/ceramide/PP2A cascade that abolish LPS/IFN-induced IKK/NF- κ B axis [89]. Moreover, Chen et al. showed that the relief of TNF- α -stimulated NF- κ B activation and then iNOS expression was due to Andro-regulated inflammation restriction in VSMCs by virtue of inhibiting JNK/Akt pathway [90].

2.4.2. Saponin. Ginsenoside F1 could upregulate zinc finger protein A20 to weaken ox-LDL-elicited LOX-1/ROS/NF- κ B pathway for lowering levels of ICAM-1, MIP-1 δ , and IL-1 α [91]. The inducible expression of VCAM-1 in response to ox-LDL is abolished in ECs with PNS and Rb1 preconditioning; this effect is primarily due to activation of Nrf2 followed by elevation of HO-1 and superoxide dismutase, causing intervention of ROS/TNF- α /p38 cascade [92]. Yu et

al. demonstrated that the therapeutic utility of XXX in AS plaques had been partially ascribed to the anti-inflammatory ability of blocking transduction of TLR4/MyD88/IKK/NF- κ B cascade in macrophages [93]. Moreover, Diosgenin preconditioning diminishes level of adhesion molecules in VSMCs exposed to TNF- α via restraining the ROS/MAPKs and ROS/Akt pathway and downstream NF- κ B activation [94].

2.4.3. Flavonoid. It is deciphered that quercetin has pharmacological efficacies on diminishing level of proinflammatory cytokines through prohibiting TLR/NF- κ B signal pathway in ECs [95]. Wogonin and Bushenningxin decoction have been indicated to activate the estrogen receptors on ECs to upregulate eNOS expression and heighten NO synthesis, leading to blockade of proinflammatory NF- κ B signaling [96, 97]. Additionally, Lee et al. reported that wogonin was effective in ameliorating inflammation state in macrophages by suppressing Ca²⁺/STAT signal axis in macrophages, as seen by decrease of cytokines including IL-1 β , MCP-1, and MIP-1 α [98]. Disrupting transcription activity of p50-p65 heterodimer, procyanidins decline macrophage-produced IL-6 and COX-2 via blocking LPS-initiated TLR4/NF- κ B pathway [99]. Ingredients in *dracocephalum moldavica* normalize the levels of VCAM-1 and ICAM-1 in VSMCs by suppressing TNF- α -triggered NF- κ B signaling [100]. Moreover, icariin reverses LPS-induced NF- κ B activation and cytokines production in macrophages via boosting PI3K/Akt pathway, whereas saponin glycyrrhizic acid and triterpenoid Ssa restrain inflammation by encumbering PI3K/Akt cascade [12, 101, 102]. This discrepancy might be attributed to the complex of signal networks and the difference of compound category, which is needed to be further elucidated.

2.4.4. Alkaloid. The reduced attachment of macrophages to ECs after pretreatment with BBR is due to diminution of ROS-inducible adhesion molecules expression caused by AMPK/nuclear respiratory factor 1/uncoupling protein 2 axis [103]. Wu et al. illustrated that the revulsive expression of IL-6 and iNOS on stimulation with LPS was abolished in macrophages with coptisine coinubation, and the mechanism underlying this effect results from inhibiting MAPKs and Akt signaling and subsequent NF- κ B activation [104].

2.4.5. Isoflavonoid. Biochanin-A, a bioactive isoflavonoid, is shown to reduce LPS-elicited TNF- α , IL-1 β , and IL-6 expression in macrophages through inhibiting TLR4-dependent p38/ATF2 and NIK/IKK/NF- κ B pathway [105]. Furthermore, red clover extracts-enhanced PPAR- α and BBR-activated PPAR- γ have the similar function in terminating transduction of inflammatory pathways, seeing that signaling of PPAR- α and PPAR- γ abolishes I κ B degradation, NF- κ B activation, and binding to DNA regions in macrophages [106, 107].

2.4.6. Monoterpenoid and Triterpenoid. The decrease of IL-6, IL-1 β , and TNF- α production in VSMCs after paeoniflorin treatment is ascribed to alleviation of TLR4/MyD88/NF- κ B cascade [108]. Gu and colleagues found that celastrol

reduced macrophages expression of iNOS, TNF- α , and IL-6 via mitigating ox-LDL-evoked LOX-1/NOX/ROS/NF- κ B cascade [109].

2.4.7. Another Class. Capsaicin augments Ca²⁺ dependent PI3K/Akt/eNOS pathway to boost NO generation, which maintain the stability of I κ B and combination with NF- κ B upon LPS stimulation, leading to levels of ICAM-1, VCAM-1, and MCP-1 back to normal [110]. Meng et al. reported that curcumin exhibited anti-inflammatory actions in VSMCs by abating LPS-provoked TLR4/NOX/ROS cascade accompanied by blockade of ERK1/2 and p38 signaling and NF- κ B inactivation, as explained by decrease of MCP-1 and TNF- α [111].

Daotan decoction has been corroborated to counteract TNF- α -induced ICAM-1 on ECs via multipathway mode, evidenced by encumbrance of p53/p21, JNK, and p38 cascades [112, 113]. Zheng et al. manifested that Longxuetongluo capsule played atheroprotective roles by curtailing contents of COX-2, VCAM-1, and MCP-1 via restraining ox-LDL-provoked ERK1/2(p38)/IKK/NF- κ B signal [114]. Sun et al. showed that Tianxiangdan granule afforded antiatherogenic actions by inhibiting inflammatory p38/NF- κ B signal pathway in ApoE^{-/-} mice [115]. With the encumbrance of IRS-1/PI3K/Akt signal flow, Shenyuandan capsule blocks activation of NF- κ B and then expression of IL-6 and TNF- α in aortas [116]. Thus, the above herb drugs execute anti-AS roles through diverse signal pathways, and NF- κ B represents the convergence of most of these cascades, hinting that NF- κ B acts as a pivotal target of CHMs for AS management.

2.5. Alleviation of Thrombogenesis

2.5.1. Terpenoid. As normal blood flow is decided by platelets, clotting factors, and fibrinolytic molecules, dysfunction of them could facilitate thrombus formation in the site of plaque lesions, contributing to atherogenesis and related complications occurrence. Fu and colleagues find that triterpenoid substances from *Callicarpa nudiflora* hook have antiplatelet roles by eliminating ADP and TXA₂-induced platelet activation and aggregation via inhibiting PI3K/Akt/GSK3 β and RhoA pathway, respectively [117] (Table 2). Chen et al. provided initial proof that Atractylodes lactone compounds were antithrombotic because of their effects on lessening platelet accumulation and secretion triggered by agonists via blocking p38 and Akt signaling [118]. After Andro administration, collagen-induced platelet activation is restrained, the mechanism is Andro-regulated augmentation of eNOS/NO/sGC signal flow, which in turn catalyze synthesis of cGMP required for reducing activities of p38, IKK β , PI3K, and PLC γ 2 implicated in the process of collagen-regulated Ca²⁺ elevation [119, 120]. Investigation into mechanisms reveals that Tan IIA suppresses platelet activation and TF expression via mediating multiple pathways including ER α /PI3K/Akt, ROS/NF- κ B, and ERK-2 [121–123]. Moreover, β -Elemene, which belongs to sesquiterpenoid, is indicated to induce the PI3K/Akt/eNOS pathway to increase NO level, thereby suppressing platelet activation and aggregation [124].

2.5.2. Saponin. Given that inflammatory factors are contributors to upregulation of TF for accelerating AS development, gypenoside XLIX and extracts of red yeast rice restore TF level and weaken AS progression via impairing NOX/ERK1/2/NF- κ B and boosting PPAR- α pathway, respectively [125, 126]. Pan et al. stated that Rb1 and Rg1 encumbered platelet accumulation and thrombosis via increasing NO synthesis by triggering PI3K/Akt and CAT-1/L-arginine cascade [127].

2.5.3. Other Compounds. Other CHMs like lignin Gomisin J, flavonoid rumex acetose L, and isoflavonoid puerarin also enhance eNOS activity and NO generation by means of mediating diverse cascades including Ca²⁺/CaMI, ER/PI3K/Akt, and CaMKII/AMPK, indicating their implication in encumbering thrombosis and AS progression [128–130]. Sal B is shown to prevent TNF- α -dependent ERK1/2/AP-1 and NF- κ B cascade, leading to decrease of PAI-1 level and restoration of malfunction of fibrinolytic system [131].

2.6. Improvement of Autophagy

2.6.1. Flavonoid. It is well established that autophagy exerts vital roles in regulating endothelial function, macrophage lipid metabolism, VSMCs phenotypic conversion, thrombosis, and angiogenesis which are involved in atheroma development [132, 133]. Jin et al. provided evidence that enhanced autophagy, resulting from AMPK/SIRT1 signal pathway induced by delphinidin-3-glucoside, attenuated ox-LDL-elicited injury in ECs [134] (Table 3). Gossypetin, a type of flavonoid, effectively weakens ox-LDL-caused ECs damage and this phenomenon is explained by drug-mediated inhibition of class I PI3K/Akt cascade and activation of class III PI3K/Beclin-1/microtubule associated protein light chain 3 (LC3) pathway, thus leading to upregulation of autophagy [135].

2.6.2. Stilbenoid. The involvement of autophagy in pterostilbene- (PT-) mitigated ECs apoptosis is crucial, because that Ca²⁺/CaMKK β /AMPK pathway induced by PT reduces TUNEL labeled cells [136]. Resveratrol, classified as a autophagy activator, is capable of boosting autophagic processes to ameliorate inflammation and injury in ECs elicited by TNF- α and ox-LDL, and the mechanism is attributed to accentuation of cAMP/AMPK/SIRT1 cascade followed by elevation of LC3II and reduction of p62 [137, 138].

2.6.3. Alkaloid and Saponin. BBR is identified to suppress level of inflammatory factors in macrophages by activating AMPK, which blocks the autophagy inhibitor mammalian target of rapamycin (mTOR), leading to initiation of autophagy responsible for inhibiting NF- κ B activity [139]. Inducing ROS to restrain PI3K/Akt/mTOR cascade, BBR-mediated sonodynamic therapeutics contribute to the autophagic processes which raise ABCA1 expression, favoring the inhibition of cholesterol uptake in macrophages [140]. Furthermore, ECs apoptosis promoted by ox-LDL is abolished in the presence of increase of Beclin-1 and LC3II

TABLE 2: The signal pathways underlying CHMs inhibit the thrombosis.

Ingredient	Herb medicine	Object	Stimulus	Role	related pathway
1, 6-di-O-caffeoyl- β -D-glucopyranoside	Callicarpa nudiflora Hook	Platelet	ADP, AA	α Ib β 3, 5-HT, TXA2, platelet aggregation \downarrow	PI3K/Akt/GSK3 β , RhoA
Atractylenolide	Atractylodes macrocephala	Platelet	ADP, collagen, thrombin	platelet aggregation and secretion \downarrow	p38, PI3K/Akt
Andrographolide	Andrographis paniculata	Platelet	Collagen	platelet aggregation, Ca ²⁺ , TxB2 \downarrow	eNOS/NO/sGC/cGMP, PI3K/Akt/p38/cPLA2, PLC γ 2/DAG/PKC
Gomisin J	Schisandra chinensis	EC	None	eNOS, NO \uparrow	eNOS/NO/sGC/cGMP, p38/ROS/IKK β /NF- κ B/ERK2
β -Elemene	Curcuma Wenyujin	EC	None	eNOS, NO \uparrow	Ca ²⁺ /CaMI, PI3K/Akt
Puerarin	Pueraria lobata	EC	TNF- α	eNOS, NO \uparrow	ER/PI3K/Akt, CaMKII/AMPK
Tanshinone IIA	Salvia miltiorrhiza Bunge	Macrophage	Ox-LDL	TF \downarrow	ROS/NF- κ B
		Platelet	None	Platelet activation \downarrow	ER α /PI3K/Akt
		Platelet	ADP	Platelet activation \downarrow	ERK-2
Xuezhikang	Red yeast rice	Macrophage	Ox-LDL	TF \downarrow , SOD \uparrow	NOX/ROS/ERK1/2/NF- κ B
Gypenoside XLIX	Gynostemma pentaphyllum	Macrophage	LPS	TF \downarrow	PPAR- α
Salvianolic acid B	Salvia miltiorrhiza Bunge	EC	TNF- α	PAI-1 \downarrow	ERK1/2/AP-1 (NF- κ B)

TABLE 3: The signal pathways responsible for CHMs-induced regulation of autophagic processes.

Agent	Herb medicine	Object	Stimulus	Role	related pathway
Delphinidin-3-glucoside	Grape seed	EC	Ox-LDL	Cell viability \uparrow , apoptosis \downarrow ; LC3II \uparrow , p62 \downarrow	AMPK/SIRT1
Gossypetin	Hibiscus	EC	Ox-LDL	LDH, cleaved caspase-3 and PARP-1 \downarrow ; LC3II and Beclin-1 \uparrow , p62 \downarrow	PTEN/class I PI3k/Akt, class III PI3K/Beclin-1
Pterostilbene	Blueberry	EC	Ox-LDL	TUNEL-positive cell \downarrow ; LC3II \uparrow , p62 \downarrow	Ca ²⁺ /CaMKK β /AMPK/mTOR
Resveratrol	Grape	EC	TNF- α	ICAM-1, COX-2, MMP-9 \downarrow ; LC3II \uparrow , p62 \downarrow	ATP/cAMP/AMPK/SIRT1
		EC	Ox-LDL	Cell vability and SOD \uparrow ; LC3II/LC3I \uparrow , p62 \downarrow	AMPK/SIRT1
Elatoside C	Aralia elata Seem	EC	Ox-LDL	TUNEL-positive nuclei, Bax, caspase-9 and -3, ROS \downarrow , Bcl-2 \uparrow ; LC3II and Beclin-1 \uparrow , p62 \downarrow	FoxO1/Beclin-1, LOX-1/NOX/ROS/Caspase
Berberine	Coptis chinensis	macrophage	Ox-LDL	MIP-1 α , RANTES \downarrow , IL-10 \uparrow ; LC3II/LC3I \uparrow , p62 \downarrow	AMPK/mTOR
			None	ABCA1, ROS \uparrow ; LC3II/LC3I \uparrow , p62 \downarrow	PI3k/Akt/mTOR
Arglabin	Artemisia glabella	macrophage	LPS	IL-1 β , IL-18 \downarrow ; LC3II \uparrow	unknown
Celosins	Celosia argentea L.	macrophage	Ox-LDL	CD36, SR-A1 \downarrow , ABCA1, ABCG1 \uparrow ; LC3II/LC3I \uparrow , Beclin-1 \uparrow	unknown

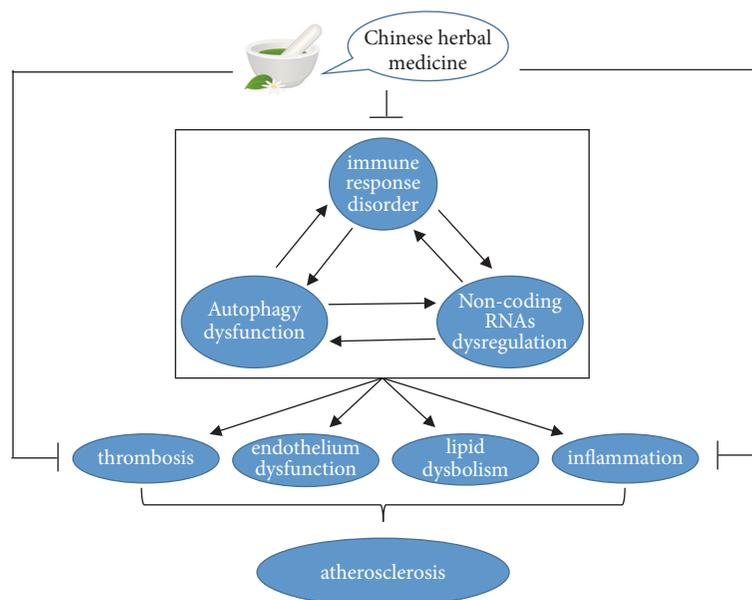


FIGURE 5: The schematic flowchart of diverse pathogenic mechanisms of AS and the intervention exerted by CHMs.

modulated by elatoside C-induced FoxO1 overexpression [141]. With autophagy activation, celastrol and polyphenolic luteolin also alleviate lipid accumulation in macrophages and then restrain AS expansion [142, 143].

2.7. Other Related Mechanisms

2.7.1. Modulation of Immune Response. Cumulative papers attempt to comprehend how immune system participates in the pathogenic processes of atherogenesis, as innate and adaptive immunity are validated to be correlated with all stages of AS [144]. It is confirmed that some CHMs impede atherogenesis via mediating immune response. For example, TXL is shown to induce the regression of AS, at least partly via inhibiting ox-LDL-evoked DCs maturation, as illustrated by reduction of membrane CD40, CD86, and CD1a [145]. Moreover, one property of baicalin and geniposide ameliorating AS is ascribed to the gathering blockade of DCs in plaque areas that launch proatherogenic immune reactions [146]. Owing that regulatory T cells (Tregs) hinder T helper (Th) cells-induced inflammation, QSYQ and amygdalin increase the content of Tregs in vascular lesions which delay the development of AS [30, 147]. Additionally, QSYQ directly inhibit Th17 cells in AS areas and then lower the release of IL-17, a proatherogenic cytokine [30].

2.7.2. Regulation of Noncoding RNAs. Noncoding RNAs (ncRNAs), a group of RNA molecules mainly containing long noncoding RNAs (lncRNAs) and microRNAs (miRNAs), are capable of affecting pathogenic processes of AS by mediating lipid metabolism, cellular apoptosis and proliferation, inflammations, etc. [148, 149]. Tan IIA accelerates the clearance of cholesterol in the vasculature by abolishing HFD-evoked miR-33a expression to elevate ABCA1 level in the liver, resulting in upregulation of HDL secretion and RCT pathway

[150]. Genipin impedes lipid deposition via boosting level of miR-142a-5p, which in turn lessen the lipogenesis pathway of SREBP-1c/ACC(FAS) in hepatocytes [151]. Following paeonol treatment, a decrease in ECs apoptosis and TNF- α production is seen, probably explained by paeonol-mediated suppression of miR-21/TNF- α axis and subsequent apoptotic cascade [152]. Moreover, TXL mitigates the inflammation via triggering Akt accompanied by reduction of miR-155 and then TNF- α expression [153]. LncRNA TUG1 overexpression is implicated in ECs apoptosis induced by ox-LDL, and tanshinol improves ECs damage via reducing TUG1 level which is followed by upregulation of miR-26a and decrease of TRPC6 responsible for calcium overload [154]. In AngII-stimulated ApoE $^{-/-}$ mice, Xiaoxianggou administration rescue miR-203 downregulation to reduce generation of Ets-2 which potentiates angiogenesis and autoimmunity, causing the regression of AS plaques [155].

3. Conclusions

As a representative of complementary and alternative medicine, CHMs have been prescribed to patients for thousands of years in Asian countries for preventing and treating diseases. In this review, we place the emphasis on the signal pathways by which CHMs produce antiatherogenic functions. We find that several herb medicines could regulate one signal pathway to provide multiple roles against AS and an herb drug is able to exhibit one anti-AS action by mediating two or more cascades, suggesting the pleiotropic and multitargeted effects of CHMs in AS alleviation. Besides, apart from directly improving the cascades of lipid dysbolism, endothelium injury, and inflammation response, herb drugs afford atheroprotective actions through mediating the processes of thrombosis, autophagy, immune reaction, and ncRNAs expression, majority of which converge on the pathways of the above three AS contributors (Figure 5). However,

there are several limitations existing in the research field of CHMs which we cannot ignore. At first, the counterevidence proving the anti-AS roles of CHMs-induced signal pathways is deficient in several literatures, especially in animal studies. So, rigorous logical thinking and experimental design are recommended. Moreover, most of the CHMs-related studies are published in Chinese journals and the theories of ancient traditional Chinese medicine are obscure to the western world, both of which impede the development of CHM research field in the globe. Thus, it is imperative to present more pharmacological and therapeutic findings of CHMs to the international anti-AS study organization and use modern scientific ways to clarify the theory of CHM. Additionally, the majority of the herb drugs like Tan IIA, icariin, BBR, paeonol, and curcumin have been proved to be effective in suppressing AS progression by preclinical experiments, but relevant clinical trials to investigate the safety and effectiveness are scarce. So, it is urgent to perform plentiful well-designed clinical studies with standard and strict procedures to afford reliable and sufficient evidences for the clinical application of these herb drugs. Furthermore, some patent drugs such as Shexiang Baoxin pill have been widely used to treat CVDs in the clinic, whereas the underlying therapeutic mechanisms are poorly understood [156]. Uncovering the antiatherogenic mechanisms of these medicines is helpful to enhance the theoretical basis of their clinic application. In addition, with the development of modern biological technology like bioinformatics analysis and network pharmacology, more and more bioactive compounds from herbs and relevant signal pathways offering protective roles against AS will be discovered.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this article.

Authors' Contributions

Li Lu and Xiaomei Guo designed and wrote the manuscript; Xiaodong Sun and Yating Qin performed the figures and tables.

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References

- [1] G. A. Roth, C. Johnson, and A. Abajobir, "Regional, and national burden of cardiovascular diseases for 10 causes," *Journal of the American College of Cardiology*, vol. 70, no. 1, pp. 1–25, 1990.
- [2] C. Weber and H. Noels, "Atherosclerosis: current pathogenesis and therapeutic options," *Nature Medicine*, vol. 17, no. 11, pp. 1410–1422, 2011.
- [3] M. D. Shapiro and S. Fazio, "From Lipids to Inflammation: New Approaches to Reducing Atherosclerotic Risk," *Circulation Research*, vol. 118, no. 4, pp. 732–749, 2016.
- [4] J. D. Spence and G. K. Dresser, "Overcoming challenges with statin therapy," *Journal of the American Heart Association*, vol. 5, no. 1, Article ID e002497, 2016.
- [5] J. M. Backes, J. F. Ruisinger, C. A. Gibson, and P. M. Moriarty, "Statin-associated muscle symptoms—Managing the highly intolerant," *Journal of Clinical Lipidology*, vol. 11, no. 1, pp. 24–33, 2017.
- [6] C. Liu and Y. Huang, "Chinese Herbal Medicine on Cardiovascular Diseases and the Mechanisms of Action," *Frontiers in Pharmacology*, vol. 7, 2016.
- [7] L. Li, X. Zhou, N. Li, M. Sun, J. Lv, and Z. Xu, "Herbal drugs against cardiovascular disease: traditional medicine and modern development," *Drug Discovery Therapy*, vol. 20, no. 9, pp. 1074–1086, 2015.
- [8] D. Wang, J. Wang, Y. Liu, Z. Zhao, and Q. Liu, "Roles of Chinese herbal medicines in ischemic heart diseases (IHD) by regulating oxidative stress," *International Journal of Cardiology*, vol. 220, pp. 314–319, 2016.
- [9] I. Tabas, G. García-Cardeña, and G. K. Owens, "Recent insights into the cellular biology of atherosclerosis," *The Journal of Cell Biology*, vol. 209, no. 1, pp. 13–22, 2015.
- [10] A. Gisterà and G. K. Hansson, "The immunology of atherosclerosis," *Nature Reviews Nephrology*, vol. 13, no. 6, pp. 368–380, 2017.
- [11] G.-X. Dong, W.-W. Li, R.-Z. Wang, W.-J. Zou, Z.-D. Zhong, and B.-G. Li, "Xinxuekang Regulates Reverse Cholesterol Transport by Improving High-density Lipoprotein Synthesis, Maturation, and Catabolism," *Journal of Cardiovascular Pharmacology*, vol. 70, no. 2, pp. 110–118, 2017.
- [12] D. He, H. Wang, L. Xu et al., "Saikosaponin-a Attenuates Oxidized LDL Uptake and Prompts Cholesterol Efflux in THP-1 Cells," *Journal of Cardiovascular Pharmacology*, vol. 67, no. 6, pp. 510–518, 2016.
- [13] Z. Liu, J. Wang, E. Huang et al., "Tanshinone IIA suppresses cholesterol accumulation in human macrophages: role of heme oxygenase-1," *Journal of Lipid Research*, vol. 55, no. 2, pp. 201–213, 2014.
- [14] Y. Yang, X. Li, L. Peng et al., "Tanshindiol C inhibits oxidized low-density lipoprotein induced macrophage foam cell formation via a peroxiredoxin 1 dependent pathway," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1864, no. 3, pp. 882–890, 2018.
- [15] F.-T. Tang, Y. Cao, T.-Q. Wang et al., "Tanshinone IIA attenuates atherosclerosis in ApoE^{-/-} mice through down-regulation of scavenger receptor expression," *European Journal of Pharmacology*, vol. 650, no. 1, pp. 275–284, 2011.
- [16] N. V. K. Pothineni, S. K. Karathanasis, Z. Ding, A. Arulandu, K. I. Varughese, and J. L. Mehta, "LOX-1 in Atherosclerosis and Myocardial Ischemia: Biology, Genetics, and Modulation," *Journal of the American College of Cardiology*, vol. 69, no. 22, pp. 2759–2768, 2017.
- [17] S. Xu, Z. Liu, and Y. Huang, "Tanshinone II-A inhibits oxidized LDL-induced LOX-1 expression in macrophages by reducing intracellular superoxide radical generation and NF- κ B activation," *Translational Research*, vol. 160, no. 2, pp. 114–124, 2012.
- [18] H.-C. Chen, P.-Y. Chen, M.-J. Wu, M.-H. Tai, and J.-H. Yen, "Tanshinone IIA modulates low density lipoprotein uptake via down-regulation of PCSK9 gene expression in HepG2 cells," *PLoS ONE*, vol. 11, no. 9, Article ID e0162414, 2016.
- [19] H. Gao, L. Li, L. Li et al., "Danshensu Promotes Cholesterol Efflux in RAW264.7 Macrophages," *Lipids*, vol. 51, no. 9, pp. 1083–1092, 2016.

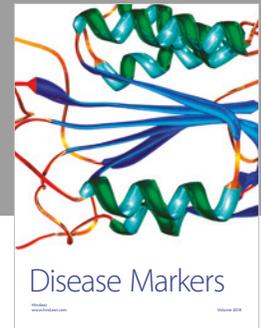
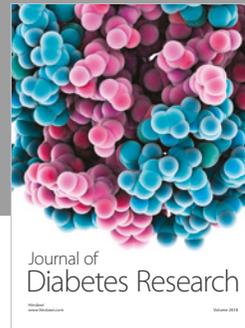
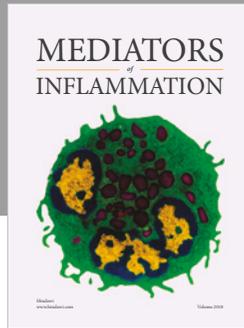
- [20] J. Wang, P. Xu, X. Xie et al., "DBZ (Danshensu Bingpian Zhi), a novel natural compound derivative, attenuates atherosclerosis in apolipoprotein E-Deficient mice," *Journal of the American Heart Association*, vol. 6, no. 10, 2017.
- [21] J.-F. Zhao, S.-J. Jim Leu, S.-K. Shyue, K.-H. Su, J. Wei, and T.-S. Lee, "Novel effect of paeonol on the formation of foam cells: promotion of LXR α -ABCA1 dependent cholesterol efflux in macrophages," *American Journal of Chinese Medicine*, vol. 41, no. 5, pp. 1079–1096, 2013.
- [22] X. Xie, S. Wang, L. Xiao et al., "DBZ blocks Lps-induced monocyte activation and foam cell formation via inhibiting nuclear factor- κ B," *Cellular Physiology and Biochemistry*, vol. 28, no. 4, pp. 649–662, 2011.
- [23] K.-J. Min, H. J. Um, K.-H. Cho, and T. K. Kwon, "Curcumin inhibits oxLDL-induced CD36 expression and foam cell formation through the inhibition of p38 MAPK phosphorylation," *Food and Chemical Toxicology*, vol. 58, pp. 77–85, 2013.
- [24] T. Liu, C. Li, H. Sun et al., "Curcumin inhibits monocyte chemoattractant protein-1 expression and enhances cholesterol efflux by suppressing the c-Jun N-terminal kinase pathway in macrophage," *Inflammation Research*, vol. 63, no. 10, pp. 841–850, 2014.
- [25] W. Kong, J. Wei, P. Abidi et al., "Berberine is a novel cholesterol-lowering drug working through a unique mechanism distinct from statins," *Nature Medicine*, vol. 10, no. 12, pp. 1344–1351, 2004.
- [26] S.-J. Jiang, H. Dong, J.-B. Li et al., "Berberine inhibits hepatic gluconeogenesis via the LKB1-AMPK-TORC2 signaling pathway in streptozotocin-induced diabetic rats," *World Journal of Gastroenterology*, vol. 21, no. 25, pp. 7777–7785, 2015.
- [27] J.-M. Brusq, N. Ancellin, P. Grondin et al., "Inhibition of lipid synthesis through activation of AMP kinase: an additional mechanism for the hypolipidemic effects of berberine," *Journal of Lipid Research*, vol. 47, no. 6, pp. 1281–1288, 2006.
- [28] D. Y. Kim, H. D. Yuan, I. K. Chung, and S. H. Chung, "Compound k, intestinal metabolite of ginsenoside, attenuates hepatic lipid accumulation via AMPK Activation in human hepatoma cells," *Journal of Agricultural and Food Chemistry*, vol. 57, no. 4, pp. 1532–1537, 2009.
- [29] X. Liu, X. Zhang, K. Wang et al., "Kuwanon G attenuates atherosclerosis by upregulation of LXR α -ABCA1/ABCG1 and inhibition of NF κ B activity in macrophages," *Toxicology and Applied Pharmacology*, vol. 341, pp. 56–63, 2018.
- [30] L. Peng, C.-S. Lv, Y. Zhao et al., "QiShenYiQi pill attenuates atherosclerosis by promoting regulatory T cells, inhibiting T helper 17 cells and accelerating cholesterol excretion," *Oncotarget*, vol. 8, no. 47, pp. 82196–82206, 2017.
- [31] U. Förstermann, N. Xia, and H. Li, "Roles of vascular oxidative stress and nitric oxide in the pathogenesis of atherosclerosis," *Circulation Research*, vol. 120, no. 4, pp. 713–735, 2017.
- [32] P. Zhou, S. Lu, Y. Luo et al., "Attenuation of TNF- α -Induced Inflammatory Injury in Endothelial Cells by Ginsenoside Rb1 via Inhibiting NF- κ B, JNK and p38 Signaling Pathways," *Frontiers in Pharmacology*, vol. 8, 2017.
- [33] K. Yang, H. Zhang, Y. Luo et al., "Gypenoside XVII Prevents Atherosclerosis by Attenuating Endothelial Apoptosis and Oxidative Stress: Insight into the ER α -Mediated PI3K/Akt Pathway," *International Journal of Molecular Sciences*, vol. 18, no. 2, p. 77, 2017.
- [34] X. Hou, Q. Tong, W. Wang, W. Xiong, C. Shi, and J. Fang, "Dihydromyricetin protects endothelial cells from hydrogen peroxide-induced oxidative stress damage by regulating mitochondrial pathways," *Life Sciences*, vol. 130, Article ID 14330, pp. 38–46, 2015.
- [35] M. Zhu, J. Li, K. Wang, X. Hao, R. Ge, and Q. Li, "Isoquercitrin inhibits hydrogen peroxide-induced apoptosis of EA.hy926 cells via the PI3K/Akt/GSK3 β signaling pathway," *Molecules*, vol. 21, no. 3, article no. 356, 2016.
- [36] Y. Luo, S. Lu, X. Dong, L. Xu, G. Sun, and X. Sun, "Dihydromyricetin protects human umbilical vein endothelial cells from injury through ERK and Akt mediated Nrf2/HO-1 signaling pathway," *Apoptosis*, vol. 22, no. 8, pp. 1013–1024, 2017.
- [37] M. Qin, Y. Luo, X.-B. Meng et al., "Myricitrin attenuates endothelial cell apoptosis to prevent atherosclerosis: an insight into PI3K/Akt activation and STAT3 signaling pathways," *Vascular Pharmacology*, vol. 70, pp. 23–34, 2015.
- [38] H. Koizumi, J. Yu, R. Hashimoto, Y. Ouchi, and T. Okabe, "Involvement of androgen receptor in nitric oxide production induced by icariin in human umbilical vein endothelial cells," *FEBS Letters*, vol. 584, no. 11, pp. 2440–2444, 2010.
- [39] T. Li, D. Li, H. Xu, H. Zhang, D. Tang, and H. Cao, "Wen-Xin Decoction ameliorates vascular endothelium dysfunction via the PI3K/AKT/eNOS pathway in experimental atherosclerosis in rats," *BMC Complementary and Alternative Medicine*, vol. 16, no. 1, article no. 27, 2016.
- [40] P. Zeng, B. Liu, Q. Wang et al., "Apigenin attenuates atherosclerosis through inducing macrophage apoptosis via inhibition of AKT Ser473 phosphorylation and downregulation of plasminogen activator inhibitor-2," *Oxidative Medicine and Cellular Longevity*, vol. 2015, Article ID 379538, 12 pages, 2015.
- [41] S. E. Lee, S. I. Jeong, H. Yang et al., "Extract of Salvia miltiorrhiza (Danshen) induces Nrf2-mediated heme oxygenase-1 expression as a cytoprotective action in RAW 264.7 macrophages," *Journal of Ethnopharmacology*, vol. 139, no. 2, pp. 541–548, 2012.
- [42] Y. Luo, G. Sun, X. Dong et al., "Isorhamnetin attenuates atherosclerosis by inhibiting macrophage apoptosis via PI3K/AKT activation and HO-1 induction," *PLoS ONE*, vol. 10, no. 3, Article ID e0120259, 19 pages, 2015.
- [43] J.-H. Chen, G. Hsiao, A.-R. Lee, C.-C. Wu, and M.-H. Yen, "Andrographolide suppresses endothelial cell apoptosis via activation of phosphatidylinositol-3-kinase/Akt pathway," *Biochemical Pharmacology*, vol. 67, no. 7, pp. 1337–1345, 2004.
- [44] P. Chan, Y.-C. Chen, L.-J. Lin et al., "Tanshinone IIA attenuates H₂O₂-induced injury in human umbilical vein endothelial cells," *American Journal of Chinese Medicine*, vol. 40, no. 6, pp. 1307–1319, 2012.
- [45] G.-B. Sun, M. Qin, J.-X. Ye et al., "Inhibitory effects of myricitrin on oxidative stress-induced endothelial damage and early atherosclerosis in ApoE^{-/-} mice," *Toxicology and Applied Pharmacology*, vol. 271, no. 1, pp. 114–126, 2013.
- [46] M.-H. Bao, Y.-W. Zhang, and H.-H. Zhou, "Paeonol suppresses oxidized low-density lipoprotein induced endothelial cell apoptosis via activation of LOX-1/p38MAPK/NF- κ B pathway," *Journal of Ethnopharmacology*, vol. 146, no. 2, pp. 543–551, 2013.
- [47] L. Zhang, Y.-H. Jia, X.-S. Zhao et al., "Trichosanatine alleviates oxidized low-density lipoprotein induced endothelial cells injury via inhibiting the LOX-1/p38 MAPK pathway," *American Journal of Translational Research*, vol. 8, no. 12, pp. 5455–5464, 2016.
- [48] K.-W. Choy, M. R. Mustafa, Y. S. Lau et al., "Paeonol protects against endoplasmic reticulum stress-induced endothelial dysfunction via AMPK/PPAR δ signaling pathway," *Biochemical Pharmacology*, vol. 116, pp. 51–62, 2016.

- [49] K. W. Choy, Y. S. Lau, D. Murugan, and M. R. Mustafa, "Chronic treatment with paeonol improves endothelial function in mice through inhibition of endoplasmic reticulum stress-mediated oxidative stress," *PLoS ONE*, vol. 12, no. 5, Article ID e0178365, 2017.
- [50] Y. Jing, D. Cai, Q. Chen et al., "Liuwei Dihuang soft capsules attenuates endothelial cell apoptosis to prevent atherosclerosis through GPR30-mediated regulation in ovariectomized ApoE-deficient mice," *Journal of Ethnopharmacology*, vol. 208, pp. 185–198, 2017.
- [51] C. Camaré, M. Pucelle, A. Nègre-Salvayre, and R. Salvayre, "Angiogenesis in the atherosclerotic plaque," *Redox Biology*, vol. 12, pp. 18–34, 2017.
- [52] A. Jeong, H.-J. Lee, S.-J. Jeong et al., "Compound K inhibits basic fibroblast growth factor-induced angiogenesis via regulation of p38 mitogen-activated protein kinase and AKT in human umbilical vein endothelial cells," *Biological & Pharmaceutical Bulletin*, vol. 33, no. 6, pp. 945–950, 2010.
- [53] Y. Qiao, P.-J. Zhang, X.-T. Lu et al., "Panax notoginseng saponins inhibits atherosclerotic plaque angiogenesis by down-regulating vascular endothelial growth factor and nicotinamide adenine dinucleotide phosphate oxidase subunit 4 expression," *Chinese Journal of Integrative Medicine*, vol. 21, no. 4, pp. 259–265, 2015.
- [54] B. N. Davis-Dusenbery, C. Wu, and A. Hata, "Micromanaging vascular smooth muscle cell differentiation and phenotypic modulation," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 31, no. 11, pp. 2370–2377, 2011.
- [55] W.-Y. Wu, H. Yan, X.-B. Wang et al., "Sodium tanshinone IIA silicate inhibits high glucose-induced vascular smooth muscle cell proliferation and migration through activation of AMP-activated protein kinase," *PLoS ONE*, vol. 9, no. 4, Article ID e94957, 2014.
- [56] U.-H. Jin, S.-J. Suh, W. C. Hyen et al., "Tanshinone IIA from *Salvia miltiorrhiza* BUNGE inhibits human aortic smooth muscle cell migration and MMP-9 activity through AKT signaling pathway," *Journal of Cellular Biochemistry*, vol. 104, no. 1, pp. 15–26, 2008.
- [57] S.-J. Suh, U.-H. Jin, H.-J. Choi et al., "Cryptotanshinone from *Salvia miltiorrhiza* BUNGE has an inhibitory effect on TNF- α -induced matrix metalloproteinase-9 production and HASMC migration via down-regulated NF- κ B and AP-1," *Biochemical Pharmacology*, vol. 72, no. 12, pp. 1680–1689, 2006.
- [58] Y. Xing, J. Tu, L. Zheng, L. Guo, and T. Xi, "Anti-angiogenic effect of tanshinone IIA involves inhibition of the VEGF/VEGFR2 pathway in vascular endothelial cells," *Oncology Reports*, vol. 33, no. 1, pp. 163–170, 2015.
- [59] L.-H. Dong, J.-K. Wen, S.-B. Miao et al., "Baicalin inhibits PDGF-BB-stimulated vascular smooth muscle cell proliferation through suppressing PDGFRB-ERK signaling and increase in p27 accumulation and prevents injury-induced neointimal hyperplasia," *Cell Research*, vol. 20, no. 11, pp. 1252–1262, 2010.
- [60] Y. Hu, K. Liu, M. Yan, Y. Zhang, Y. Wang, and L. Ren, "Icariin inhibits oxidized low-density lipoprotein-induced proliferation of vascular smooth muscle cells by suppressing activation of extracellular signal-regulated kinase 1/2 and expression of proliferating cell nuclear antigen," *Molecular Medicine Reports*, vol. 13, no. 3, pp. 2899–2903, 2016.
- [61] Z. Zhang, D. Zhang, B. Du, and Z. Chen, "Hyperoside inhibits the effects induced by oxidized low-density lipoprotein in vascular smooth muscle cells via oxLDL-LOX-1-ERK pathway," *Molecular and Cellular Biochemistry*, vol. 433, no. 1-2, pp. 169–176, 2017.
- [62] W. Li, W. Zhi, F. Liu, J. Zhao, Q. Yao, and X. Niu, "Paeoniflorin inhibits VSMCs proliferation and migration by arresting cell cycle and activating HO-1 through MAPKs and NF- κ B pathway," *International Immunopharmacology*, vol. 54, pp. 103–111, 2018.
- [63] K.-W. Liang, S.-C. Yin, C.-T. Ting et al., "Berberine inhibits platelet-derived growth factor-induced growth and migration partly through an AMPK-dependent pathway in vascular smooth muscle cells," *European Journal of Pharmacology*, vol. 590, no. 1-3, pp. 343–354, 2008.
- [64] K.-W. Liang, C.-T. Ting, S.-C. Yin et al., "Berberine suppresses MEK/ERK-dependent Egr-1 signaling pathway and inhibits vascular smooth muscle cell regrowth after in vitro mechanical injury," *Biochemical Pharmacology*, vol. 71, no. 6, pp. 806–817, 2006.
- [65] L. Yu, X. Huang, K. Huang, C. Gui, Q. Huang, and B. Wei, "Ligustrazine attenuates the platelet-derived growth factor-BB-induced proliferation and migration of vascular smooth muscle cells by interrupting extracellular signal-regulated kinase and P38 mitogen-activated protein kinase pathways," *Molecular Medicine Reports*, vol. 12, no. 1, pp. 705–711, 2015.
- [66] Y. Song, L. Long, N. Zhang, and Y. Liu, "Inhibitory effects of hydroxysafflor yellow A on PDGF-BB-induced proliferation and migration of vascular smooth muscle cells via mediating Akt signaling," *Molecular Medicine Reports*, vol. 10, no. 3, pp. 1555–1560, 2014.
- [67] S. J. Lin, I. T. Lee, Y. H. Chen et al., "Salvianolic acid B attenuates MMP-2 and MMP-9 expression in vivo in apolipoprotein-E-deficient mouse aorta and in vitro in LPS-treated human aortic smooth muscle cells," *Journal of Cellular Biochemistry*, vol. 100, no. 2, pp. 372–384, 2007.
- [68] L.-H. Zhu, L. Wang, D. Wang et al., "Puerarin attenuates high-glucose and diabetes-induced vascular smooth muscle cell proliferation by blocking PKC β 2/Rac1-dependent signaling," *Free Radical Biology & Medicine*, vol. 48, no. 4, pp. 471–482, 2010.
- [69] J. Chen, M. Dai, and Y. Wang, "Paeonol Inhibits Proliferation of Vascular Smooth Muscle Cells Stimulated by High Glucose via Ras-Raf-ERK1/2 Signaling Pathway in Coculture Model," *Evidence-Based Complementary and Alternative Medicine*, vol. 2014, 2014.
- [70] H. R. Bateman, Q. Liang, D. Fan, V. Rodriguez, and S. M. Lessner, "Sparstolonin B Inhibits Pro-Angiogenic Functions and Blocks Cell Cycle Progression in Endothelial Cells," *PLoS ONE*, vol. 8, no. 8, Article ID e70500, 2013.
- [71] H.-L. Yang, H. C. Chang, S.-W. Lin et al., "Antrodia salmonea inhibits TNF- α -induced angiogenesis and atherogenesis in human endothelial cells through the down-regulation of NF- κ B and up-regulation of Nrf2 signaling pathways," *Journal of Ethnopharmacology*, vol. 151, no. 1, pp. 394–406, 2014.
- [72] J. Cao, Z. Han, L. Tian et al., "Curcumin inhibits EMMPRIN and MMP-9 expression through AMPK-MAPK and PKC signaling in PMA induced macrophages," *Journal of Translational Medicine*, vol. 12, no. 1, article no. 266, 2014.
- [73] L. Ma, X. Liu, H. Lu et al., "Traditional Chinese medication Tongxinluo inhibits inflammatory angiogenesis via Bmx/NF-B/MAPK pathways," *European Heart Journal Supplements*, vol. 17, no. Supplement B, pp. B13–B22, 2015.

- [74] W. Zhao, C. Wu, and X. Chen, "Cryptotanshinone inhibits oxidized LDL-induced adhesion molecule expression via ROS dependent NF- κ B pathways," *Cell Adhesion & Migration*, 2015.
- [75] W. Zhao, H. Feng, S. Guo, Y. Han, and X. Chen, "Danshenol A inhibits TNF- α -induced expression of intercellular adhesion molecule-1 (ICAM-1) mediated by NOX4 in endothelial cells," *Scientific Reports*, vol. 7, no. 1, 2017.
- [76] W. Zhao, C. Li, H. Gao, Q. Wu, J. Shi, and X. Chen, "Dihydro-tanshinone I attenuates atherosclerosis in ApoE-deficient mice: Role of NOX4/NF- κ B mediated lectin-like oxidized LDL receptor-1 (LOX-1) of the Endothelium," *Frontiers in Pharmacology*, vol. 7, 2016.
- [77] W. Li, W. Sun, C.-H. Yang, H.-Z. Hu, and Y.-H. Jiang, "Tanshinone II A protects against lipopolysaccharides-induced endothelial cell injury via Rho/Rho kinase pathway," *Chinese Journal of Integrative Medicine*, vol. 20, no. 3, pp. 216–223, 2014.
- [78] S. Zhuang, T.-H. Cheng, N.-L. Shih et al., "Tanshinone IIA Induces Heme Oxygenase 1 Expression and Inhibits Cyclic Strain-Induced Interleukin 8 Expression in Vascular Endothelial Cells," *American Journal of Chinese Medicine*, vol. 44, no. 2, pp. 377–388, 2016.
- [79] I. T. Nizamutdinova, Y. M. Kim, H. Jin et al., "Tanshinone IIA inhibits TNF- α -mediated induction of VCAM-1 but not ICAM-1 through the regulation of GATA-6 and IRF-1," *International Immunopharmacology*, vol. 14, no. 4, pp. 650–657, 2012.
- [80] C.-C. Chang, C.-F. Chu, C.-N. Wang et al., "The anti-atherosclerotic effect of tanshinone IIA is associated with the inhibition of TNF- α -induced VCAM-1, ICAM-1 and CX3CL1 expression," *Phytomedicine*, vol. 21, no. 3, pp. 207–216, 2014.
- [81] F.-Q. Li, D.-K. Zeng, C.-L. Jia et al., "The effects of sodium tanshinone iia sulfonate pretreatment on high glucose-induced expression of fractalkine and apoptosis in human umbilical vein endothelial cells," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 4, pp. 5279–5286, 2015.
- [82] G. Fan, X. Jiang, X. Wu et al., "Anti-inflammatory activity of tanshinone IIA in LPS-stimulated RAW264.7 macrophages via miRNAs and TLR4–NF- κ B Pathway," *Inflammation*, vol. 39, no. 1, pp. 375–384, 2016.
- [83] X. Li, L.-H. Lian, T. Bai et al., "Cryptotanshinone inhibits LPS-induced proinflammatory mediators via TLR4 and TAK1 signaling pathway," *International Immunopharmacology*, vol. 11, no. 11, pp. 1871–1876, 2011.
- [84] S. Tang, X.-Y. Shen, H.-Q. Huang et al., "Cryptotanshinone suppressed inflammatory cytokines secretion in RAW264.7 macrophages through inhibition of the NF- κ B and MAPK signaling pathways," *Inflammation*, vol. 34, no. 2, pp. 111–118, 2011.
- [85] S. I. Jang, H. J. Kim, Y. J. Kim, S. I. Jeong, and Y. O. You, "Tanshinone IIA inhibits LPS-induced NF- κ B activation in RAW 264.7 cells: possible involvement of the NIK-IKK, ERK1/2, p38 and JNK pathways," *European Journal of Pharmacology*, vol. 542, no. 1–3, pp. 1–7, 2006.
- [86] T.-H. Chen, Y.-T. Hsu, C.-H. Chen, S.-H. Kao, and H.-M. Lee, "Tanshinone IIA from *Salvia miltiorrhiza* induces heme oxygenase-1 expression and inhibits lipopolysaccharide-induced nitric oxide expression in RAW 264.7 cells," *Mitochondrion*, vol. 7, no. 1–2, pp. 101–105, 2007.
- [87] X.-H. Liu, X.-L. Wang, H. Xin et al., "Induction of Heme Oxygenase-1 by Sodium 9-Hydroxytanshinone IIA Sulfonate Derivative Contributes to Inhibit LPS-Mediated Inflammatory Response in Macrophages," *Cellular Physiology and Biochemistry*, vol. 36, no. 4, pp. 1316–1330, 2015.
- [88] Y. Joe, M. Zheng, H. J. Kim et al., "Salvianolic acid B exerts vasoprotective effects through the modulation of heme oxygenase-1 and arginase activities," *The Journal of Pharmacology and Experimental Therapeutics*, vol. 341, no. 3, pp. 850–858, 2012.
- [89] C. Y. Hsieh, M. J. Hsu, G. Hsiao et al., "Andrographolide enhances nuclear factor- κ B subunit p65 Ser536 dephosphorylation through activation of protein phosphatase 2A in vascular smooth muscle cells," *The Journal of Biological Chemistry*, vol. 286, no. 8, pp. 5942–5955, 2011.
- [90] Y. Chen, M. Hsu, C. Hsieh, L. Lee, Z. Chen, and J. Sheu, "Andrographolide Inhibits Nuclear Factor- κ B," *The Scientific World Journal*, vol. 2014, pp. 1–10, 2014.
- [91] M. Qin, Y. Luo, S. Lu et al., "Ginsenoside F1 ameliorates endothelial cell inflammatory injury and prevents atherosclerosis in mice through A20-mediated suppression of NF- κ B signaling," *Frontiers in Pharmacology*, vol. 8, 2017.
- [92] J. Fan, D. Liu, C. He, X. Li, and F. He, "Inhibiting adhesion events by Panax notoginseng saponins and Ginsenoside Rb1 protecting arteries via activation of Nrf2 and suppression of p38 – VCAM-1 signal pathway," *Journal of Ethnopharmacology*, vol. 192, pp. 423–430, 2016.
- [93] Y. Yu, X. Li, L. Qu et al., "DXKK exerts anti-inflammatory effects by inhibiting the lipopolysaccharide-induced NF- κ B/COX-2 signalling pathway and the expression of inflammatory mediators," *Journal of Ethnopharmacology*, vol. 178, pp. 199–208, 2016.
- [94] K.-W. Choi, H.-J. Park, D.-H. Jung et al., "Inhibition of TNF- α -induced adhesion molecule expression by diosgenin in mouse vascular smooth muscle cells via downregulation of the MAPK, Akt and NF- κ B signaling pathways," *Vascular Pharmacology*, vol. 53, no. 5–6, pp. 273–280, 2010.
- [95] S. Bhaskar, P. R. Sudhakaran, and A. Helen, "Quercetin attenuates atherosclerotic inflammation and adhesion molecule expression by modulating TLR-NF- κ B signaling pathway," *Cellular Immunology*, vol. 310, pp. 131–140, 2016.
- [96] L. Wang, X. M. Qiu, Q. Hao, and D. J. Li, "Anti-inflammatory effects of a Chinese herbal medicine in atherosclerosis via estrogen receptor β mediating nitric oxide production and NF- κ B suppression in endothelial cells," *Cell Death & Disease*, vol. 4, no. 3, pp. e551–e551, 2013.
- [97] B. Oche, L. Chen, Y.-K. Ma et al., "Cryptotanshinone and wogonin up-regulate eNOS in vascular endothelial cells via ER α and down-regulate iNOS in LPS stimulated vascular smooth muscle cells via ER β ," *Archives of Pharmacological Research*, vol. 39, no. 2, pp. 249–258, 2016.
- [98] J. Y. Lee and W. Park, "Anti-inflammatory effect of wogonin on RAW 264.7 mouse macrophages induced with polyinosinic-polycytidylic acid," *Molecules*, vol. 20, no. 4, pp. 6888–6900, 2015.
- [99] N. Martínez-Micaelo, N. González-Abuín, X. Terra et al., "Omega-3 docosahexaenoic acid and procyanidins inhibit cyclo-oxygenase activity and attenuate NF- κ B activation through a p105/p50 regulatory mechanism in macrophage inflammation," *Biochemical Journal*, vol. 441, no. 2, pp. 653–663, 2012.
- [100] J. Xing, K. Peng, W. Cao, X. Lian, Q. Wang, and X. Wang, "Effects of total flavonoids from *Dracocephalum moldavica* on the proliferation, migration, and adhesion molecule expression of rat vascular smooth muscle cells induced by TNF- α ," *Pharmaceutical Biology*, vol. 51, no. 1, pp. 74–83, 2013.
- [101] C.-Q. Xu, B.-J. Liu, J.-F. Wu et al., "Icariin attenuates LPS-induced acute inflammatory responses: involvement of

- PI3K/Akt and NF κ B signaling pathway," *European Journal of Pharmacology*, vol. 642, no. 1–3, pp. 146–153, 2010.
- [102] C.-Y. Wang, T.-C. Kao, W.-H. Lo, and G.-C. Yen, "Glycyrrhizic acid and 18 β -glycyrrhetic acid modulate lipopolysaccharide-induced inflammatory response by suppression of NF- κ B through PI3K p110 δ and p110 γ inhibitions," *Journal of Agricultural and Food Chemistry*, vol. 59, no. 14, pp. 7726–7733, 2011.
- [103] Q. Wang, M. Zhang, B. Liang, N. Shirwany, Y. Zhu, and M.-H. Zou, "Activation of AMP-activated protein kinase is required for berberine-induced reduction of atherosclerosis in mice: the role of uncoupling protein 2," *PLoS ONE*, vol. 6, no. 9, Article ID e25436, 2011.
- [104] J. Wu, H. Zhang, B. Hu et al., "Coptisine from *Coptis chinensis* inhibits production of inflammatory mediators in lipopolysaccharide-stimulated RAW 264.7 murine macrophage cells," *European Journal of Pharmacology*, vol. 780, pp. 106–114, 2016.
- [105] L. Kole, B. Giri, S. K. Manna, B. Pal, and S. Ghosh, "Biochanin-A, an isoflavon, showed anti-proliferative and anti-inflammatory activities through the inhibition of iNOS expression, p38-MAPK and ATF-2 phosphorylation and blocking NF κ B nuclear translocation," *European Journal of Pharmacology*, vol. 653, no. 1–3, pp. 8–15, 2011.
- [106] M. Mueller, S. Hobiger, and A. Jungbauer, "Red clover extract: A source for substances that activate peroxisome proliferator-activated receptor α and ameliorate the cytokine secretion profile of lipopolysaccharide-stimulated macrophages," *Menopause*, vol. 17, no. 2, pp. 379–387, 2010.
- [107] F. L. Chen, Z. H. Yang, Y. Liu et al., "Berberine inhibits the expression of TNF α , MCP-1, and IL-6 in AcLDL-stimulated macrophages through PPAR γ pathway," *Endocrine Journal*, vol. 33, no. 3, pp. 331–337, 2008.
- [108] H. Li, Y. Jiao, and M. Xie, "Paeoniflorin Ameliorates Atherosclerosis by Suppressing TLR4-Mediated NF- κ B Activation," *Inflammation*, vol. 40, no. 6, pp. 2042–2051, 2017.
- [109] L. Gu, W. Bai, S. Li et al., "Celastrol prevents atherosclerosis via inhibiting LOX-1 and oxidative stress," *PLoS ONE*, vol. 8, no. 6, Article ID e65477, 2013.
- [110] Y. Wang, L. Cui, H. Xu et al., "TRPV1 agonism inhibits endothelial cell inflammation via activation of eNOS/NO pathway," *Atherosclerosis*, vol. 260, pp. 13–19, 2017.
- [111] Z. Meng, C. Yan, Q. Deng, D.-F. Gao, and X.-L. Niu, "Curcumin inhibits LPS-induced inflammation in rat vascular smooth muscle cells in vitro via ROS-relative TLR4-MAPK/NF- κ B pathways," *Acta Pharmacologica Sinica*, vol. 34, no. 7, pp. 901–911, 2013.
- [112] X. Huang, F. Wang, W. Chen, N. Wang, Y. Chen, and L. Sun, "DaoTan decoction (DTD) inhibits tumor necrosis factor- α (TNF- α)-induced expression of intercellular adhesion molecule-1 (ICAM-1), p53 and p21, in human umbilical vein endothelial cells (HUVECs)," *Pharmaceutical Biology*, vol. 52, no. 10, pp. 1320–1326, 2014.
- [113] X. Huang, F. Wang, W. Chen et al., "Dao-Tan decoction inhibits tumor necrosis factor- α -induced intercellular adhesion molecule-1 expression by blocking JNK and p38 signaling pathways in human umbilical vein endothelial cells," *Pharmaceutical Biology*, vol. 50, no. 9, pp. 1111–1117, 2012.
- [114] J. Zheng, B. Liu, Q. Lun et al., "Longxuetongluo capsule inhibits atherosclerosis progression in high-fat diet-induced ApoE $^{-/-}$ mice by improving endothelial dysfunction," *Atherosclerosis*, vol. 255, pp. 156–163, 2016.
- [115] L. F. Sun, D. Q. An, and G. L. Niyazi, "Effects of Tianxiangdan Granule treatment on atherosclerosis via NF κ B and p38 MAPK signaling pathways," *Molecular Medicine Reports*, vol. 17, no. 1, pp. 1642–1650, 2018.
- [116] M. Zhou, P. Li, Q. Kang et al., "Shen-Yuan-Dan Capsule inhibiting inflammatory reaction by regulating insulin receptor substrate 1/PI3K/Akt/NF- κ B signaling pathway in apolipoprotein e knockout mice fed with a high-fat diet," *Acta Cardiologica Sinica*, vol. 33, no. 3, pp. 285–291, 2017.
- [117] J. Fu, X. Zhu, W. Wang et al., "1, 6-di-O-caffeoyl- β -D-glucopyranoside, a natural compound from *Callicarpa nudiflora* Hook impairs P2Y12 and thromboxane A2 receptor-mediated amplification of platelet activation and aggregation," *Phytomedicine*, vol. 36, pp. 273–282, 2017.
- [118] Y. Chen, W. Yang, L. Guo et al., "Atractylodes lactone compounds inhibit platelet activation," *Platelets*, vol. 28, no. 2, pp. 194–202, 2017.
- [119] W. Lu, J. Lee, D. Chou et al., "A novel role of andrographolide, an NF-kappa B inhibitor, on inhibition of platelet activation: the pivotal mechanisms of endothelial nitric oxide synthase/cyclic GMP," *Journal of Molecular Medicine*, vol. 89, no. 12, pp. 1261–1273, 2011.
- [120] W. J. Lu, K. H. Lin, M. J. Hsu, D. S. Chou, G. Hsiao, and J. R. Sheu, "Suppression of NF- κ B signaling by andrographolide with a novel mechanism in human platelets: Regulatory roles of the p38 MAPK-hydroxyl radical-ERK2 cascade," *Biochemical Pharmacology*, vol. 84, no. 7, pp. 914–924, 2012.
- [121] Y. Wang, Z.-Y. Fang, S.-A. Huang, and L. Cui, "Tanshinone IIA down-regulates the expression of MMP-12 and TF in RAW 264.7 cells," *Journal of Southern Medical University*, vol. 29, no. 7, pp. 1317–1320, 2009.
- [122] C. Shi, X. Zhu, J. Wang, and D. Long, "Tanshinone IIA promotes non-amyloidogenic processing of amyloid precursor protein in platelets via estrogen receptor signaling to phosphatidylinositol 3-kinase/Akt," *Biomedical Reports*, vol. 2, no. 4, pp. 500–504, 2014.
- [123] F. Maione, V. de Feo, E. Caiazza, L. de Martino, C. Cicala, and N. Mascolo, "Tanshinone IIA, a major component of *Salvia miltiorrhiza* Bunge, inhibits platelet activation via Erk-2 signaling pathway," *Journal of Ethnopharmacology*, vol. 155, no. 2, pp. 1236–1242, 2014.
- [124] M. Liu, X. Chen, J. Ma et al., " β -Elemene attenuates atherosclerosis in apolipoprotein E-deficient mice via restoring NO levels and alleviating oxidative stress," *Biomedicine & Pharmacotherapy*, vol. 95, pp. 1789–1798, 2017.
- [125] P. Li, Y. Yang, and M. Liu, "Xuezhikang, Extract of Red Yeast Rice, Inhibited Tissue Factor and Hypercoagulable State Through Suppressing Nicotinamide Adenine Dinucleotide Phosphate Oxidase and Extracellular Signal-regulated Kinase Activation," *Journal of Cardiovascular Pharmacology*, vol. 58, no. 3, pp. 307–318, 2011.
- [126] T. H.-W. Huang, V. H. Tran, B. D. Roufogalis, and Y. Li, "Gypenoside XLIX, a naturally occurring gynosaponin, PPAR-alpha dependently inhibits LPS-induced tissue factor expression and activity in human THP-1 monocytic cells," *Toxicology and Applied Pharmacology*, vol. 218, no. 1, pp. 30–36, 2007.
- [127] C. Pan, Y. Huo, X. An et al., "Panax notoginseng and its components decreased hypertension via stimulation of endothelial-dependent vessel dilatation," *Vascular Pharmacology*, vol. 56, no. 3–4, pp. 150–158, 2012.
- [128] J. Y. Park, Y. W. Choi, J. W. Yun et al., "Gomisin J from *Schisandra chinensis* induces vascular relaxation via activation

- of endothelial nitric oxide synthase," *Vascular Pharmacology*, vol. 57, no. 2-4, pp. 124-130, 2012.
- [129] Y. Y. Sun, X. H. Su, J. Y. Jin et al., "Rumex acetosa L. Induces vasorelaxation in rat aorta via activation of PI3-kinase/Akt- and Ca²⁺-eNOS-NO signaling in endothelial cells," *Journal of Physiology and Pharmacology*, vol. 66, no. 6, pp. 907-915, 2015.
- [130] Y. P. Hwang, H. G. Kim, T. T. Hien, M. H. Jeong, T. C. Jeong, and H. G. Jeong, "Puerarin activates endothelial nitric oxide synthase through estrogen receptor-dependent PI3-kinase and calcium-dependent AMP-activated protein kinase," *Toxicology and Applied Pharmacology*, vol. 257, no. 1, pp. 48-58, 2011.
- [131] Z. Zhou, Y. Liu, A.-D. Miao, and S.-Q. Wang, "Salvianolic acid B attenuates plasminogen activator inhibitor type 1 production in TNF- α treated human umbilical vein endothelial cells," *Journal of Cellular Biochemistry*, vol. 96, no. 1, pp. 109-116, 2005.
- [132] G. R. Y. De Meyer, M. O. J. Grootaert, C. F. Michiels, A. Kurdi, D. M. Schrijvers, and W. Martinet, "Autophagy in vascular disease," *Circulation Research*, vol. 116, pp. 468-479, 2015.
- [133] S. C. Nussenzweig, S. Verma, and T. Finkel, "The role of autophagy in vascular biology," *Circulation Research*, vol. 116, no. 3, pp. 480-488, 2015.
- [134] X. Jin, M. Chen, L. Yi et al., "Delphinidin-3-glucoside protects human umbilical vein endothelial cells against oxidized low-density lipoprotein-induced injury by autophagy upregulation via the AMPK/SIRT1 signaling pathway," *Molecular Nutrition & Food Research*, vol. 58, no. 10, pp. 1941-1951, 2014.
- [135] H.-H. Lin, "In Vitro and in Vivo Atheroprotective Effects of Gossypetin against Endothelial Cell Injury by Induction of Autophagy," *Chemical Research in Toxicology*, vol. 28, no. 2, pp. 202-215, 2016.
- [136] L. Zhang, L. Cui, G. Zhou, H. Jing, Y. Guo, and W. Sun, "Pterostilbene, a natural small-molecular compound, promotes cytoprotective macroautophagy in vascular endothelial cells," *The Journal of Nutritional Biochemistry*, vol. 24, no. 5, pp. 903-911, 2013.
- [137] M.-L. Chen, L. Yi, X. Jin et al., "Resveratrol attenuates vascular endothelial inflammation by inducing autophagy through the cAMP signaling pathway," *Autophagy*, vol. 9, no. 12, pp. 2033-2045, 2013.
- [138] H. Guo, Y. Chen, L. Liao, and W. Wu, "Resveratrol protects HUVECs from oxidized-LDL induced oxidative damage by autophagy upregulation via the AMPK/SIRT1 pathway," *Cardiovascular Drugs and Therapy*, vol. 27, no. 3, pp. 189-198, 2013.
- [139] X. Fan, J. Wang, J. Hou et al., "Berberine alleviates ox-LDL induced inflammatory factors by up-regulation of autophagy via AMPK/mTOR signaling pathway," *Journal of Translational Medicine*, vol. 13, no. 1, article 92, 2015.
- [140] J. Y. Kou, Y. Li, and Z. Y. Zhong, "Berberine-sonodynamic therapy induces autophagy and lipid unloading in macrophage," *Cell Death Disease*, vol. 8, no. 1, p. e2558, 2017.
- [141] Y. Luo, X. Meng, P. Zhou et al., "Elatoside C protects against ox-LDL-induced HUVECs injury by FoxO1-mediated autophagy induction," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1863, no. 6, pp. 1654-1665, 2017.
- [142] B.-C. Zhang, C.-W. Zhang, C. Wang, D.-F. Pan, T.-D. Xu, and D.-Y. Li, "Luteolin Attenuates Foam Cell Formation and Apoptosis in Ox-LDL-Stimulated Macrophages by Enhancing Autophagy," *Cellular Physiology and Biochemistry*, vol. 39, no. 5, pp. 2065-2076, 2016.
- [143] Y. Tang, H. Wu, B. Shao, Y. Wang, C. Liu, and M. Guo, "Celosins inhibit atherosclerosis in ApoE^{-/-} mice and promote autophagy flow," *Journal of Ethnopharmacology*, vol. 215, pp. 74-82, 2017.
- [144] I. Tabas and A. H. Lichtman, "Monocyte-Macrophages and T Cells in Atherosclerosis," *Immunity*, vol. 47, no. 4, pp. 621-634, 2017.
- [145] W. Su, A. Sun, D. Xu et al., "Tongxinluo inhibits oxidized low-density lipoprotein-induced maturation of human dendritic cells via activating peroxisome proliferator-activated receptor gamma pathway," *Journal of Cardiovascular Pharmacology*, vol. 56, no. 2, pp. 177-183, 2010.
- [146] L. Liu, P. Liao, B. Wang, X. Fang, W. Li, and S. Guan, "Oral administration of baicalin and geniposide induces regression of atherosclerosis via inhibiting dendritic cells in ApoE-knockout mice," *International Immunopharmacology*, vol. 20, no. 1, pp. 197-204, 2014.
- [147] J. Lv, W. Xiong, T. Lei et al., "Amygdalin ameliorates the progression of atherosclerosis in LDL receptor-deficient mice," *Molecular Medicine Reports*, vol. 16, no. 6, pp. 8171-8179, 2017.
- [148] Y. Liu, L. Zheng, Q. Wang, and Y.-W. Hu, "Emerging roles and mechanisms of long noncoding RNAs in atherosclerosis," *International Journal of Cardiology*, vol. 228, pp. 570-582, 2017.
- [149] M. W. Feinberg and K. J. Moore, "MicroRNA regulation of atherosclerosis," *Circulation Research*, vol. 118, no. 4, pp. 703-720, 2016.
- [150] L. Jia, N. Song, G. Yang et al., "Effects of Tanshinone IIA on the modulation of MIR-33a and the SREBP-2/Pcsk9 signaling pathway in hyperlipidemic rats," *Molecular Medicine Reports*, vol. 13, no. 6, pp. 4627-4635, 2016.
- [151] H. Zhong, K. Chen, M. Feng et al., "Genipin alleviates high-fat diet-induced hyperlipidemia and hepatic lipid accumulation in mice via miR-142a-5p/SREBP-1c axis," *FEBS Journal*, 2017.
- [152] Y.-R. Liu, J.-J. Chen, and M. Dai, "Paeonol protects rat vascular endothelial cells from ox-LDL-induced injury in vitro via downregulating microRNA-21 expression and TNF- α release," *Acta Pharmacologica Sinica*, vol. 35, no. 4, pp. 483-488, 2014.
- [153] R.-N. Zhang, B. Zheng, L.-M. Li, J. Zhang, X.-H. Zhang, and J.-K. Wen, "Tongxinluo inhibits vascular inflammation and neointimal hyperplasia through blockade of the positive feedback loop between miR-155 and TNF- α ," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 307, no. 4, pp. H552-H562, 2014.
- [154] C. Chen, G. Cheng, X. Yang, C. Li, R. Shi, and N. Zhao, "Tanshinol suppresses endothelial cells apoptosis in mice with atherosclerosis via lncRNA TUG1 up-regulating the expression of miR-26a," *American Journal of Translational Research*, vol. 8, no. 7, pp. 2981-2991, 2016.
- [155] W. Nie, X. Zhang, H. Yan et al., "Xiaoxianggou attenuates atherosclerotic plaque formation in endogenous high Ang II ApoE^{-/-} mice via the inhibition of miR-203 on the expression of Ets-2 in endothelial cells," *Biomedicine & Pharmacotherapy*, vol. 82, pp. 173-179, 2016.
- [156] Z. Zhou, W. Shen, L. Yu, C. Xu, and Q. Wu, "A Chinese patent medicine, Shexiang Baoxin Pill, for Non-ST-elevation acute coronary syndromes: A systematic review," *Journal of Ethnopharmacology*, vol. 194, pp. 1130-1139, 2016.



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