A Reasonable Diet Promotes Balance of Intestinal Microbiota: Prevention of Precolorectal Cancer

Pan Huang and Yi Liu

Key Laboratory of Agro-Ecological Processes in Subtropical Regions and Taoyuan Station of Agro-Ecology Research, Institute of Subtropical Agriculture, Chinese Academy of Sciences, Changsha 410125, China

Correspondence should be addressed to Yi Liu; liuyi@isa.ac.cn

Received 15 May 2019; Accepted 9 July 2019; Published 25 July 2019

Guest Editor: Qiang Tong

Colorectal cancer (CRC) is a multifactorial disease and the second leading cause of cancer death worldwide. The pathogenesis of colorectal cancer includes genetics, age, chronic inflammation, and lifestyle. Increasing attention has recently been paid to dietary factors. Evidence from epidemiological studies and clinical research suggests that high-fibre diets can significantly reduce the incidence of CRC, whilst the consumption of high-fat diets, high-protein diets, red meat, and processed meat is high-risk factors for tumorigenesis. Fibre is a regulator of intestinal microflora and metabolism and is thus a key dietary component for maintaining intestinal health. Intestinal microbes are closely linked to CRC, with the growth of certain microbiota (such as \textit{Fusobacterium nucleatum}, \textit{Escherichia coli}, or \textit{Bacteroides fragilis}) favouring carcinogenesis, whilst the dominant microbiota population of the intestine, such as \textit{Bacteroidetes}, \textit{Firmicutes}, \textit{Actinobacteria}, and \textit{Proteobacteria}, have multiple mechanisms of antitumour activity. Various dietary components have direct effects on the types of intestinal microflora: in the Western diet mode (high-fat, high-protein, and red meat), the proportion of conditional pathogens in the intestinal flora increases, the proportion of commensal bacteriadecreases, and the occurrence of colorectal cancer is promoted. Conversely, a high-fibre diet can increase the abundance of \textit{Firmicutes} and reduce the abundance of \textit{Bacteroides} and consequently increase the concentration of short-chain fatty acids (SCFAs) in the intestine, inhibiting the development of CRC. This article reviews the study of the relationship between diet, intestinal microbes, and the promotion or inhibition of CRC and analyses the relevant molecular mechanisms to provide ideas for the prevention and treatment of CRC.

1. Introduction

Colorectal cancer (CRC) is one of the world’s most common cancers. The incidence of CRC in the United States has ranked in the top three for several consecutive years, with the American Cancer Society estimating that in 2019 CRC will constitute the third most common cause of cancer cases and deaths for both sexes [1].

CRC is not only high in incidence but also high in mortality rate. In the 2018 Global Cancer Survey, CRC mortality ranks second (9.2%) [2], and the age of onset is gradually decreasing [3, 4]. Early CRC is generally asymptomatic, and once symptoms appear the cancer is usually well advanced, so it can only be prevented by regular screening [5]. CRC has many causes, and it is generally believed that genetics, age, environment, lifestyle, and other factors affect its occurrence [6–8]. Among these other factors, dietary patterns have received the most recent attention [9–12].

Different dietary components have different effects on the potential to develop CRC. It is generally believed that a high-fat, high-protein diet including red meat and processed meats promotes the development of CRC, whilst a high-fibre diet inhibits its occurrence [13–17]. High-fat consumption increases the synthesis of intrahepatic bile acids, which are transported to the colon and metabolised by the microbial community into products with tumorigenic activity (secondary bile acids) in the colon [18–20]. Red meat, processed meat, and protein are enzymatically digested by the gut into toxic nitrogen and sulfur-containing substances in the large intestine, which promote CRC [11, 21].

In contrast, soluble fibre is fermented into short-chain fatty acids (SCFAs) by bacteria in the large intestine, and
SCFAs including butyrate serve as the main source of energy for colon cells and play an important role in the energy homeostasis of colon tissue [22]. Moreover, a high-fibre intake can increase the number of butyrate-producing bacteria in the gut, such as Clostridium, Anaerostipes, and Eubacterium species [23].

Human studies, preclinical studies, and epidemiological studies have demonstrated a correlation between gut microbiota and CRC [24–28], with the composition of microbes in the intestines or faeces of patients with CRC and precancerous lesions being different from that of healthy control subjects [29, 30]. Consistent data show that the microbiota of patients with CRC is rich in Fusobacterium nucleatum, Bacteroides fragilis, and Escherichia coli, whereas butyrate-producing bacteria such as Bacteroidetes, Fimicutes, Echinococcus, and Proteobacteria are depleted in the microbiota of cancer patients [24, 31].

Herein, we provide an overview of the effects of diet on CRC and its possible molecular mechanisms, review the evidence that the intestinal microbiota affects CRC, and discuss potential interactions between diet, the gut microbiota, and CRC (Figure 1).

2. Effects of Different Dietary Patterns on the Occurrence of Colorectal Cancer

2.1. High-Fat Diet. In 1973, Drasar and Irving first proposed that colorectal cancer and fat are highly correlated [32], which was consistent with the earlier experimental results of Gregor et al. [33] and Wyndr and Shigematsu [34]. Gregor’s study highlighted that the mortality of gastric cancer and colorectal cancer was related to animal protein intake, and Wyndr and Shigematsu found that the inhabitants of areas with a high incidence of colorectal cancer usually had a high-fat diet pattern. Related research in this area has since burgeoned, confirming Drasar and Irving’s original hypothesis.

We know that fat absorption needs to increase bile flow and that a high-fat diet affects bile metabolism. Data indicate that a high-fat diet increases bile acid concentrations in the faeces [35], and Narisawa et al. [36] reported in 1974 the role of bile acids in the colon promoting cancer development in rats. In 1990, Walter C. Willett et al. [37] conducted used questionnaires to survey and follow up more than 80,000 women of different ages and found that within this population, a large intake of animal fat increased the risk of colon cancer. A study in 2000 confirmed that increased deoxybile acid levels in bile acids were associated with an increased risk of colon cancer, and the transporter gene of bile acid was identified in Clostridium sp. strain TO-93 isolated from human faeces [38].

In 2011, researchers conducted experiments on wild-type mice to prove that deoxycholic acid derived from components of a high-fat diet is a colon carcinogen and that some dietary antioxidants can enhance this carcinogenicity [39]. High-fat diets can cause intestinal flora imbalance, increase intestinal permeability and reduce intestinal barrier function [40]. In this way, harmful bacterial products such as lipopolysaccharides (LPS) can enter the intestinal circulation and cause inflammation [41, 42]. Viggiano et al. [43] found that adenosine monophosphate-dependent kinase (AMPK) activation, inflammation and oxidative stress were increased in the hypothalamus of lard-fed rats compared with those fed a control diet, whereas no changes were observed in rats fed fish oil, suggesting that a saturated fat-based diet promotes hypothalamic inflammation and oxidative stress. Studies have also shown that, in an inflammatory environment, a high-fat diet can cause an increase in the concentration of E. coli, an increase in the thickness of the mucus layer, an increase in intestinal permeability, upregulation of Nos2 and Tlr5 expression, and promotion of TNF-α secretion. These changes lead to the colonisation of the gut by adherent-invasive Escherichia coli (AIEC) bacteria, which induce inflammation in the intestinal mucosa and promote the development of CRC [44].

In summary, high-fat diets promote CRC by affecting bile acid metabolism, damaging the integrity of the intestinal barrier and perturbing the intestinal microbiota.

2.2. Red Meat. Although academics still dispute whether typical consumption of red meat causes cancer [45], a growing number of convincing studies [21, 46] have shown that red meat and processed meat increase the risk of colon cancer.

In 2015, the International Agency for Research on Cancer (IARC) [47] evaluated red meat as “probably carcinogenic to humans” and processed meat as “carcinogenic to humans”, indicating that red meat consumption may be associated with CRC development. Researchers have studied the relationship between colon cancer and meat intake in more than 600 colon cancer cases and 1,000 controls collected from 1996 to 2000 [48] and found that when people eat more high-temperature-cooked red meat, they are more likely to develop colon cancer. This is suggested be at least partly due to the heterocyclic amines (HCAs) produced in this cooking mode, of which 2-amino-3,4,8-trimethylimidazo[4,5-f]quinazoline (DiMeIQx) is the most common.

Red meat is typically red because it is rich in myoglobin, and haem iron is extremely high in red meat myoglobin [49]. Corpet’s study [50] described the dose-response relationship between haem iron levels and colon cancer promotion, and their results in rat experiments suggest that haem iron promotes rectal cancer through direct or indirect effects. In addition, studies have shown that [51] trimethylamine N-oxide (TMAO) in high-fat diet mice can impair glucose tolerance, impede the hepatic insulin-signalling pathway, and lead to inflammation of adipose tissue. Studies by Xu et al. [52] have shown that TMAO is genetically associated with colorectal cancer and that TMAO may be an important intermediate marker linking the metabolism of meat and fat in the diet and gut microbiota to CRC risk.

A large number of previous studies have found that TMAO is produced by gut microbiota metabolism of dietary l-carnitine (a trimethylamine that is present in high levels in red meat) and phosphatidylcholine and is mechanistically linked to the risk of cardiovascular disease (CVD) [53–56]. Koeth et al. [55] showed that the intestinal microbial metabolism of l-carnitine in the diet also produces TMAO, which accelerates atherosclerosis in mice. Wilson Tang et al. proposed that dietary phosphatidylcholine conversion to
TMAO depends on the metabolism of intestinal microbiota [53].

In summary, the possible mechanisms of CRC caused by red meat-rich diet primarily involve HCAAs generated in the cooking process, haem iron, intestinal microorganisms, and their metabolites, such as TMAO.

2.3. High-Protein Diet. Epidemiological studies suggest that high-protein levels in the diet may increase the risk of colorectal cancer [57, 58], but this conclusion remains controversial [59, 60]. Shusuke Toden et al. first studied [61] the association between dietary-resistant starch (RS) and protein and genetic damage of colon cells in rats and obtained results that suggested that increased intake of protein (such as casein) may have deleterious effects on the intestine, whilst fermented complex carbohydrates (such as RS) can counteract these adverse effects in the intestine. Subsequent research by the same group [62] also demonstrated that increased DNA damage caused by high-protein diets (such as cooked red meat or casein) may increase the risk of CRC. Mireille Andriamihaja et al. [63] found that a high-protein diet caused a significant change in the luminal environment of colon cells and the characteristics of these cells, which suggests that a high-protein diet interferes with the metabolism and morphology of colon cells and that it is recommended to consider a high-protein diet cautiously.

Conversely, Sun et al. [64] found that a high-protein diet may actually reduce the risk of cancer. One possible explanation for this finding is that low protein intake may lead to abnormal DNA methylation [65], triggering deactivation of tumour-suppressor genes or related deleterious epigenetic changes.

The small intestine digests and absorbs protein more efficiently than other parts of the intestine (i.e., >95% of the protein that passes through the stomach) [66], so only a small number of proteins that are not completely digested by enzymes enter the large intestine and are microbially hydrolysed [67]. Bacteria such as *Bacillus*, *Streptococcus*, *Propionibacterium*, *Clostridium*, and *Bacteroides* can degrade proteins in the large intestine in addition to producing many SCFAs, and the large intestine also metabolises nitrogen and sulfur metabolites such as ammonia, amines, nitrates, nitrites, and hydrogen sulfide, all of which are strong carcinogens or genotoxic substances [11, 68].

In summary, the mechanisms by which a high-protein diet may cause CRC may involve DNA damage and epigenetic changes, interference with the metabolism and morphology of colon cells and toxic metabolites produced by intestinal microorganisms during protein digestion.

2.4. High-Fibre Diet. Most studies on dietary fibre and CRC risk have suggested that dietary fibre intake may reduce the risk of CRC [69–71]. In 1971, based on a survey of the prevalence of a high-fibre diet and of CRC in Africa, Burkitt [72] speculated that a high-fibre diet may reduce the incidence of colorectal cancer by swelling the faeces and promoting intestinal peristalsis. Hill [73] also determined that when fibre is not digested in the small intestine, bacteria...
ferment fibre in the colon, leading to an increase in faecal expansion weight and a shortening of the intestinal transit time, thereby accelerating the excretion of toxic substances.

A high-fibre diet also promotes the growth of beneficial flora in the gut, which binds to bile acids in the intestines and reduces levels of carcinogenic secondary bile acid production [74, 75]. In addition, dietary fibre is enzymatically hydrolysed in the colon to produce SCFAs, such as acetic acid, propionic acid and butyric acid. SCFAs have the function of keeping the intestinal flora stable, inhibiting the proliferation of cancer cells and promoting apoptosis [76, 77]. Butyrate acts as a major source of energy for intestinal epithelial cells and exerts anticancer effects via various pathways, such as upregulation of Bak gene expression and downregulation of Bcl-XL gene expression to induce apoptosis in human colon cancer cells [78]. Butyrate also increases antioxidant activity, thereby inhibiting formation of reactive nitrogen fragments (RONS), an important pathogenic factor in intestinal inflammation and CRC [79, 80], which functions by activating histone deacetylases (HDACs), leading to the proliferation of undifferentiated, highly proliferating colonic adenocarcinoma cells and inhibition of cell differentiation and apoptosis [81].

In summary, the possible mechanisms by which a high-fibre diet can prevent CRC may involve enhancing water-absorbing expansion of faecal matter to promote intestinal movement and accelerate the expulsion of carcinogens, promotion of the growth of beneficial flora to reduce the level of secondary bile acid, and the protective effect of SCFAs generated by fibre-processing by intestinal flora.

3. Relationship between Intestinal Microbes and CRC

The human intestine has a large number of microorganisms (78% of the total number of microorganisms in and on the human body) [82–84], and they have a symbiotic relationship with the human host. The microbial population promotes the growth and development of intestinal epithelial cells by participating in the metabolism of the three major nutrients (sugar, protein, fat) in the host to maintain the stability of the alimentary environment [85, 86], and the balance of intestinal microecology is the basis for the normal functioning of other organs [87, 88]. When infection, stress, eating habits, and other factors change, the intestinal microecological balance is perturbed, and the type, quantity, and structure of the intestinal flora changes, leading to an intestinal flora imbalance. This in turn affects the host’s health, potentially leading to disease such as CRC, liver cancer, and diabetes [89, 90].

3.1. Imbalance of Intestinal Microflora Induces Colorectal Cancer

The interaction between gut microbiota and CRC depends mainly on the balance of microbial populations. Microbial studies have shown that microbial composition is disturbed in colorectal cancer and precancerous lesions [29, 30]. Several studies have shown that Fusobacterium nucleatum is closely tied to human CRC.

Kostic et al. [91] described the composition of microflora in colorectal cancer by using the whole genome sequence of nine pairs of tumour group/normal group subjects. Castellari et al. [92] verified the overexpression of the Fusobacterium sequence in tumours relative to normal control tissues by quantitative PCR analysis of 99 subjects, observing a positive correlation of overexpression with lymph node metastasis. Related studies suggest that the abundance of Fusobacterium nucleatum (Fn) in stool samples of colon cancer patients is significantly higher than that in a healthy control group, and an enrichment of Fn is also found in colon cancer adenomas [93]. Hashemi Goradel et al. [94] summarised the possible mechanisms of Fn action in CRC in a 2018 publication, listing these as immunomodulation, virulence factors, miRNAs, and bacterial metabolism.

E. coli promotes the development of intestinal malignancy by inducing local inflammation. Lipopolysaccharides (LPS) produced by Gram-negative bacteria such as E. coli increase the expression of Toll-like receptor-4 (TLR4). Activation of TLR4 may promote the occurrence of CRC via Cox-2 expression enhancement and epidermal growth factor receptor (EGFR) signal transduction and other mechanisms [95]. Bacteriocin, a bactericidal protein produced by Enterobacteriaceae bacteria, is generally thought to be associated with some intestinal diseases, and the levels of Enterobacteriaceae are significantly elevated in patients with CRC [96]. Raisch et al. [88] showed by in vivo experiments with a model of chronic infection of CEACAM6-expressing mice that a B2 E. coli strain 11G5 was isolatable from colon cancer and persists in the intestine and induces colonic inflammation, epithelial damage and cell proliferation [97]. Clostridium coli and E. coli induce CRC mainly by inducing the inflammatory mode of the tumour microenvironment and producing carcinogenic substances [98, 99].

The study found that the colonic commensal bacteria Bacteroides fragilis [100], which is closely related to mucosal colonisation, is associated with acute diarrheal disease, inflammatory bowel disease and CRC [101–103]. In particular, high levels of Bacteroides fragilis toxin genes (BFT) have been found in advanced CRC [104], and this toxin can stimulate the production of interleukin 17 (IL-17), a proinflammatory cytokine, and thus induce inflammation. In addition, BFT degrades E-cadherin, disrupting epithelial homeostasis and leading to colonic epithelial cell proliferation and possibly CRC [104, 105]. A number of studies have shown that the synergistic effects of various bacterial strains in immune regulation may be involved in the promotion of colon tumorigenesis.

3.2. Intestinal Probiotics Inhibit Inflammation and Tumours

Intestinal probiotics can promote the maintenance of normal intestinal microecology and protect the host against CRC by generation of beneficial metabolites, immune tolerance induction and pathogen resistance [89, 106]. The dominant intestinal flora is Bacteroidetes, Firmicutes, Echinococcus, and Proteobacteria. The most common groups of probiotics are Lactobacillus and Bifidobacterium [107]. Lactobacillus can inhibit the expression of tumour-specific proteins and polysaccharides. Orlando et al. [108] found that the increase of Lactobacillus rhamnosus strain GG (L. GG) concentration has an effect on the growth and proliferation of HGC-27 (human
gastric cancer) and DLD-1 (human colonic cancer) cells and that L. GG can significantly reduce synthesis of polyamine metabolites.

Shida et al. [109] tested the interleukin 12 (IL-12) induction ability of 47 strains of Lactobacillus present in ten kinds of mouse peritoneal macrophages (IL-12 is a cytokine that plays a key role in activating innate immunity), and it was found that almost all strains belonged to the Lactobacillus casei group (L. casei, Lactobacillus rhamnosus, and Lactobacillus zae) and that Lactobacillus fermentum can induce high levels of IL-12. Ma et al. [110] studied the effect of Bacillus on tumour growth and found that Bacillus inhibits tumour growth by inhibiting ErbB2 and ErbB3, suggesting that Bacillus could be a clinical preventive measure for colon cancer. Kim et al. [111] evaluated the anticancer activity and bacterial enzyme inhibition of Bifidobacterium sp. SPM0212, which inhibited the proliferation of three human colon cancer cell lines (HT-29, SW480, and Caco-2), and found that B. adolescentis SPM0212 also inhibits harmful faecal enzymes.

4. Effects of Dietary Ingredients on Intestinal Microbes

Dietary factors have regulatory effects on intestinal symbiotic bacteria as a whole and on bacteria such as Lactobacillus and Clostridium. For example, long-term consumption of foods high in fat, sugar, protein, red meat, and processed meat can increase the proportion of opportunistic pathogens in intestinal flora and decrease the proportion of symbiotic bacteria [112]. However, a high-fibre diet can increase the abundance of Firmicutes, reduce the abundance of Bacteroidetes and increase the SCFA concentration in the intestinal tract, all of which can promote the efficacy of immunotherapy [113].

High-fat diets stimulate the metabolism of bile acids by microorganisms into carcinogens (such as secondary bile acids) or promote inflammation, increasing the risk of CRC [11, 59]. Bile acids are produced in the liver and are metabolised by microbes in the gut and regulate many metabolic pathways in the host. In addition, bile acids regulate the composition of gut microbes by activating the innate immune genes in the small intestine [114]. Higashimura et al. [115] reported the effects of intestinal dysregulation induced by a high-fat diet in mice, and the use of terminal restriction fragment-length polymorphism (t-rflp) to analyse faecal microbiota showed a decrease in lactic acid bacteria in the HFD group and an increase in the Clostridium difficile subgroup XIVa, and HPLC-MS determination of cecal organic acid and serum bile acid content found that secondary bile acid levels had increased. Similarly, another mouse study showed that the relative proportion of faecal F/B (the relative proportion of Firmicutes and Bacteroidetes) in the high-fat diet group was significantly higher than that in the low-fat diet group [116] and that the F/B ratio was considered to be a good indicator of significant changes in the composition of intestinal microbial community [117].

Earlier studies compared the faecal microbiota in African rural children and that in European children. High-throughput 16S rDNA sequencing and biochemical analysis revealed significant differences in the intestinal flora between the two groups: African children had significant enrichment in Bacteroides and reductions in Firmicutes and significantly higher expression of SCFAs than EU children [118]. Moreover, Enterobacteriaceae levels were significantly lower than in EU children. Further analysis showed that the Bacteroides levels in the intestine were associated with the consumption of animal protein and saturated fat level, supporting the contention that meat consumption in the Western diet is driving the intestinal phenotype. The researchers also noted that changes in the proportion of intestinal microbes may be related to a decrease in dietary fibre content and that the lack of colonic butyrate enhances the carcinogenic potential of meat and fat-stimulated metabolites [62, 119, 120].

Dietary fibre refers to the edible part of a plant or its extract or similar carbohydrate that is not absorbed and use in the small intestine, but is utilised by the microbial population that is resident after partial or complete fermentation in the large intestine [17]. Foods high in dietary fibre reduce the bacterial abundance of Bacteroides and Helicobacter species and enrich the levels of Ruminococi, which degrade dietary fibre and produce short-chain fatty acids [74].

O’Keefe et al. [121] conducted a 2-week dietary intervention in African Americans and rural occupants in South Africa. South African aborigines switched from a high-fibre diet to a low-fibre diet, whilst the African Americans switched from a low-fibre diet to a high-fibre diet. This resulted in an altered metabolism and metabolome in both groups, including a change in levels of SCFAs such as butyrate. Koropatkin et al. [122] found that adjusting the intake of RS or nonstarch polysaccharides in the diet can change the distribution of bacteria such as Ruminococcus and Eubacterium rectale, and analysis of faecal samples in vitro confirmed that these bacteria could selectively metabolise insoluble carbohydrates and metabolise polysaccharides by fermentation to produce SCFAs.

A mixture of inulin and oligofructose supplements in human diet experiments can stimulate Bifidobacteria, especially the proliferation of Bifidobacterium adolescentis and Faecalibacterium prausnitzii [123]. In addition, the SCFAs produced by intestinal flora metabolism can activate G-protein coupled receptors and regulatory T cells, increasing mucosal immune tolerance [124, 125]. Various dietary components can fuel different intestinal microflora and thus affect the metabolic function of the microorganisms, thereby regulating the hosts’ physiological functions.

5. Conclusions and Outlook

Diet is closely related to CRC pathogenesis: a high-fat diet increases bile acid concentration and damages the intestinal barrier and its function. Red meat and a high-protein diet favour the formation of nitrogen and sulfur metabolites, which are beneficial to the growth of proinflammatory intestinal bacteria. However, a high-fibre diet stimulates production of protective or anti-inflammatory components, such as SCFAs, that protect the gut.
Dietary changes can significantly affect gut health, and people with different dietary patterns will have different gut microbiota, and dietary components have been convincingly demonstrated to shape the gut microbiota throughout life. The microbiota can also affect the health of the host intestinal epithelial cells, and evidence even shows that the gut microbiota regulate the host’s satiety and brain function (denoted the visceral-cerebral axis) [126,127].

A series of interactions exist between dietary factors, gut microbiota and CRC. Excessive intake of saturated fats and processed meat lead to an increase in harmful bacteria such as *Fn* and *Ec*, which changes the permeability of intestinal mucosa, induces inflammation and promotes the development of CRC. People with a high-fibre diet therefore have a healthier intestinal microecology. Numerous studies have shown that dietary intervention and regulation of intestinal flora are the most cost-effective strategies for the prevention of CRC, and we strongly recommend a balanced diet rich in fibre.

**Conflicts of Interest**

The authors declare no conflicts of interest on this manuscript.

**Acknowledgments**

This work was financially supported by the Natural Science Foundation of China (41771300 and 41301274) and the Youth Innovation Team Project of Institute of Subtropical Agriculture, Chinese Academy of Sciences (2017QNCXTD.GTD).

**References**


