

Review Article

Current Status and Prospects of Spontaneous Peritonitis in Patients with Cirrhosis

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Spontaneous bacterial peritonitis (SBP) is a common cirrhotic ascites complication which exacerbates the patient's condition. SBP is caused by gram-negative bacilli and, to a lesser extent, gram-positive cocci. Hospital-acquired infections show higher levels of drug-resistant bacteria. Geographical location influences pathogenic bacteria distribution; therefore, different hospitals in the same country record different bacteria strains. Intestinal changes and a weak immune system in patients with liver cirrhosis lead to bacterial translocation thus causing SBP. Early diagnosis and timely treatment are important in SBP management. When the treatment effect is not effective, other rare pathogens should be explored.

1. Introduction

Spontaneous bacterial peritonitis (SBP) is a common complication in patients with liver cirrhosis and is recorded in 10–30% of hospitalized patients with cirrhotic ascites leading to sepsis or even death [1–4]. Studies show that bacterial translocation plays a key role in the occurrence and development of SBP [5, 6]. Bacterial translocation is caused by disorder of gut microflora, increased intestinal permeability, and host immunodeficiency [7, 8]. Although gram-negative bacilli are the main cause of SBP, infections due to gram-positive bacteria drug-resistant bacteria have been reported [9–11]. Therefore, it is important to understand the epidemiology and pathogenesis of SBP and develop effective therapy approaches.

2. Epidemiology

Geographical location affects SBP pathogen distribution with variations recorded among different hospitals in the same country. Gram-negative bacilli are the main SBP-causing pathogens, but infections of gram-positive cocci [12, 13], fungi, and some other rare pathogens cannot be ignored [14–18]. Increased use of broad-spectrum antibiotics and

prophylactic quinolones has led to the emergence of multidrug-resistant bacteria, especially in hospital-acquired infections [19–22]. Only 50–60% of SBP patients have positive ascites culture; therefore, pathogen identification is challenging [23]. These limitations hamper development of effective anti-infection therapy.

2.1. Asia. Li et al. [24] retrospectively analyzed 288 Chinese patients with spontaneous peritonitis from 2011 to 2013 and isolated 306 pathogenic bacteria, among which gram-negative bacteria, gram-positive bacteria, and fungi accounted for 58.2%, 27.8%, and 2.9% of the isolates. The main pathogenic bacteria were *Escherichia coli*, *Klebsiella pneumoniae*, *Enterococcus*, and *Staphylococcus aureus*. Of the 306 pathogenic bacteria, 99 cause nosocomial infections and 207 were community-acquired and play a role in other infection pathogenesis. *Escherichia coli* and *K. pneumoniae* produce more broad-spectrum β -lactamase in nosocomial infections compared with nonnosocomial infections. Piperacillin/tazobactam combination is a more effective therapy for nonhospital infections than nosocomial infections caused by *E. coli*. The authors reported that the pathogenic bacteria causing abdominal infection in patients with liver cirrhosis were mainly gram-negative, and the drug resistance rate of

nosocomial infection was significantly higher compared with the rate for nonnosocomial infection.

In another retrospective study, Ding et al. [25] analyzed the etiology of 334 Chinese patients with SBP from 2012 to 2016 and arrived at a similar conclusion. A total of 334 pathogenic bacteria were isolated, including 178 gram-negative bacteria and 138 gram-positive bacteria. The main pathogens were *E. coli*, *K. pneumoniae*, and *Enterococcus faecium*. The proportion of *Enterococci* in patients with hospital-acquired SBP was significantly higher than in those with community-acquired SBP. Pathogens isolated from nosocomial infections showed significantly higher resistance to first-line recommended drugs and were associated with poor prognosis.

In a retrospective cohort study in South Korea, Cheong et al. [21] analyzed the microbial characteristics of 236 patients with SBP from 2000 to 2007: *E. coli* accounted for 43.2%, *Klebsiella* accounted for 14.0% while *Streptococcus* accounted for 9.8% of the total bacteria population. The resistance rate of G⁻ to third generation cephalosporins and quinolones for hospital-acquired infections was significantly higher compared with that for community-acquired infections. In another study, Choi et al. [15] found 43 cases of SBP caused by *Aeromonas aerobius* as a result of weather changes between 1997 and 2006. Hwang et al. [26] reported that *Candida* infection was the main causative agent of fungal spontaneous peritonitis in Korea from 2000 to 2005.

2.2. Europe. In a Spanish retrospective study from 2001 to 2009, 34.6% of the 200 SBP patients showed community-acquired infections while 26.8% of these infections were hospital acquired. The third-generation cephalosporin resistance rate was 7.1% for the community-acquired infections and 40.9% for the hospital-acquired infections. These drug-resistant cases were mainly a result of gram-negative bacilli and *Enterococci* that produce extended-spectrum β -lactamases. Previous use of cephalosporins, diabetes, upper gastrointestinal bleeding, and nosocomial-acquired infections are risk factors for the development of drug-resistant bacterial infections [27]. Fernandez et al. [28] analyzed bacterial infection in 507 Spanish patients with liver cirrhosis and ascites admitted to hospital during 2005–2007 and 2010–2011 in a prospective study. 35% of hospital-acquired patients had higher number of drug-resistant strains compared with those with community-acquired infections (4%). Moreover, SBP mortality caused by drug-resistant bacteria was significantly higher.

Friedrich et al. [29] retrospectively analyzed the etiology of the first occurrence of SBP in 311 German patients with liver cirrhosis from 2007 to 2013. Gram-positive bacteria accounted for 47.8% of the total infections, gram-negative bacteria accounted for 44.9% while fungi accounted for 7.2% of the infections. In this study, *Enterobacter*, *Enterococcus*, and *Staphylococcus* were the most common isolates. Third-generation cephalosporins were effective in 70.2% of non-hospital-acquired SBP patients and in 56.3% of hospital-acquired SBP patients. In another prospective study from Germany, Lutz et al. [30] analyzed 86 German SBP patients from 2012 to 2016 and obtained similar results. *E.*

coli, *Klebsiella*, *Enterococcus*, and *Streptococcus* were the most common isolates. The resistance rate of nosocomial bacteria was higher than that of healthcare-related bacteria.

Bert et al. [31] analyzed 95 cases of hospital-acquired and community-acquired bacterial peritonitis in France from 1998 to 1999. A total of 78 pathogenic bacteria were isolated, of which 34 were *Streptococcus* spp. and 23 were *E. coli*. Streptococci are more common in community-acquired infections while gram-negative bacteria are more common in hospital-acquired infections. Another prospective observational study in France in 2005, involving 331 patients with SBP at 25 medical centers, revealed 222 gram-negative bacilli, mainly *E. coli*, *Enterobacter*, *K. pneumoniae*, and *P. aeruginosa*; 148 gram-positive cocci, mainly *Streptococcus*, *Enterococcus faecalis*, *Enterococcus faecium*, and *Staphylococcus aureus* while all 19 strains of fungi were *Candida albicans* [32]. Imipenem is an effective treatment for *P. aeruginosa* hospital-acquired infections [32].

Piroth et al. [33] retrospectively analyzed 114 strains of SBP in five hospitals in France from 2006 to 2007. *Staphylococci* and *E. coli* were the most common pathogens. Notably, 28% patients infected by the *E. coli* strain showed resistance to amoxicillin+clavulanic acid, and 27% of patients infected with *S. aureus* were resistant to methicillin. An observational study carried out in France in 2010 and 2011 showed that of the 57 confirmed SBP cases, gram-positive cocci (64.9%) were the main causative pathogens, including coagulase-negative *Staphylococci*, *Enterococci*, *Streptococci*, *Staphylococcus aureus*, and *Streptococcus pneumoniae* [13]. Another study on SBP patients in France reported that gram-positive bacteria were the dominant strains, accounting for 70% of nosocomial infections [34].

Gunjaca and Francetić [35] prospectively studied 108 cases of cirrhosis in Croatia, where SBP prevalence was 21% and the mortality was 26%. The pathogens causing SBP were mainly gram-negative bacteria such as *E. coli*, methicillin-resistant *S. aureus* (MRSA), and *Acinetobacter*.

Alexopoulou et al. [36] retrospectively carried out a study on 47 SBP patients in Greece from 2008 to 2011. Twenty-eight patients had medically related infections and 15 were treated with quinolone prophylaxis. Gram-positive coccus was the most commonly isolated pathogen. Nine isolates were multidrug-resistant bacteria, including *K. pneumoniae*-producing carbapenemase and *E. coli*- and *P. aeruginosa*-producing ultrabroad spectrum β -lactamase. Higher number of gram-negative bacteria was reported in hospital-associated infections compared with gram-positive cocci. Another Greek prospective study from 2012 to 2014 included 130 SBP patients with a 30-day follow-up. The results showed that gram-positive cocci (GPC) were the causative agents for half of the cases. Multidrug-resistant (MDR) strains comprised 20.8% of the total cases while 10% were extensively drug resistant (XDR). Drug-resistant bacteria showed a significant increase in mortality rates [37].

2.3. America. Chaulk et al. [38] retrospectively analyzed 192 Canadian SBP patients from 2003 to 2011. Among them, 77 patients had culture-positive infection with gram-positive bacteria causing 57% of these cases. The antibiotic

TABLE 1: Pathogens associated with spontaneous peritonitis in cirrhosis.

Country/author/year	Pathogens	Type of study	G ⁻	G ⁺	HA SBP	CA SBP
China/Li et al./2011-2013	306	Retrospective	58.2%	27.8%	99	207
China/Ding et al./2102-2016	334	Retrospective	52.3%	41.3%	155	179
Korea/Cheong/2000-2007	236	Retrospective	72.9%	22.9%	126	110
Germany/Friedrich/2007-2013	114	Retrospective	44.9%	47.8%	—	—
France/Bert/1998-1999	78	Retrospective	44.9%	51.3%	39	39
France/Montravers/2005.1-2005.7	829	Prospective	41%	27%	540	289
France/Piroch/2010-2011	268	Prospective	34%	64.9%	109	159
Canada/Chaulk/2003-2011	77	Retrospective	27%	57%	52	25

G⁻: gram-negative bacteria; G⁺: gram-positive bacteria; HA: hospital acquired; CA: community acquired; SBP: spontaneous bacterial peritonitis.

resistance rate was 8% in community-acquired infections and 41% in hospital-acquired infections (Table 1).

Ardolino et al. [39] retrospectively studied 160 SBP cases in the United States from 2005 to 2015. This study reports that gram-negative bacteria were mainly *E. coli*. The sensitivity rate to ceftriaxone was 71%. Gram-positive cocci including *Enterococci*, *Streptococcus*, and *Staphylococcus* accounted for 37.5% of the cases. 71% of *Enterococci* were resistant to vancomycin, and MRSA accounted for 80% of the infections.

Reddy et al. [40] reported a rare case of SBP caused by the *Salmonella enteritidis* group b in a patient with liver cirrhosis in the United States. Wu and Giri [41] first reported a case of SBP caused by *Haemophilus paraphilus*. Later, the patient also developed tuberculous peritonitis, a combination that had not been reported before. Emily and Maraj [42] reported cases of SBP with *Lactobacillus* as the pathogen. *Lactobacillus paracasei* was isolated from the abdominal cavity of a 73-year-old American man with liver cirrhosis. This strain was resistant to carbapenem antibiotics. Further, the patient eventually developed hepatorenal syndrome and succumbed to acute renal failure. Toyoshima et al. [43] reported SBP cases caused by *Listeria monocytogenes* in two patients with liver cirrhosis in Brazil. Third-generation cephalosporins are not effective for *Listeria* infections.

2.4. Africa. Oladimeji et al. [44] conducted a retrospective analysis of 31 patients with ascites in Nigeria from 2009 to 2010. In these SBP patients, the main pathogens were *E. coli* and *Klebsiella*. The gram-positive bacteria implicated in SBP infections were mainly *Streptococcus* and *Staphylococcus aureus*. Zaki et al. [45] explored the bacterial and fungal causes of SBP in an Egyptian population comprising 100 SBP patients. In this population, the pathogens were mainly gram-positive coccus (48.8%), gram-negative bacillus (12.2%), and 7.3% were *Mycobacterium tuberculosis*. Mohamed et al. [46] performed SBP screening on 3000 cirrhosis patients with ascites and pleural effusion in Egypt. SBP prevalence in patients with cirrhosis was reported to be 1.6% with the main causative pathogens being *E. coli* and *K. pneumoniae*.

3. Pathogenesis

Intestinal flora is considered as an important component of the intestinal barrier [47]. Changes to the gut microbiota are implicated in the SBP occurrence and progression [48–51].

Therefore, exploring the role of intestinal flora on SBP pathogenesis is the key in development of effective prevention and treatment strategies. For patients with liver cirrhosis, bacterial translocation (BT) as a result of intestinal gram-negative *Enterobacteriaceae* infections is the main cause of SBP occurrence and development [6, 52, 53]. Previous studies have shown that gastrointestinal stasis due to portal hypertension in patients with liver cirrhosis, intestinal bacterial overgrowth due to low levels of bile acid and gastric acid, delayed intestinal transport, altered intestinal permeability, and immune dysfunction promote BT and ultimately SBP [5, 7, 8] (Figure 1).

3.1. Small Intestinal Bacterial Overgrowth (SIBO). Cirrhosis results in small intestinal bacterial overgrowth [54–56], especially in patients with ascites and SBP history [57]. Overgrowth of small intestinal bacteria is implicated in bacterial translocation and SBP [58]. In a previous study, Bauer et al. reported that small intestinal bacterial overgrowth (SIBO) in patients with cirrhosis has no effect on spontaneous bacterial peritonitis [59]. However, in a subsequent study, he carried out quantitative culturing of jejunal secretion in 53 cirrhosis patients with a 1-year follow-up. In his findings, he reported that SIBO was present in 59% of the cirrhosis patients he examined and was associated with systemic endotoxemia [60]. Fukui et al. [61] also reported an increase in gram-negative bacteria represented by *E. coli* resulting in high levels of lipopolysaccharides (LPS) and endotoxemia in patients with liver disease. BT or microbial translocation is defined as the migration of surviving microorganisms or bacterial products (i.e., bacterial LPS, peptidoglycans, and lipopeptides) from the intestinal lumen to the mesenteric lymph nodes and other external intestinal sites [62–66]. In addition, studies have shown that small bowel transport is significantly longer in patients with SIBO [67]. Animal experiments by Pérez-Paramo et al. [68] reported that intestinal overgrowth and severe impairment of intestinal permeability in cirrhotic rats with ascites cause bacterial translocation and SIBO was associated with insufficient intestinal motility. In recent studies, gastrointestinal stasis due to portal hypertension, relative lack of bile and gastric acid secretion, intestinal dyskinesia, and long-term use of broad-spectrum antibiotics in patients with liver cirrhosis are implicated in increased intestinal aerobic bacteria and colonic bacterial migration to the jejunum and duodenum.

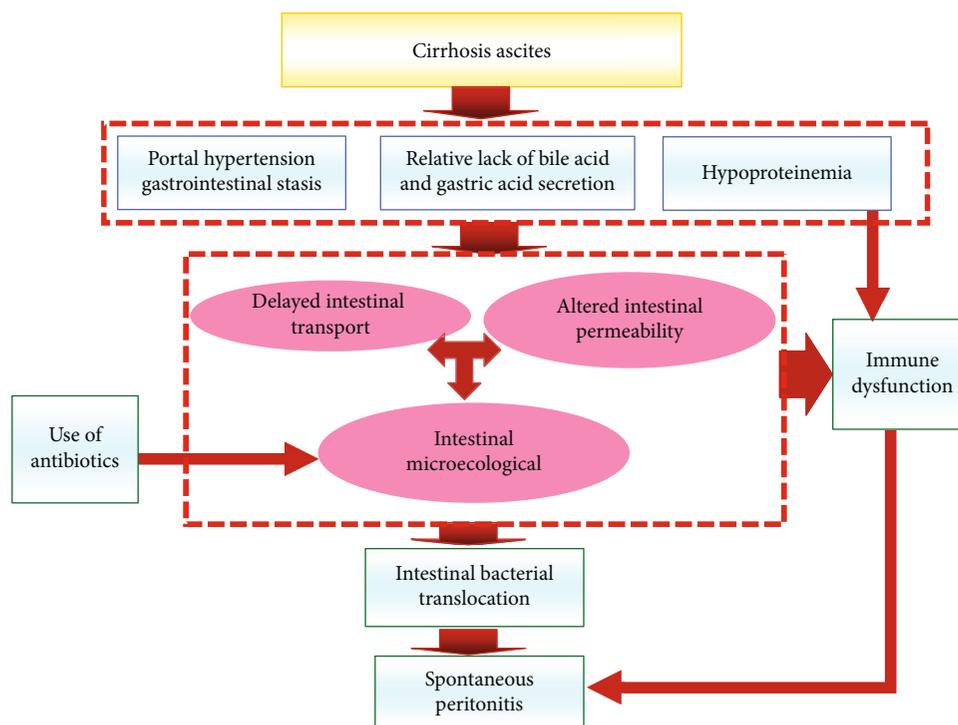


FIGURE 1: The pathogenesis of spontaneous peritonitis.

These changes further cause SIBO and promote BT, which is implicated in SBP prognosis in patients with liver cirrhosis [7]. Notably, the most common pathogenic microorganisms were isolated from the intestinal flora of cirrhotic ascites in SBP patients [69]. Interestingly, quantitative metagenomics analysis showed that some of the bacteria in SIBO were oral strains. Qin et al. [70] proposed that oral symbiotic bacteria in liver cirrhosis patients invaded the intestine as a result of bile secretion changes in these patients. The changes in bile secretion created a more favorable environment for the survival of foreign bacteria in the intestinal tract. Pardo et al. [54] also reported that cisapride increases BT from the oral cavity to the cecum. The use of cisapride in cirrhotic rats showed reduction of SIBO and occurrence of BT.

3.2. Altered Intestinal Permeability. The human intestinal mucosa mechanical barrier is the first barrier against BT and consists of intestinal epithelial cells and cell-to-cell connections [71–73]. The intestinal barrier system of intestinal epithelial cells prevents the transportation of a large number of bacteria and bacterial products; therefore, few bacteria and bacterial products reach the liver [74]. Tight junctions between cells are the key in maintaining integrity of the intestinal barrier, and reduction in density of these tight junctions impairs the function of the intestinal barrier [75, 76]. Assimakopoulos et al. [77] reported that expression levels of proteins associated with tight junctions in intestinal epithelial cells were lower in cirrhosis patients compared with patients with decompensated cirrhosis. Animal experiments [78] show that the intestinal mucosa of rats with liver cirrhosis shows signs of atrophy, shortening, and villus rupture. Capsule endoscopy studies show abnormal changes in the

mucosa of the small intestine in cirrhosis patients [79] while pathological examination shows shortening and atrophy of the small intestine [80, 81]. However, Du Plessis et al. [82] reported that electron microscopy showed complete epithelial barriers in patients with decompensated cirrhosis, implying that the epithelial barrier was functionally altered but structurally normal in cirrhosis. The contrasting findings may be due to differences in methodology and the relatively small number of studies/patients [83]. Assimakopoulos et al. [84] performed duodenal biopsies on healthy controls and patients with cirrhosis and decompensated cirrhosis. In this study, patients with decompensated and decompensated cirrhosis had decreased intestinal mucosa mitosis and increased cell apoptosis compared with the control group. Intestinal permeability changes with progression of cirrhosis and occurrence of SIBO, with increased intestinal permeability of bacteria and their products resulting in BT [83, 85, 86]. Several studies report that cirrhosis and ascites patients have significantly high intestinal permeability, while the intestinal permeability of patients with Child–Pugh C is significantly higher than the permeability of those with Child–Pugh with A and B cirrhosis [87, 88]. For patients with SBP history, intestinal permeability is higher and can lead to severe sepsis complications [89, 90].

3.3. Delayed Bowel Transit. Studies show that liver cirrhosis changes intestinal motility [91]. Delayed movements of the small intestine can lead to SIBO and eventually cause BT [92]. A radiological examination by Kalaitzakis et al. [93] showed that intestinal transit time was prolonged in 38% patients with liver cirrhosis. Chen et al. [94] used a noninvasive hydrogen breath test and found that the intestinal transit

time of patients with decompensated cirrhosis was significantly longer compared with that of patients with decompensated cirrhosis. Further, the intestinal transit time was positively correlated with the severity of cirrhosis [95]. The small intestine transit delay and SIBO interact and activate each other [71]. Perez-Paramo et al. [68] reported that nonselective beta blocker (NSBB (propranolol)) treatment in cirrhotic animals significantly reduces portal vein pressure and accelerates intestinal transport. The rate of bacterial overgrowth and metastasis in liver cirrhosis cases is low; therefore, intestinal bacteria overgrowth is positively correlated with insufficient intestinal motility. Propranolol accelerates intestinal transport and reduces bacterial overgrowth and transfer rates. However, Mandorfer et al. [96] found that although NSBB can reduce the risk of portal vein pressure and esophageal varix bleeding in patients with liver cirrhosis, it can increase the rate of hemodynamic disorders and liver-renal syndrome in patients with liver cirrhosis and SBP. Animal experiment results show that cisapride accelerates the transit time, improves the permeability of the small intestine, and reduces BT [97].

3.4. Impaired Local and Systemic Immune Function. Although the intestinal immune system is the last line of defense in microbial invasion, it is the most important line of defense against intestinal microbial invasion. The interaction between intestinal flora and mucosal immune system is dynamic and complex [98]. Under normal physiological conditions, the microbiome can maintain a delicate balance with the mucosal immune system, which is extremely important for the host health [99]. Changes in the intestinal microenvironment causes excessive growth of opportunistic pathogenic bacteria and the reduction of symbiotic bacteria in critically ill patients. The changes aggravate mucosal immune dysfunction, promote the increase of intestinal BT, and eventually lead to intestinal infection [100–103].

Bacteria occur in the intestinal lymphoid tissue but do not harm the body, as they are usually effectively cleared by phagocytes [104]. Damage to the body's defense mechanisms also promotes subsequent infection of fluid in the peritoneal cavity [54]. Immune disorders in patients with cirrhosis are known as cirrhosis-associated immune dysfunction (CAID) [105]. Cirrhosis-related immune dysfunction and immunodeficiency are dynamic and result from liver inflammation driven primarily by monocytes/macrophages. The liver's mononuclear-phagocytic system function in patients with cirrhosis is impaired, leading to a decrease in the body's immune function and opsonin activity in the ascites [106]. This further reduces the level of bacteria removal leading to the body's inability to effectively remove pathogenic bacteria eventually causing bacterial translocation and ultimately results in SBP. Phagocytosis of hepatic macrophages in cirrhosis patients is lower compared with that in the healthy control group and is correlated with the severity of liver disease [107–110]. In addition, severe malnutrition in patients with cirrhosis also affects their immune system. Diet and nutrition are key factors in host-microbe interactions while starvation adversely affects intestinal mucosal integrity, epithelial cell proliferation, and mucin and anti-

microbial peptide synthesis. Hodin et al. [111] observed autophagy of Paneth cells in starved mice due to lack of enteral nutrition and decreased expression of antibacterial products. The poor nutrition weakened the protective effect on BT, thereby causing BT. Therefore, improving the nutritional status of patients with advanced cirrhosis improves the body's immune function and reduces the BT and SBP incidences. Albumin is specifically synthesized in the liver and is implicated in a myriad of functions such as the binding and transport of substances, the regulation of endothelial function, antioxidant and clearance properties, and the regulation of inflammatory responses. Serum albumin levels are low in liver cirrhosis patients due to synthetic defects, and structural and functional changes due to posttranscriptional modifications hinder their ability to perform physiological functions [112, 113].

4. Treatment

For patients with decompensated liver cirrhosis, spontaneous peritonitis can lead to further decompensation and multiple organ failure; therefore, SBP therapy is important for these patients. However, current methods are limited to antibiotic treatment, which leads to increases in drug-resistant bacteria and nonclassical pathogen infections [9–11]. Therefore, understanding the mechanism of SBP development, antibiotic treatment, new adjuvant treatment methods, and multiple treatment coordination are needed to minimize the occurrence of infection, reduce bacterial resistance, and improve survival.

4.1. Antibiotic Treatment. If the patient is clinically suspected of developing SBP, ascites culture should be performed immediately along with initiation of antibiotic treatment to reduce complications and improve survival [114, 115]. Third-generation broad-spectrum cephalosporin, cefixime, is the first-line treatment option for out-of-hospital SBP infection, with a recommended dose of 2 g/8 h (6 g/day) for 5 days [116, 117], which can be extended to 7 days [118]. Fluoroquinolones have good oral bioavailability and can be used as therapy for uncomplicated SBP [119]. Third-generation cephalosporin antibiotics and quinolones have been used to control SBP infection with high levels of clinical efficacy. However, long-term application increases the risk of bacterial resistance and double infection. Notably, *Enterobacteriaceae* family shows increased resistance to cephalosporins, particularly in nosocomial infections [120, 121]. Long-term preventive norfloxacin treatment reduces the risk of gram-negative infections but increases the risk of hospital-acquired *Staphylococcal* infections [122]. Therefore, considering that the distribution of SBP varies with geographic region and the proportion of drug-resistant pathogens is high, when selecting first-line empirical antibiotic treatment, the epidemic situation of drug-resistant bacteria should be based on the local situation [10]. Piperacillin/tazobactam is the first-line treatment for nosocomial SBP infection in areas with low resistance. Meropenem is recommended in hospitals with a high positive rate of ESBLs produced by *Enterobacteria* [30]. In areas with high prevalence of MRSA and

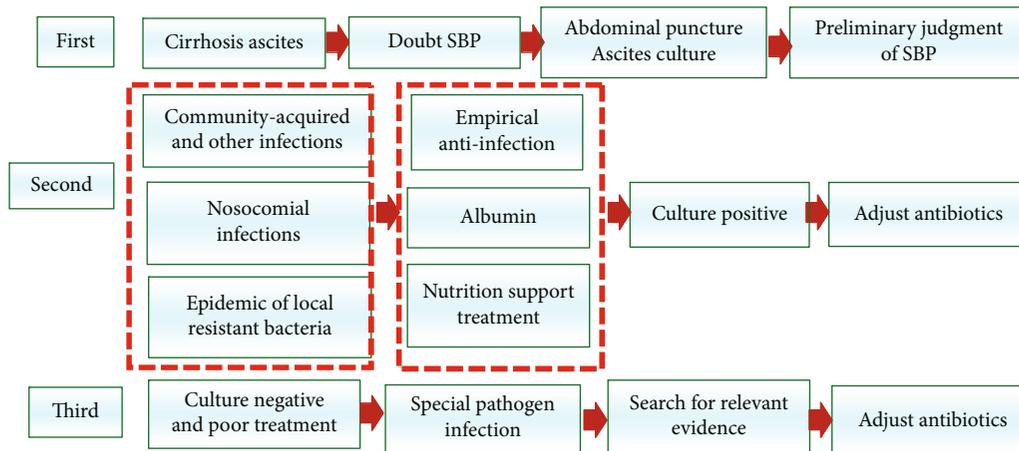


FIGURE 2: Treatment procedure of spontaneous peritonitis.

vancomycin-sensitive *Enterococcus* (VSE), a combination of meropenem and vancomycin or teicoplanin is recommended, while linezolid is recommended in case of vancomycin-resistant *Enterococcus* (VRE) [19]. In areas with high resistance to third-generation cephalosporins, meropenem combined with daptomycin can be used to improve patient survival of the nosocomial SBP [123]. If the ascites culture is positive, non-broad-spectrum antibiotics should be selected according to the drug sensitivity results to reduce the emergence of drug-resistant bacteria [115]. When antibiotic therapy fails in patients with spontaneous peritonitis, the possibility of fungal or other rare pathogens should be considered [14, 26, 124].

4.2. Gut Microecological Intervention. Intestinal bacteria are the main source of infections in patients with decompensated cirrhosis; therefore, norfloxacin is often used to clear the intestines for preventive treatment. However, antibiotic prevention can lead to increase in drug-resistant bacteria [125, 126]. Therefore, prevention is limited to a small number of patients with a high risk of infection. Probiotics can competitively inhibit adhesion to epithelial cells through competitive nutrients, reduce intestinal pH, and secrete antibacterial compounds to inhibit the growth of harmful pathogenic microorganisms. On the contrary, probiotics improve the intestinal mucosal barrier function and regulate the liver's natural killing of T lymphocytes [127]. Studies have reported that probiotics can reduce BT and effectively prevent the occurrence of hepatic encephalopathy [128]. Rat models with cirrhosis show that probiotics reduce BT, proinflammatory response status, formation of ascites, and oxidative damage in the ileum [129]. In a previous study, *Bifidobacterium* was shown to reduce the expression of proinflammatory chemokine receptors in the lymphocytes of mice with liver cirrhosis. Thus, the intestinal permeability of mice treated with *Bifidobacterium* was reduced while the liver function and inflammatory response improved [65]. The use of probiotics in liver-damaged rats alters the host's intestinal environment and reduces the occurrence of BTs [6, 130]. In a randomized double-blind controlled experiment, Gupta et al. [66]

reported that the hepatic vein pressure gradient in the probiotic group was significantly lower compared with the propranolol group and that the addition of probiotics increased the effectiveness of propranolol treatment. However, a randomized controlled trial by Pande et al. [131] showed that the addition of probiotics to norfloxacin had no significant effect on SBP prevention in cirrhosis and ascites patients. Although more studies should be carried out needed to support the application of probiotic therapy in the prevention or management of SBP, previous studies report that probiotic therapy is effective in managing gastrointestinal diseases.

4.3. Immunity Therapy. In addition to intestinal targeting methods, immunotherapy methods have been developed to reduce the susceptibility of patients with decompensated cirrhosis to infection. In addition to antibiotics, albumin is a key therapy for SBP patients as it restores the immune function and improves survival [132]. Studies have found that infusion of human albumin reduces immunosuppression and the risk of infection in patients with acute decompensated cirrhosis [9, 133]. Combination of antibiotics and albumin significantly reduces serum and ascites cytokines and LPS levels in patients with SBP [134]. Caraceni et al. [135] evaluated 440 patients with decompensated liver cirrhosis who received standard treatment or standard treatment plus albumin. The 18-month survival rate of the treatment group was significantly higher compared with that of the standard treatment group. Sort et al. [136] randomly divided 126 patients with SBP; the mortality rate of the antibiotic plus albumin group was lower compared with that of the antibiotic group. Although the role of albumin is beneficial, not all patients with SBP can be treated with albumin, and patients with bile $< 68.4 \mu\text{mol/L}$ and creatinine $< 88.4 \mu\text{mol/L}$ cannot receive albumin treatment [136, 137]. Most patients with advanced liver cirrhosis are malnourished, which can easily lead to BT and SBP [138]. Patients with liver cirrhosis should optimize nutrition, avoid raw foods and coarse superfoods, limit sodium intake, eat small meals, and include 1.2-1.5 g of protein daily [139]. Cytokine treatments can improve the function of existing immune cells, significantly increase peripheral

white blood cell counts, and improve the prognosis of patients with decompensated cirrhosis [140, 141]; however, more experimental and clinical evidence is needed.

5. Conclusion

Spontaneous bacterial peritonitis causes high mortality rates and occurs in 7-31% of hospitalized patients with cirrhosis and ascites [142]. Patients susceptible to SBP need rigorous evaluation to optimize nutrition and avoid unnecessary drug treatment [12]. When patients with cirrhosis and ascites are hospitalized for gastrointestinal and parenteral diseases, ascites analysis should be performed whether symptoms are present or not. The long-term use of antibiotics has led to the emergence of multidrug-resistant bacteria and recent changes in the bacterial spectrum, including increased incidence of SBP associated with gram-positive cocci. Therefore, patients with cirrhosis and ascites should be monitored keenly and early diagnosis and treatment of SBP are important to prevent poor prognosis. A good understanding of the epidemiology of the region is the key to the correct choice of antibiotics. When encountering cases with poor treatment results, it is necessary to consider the possibility of other rare pathogens such as fungi and adjust the treatment strategy. Therapy approaches should include improved nutrition support to enhance the immunity of patients and comprehensive treatment should be considered for better results (Figure 2). SBP prevention should focus on stabilizing the intestinal environment, restoring the balance of intestinal flora, and reducing the occurrence of BT.

Conflicts of Interest

The authors declare no competing interests.

Authors' Contributions

Li YT wrote the paper. Huang JR and Peng ML have revised the paper for final approval.

References

- [1] A. A. De Mattos, A. M. Costabeber, L. C. Lionço, and C. V. Tovo, "Multi-resistant bacteria in spontaneous bacterial peritonitis: new step in management?," *World Journal of Gastroenterology*, vol. 20, no. 39, pp. 14079–14086, 2014.
- [2] P. Ginès, P. Angeli, K. Lenz et al., "EASL clinical practice guidelines on the management of ascites, spontaneous bacterial peritonitis, and hepatorenal syndrome in cirrhosis," *Journal of Hepatology*, vol. 53, no. 3, pp. 397–417, 2010.
- [3] K. H. J. Lim, J. R. Potts, J. Chetwood, S. Goubet, and S. Verma, "Long-term outcomes after hospitalization with spontaneous bacterial peritonitis," *Journal of Digestive Diseases*, vol. 16, no. 4, pp. 228–240, 2015.
- [4] L. Titó, A. Rimola, P. Ginès, J. Llach, V. Arroyo, and J. Rodés, "Recurrence of spontaneous bacterial peritonitis in cirrhosis: Frequency and predictive factors," *Hepatology*, vol. 8, no. 1, pp. 27–31, 1988.
- [5] P. Bellot, R. Francés, and J. Such, "Pathological bacterial translocation in cirrhosis: pathophysiology, diagnosis and clinical implications," *Liver International*, vol. 33, no. 1, pp. 31–39, 2013.
- [6] R. Wiest and G. Garcia-Tsao, "Bacterial translocation (BT) in cirrhosis," *Hepatology*, vol. 41, no. 3, pp. 422–433, 2005.
- [7] G. Ghosh and A. B. Jesudian, "Small intestinal bacterial overgrowth in patients with cirrhosis," *Journal of Clinical and Experimental Hepatology*, vol. 9, no. 2, pp. 257–267, 2019.
- [8] C. Wang, Q. Li, and J. Ren, "Microbiota-immune interaction in the pathogenesis of gut-derived infection," *Frontiers in Immunology*, vol. 10, 2019.
- [9] J. Fernández, V. Prado, J. Trebicka et al., "Multidrug-resistant bacterial infections in patients with decompensated cirrhosis and with acute-on-chronic liver failure in Europe," *Journal of Hepatology*, vol. 70, no. 3, pp. 398–411, 2019.
- [10] S. Piano, V. Singh, P. Caraceni et al., "Epidemiology and effects of bacterial infections in patients with cirrhosis worldwide," *Gastroenterology*, vol. 156, no. 5, pp. 1368–1380.e10, 2019.
- [11] J. Fernández, M. Navasa, J. Gómez et al., "Bacterial infections in cirrhosis: epidemiological changes with invasive procedures and norfloxacin prophylaxis," *Hepatology*, vol. 35, no. 1, pp. 140–148, 2002.
- [12] J. B. Dever and M. Y. Sheikh, "Review article: Spontaneous bacterial peritonitis - bacteriology, diagnosis, treatment, risk factors and prevention," *Alimentary Pharmacology & Therapeutics*, vol. 41, no. 11, pp. 1116–1131, 2015.
- [13] L. Piroth, A. Pechinot, V. Di Martino et al., "Evolving epidemiology and antimicrobial resistance in spontaneous bacterial peritonitis: a two-year observational study," *BMC Infectious Diseases*, vol. 14, no. 1, 2014.
- [14] M. Fiore and S. Leone, "Spontaneous fungal peritonitis: epidemiology, current evidence and future prospective," *World Journal of Gastroenterology*, vol. 22, no. 34, pp. 7742–7747, 2016.
- [15] J. P. Choi, S. O. Lee, H. H. Kwon et al., "Clinical significance of spontaneous aeromonas bacterial peritonitis in cirrhotic patients: a matched case-control study," *Clinical Infectious Diseases*, vol. 47, no. 1, pp. 66–72, 2008.
- [16] S. Jayasinghe, M. Connor, S. Donaldson, H. Austin, and A. Foster, "Spontaneous bacterial peritonitis due to *Listeria monocytogenes*: importance of enrichment culture," *Journal of Clinical Pathology*, vol. 63, no. 9, pp. 835–836, 2010.
- [17] R. Hörner, A. Salla, L. O. de Oliveira et al., "Spontaneous bacterial peritonitis caused by *Streptococcus bovis*: case report and review of the literature," *Brazilian Journal of Infectious Diseases*, vol. 14, no. 3, pp. 294–296, 2010.
- [18] N. R. Dlamini, A. Bhamjee, P. Levick, E. Uniacke, H. Ismail, and A. Smith, "Spontaneous bacterial peritonitis and pneumonia caused by *Bordetella bronchiseptica*," *The Journal of Infection in Developing Countries*, vol. 607, pp. 588–591, 2012.
- [19] K. A. Rostkowska, A. Szymanek-Pasternak, and K. A. Simon, "Spontaneous bacterial peritonitis – therapeutic challenges in the era of increasing drug resistance of bacteria," *Clinical and Experimental Hepatology*, vol. 4, no. 4, pp. 224–231, 2018.
- [20] N. Singh, M. M. Wagener, and T. Gayowski, "Changing epidemiology and predictors of mortality in patients with spontaneous bacterial peritonitis at a liver transplant unit¹," *Clinical Microbiology and Infection*, vol. 9, no. 6, pp. 531–537, 2003.

- [21] H. S. Cheong, C.-I. Kang, J. A. Lee et al., "Clinical significance and outcome of nosocomial acquisition of spontaneous bacterial peritonitis in patients with liver cirrhosis," *Clinical Infectious Diseases*, vol. 48, no. 9, pp. 1230–1236, 2009.
- [22] J. D. Pitout and K. B. Laupland, "Extended-spectrum β -lactamase-producing Enterobacteriaceae: an emerging public-health concern," *The Lancet Infectious Diseases*, vol. 8, no. 3, pp. 159–166, 2008.
- [23] Y. H. Park, H. C. Lee, H. G. Song et al., "Recent increase in antibiotic-resistant microorganisms in patients with spontaneous bacterial peritonitis adversely affects the clinical outcome in Korea," *Journal of Gastroenterology and Hepatology*, vol. 18, no. 8, pp. 927–933, 2003.
- [24] Y.-T. Li, "Pathogen profile and drug resistance analysis of spontaneous peritonitis in cirrhotic patients," *World Journal of Gastroenterology*, vol. 21, no. 36, pp. 10409–10417, 2015.
- [25] X. Ding, Y. H. Yu, M. Chen, C. Wang, Y. F. Kang, and J. L. Lou, "Causative agents and outcome of spontaneous bacterial peritonitis in cirrhotic patients: community-acquired versus nosocomial infections," *BMC Infectious Diseases*, vol. 19, no. 1, 2019.
- [26] S. Y. Hwang, S. J. Yu, J. H. Lee et al., "Spontaneous fungal peritonitis: a severe complication in patients with advanced liver cirrhosis," *European Journal of Clinical Microbiology & Infectious Diseases*, vol. 33, no. 2, pp. 259–264, 2014.
- [27] X. Ariza, J. Castellote, J. Lora-Tamayo et al., "Risk factors for resistance to ceftriaxone and its impact on mortality in community, healthcare and nosocomial spontaneous bacterial peritonitis," *Journal of Hepatology*, vol. 56, no. 4, pp. 825–832, 2012.
- [28] J. Fernández, J. Acevedo, M. Castro et al., "Prevalence and risk factors of infections by multiresistant bacteria in cirrhosis: a prospective study," *Hepatology*, vol. 55, no. 5, pp. 1551–1561, 2012.
- [29] K. Friedrich, S. Nüssle, T. Rehlen, W. Stremmel, A. Mischnik, and C. Eisenbach, "Microbiology and resistance in first episodes of spontaneous bacterial peritonitis: implications for management and prognosis," *Journal of Gastroenterology and Hepatology*, vol. 31, no. 6, pp. 1191–1195, 2016.
- [30] P. Lutz, H. D. Nischalke, B. Krämer et al., "Antibiotic resistance in healthcare-related and nosocomial spontaneous bacterial peritonitis," *European Journal of Clinical Investigation*, vol. 47, no. 1, pp. 44–52, 2017.
- [31] F. Bert, M. Andreu, F. Durand et al., "Nosocomial and community-acquired spontaneous bacterial peritonitis: comparative microbiology and therapeutic implications," *European Journal of Clinical Microbiology and Infectious Diseases*, vol. 22, no. 1, pp. 10–15, 2003.
- [32] P. Montravers, A. Lepape, L. Dubreuil et al., "Clinical and microbiological profiles of community-acquired and nosocomial intra-abdominal infections: results of the French prospective, observational EBIIA study," *Journal of Antimicrobial Chemotherapy*, vol. 63, no. 4, pp. 785–794, 2009.
- [33] L. Piroth, A. Pechinot, A. Minello et al., "Bacterial epidemiology and antimicrobial resistance in ascitic fluid: a 2-year retrospective study," *Scandinavian Journal of Infectious Diseases*, vol. 41, no. 11-12, pp. 847–851, 2009.
- [34] B. Campillo, J.-P. Richardet, T. Kheo, and C. Dupeyron, "Nosocomial spontaneous bacterial peritonitis and bacteremia in cirrhotic patients: impact of isolate type on prognosis and characteristics of infection," *Clinical Infectious Diseases*, vol. 35, no. 1, pp. 1–10, 2002.
- [35] I. Gunjača and I. Francetić, "Prevalence and clinical outcome of spontaneous bacterial peritonitis in hospitalized patients with liver cirrhosis: a prospective observational study in central part of Croatia," *Acta Clinica Croatica*, vol. 49, no. 1, pp. 11–18, 2010.
- [36] A. Alexopoulou, N. Papadopoulos, D. G. Eliopoulos et al., "Increasing frequency of gram-positive cocci and gram-negative multidrug-resistant bacteria in spontaneous bacterial peritonitis," *Liver International*, vol. 33, no. 7, pp. 975–981, 2013.
- [37] A. Alexopoulou, L. Vasilieva, D. Agiasotelli et al., "Extensively drug-resistant bacteria are an independent predictive factor of mortality in 130 patients with spontaneous bacterial peritonitis or spontaneous bacteremia," *World Journal of Gastroenterology*, vol. 22, no. 15, pp. 4049–4056, 2016.
- [38] J. Chaulk, M. Carbonneau, H. Qamar et al., "Third-generation cephalosporin-resistant spontaneous bacterial peritonitis: a single-centre experience and summary of existing studies," *Canadian Journal of Gastroenterology and Hepatology*, vol. 28, no. 2, pp. 83–88, 2014.
- [39] E. Ardolino, S. S. Wang, and V. R. Patwardhan, "Evidence of significant ceftriaxone and quinolone resistance in cirrhotics with spontaneous bacterial peritonitis," *Digestive Diseases and Sciences*, vol. 64, no. 8, pp. 2359–2367, 2019.
- [40] K. R. Reddy, J. C. Chan, D. Smiley, L. J. Jeffers, and E. R. Schiff, "Spontaneous group B Salmonella enteritidis peritonitis in cirrhotic ascites and acquired immune deficiency syndrome," *The American Journal of Gastroenterology*, vol. 83, no. 8, pp. 882–884, 1988.
- [41] D. C. Wu and B. Giri, "*Haemophilus paraprophilus* Peritonitis Followed by Tuberculous Peritonitis and Pott's Disease," *The American Journal of the Medical Sciences*, vol. 340, no. 6, pp. 511–513, 2010.
- [42] E. Harding-Theobald and B. Maraj, "Spontaneous bacterial peritonitis due to *Lactobacillus paracasei* in cirrhosis," *Case Reports in Gastrointestinal Medicine*, vol. 2018, 2 pages, 2018.
- [43] M. T. K. Toyoshima, A. Apanavicius, A. de Matos Soeiro, G. M. D. de Almeida, and M. H. Arai, "*Listeria monocytogenes* peritonitis in cirrhotic patients: first description in Brazil," *Revista do Instituto de Medicina Tropical de São Paulo*, vol. 48, no. 5, pp. 291–293, 2006.
- [44] A. O. Ajayi, P. T. Adegun, E. A. Ajayi, H. T. Raimi, and S. A. Dada, "Prevalence of spontaneous bacterial peritonitis in liver cirrhosis with ascites," *Pan African Medical Journal*, vol. 15, 2013.
- [45] M. E. S. Zaki, W. O. El Shabrawy, M. M. El-Eshmawy, and S. A. Eletreby, "The high prevalence of *Listeria monocytogenes* peritonitis in cirrhotic patients of an Egyptian Medical Center," *Journal of Infection and Public Health*, vol. 4, no. 4, pp. 211–216, 2011.
- [46] A. Mohamed, M. Atef, A. Alsebaey, M. Musa Elhabshy, and M. Salama, "Combined spontaneous bacterial empyema and peritonitis in cirrhotic patients with ascites and hepatic hydrothorax," *Arab Journal of Gastroenterology*, vol. 18, no. 2, pp. 104–107, 2017.
- [47] P. B. Eckburg, E. M. Bik, C. N. Bernstein et al., "Diversity of the human intestinal microbial flora," *Science*, vol. 308, no. 5728, pp. 1635–1638, 2005.
- [48] J. P. Nolan, "The role of intestinal endotoxin in liver injury: a long and evolving history," *Hepatology*, vol. 52, no. 5, pp. 1829–1835, 2010.

- [49] S. R. Gill, M. Pop, R. T. Deboy et al., "Metagenomic analysis of the human distal gut microbiome," *Science*, vol. 312, no. 5778, pp. 1355–1359, 2006.
- [50] R. Wiest, A. Krag, and A. Gerbes, "Spontaneous bacterial peritonitis: recent guidelines and beyond," *Gut*, vol. 61, no. 2, pp. 297–310, 2011.
- [51] D. Benten and R. Wiest, "Gut microbiome and intestinal barrier failure - The "Achilles heel" in hepatology?," *Journal of Hepatology*, vol. 56, no. 6, pp. 1221–1223, 2012.
- [52] G. Garcia-Tsao and R. Wiest, "Gut microflora in the pathogenesis of the complications of cirrhosis," *Best Practice & Research Clinical Gastroenterology*, vol. 18, no. 2, pp. 353–372, 2004.
- [53] C. Guarner and G. Soriano, "Bacterial translocation and its consequences in patients with cirrhosis," *European Journal of Gastroenterology & Hepatology*, vol. 17, no. 1, pp. 27–31, 2005.
- [54] A. Pardo, R. Bartolí, V. Lorenzo-Zúñiga et al., "Effect of cisapride on intestinal bacterial overgrowth and bacterial translocation in cirrhosis," *Hepatology*, vol. 31, no. 4, pp. 858–863, 2000.
- [55] T. M. Bauer, H. Schwacha, B. Steinbrückner et al., "Diagnosis of small intestinal bacterial overgrowth in patients with cirrhosis of the liver: poor performance of the glucose breath hydrogen test," *Journal of Hepatology*, vol. 33, no. 3, pp. 382–386, 2000.
- [56] C. Pande, A. Kumar, and S. K. Sarin, "Small-intestinal bacterial overgrowth in cirrhosis is related to the severity of liver disease," *Alimentary Pharmacology & Therapeutics*, vol. 29, no. 12, pp. 1273–1281, 2009.
- [57] F. C. Morencos, G. de Las Heras Castaño, L. M. Ramos, M. J. López Arias, F. Ledesma, and F. P. Romero, "Small bowel bacterial overgrowth in patients with alcoholic cirrhosis," *Digestive Diseases and Sciences*, vol. 41, no. 3, pp. 552–556, 1996.
- [58] C. S. Chang, G. H. Chen, H. C. Lien, and H. Z. Yeh, "Small intestine dysmotility and bacterial overgrowth in cirrhotic patients with spontaneous bacterial peritonitis," *Hepatology*, vol. 28, no. 5, pp. 1187–1190, 1998.
- [59] T. M. Bauer, B. Steinbrückner, F. E. Brinkmann et al., "Small intestinal bacterial overgrowth in patients with cirrhosis: prevalence and relation with spontaneous bacterial peritonitis," *The American Journal of Gastroenterology*, vol. 96, no. 10, pp. 2962–2967, 2001.
- [60] T. M. Bauer, H. Schwacha, B. Steinbrückner et al., "Small intestinal bacterial overgrowth in human cirrhosis is associated with systemic endotoxemia," *The American Journal of Gastroenterology*, vol. 97, no. 9, pp. 2364–2370, 2002.
- [61] H. Fukui, B. Brauner, J. C. Bode, and C. Bode, "Plasma endotoxin concentrations in patients with alcoholic and non-alcoholic liver disease: reevaluation with an improved chromogenic assay," *Journal of Hepatology*, vol. 12, no. 2, pp. 162–169, 1991.
- [62] H. Fukui, "Gut-liver axis in liver cirrhosis: how to manage leaky gut and endotoxemia," *World Journal of Hepatology*, vol. 7, no. 3, pp. 425–442, 2015.
- [63] R. Wiest and H. C. Rath, "Bacterial translocation in the gut," *Best Practice & Research Clinical Gastroenterology*, vol. 17, no. 3, pp. 397–425, 2003.
- [64] M. R. Pinzone, B. M. Celesia, M. Di Rosa, B. Cacopardo, and G. Nunnari, "Microbial translocation in chronic liver diseases," *International Journal of Microbiology*, vol. 2012, Article ID 694629, 12 pages, 2012.
- [65] A. Moratalla, I. Gómez-Hurtado, Á. Moya-Pérez et al., "*Bifidobacterium pseudocatenulatum* CECT₇₇₆₅ promotes a TLR₂-dependent anti-inflammatory response in intestinal lymphocytes from mice with cirrhosis," *European Journal of Nutrition*, vol. 55, no. 1, pp. 197–206, 2016.
- [66] N. Gupta, A. Kumar, P. Sharma, V. Garg, B. C. Sharma, and S. K. Sarin, "Effects of the adjunctive probiotic VSL#3 on portal haemodynamics in patients with cirrhosis and large varices: a randomized trial," *Liver International*, vol. 33, no. 8, pp. 1148–1157, 2013.
- [67] A. Gupta, R. K. Dhiman, S. Kumari et al., "Role of small intestinal bacterial overgrowth and delayed gastrointestinal transit time in cirrhotic patients with minimal hepatic encephalopathy," *Journal of Hepatology*, vol. 53, no. 5, pp. 849–855, 2010.
- [68] M. Pérez-Paramo, J. Muñoz, A. Albillos et al., "Effect of propranolol on the factors promoting bacterial translocation in cirrhotic rats with ascites," *Hepatology*, vol. 31, no. 1, pp. 43–48, 2000.
- [69] G. B. Rogers, C. J. van der Gast, K. D. Bruce et al., "Ascitic microbiota composition is correlated with clinical severity in cirrhosis with portal hypertension," *PLoS One*, vol. 8, no. 9, p. e74884, 2013.
- [70] N. Qin, F. Yang, A. Li et al., "Alterations of the human gut microbiome in liver cirrhosis," *Nature*, vol. 513, no. 7516, pp. 59–64, 2014.
- [71] E. Kalaitzakis, "Gastrointestinal dysfunction in liver cirrhosis," *World Journal of Gastroenterology*, vol. 20, no. 40, pp. 14686–14695, 2014.
- [72] D. Artis, "Epithelial-cell recognition of commensal bacteria and maintenance of immune homeostasis in the gut," *Nature Reviews Immunology*, vol. 8, no. 6, pp. 411–420, 2008.
- [73] F. Bäckhed, R. E. Ley, J. L. Sonnenburg, D. A. Peterson, and J. I. Gordon, "Host-bacterial mutualism in the human intestine," *Science*, vol. 307, no. 5717, pp. 1915–1920, 2005.
- [74] Y. S. Seo and V. H. Shah, "The role of gut-liver axis in the pathogenesis of liver cirrhosis and portal hypertension," *Clinical and Molecular Hepatology*, vol. 18, no. 4, pp. 337–346, 2012.
- [75] M. Cereijido, R. G. Contreras, D. Flores-Benítez et al., "New diseases derived or associated with the tight junction," *Archives of Medical Research*, vol. 38, no. 5, pp. 465–478, 2007.
- [76] N. Sonoda, M. Furuse, H. Sasaki et al., "*Clostridium perfringens* enterotoxin fragment removes specific claudins from tight junction strands: evidence for direct involvement of claudins in tight junction barrier," *Journal of Cell Biology*, vol. 147, no. 1, pp. 195–204, 1999.
- [77] S. F. Assimakopoulos, A. C. Tsamandas, G. I. Tsiaoussis et al., "Altered intestinal tight junctions' expression in patients with liver cirrhosis: a pathogenetic mechanism of intestinal hyperpermeability," *European Journal of Clinical Investigation*, vol. 42, no. 4, pp. 439–446, 2012.
- [78] J. B. Wen, F. Q. Zhu, W. G. Chen et al., "Oxymatrine improves intestinal epithelial barrier function involving NF- κ B-mediated signaling pathway in CCl₄-induced cirrhotic rats," *PLoS One*, vol. 9, no. 8, article e106082, 2014.
- [79] G. D. De Palma, M. Rega, S. Masone et al., "Mucosal abnormalities of the small bowel in patients with cirrhosis and

- portal hypertension: a capsule endoscopy study," *Gastrointestinal Endoscopy*, vol. 62, no. 4, pp. 529–534, 2005.
- [80] M. Barakat, M. Mostafa, Z. Mahran, and A. G. Soliman, "Portal hypertensive duodenopathy: clinical, endoscopic, and histopathologic profiles," *The American Journal of Gastroenterology*, vol. 102, no. 12, pp. 2793–2802, 2007.
- [81] V. Misra, S. P. Misra, M. Dwivedi, and S. C. Gupta, "Histomorphometric study of portal hypertensive enteropathy," *American Journal of Clinical Pathology*, vol. 108, no. 6, pp. 652–657, 1997.
- [82] J. Du Plessis, H. Vanheel, C. E. I. Janssen et al., "Activated intestinal macrophages in patients with cirrhosis release NO and IL-6 that may disrupt intestinal barrier function," *Journal of Hepatology*, vol. 58, no. 6, pp. 1125–1132, 2013.
- [83] K. E. Pijls, D. M. A. E. Jonkers, E. E. Elamin, A. A. M. Masclee, and G. H. Koek, "Intestinal epithelial barrier function in liver cirrhosis: an extensive review of the literature," *Liver International*, vol. 33, 2013.
- [84] S. F. Assimakopoulos, A. C. Tsamandas, G. I. Tsiaoussis et al., "Intestinal mucosal proliferation, apoptosis and oxidative stress in patients with liver cirrhosis," *Annals of Hepatology*, vol. 12, no. 2, pp. 301–307, 2013.
- [85] P. Palma, N. Mihaljevic, T. Hasenberg, M. Keese, and T. A. Koepfel, "Intestinal barrier dysfunction in developing liver cirrhosis: an in vivo analysis of bacterial translocation," *Hepatology Research*, vol. 37, no. 1, pp. 6–12, 2007.
- [86] K. E. Pijls, G. H. Koek, E. E. Elamin, H. de Vries, A. A. M. Masclee, and D. M. A. E. Jonkers, "Large intestine permeability is increased in patients with compensated liver cirrhosis," *American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 306, no. 2, pp. G147–G153, 2014.
- [87] S. Pascual, J. Such, A. Esteban et al., "Intestinal permeability is increased in patients with advanced cirrhosis," *Hepatology*, vol. 50, no. 5, pp. 1482–1486, 2003.
- [88] S. Lee, S. C. Son, M. J. Han et al., "Increased intestinal macromolecular permeability and urine nitrite excretion associated with liver cirrhosis with ascites," *World Journal of Gastroenterology*, vol. 14, no. 24, pp. 3884–3890, 2008.
- [89] E. Scarpellini, V. Valenza, M. Gabrielli et al., "Intestinal permeability in cirrhotic patients with and without spontaneous bacterial peritonitis: is the ring closed?," *American Journal of Gastroenterology*, vol. 105, no. 2, pp. 323–327, 2010.
- [90] B. Campillo, P. Pernet, P. N. Bories, J. P. Richardet, M. Devanlay, and C. Aussel, "Intestinal permeability in liver cirrhosis: relationship with severe septic complications," *European Journal of Gastroenterology & Hepatology*, vol. 11, no. 7, pp. 755–760, 1999.
- [91] S. A. Gunnarsdottir, R. Sadik, S. Shev et al., "Small intestinal motility disturbances and bacterial overgrowth in patients with liver cirrhosis and portal hypertension," *The American Journal of Gastroenterology*, vol. 98, no. 6, pp. 1362–1370, 2003.
- [92] U. Thalheimer, F. De Iorio, F. Capra et al., "Altered intestinal function precedes the appearance of bacterial DNA in serum and ascites in patients with cirrhosis: a pilot study," *European Journal of Gastroenterology & Hepatology*, vol. 22, no. 10, pp. 1228–1234, 2010.
- [93] E. Kalaitzakis, R. Sadik, J. J. Holst, L. Ohman, and E. Björnsson, "Gut transit is associated with gastrointestinal symptoms and gut hormone profile in patients with cirrhosis," *Clinical Gastroenterology and Hepatology*, vol. 7, no. 3, pp. 346–352, 2009.
- [94] C. Y. Chen, C. L. Lu, F. Y. Chang et al., "The impact of chronic hepatitis B viral infection on gastrointestinal motility," *European Journal of Gastroenterology & Hepatology*, vol. 12, no. 9, pp. 995–1000, 2000.
- [95] B. C. Roland, G. Garcia-Tsao, M. M. Ciarleglio, Y. Deng, and A. Sheth, "Decompensated cirrhotics have slower intestinal transit times as compared with compensated cirrhotics and healthy controls," *Journal of Clinical Gastroenterology*, vol. 47, no. 10, pp. 888–893, 2013.
- [96] M. Mandorfer, S. Bota, P. Schwabl et al., "Nonselective β blockers increase risk for hepatorenal syndrome and death in patients with cirrhosis and spontaneous bacterial peritonitis," *Gastroenterology*, vol. 146, no. 7, pp. 1680–1690.e1, 2014.
- [97] S. C. Zhang, W. Wang, W. Y. Ren, B. M. He, K. Zhou, and W. N. Zhu, "Effect of cisapride on intestinal bacterial and endotoxin translocation in cirrhosis," *World Journal of Gastroenterology*, vol. 9, no. 3, pp. 534–538, 2003.
- [98] C. L. Maynard, C. O. Elson, R. D. Hatton, and C. T. Weaver, "Reciprocal interactions of the intestinal microbiota and immune system," *Nature*, vol. 489, no. 7415, pp. 231–241, 2012.
- [99] J. L. Round and S. K. Mazmanian, "The gut microbiota shapes intestinal immune responses during health and disease," *Nature Reviews Immunology*, vol. 9, no. 5, pp. 313–323, 2009.
- [100] M. Ojima, D. Motooka, K. Shimizu et al., "Metagenomic analysis reveals dynamic changes of whole gut microbiota in the acute phase of intensive care unit patients," *Digestive Diseases and Sciences*, vol. 61, no. 6, pp. 1628–1634, 2016.
- [101] J. M. Lankelma, L. A. van Vught, C. Belzer et al., "Critically ill patients demonstrate large interpersonal variation in intestinal microbiota dysregulation: a pilot study," *Intensive Care Medicine*, vol. 43, no. 1, pp. 59–68, 2017.
- [102] P. T. McKenney and E. G. Pamer, "From hype to hope: the gut microbiota in enteric infectious disease," *Cell*, vol. 163, no. 6, pp. 1326–1332, 2015.
- [103] M. C. Jacobs, B. W. Haak, F. Hugenholtz, and W. J. Wiersinga, "Gut microbiota and host defense in critical illness," *Current Opinion in Critical Care*, vol. 23, no. 4, pp. 257–263, 2017.
- [104] C. J. O'Boyle, J. MacFie, C. J. Mitchell, D. Johnstone, P. M. Sagar, and P. C. Sedman, "Microbiology of bacterial translocation in humans," *Gut*, vol. 42, no. 1, pp. 29–35, 1998.
- [105] K. M. Irvine, I. Ratnasekera, E. E. Powell, and D. A. Hume, "Causes and consequences of innate immune dysfunction in cirrhosis," *Frontiers in Immunology*, vol. 10, 2019.
- [106] F. Cereto, I. Molina, A. González et al., "Role of immunosuppression in the development of quinolone-resistant *Escherichia coli* spontaneous bacterial peritonitis and in the mortality of *E. coli* spontaneous bacterial peritonitis," *Alimentary Pharmacology and Therapeutics*, vol. 17, no. 5, pp. 695–701, 2003.
- [107] J. C. Nieto, E. Sánchez, C. Romero et al., "Impaired innate immune response of leukocytes from ascitic fluid of patients with spontaneous bacterial peritonitis," *Journal of Leukocyte Biology*, vol. 98, no. 5, pp. 819–825, 2015.
- [108] K. M. Irvine, X. Banh, V. L. Gadd et al., "CR1g-expressing peritoneal macrophages are associated with disease severity in patients with cirrhosis and ascites," *JCI Insight*, vol. 1, no. 8, p. e86914, 2016.

- [109] C. H. Huang, W. J. Jeng, Y. P. Ho et al., "Increased EMR2 expression on neutrophils correlates with disease severity and predicts overall mortality in cirrhotic patients," *Scientific Reports*, vol. 6, no. 1, 2016.
- [110] H. Fukui and R. Wiest, "Changes of intestinal functions in liver cirrhosis," *Inflammatory Intestinal Diseases*, vol. 1, no. 1, pp. 24–40, 2016.
- [111] C. M. Hodin, K. Lenaerts, J. Grootjans et al., "Starvation compromises Paneth cells," *The American Journal of Pathology*, vol. 179, no. 6, pp. 2885–2893, 2011.
- [112] M. Domenicali, M. Baldassarre, F. A. Giannone et al., "Post-transcriptional changes of serum albumin: clinical and prognostic significance in hospitalized patients with cirrhosis," *Hepatology*, vol. 60, no. 6, pp. 1851–1860, 2014.
- [113] F. A. Giannone, M. Domenicali, M. Baldassarre et al., "Ischaemia-modified albumin: a marker of bacterial infection in hospitalized patients with cirrhosis," *Liver International*, vol. 35, no. 11, pp. 2425–2432, 2015.
- [114] J. Felisart, A. Rimola, V. Arroyo et al., "Cefotaxime is more effective than is ampicillin–tobramycin in cirrhotics with severe infections," *Hepatology*, vol. 5, no. 3, pp. 457–462, 1985.
- [115] R. Jalan, J. Fernandez, R. Wiest et al., "Bacterial infections in cirrhosis: a position statement based on the EASL Special Conference 2013," *Journal of Hepatology*, vol. 60, no. 6, pp. 1310–1324, 2014.
- [116] B. A. Runyon, J. G. McHutchison, M. R. Antillon, E. A. Akriviadis, and A. A. Montano, "Short-course versus long-course antibiotic treatment of spontaneous bacterial peritonitis: A randomized controlled study of 100 patients," *Gastroenterology*, vol. 100, no. 6, pp. 1737–1742, 1991.
- [117] B. A. Runyon, "management of adult patients with ascites due to cirrhosis: an update," *Hepatology*, vol. 49, no. 6, pp. 2087–2107, 2009.
- [118] J. Castellote, A. Girbau, X. Ariza et al., "Usefulness of reagent strips for checking cure in spontaneous bacterial peritonitis after short-course treatment," *Alimentary Pharmacology & Therapeutics*, vol. 31, no. 1, pp. 125–130, 2010.
- [119] B. A. Runyon, "Introduction to the revised American Association for the Study of Liver Diseases Practice Guideline management of adult patients with ascites due to cirrhosis 2012," *Hepatology*, vol. 57, no. 4, pp. 1651–1653, 2013.
- [120] C. Dupeyron, B. Campillo, N. Mangeney, M. Bordes, J.-P. Richardet, and G. Leluan, "Carriage of *Staphylococcus aureus* and of Gram-Negative bacilli resistant to Third-Generation cephalosporins in cirrhotic Patients a prospective assessment of Hospital-Acquired infections," *Infection Control & Hospital Epidemiology*, vol. 22, no. 7, pp. 427–432, 2001.
- [121] M. Fiore, I. Gentile, A. E. Maraolo et al., "Are third-generation cephalosporins still the empirical antibiotic treatment of community-acquired spontaneous bacterial peritonitis? A systematic review and meta-analysis," *European Journal of Gastroenterology & Hepatology*, vol. 30, no. 3, pp. 329–336, 2018.
- [122] B. Campillo, C. Dupeyron, J. P. Richardet, N. Mangeney, and G. Leluan, "Epidemiology of severe Hospital-Acquired infections in patients with liver cirrhosis: effect of long-term administration of norfloxacin," *Clinical Infectious Diseases*, vol. 26, no. 5, pp. 1066–1070, 1998.
- [123] S. Piano, S. Fasolato, F. Salinas et al., "The empirical antibiotic treatment of nosocomial spontaneous bacterial peritonitis: results of a randomized, controlled clinical trial," *Hepatology*, vol. 63, no. 4, pp. 1299–1309, 2016.
- [124] T. Tariq, F. B. Irfan, M. Farishta, B. Dykstra, E. M. Sieloff, and A. P. Desai, "Spontaneous fungal peritonitis: micro-organisms, management and mortality in liver cirrhosis—a systematic review," *World Journal of Hepatology*, vol. 11, no. 7, pp. 596–606, 2019.
- [125] K. Yan and G. Garcia-Tsao, "Novel prevention strategies for bacterial infections in cirrhosis," *Expert Opinion on Pharmacotherapy*, vol. 17, no. 5, pp. 689–701, 2016.
- [126] P. Zapater, J. M. Gonzalez-Navajas, J. Such, and R. Frances, "Immunomodulating effects of antibiotics used in the prophylaxis of bacterial infections in advanced cirrhosis," *World Journal of Gastroenterology*, vol. 21, no. 41, pp. 11493–11501, 2015.
- [127] S. Liang, T. Webb, and Z. Li, "Probiotic antigens stimulate hepatic natural killer T cells," *Immunology*, vol. 141, no. 2, pp. 203–210, 2014.
- [128] M. K. Lunia, B. C. Sharma, P. Sharma, S. Sachdeva, and S. Srivastava, "Probiotics prevent hepatic encephalopathy in patients with cirrhosis: a randomized controlled trial," *Clinical Gastroenterology and Hepatology*, vol. 12, no. 6, pp. 1003–1008.e1, 2014.
- [129] E. Sánchez, J. C. Nieto, A. Boullosa et al., "VSL#3 probiotic treatment decreases bacterial translocation in rats with carbon tetrachloride-induced cirrhosis," *Liver International*, vol. 35, no. 3, pp. 735–745, 2015.
- [130] Y. T. Li, L. Wang, Y. Chen et al., "Effects of gut microflora on hepatic damage after acute liver injury in rats," *The Journal of Trauma: Injury, Infection, and Critical Care*, vol. 68, no. 1, pp. 76–83, 2010.
- [131] C. Pande, A. Kumar, and S. K. Sarin, "Addition of probiotics to norfloxacin does not improve efficacy in the prevention of spontaneous bacterial peritonitis," *European Journal of Gastroenterology & Hepatology*, vol. 24, no. 7, pp. 831–839, 2012.
- [132] C. Bernsmeier, A. Singanayagam, V. C. Patel, J. Wendon, and C. G. Antoniades, "Immunotherapy in the treatment and prevention of infection in acute-on-chronic liver failure," *Immunotherapy*, vol. 7, no. 6, pp. 641–654, 2015.
- [133] L. China, S. S. Skene, Z. Shabir et al., "Administration of albumin solution increases serum levels of albumin in patients with chronic liver failure in a single-arm feasibility trial," *Clinical Gastroenterology Hepatology*, vol. 16, no. 5, pp. 748–755.e6, 2018.
- [134] T. A. Chen, Y. C. Tsao, A. Chen et al., "Effect of intravenous albumin on endotoxin removal, cytokines, and nitric oxide production in patients with cirrhosis and spontaneous bacterial peritonitis," *Scandinavian Journal of Gastroenterology*, vol. 44, no. 5, pp. 619–625, 2009.
- [135] P. Caraceni, O. Riggio, P. Angeli et al., "Long-term albumin administration in decompensated cirrhosis (ANSWER): an open-label randomised trial," *Lancet*, vol. 391, no. 10138, pp. 2417–2429, 2018.
- [136] P. Sort, M. Navasa, V. Arroyo et al., "Effect of intravenous albumin on renal impairment and mortality in patients with cirrhosis and spontaneous bacterial peritonitis," *New England Journal of Medicine*, vol. 341, no. 6, pp. 403–409, 1999.

- [137] S. H. Sigal, C. M. Stanca, J. Fernandez, V. Arroyo, and M. Navasa, "Restricted use of albumin for spontaneous bacterial peritonitis," *Gut*, vol. 56, no. 4, pp. 597–599, 2007.
- [138] B. Campillo, J. P. Richardet, E. Scherman, and P. N. Bories, "Evaluation of nutritional practice in hospitalized cirrhotic patients: results of a prospective study," *Nutrition*, vol. 19, no. 6, pp. 515–521, 2003.
- [139] A. O'Brien and R. Williams, "Nutrition in end-stage liver disease: principles and practice," *Gastroenterology*, vol. 134, no. 6, pp. 1729–1740, 2008.
- [140] V. Garg, H. Garg, A. Khan et al., "Granulocyte colony-stimulating factor mobilizes CD34⁺ cells and improves survival of patients with acute-on-chronic liver failure," *Gastroenterology*, vol. 142, no. 3, pp. 505–512.e1, 2012.
- [141] C. K. Kedarisetty, L. Anand, A. Bhardwaj et al., "Combination of granulocyte colony-stimulating factor and erythropoietin improves outcomes of patients with decompensated cirrhosis," *Gastroenterology*, vol. 148, no. 7, pp. 1362–1370.e7, 2015.
- [142] M. Borzio, F. Salerno, L. Piantoni et al., "Bacterial infection in patients with advanced cirrhosis: a multicentre prospective study," *Digestion of Liver Disease*, vol. 33, no. 1, pp. 41–48, 2001.