

Research Article

Lack of Association between Common Polymorphisms in Selenoprotein P Gene and Susceptibility to Colorectal Cancer, Breast Cancer, and Prostate Cancer: A Meta-Analysis

Hanjiang Xu,^{1,2,3} Fan Mo^(b),^{1,2,3} Jun Zhou,^{1,2,3} Zongyao Hao,^{1,2,3} Xianguo Chen^(b),^{1,2,3} and Chaozhao Liang^(b),^{1,2,3}

¹Department of Urology, The First Affiliated Hospital of Anhui Medical University, Hefei 230000, China ²Institute of Urology, Anhui Medical University, Hefei 230000, China ³Anhui Province Key Laboratory of Genitourinary Diseases, Anhui Medical University, Hefei 230000, China

Correspondence should be addressed to Xianguo Chen; cxg7866186@126.com and Chaozhao Liang; liang_chaozhao@163.com

Received 12 May 2021; Revised 30 August 2021; Accepted 7 September 2021; Published 27 September 2021

Academic Editor: Syed Sameer Aga

Copyright © 2021 Hanjiang Xu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background and Objective. Selenoprotein P (SEPP1) is the major selenoprotein in plasma. Previous studies have demonstrated that SEPP1 expression was reduced in human prostate and colon tumors. Nowadays, studies concerning *SEPP1* gene polymorphisms and cancer susceptibility have been extensively investigated, whereas results from these studies remain debatable rather than conclusive. Thus, we performed the present meta-analysis to comprehensively assess the association between two common polymorphisms (rs3877899 and rs7579) in *SEPP1* and cancer susceptibility. *Method.* We search the PubMed, Embase, Google Scholar, and Wanfang (China) databases (up to December 1, 2020) to identify all eligible publications. The pooled odds ratio (OR) correspondence with 95% confidence interval (CI) was calculated to evaluate the associations. *Results.* Finally, nine eligible studies with 7,157 cases and 6,440 controls and five studies with 2,278 cases and 2,821 controls were enrolled in rs3877899 and rs7579 polymorphisms, individually. However, a null significant association was detected between the two polymorphisms in *SEPP1* and susceptibility to colorectal, breast, and prostate cancer in all comparison models. Subsequently, subgroup analysis based on tumor type, no significant association was identified for prostate, breast, and colorectal cancer. In addition, when the stratification analyses were conducted by the source of control, HWE status, and ethnicity, yet no significant association was found. *Conclusions.* The current meta-analysis shows that SEPP1 rs3877899 and rs7579 polymorphisms and succeptibility to colon cancer, breast cancer, and prostate cancer.

1. Introduction

There has been a progressive increase in the global incidence of malignancies, causing a serious threat to human health, presently, among the main causes of death [1]. Increasing evidence suggests that cancers are multifactorial diseases, which derive from complex coactions between genetic and environmental factors [2].

Oxidative stress, which causes mitochondrial damage and DNA breakage by reactive oxygen species (ROS), is closely related to tumor progression [3, 4]. ROS, like hydrogen peroxide (H_2O_2), can cause DNA damage due to the continuous production of various cellular metabolic processes in the body, which may lead to malignant transformation of cells [5]. Selenoprotein P (SEPP1) is the dominant selenoprotein in plasma as two isoforms (~50 kDa and ~60 kDa) and is believed to have two main functions: providing tissues with selenium for tissues and exerting antioxidant defense capabilities [6]. The insufficiency of SEPP1 may participate in the occurrence and progression of cancer. Earlier studies have testified high expression of SEPP1 in colonic mucosa and relatively lower expression of SEPP1 in human colon tumors. Moreover, SEPP1 affects colitisinduced tumorigenesis through regulating stemness and oxidative damage also has been confirmed in the study [7]. In Calvo et al.'s research, they advanced pointed that the SEPP1 was reduced in prostate cancer (PCa) [8]. Gonzalez-Moreno et al.'s study has shown that knockdown of SEPP1 expression in prostate epithelial neoplasia lesion cell lines and invasive tumors significantly increased ROS and cell growth inhibition after exposure to H_2O_2 [9].

Nowadays, more and more studies have demonstrated that several polymorphisms of the selenoprotein P gene (*SEPP1*) were associated with susceptibility of tumors, including breast cancer (BC) [10], colorectal cancer (CRC) [11], and PCa [12]. However, results from these studies remain inconclusive. In order to yield a more accurate and robust estimation, we conducted this meta-analysis trying to comprehensively analyze the connection between two common polymorphisms (rs3877899 and rs7579) in *SEPP1* and cancer susceptibility.

2. Materials and Methods

2.1. Search Strategy. We conducted this meta-analysis on the basis of the PRISMA meta-analysis guidelines [13]. A comprehensively retrieve of the literature concerning relationships between the *SEPP1* polymorphisms and cancer susceptibility was performed on PubMed, Embase, and Google Scholar databases (up to December 1, 2020) by using the following searching terms: "SEPP1 OR Selenoprotein P" AND "polymorphism OR variation OR SNP OR genotype OR allele OR mutation" AND "cancer OR malignancy OR tumor OR neoplasm OR carcinoma". We also conducted manual searches on the references of these selected original studies to identify other eligible studies.

2.2. Inclusion and Exclusion Criteria. Included literature should be in line with the following criteria: (1) studies that evaluated the relationship between SEPP1 polymorphisms (rs3877899 and rs7579) and cancer susceptibility; (2) sufficient genotype data from the text or the supporting information; (3) case-control studies. Moreover, these studies should also be excluded when they were as follows: (1) insufficient data; (2) not a case-control study, such as Comments, Case Reports, and Reviews; (3) the total scores of Newcastle-Ottawa Scale (NOS) is less than 5 (The quality of the enrolled studies was assessed by NOS (Newcastle-Ottawa Scale), which is presented in Table 1. In addition, the specific scoring rules of NOS are listed in Table S1.).

2.3. Data Extraction. Two reviewers (Hanjiang Xu and Fan Mo) have devoted themselves to the data extraction process referring to the predetermined criteria. All the discrepancies were settled through discussion till all consensus was settled. Furthermore, the following details should also be extracted: name of the first author, publication year, source of controls, ethnicity of a case-control study, genotype frequencies of cases and controls, and so on.

2.4. Statistical Analysis. We calculated the odds ratio (OR) with 95% CI confidence interval (CI) to appraise the intensity of relationships between *SEPP1* polymorphisms (rs3877899 and rs7579) and cancer susceptibility in the fol-

lowing genetic models: allele contrast (A vs. G), recessive (AA vs. AG+GG), dominant (AA+AG vs. GG), heterozygous (AG vs. GG), and homozygous (AA vs. GG) models (G: wild allele; A: variant allele). We assessed the statistical heterogeneity hypothesis through I^2 statistics to quantify the inconsistency, which represents the proportion of variability between studies that potentially arose from heterogeneity instead of contingency. I^2 values greater than 50% are considered to have significant heterogeneity [14], indicating the random-effects model would be selected to calculate the pooled OR estimated value of individual study; if not, the fixed-effects model was obtained. Our current study also assessed sensitivity analysis as well as publication bias [15]. We use Stata software for all statistical analysis (version 12.0; STATA Corp, College Station, TX). $P \leq 0.05$ was considered statistically significant.

3. Results

3.1. Characteristics of Studies. Overall, 10 publications with 14 independent studies on SEPP1 polymorphisms (rs3877899 and rs7579) and cancer susceptibility were available for our meta-analysis [10, 11, 16-23], and the publication selection process is displayed in Figure 1. For rs3877899 polymorphism, nine case-control studies with 7,157 cases and 6,440 controls met the inclusion criteria, including three BC, two CRC, and four PCa studies. For rs7579 polymorphism, there were five studies (one BC study, one PCa study, and three CRC studies) with 2,278 cases and 2,821 controls that met the eligibility criteria. Cancers were confirmed pathologically or histologically in most studies. The authors of included studies used a variety of genotyping methods, including PCR-RFLP and TaqMan. We think the earlier sentence was correct regarding the genotyping methods used in the included studies. Except for these studies [10, 11, 20, 23], the genotype distribution in control groups of the enrolled studies was in line with HWE. The selected study characteristics are enumerated in Table 2.

3.2. Pooled Analysis. A result of the detailed associations of *SEPP1* polymorphisms with cancer susceptibility in all of the genetic models is presented in Table 3. And the results demonstrated that no evidence of the relevance between the two polymorphisms (rs3877899 and rs7579) and susceptibility to CRC, BC, and PCa was found in each genetic model (Table 3).

For rs3877899 polymorphism, no significant association was found when pooling all the eligible studies (A vs. G: OR = 1.099, 95% CI: 0.938-1.287, P = 0.243; AA vs. GG: OR = 1.129, 95% CI: 0.794-1.605, P = 0.498; AG vs. GG: OR = 1.035, 95% CI: 0.909-1.179, P = 0.603; AA+AG vs. GG: OR = 1.079, 95% CI: 0.919-1.267, P = 0.356; AA vs. AG+GG: OR = 1.017, 95% CI: 0.871-1.189, P = 0.555, Table 3). In addition, there was also no significant relationship between rs7579 polymorphism and cancer risk in each genetic models (A vs. G: OR = 1.090, 95% CI: 0.923-1.286, P = 0.309; AA vs. GG: OR = 1.022, 95% CI: 0.908-1.151, P = 0.715; AA+AG vs. GG: OR = 1.075, 95% CI: 0.897-1.287, P = 0.434; AA vs. AG+GG: OR = 1.209, 95% CI: 0.878-1.666, P = 0.245, Table 3).

Variants	Author	Representativeness of cases	Source of controls	HWE in controls	Genotyping examination	Association assessment	Total scores
rs7579	Meplan et al.	* *	* *	* *	0	**	8
	Steinbrecher et al.	* *	* *	* *	0	* *	8
	Meplan et al.	* *	*	*	0	*	5
	Sutherland et al.	* *	*	*	0	* *	6
	Amini et al.	* *	* *	*	0	* *	7
rs3877899	Steinbrecher et al.	* *	* *	* *	0	**	8
	Cooper et al.	* *	* *	* *	0	* *	8
	Geybels et al.	* *	* *	* *	0	* *	8
	Karunasinghe et al.	* *	* *	* *	0	* *	8
	Meplan et al.	* *	*	*	0	*	5
	Sutherland et al.	* *	*	* *	0	* *	7
	Meplan et al.	* *	* *	* *	0	* *	8
	Jablonska et al.	* *	* *	* *	0	* *	8
	Mohammaddoust et al.	**	* *	*	0	* *	7

TABLE 1: Methodological quality of the included studies according to the Newcastle-Ottawa Scale.

This table identifies "high" quality choices with a "star." A study can be awarded a maximum of one star for each numbered item within the selection and exposure categories. A maximum of 2 stars can be given for comparability.



FIGURE 1: The eligible study selection process.

3.3. Subgroup Analysis. As to a stratification analysis conducted by cancer type, no association was identified for PCa, BC, and CRC of rs3877899 polymorphism in all five genetic models (Table 3). In addition, we also conducted

stratification analyses by the source of control, ethnicity, and HWE status for both two polymorphisms; the null association was detected (Table 3). In all subgroups, the number of included studies is not less than 3 ($n \ge 3$).

	Author		Tumor		Constrains	Source of	Case				Control		
Variants		Year	type	Ethnicity	method	controls	GG	GA	AA	GG	GA	AA	P (HWE)
	Méplan et al.	2013	BC	European	TaqMan	PB	455	396	86	436	420	101	0.992
rs7579	Steinbrecher et al.	2010	PCa	European	MALDI-TOF	PB	116	105	27	250	209	33	0.224
	Méplan et al.	2010	CRC	European	KASPar	HB	260	369	61	269	323	37	≤ 0.001
	Sutherland et al.	2010	CRC	Asian	PCR-RFLP	HB	190	121	32	363	239	67	0.004
	Amini et al.	2019	CRC	Asian	PCR-HRM	PB	24	22	14	40	23	11	0.022
	Steinbrecher et al.	2010	PCa	European	MALDI-TOF	РВ	152	86	10	271	194	27	0.309
	Cooper et al.	2008	PCa	European	TaqMan	PB	1522	949	172	878	595	97	0.775
	Geybels et al.	2013	PCa	European	MALDI-TOF	PB	758	441	53	739	426	67	0.585
	Karunasinghe et al.	2012	PCa	European	TaqMan	PB	153	88	18	255	162	19	0.286
rs3877899	Méplan et al.	2010	CRC	European	KASPar	HB	427	258	47	409	204	44	0.009
183077077	Sutherland et al.	2010	CRC	Asian	PCR-RFLP	HB	797	5	0	710	1	0	0.985
	Méplan et al.	2013	BC	European	TaqMan	PB	586	321	30	594	317	48	0.499
	Jablonska et al.	2015	BC	European	TaqMan	PB	81	44	9	122	55	6	0.948
	Mohammaddoust et al.	2017	BC	Asian	PCR-RFLP	РВ	80	37	33	151	33	16	≤0.001

TABLE 2: Characteristics of case-control studies on SEPP1 polymorphisms and cancer risk included in the meta-analysis.

BC: breast cancer; PCa: prostate cancer; CRC: colorectal cancer; PB: population-based; HB: hospital-based; MALDI-TOF: matrix-assisted laser desorption/ionizing time-of-flight mass spectrometry; PCR-RFLP: polymerase chain reaction-restriction fragment length polymorphism; PCR-HRM: polymerase chain reaction high-resolution melting method; KASPar: KBiosciences' Competitive Allele-Specific PCR; TaqMan: TaqMan fluorescent probe method.

3.4. Heterogeneity Evaluation. Table 3 shows that the statistical heterogeneity within studies was evaluated by a chisquared-based Q-statistic test. When P > 0.10, the fixedeffects model (the Mantel-Haenszel model) was used; else, the random-effects model (the DerSimonian-Laird model) was adopted.

3.5. Sensitivity Analysis and Publication Bias. We explored the impact of each study on the pooled OR by excluding one study from the pooled analysis, thereby performing a sensitivity analysis. It turns out there is no material influence on the stability of the results. So as to assess the publication bias of the existing literature, we performed Begg's funnel plot as well as Egger's test. In all comparison models, the pattern of the funnel chart was roughly symmetrical (Figures 2(a) and 2(b)). In addition, we also used the Egger test, and these results indicated no publication bias (Table 3).

4. Discussion

SEPP1 plays an important role in both supplying selenium to tissues and exerting antioxidant defenses. The delivery of selenium is accomplished by the C-terminal domain of SEPP1, which includes nine selenocysteine residues, while antioxidation is accomplished by selenocysteine that has been shown to have peroxidase activity [24]. The antioxidant function of SEPP1 suggests that it has the effect of preventing cancer, especially in inflammatory cancer characterized by increased oxidative stress [25]. Dysfunction of SEPP1 may contribute to the occurrence and progression of cancer. The human SEPP gene (SEPP1) contains two functional polymorphisms, rs3877899 (Ala234Thr) and rs7579 (Gram A base mutation in SEPP1 mRNA 3'UTR), which affect the selenoprotein activity of plasma and lymphocytes and the relative proportion of plasma SEPP isotypes in vivo experiments [26, 27]. Therefore, the mutation of *SEPP1* will produce some nonfunctional or low-functional protein subtypes, reducing the antioxidant activity of SEPP1. At the same time, the accumulation of peroxide is conducive to the production and development of cancer. Therefore, mutations in *SEPP1* theoretically increase the susceptibility of tumors.

Both polymorphisms of SEPP1 have been reported to be related to the risk of PCa [19] and CRC [20]. Furthermore, in the study conducted by Meplan et al. [18], a connection between the SEPP1 rs3877899 mutation and the risk of BC was also found. However, in a study by Jablonska et al. [22] in Polish women, there was no evidence of a relation between BC risk and the rs3877899 polymorphism. In fact, many related epidemiological studies have been carried out so far, but no definite conclusions have been obtained, and some results are even controversial. Therefore, in order to clarify the relationship between cancer risk and the SEPP1 polymorphism, we conducted this meta-analysis. After pooling all data from 7,157 cases and 6,440 controls for the rs3877899 polymorphism and 2,278 cases and 2,821 controls for the rs7579 polymorphism, a null significant association was identified between SEPP1 polymorphism and cancers (prostate, breast, and colorectal cancer) in all comparative models. We subsequently did subgroup analyses based on cancer type control source HWE status and race for both polymorphisms and found no significant association.

Manianta	Comparison	Subgroup	Ν	P value			Regression model			
v al lallts				$P_{\rm H}$	$P_{\rm Z}$	$P_{\rm E}$	Random	Fixed		
	A vs. G	Overall	9	≤0.001	0.243	0.104	1.099 (0.938-1.287)	1.008 (0.948-1.071)		
	A vs. G	PCa	4	0.514	0.206		0.954 (0.885-1.027)	0.953 (0.885-1.027)		
	A vs. G	BC	3	≤ 0.001	0.240		1.480 (0.770-2.844)	1.134 (0.992-1.295)		
	A vs. G	PB	7	≤ 0.001	0.335		1.093 (0.912-1.310)	0.993 (0.931-1.059)		
	A vs. G	Y	7	0.293	0.231		0.962 (0.888-1.042)	0.961 (0.899-1.026)		
	A vs. G	European	7	0.269	0.434		0.979 (0.910-1.055)	0.976 (0.917-1.038)		
	AA vs. GG	Overall	8	≤0.001	0.498	0.337	1.129 (0.794-1.605)	1.021 (0.871-1.196)		
	AA vs. GG	PCa	4	0.206	0.604		0.946 (0.718-1.246)	0.949 (0.780-1.155)		
	AA vs. GG	BC	3	≤0.001	0.405		1.726 (0.478-6.237)	1.269 (0.902-1.784)		
	AA vs. GG	PB	7	≤0.001	0.483		1.162 (0.765-1.765)	1.020 (0.861-1.210)		
	AA vs. GG	Y	6	0.081	0.599		0.925 (0.693-1.236)	0.917 (0.767-1.095)		
	AA vs. GG	European	7	0.124	0.397		0.936 (0.738-1.187)	0.931 (0.790-1.098)		
	AG vs. GG	Overall	9	0.036	0.603	0.109	1.035 (0.909-1.179)	1.003 (0.928-1.084)		
	AG vs. GG	PCa	4	0.585	0.156		0.934 (0.850-1.027)	0.934 (0.850-1.026)		
ro3877800	AG vs. GG	BC	3	0.046	0.203		1.300 (0.868-1.947)	1.123 (0.948-1.331)		
1830//099	AG vs. GG	PB	7	0.072	0.975		0.998 (0.875-1.138)	0.976 (0.898-1.060)		
	AG vs. GG	Y	7	0.478	0.351		0.960 (0.883-1.044)	0.961 (0.884-1.045)		
	AG vs. GG	European	7	0.308	0.703		0.989 (0.904-1.082)	0.985 (0.911-1.065)		
	AG+AA vs. GG	Overall	9	0.001	0.356	0.098	1.079 (0.919-1.267)	1.008 (0.936-1.085)		
	AG+AA vs. GG	PCa	4	0.623	0.152		0.936 (0.855-1.025)	0.936 (0.855-1.025)		
	AG+AA vs. GG	BC	3	≤0.001	0.210		1.474 (0.804-2.704)	1.150 (0.980-1.350)		
	AG+AA vs. GG	PB	7	0.001	0.551		1.056 (0.884-1.261)	0.984 (0.909-1.064)		
	AG+AA vs. GG	Y	7	0.452	0.266		0.955 (0.881-1.034)	0.956 (0.882-1.035)		
	AG+AA vs. GG	European	7	0.317	0.553		0.981 (0.901-1.068)	0.978 (0.907-1.054)		
	AA vs. AG+GG	Overall	8	0.001	0.555	0.371	1.103 (0.797-1.526)	1.017 (0.871-1.189)		
	AA vs. AG+GG	PCa	4	0.176	0.811		0.974 (0.733-1.295)	0.977 (0.806-1.184)		
	AA vs. AG+GG	BC	3	≤ 0.001	0.445		1.580 (0.489-5.108)	1.199 (0.856-1.681)		
	AA vs. AG+GG	PB	7	0.001	0.497		1.143 (0.777-1.681)	1.027 (0.869-1.215)		
	AA vs. AG+GG	Y	6	0.067	0.699		0.944 (0.704-1.265)	0.936 (0.786-1.115)		
	AA vs. AG+GG	European	7	0.113	0.444		0.940 (0.741-1.192)	0.939 (0.799-1.103)		
	A vs. G	Overall	5	0.015	0.309	0.233	1.090 (0.923-1.286)	1.044 (0.958-1.137)		
	A vs. G	CRC	3	0.079	0.236		1.148 (0.914-1.443)	1.125 (0.995-1.273)		
	A vs. G	European	3	0.013	0.449		1.084 (0.880-1.335)	1.047 (0.952-1.152)		
	A vs. G	PB	3	0.015	0.405		1.137 (0.840-1.539)	1.01 (0.892-1.123)		
	A vs. G	Ν	3	0.079	0.236		1.148 (0.914-1.443)	1.125 (0.995-1.273)		
	AA vs. GG	Overall	5	0.014	0.230	0.157	1.267 (0.861-1.866)	1.128 (0.923-1.379)		
	AA vs. GG	CRC	3	0.091	0.204		1.387 (0.837-2.296)	1.329 (0.989-1.786)		
	AA vs. GG	European	3	0.007	0.337		1.310 (0.755-2.274)	1.147 (0.910-1.445)		
rs7579	AA vs. GG	PB	3	0.019	0.383		1.334 (0.698-2.548)	1.043 (0.802-1.356)		
	AA vs. GG	Ν	3	0.091	0.204		1.387 (0.837-2.296)	1.329 (0.989-1.786)		
	AG vs. GG	Overall	5	0.315	0.715	0.264	1.029 (0.900-1.177)	1.022 (0.908-1.151)		
	AG vs. GG	CRC	3	0.357	0.223		1.112 (0.932-1.327)	1.112 (0.937-1.321)		
	AG vs. GG	European	3	0.190	0.753		1.034 (0.866-1.234)	1.022 (0.895-1.166)		
	AG vs. GG	PB	3	0.276	0.693		0.994 (0.808-1.224)	0.968 (0.825-1.136)		
	AG vs. GG	Ν	3	0.357	0.223		1.112 (0.932-1.327)	1.112 (0.937-1.321)		
	AG+AA vs. GG	Overall	5	0.071	0.434	0.210	1.075 (0.897-1.287)	1.038 (0.927-1.162)		
	AG+AA vs. GG	CRC	3	0.149	0.116		1.158 (0.897-1.496)	1.140 (0.968-1.342)		

Variants	Comparison	Subgroup	Ν	P value			Regression model			
				$P_{\rm H}$	$P_{\rm Z}$	$P_{\rm E}$	Random	Fixed		
	AG+AA vs. GG	European	3	0.051	0.551		1.072 (0.852-1.350)	1.039 (0.915-1.180)		
	AG+AA vs. GG	PB	3	0.069	0.581		1.093 (0.798-1.497)	0.983 (0.845-1.144)		
	AG+AA vs. GG	Ν	3	0.149	0.116		1.158 (0.897-1.287)	1.140 (0.968-1.342)		
	AA vs. AG+GG	Overall	5	0.053	0.245	0.180	1.209 (0.878-1.666)	1.117 (0.921-1.355)		
	AA vs. AG+GG	CRC	3	0.189	0.109		1.280 (0.861-1.903)	1.261 (0.949-1.676)		
	AA vs. AG+GG	European	3	0.022	0.316		1.268 (0.797-2.019)	1.139 (0.912-1.422)		
	AA vs. AG+GG	PB	3	0.046	0.396		1.264 (0.736-2.169)	1.054 (0.820-1.355)		
	AA vs. AG+GG	Ν	3	0.189	0.109		1.280 (0.861-1.903)	1.261 (0.949-1.676)		

TABLE 3: Continued.

PCa: prostate cancer; BC: breast cancer; CRC: colorectal cancer; HWE: Hardy-Weinberg Equilibrium; Y: study conformed to HWE; N: study did not conform to HWE; P-B: population-based; H-B: hospital-based; $P_E = P$ value of Egger test; $P_H = P$ value of heterogeneity test; $P_Z = P$ value of Z test.



Begg's funnel plot with pseudo 95% confidence limits



FIGURE 2: (a) Begg's funnel plot of publication bias (for rs7579 polymorphism, A vs. G). Each point represents a separate study for the indicated association. Log (OR), natural logarithm of OR; horizontal line, mean effect size. CI = confidence interval; OR = odds ratio. (b) Begg's funnel plot of publication bias (for rs3877899 polymorphism, GA vs. GG). Each point represents a separate study for the indicated association. Log (OR), natural logarithm of OR; horizontal line, mean effect size. CI = confidence interval; OR = odds ratio.

This result contradicts our previous theoretical speculation. In fact, *SEPP1* polymorphism affects tumor susceptibility through the antioxidant activity of SEPP1 protein. However, there may be more than one factor that can change the antioxidant activity of SEPP1 protein. The study by Sutherland et al. [23] showed that the two polymorphisms of *SEPP1* (rs3877899 and rs7579) are not associated with the risk of CRC, which may be due to the inconsistent dietary selenium intake of the subjects. It is possible that when the plasma selenium level is low, the difference in the plasma level of SEPP1 caused by genetic polymorphism can be reflected, and this difference will disappear after selenium supplementation [26]. Furthermore, our results can be analyzed more accurately by age, cancer grade, and environmental factors (such as selenium status related to SEPP1 expression). For example, in the study of Cooper et al. [21], cancer was divided into two groups of nonprogressive and progressive, and even the factor of smoking was included. Therefore, to explore the impact of genotype on cancer susceptibility, environmental and nutritional factors should be strictly controlled; otherwise, the results may be biased.

In conclusion, despite providing a sufficient statistical sample size to enhance the reliability of our findings, there are some shortcomings of the study. Firstly, the relatively small number of included studies would be a limitation and may constrain our conclusions. Secondly, we only searched papers published in a limited number of databases and some studies may have been overlooked. Finally, the results may be false negative, because some included studies show a significant relation between *SEPP1* polymorphism and cancer susceptibility.

5. Conclusions

In this meta-analysis, our results find no association between *SEPP1* rs3877899 and rs7579 polymorphisms and susceptibility to CRC, BC, and PCa. Taking into account the complex interactions between genes and the environment, it is necessary to conduct unbiased studies with more sample sizes and more cancer types in different ethnic groups.

Data Availability

The dataset can be accessed from the corresponding author upon reasonable request.

Disclosure

The funders had no roles in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Conflicts of Interest

All authors declared that there is no competing interest.

Authors' Contributions

H.X. and F.M. accessed information from the literature for this article. J.Z., Z.H., X.C., and C.L. contributed towards writing, discussing, and editing the manuscript. Hanjiang Xu and Fan Mo contributed equally to the work.

Acknowledgments

This work was supported by the Clinical Key Subjects Program of the Ministry of Public Health (Urology), the National Natural Science Foundation of China (81170698, 81370856, and 81401518), the Key Science and Technology Program of Anhui Province (12010402128), the Anhui Provincial Natural Science Foundation (1408085QH180), and the cultivation project for NSFC at Anhui Medical University (2013KJ14).

Supplementary Materials

 Table S1: scale for quality assessment. (Supplementary Materials)

References

- F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," *CA: a Cancer Journal for Clinicians*, vol. 68, no. 6, pp. 394–424, 2018.
- [2] M. Carbone, S. T. Arron, B. Beutler et al., "Tumour predisposition and cancer syndromes as models to study gene- environment interactions," *Nature Reviews. Cancer*, vol. 20, no. 9, pp. 533–549, 2020.
- [3] U. S. Srinivas, B. W. Q. Tan, B. A. Vellayappan, and A. D. Jeyasekharan, "ROS and the DNA damage response in cancer," *Redox Biology*, vol. 25, article 101084, 2019.
- [4] Y. Yang, S. Karakhanova, W. Hartwig et al., "Mitochondria and mitochondrial ROS in cancer: novel targets for anticancer therapy," *Journal of Cellular Physiology*, vol. 231, no. 12, pp. 2570–2581, 2016.
- [5] Ö. Canli, A. M. Nicolas, J. Gupta et al., "Myeloid cell-derived reactive oxygen species induce epithelial mutagenesis," *Cancer Cell*, vol. 32, no. 6, pp. 869–883.e5, 2017.
- [6] Y. Saito, "Selenoprotein P as an <i>in vivo</i> redox regulator: disorders related to its deficiency and excess," *Journal of Clinical Biochemistry and Nutrition*, vol. 66, no. 1, pp. 1–7, 2020.
- [7] O. H. al-Taie, N. Uceyler, U. Eubner et al., "Expression profiling and genetic alterations of the selenoproteins GI-GPx and SePP in colorectal carcinogenesis," *Nutrition and Cancer*, vol. 48, no. 1, pp. 6–14, 2004.
- [8] A. Calvo, N. Xiao, J. Kang et al., "Alterations in gene expression profiles during prostate cancer progression: functional correlations to tumorigenicity and down-regulation of selenoprotein-P in mouse and human tumors," *Cancer Research*, vol. 62, no. 18, pp. 5325–5335, 2002.
- [9] O. Gonzalez-Moreno, N. Boque, M. Redrado et al., "Selenoprotein-P is down-regulated in prostate cancer, which results in lack of protection against oxidative damage," *The Prostate*, vol. 71, no. 8, pp. 824–834, 2011.
- [10] S. Mohammaddoust, Z. Salehi, and H. Saeidi Saedi, "SEP-PlandSEP15gene polymorphisms and susceptibility to breast cancer," *British Journal of Biomedical Science*, vol. 75, no. 1, pp. 36–39, 2018.
- [11] G. Amini, R. Salehi, A. Moshtaghi, M. Kazemi, M. Behjati, and S. Khosravi, "Evaluation of SEPP1 and selenoprotein S gene polymorphisms (rs 7579 and rs 34713741) in relation to colorectal cancer susceptibility in subset of Iranian population: a case-control study," *Advanced Biomedical Research*, vol. 8, no. 1, p. 47, 2019.

- [12] W. Xie, M. Yang, J. Chan et al., "Association of genetic variations of selenoprotein genes, plasma selenium levels, and prostate cancer aggressiveness at diagnosis," *The Prostate*, vol. 76, no. 7, pp. 691–699, 2016.
- [13] D. Moher, A. Liberati, J. Tetzlaff, D. G. Altman, and PRISMA Group, "Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement," *Journal of Clinical Epidemiology*, vol. 62, no. 10, pp. 1006–1012, 2009.
- [14] J. P. T. Higgins and S. G. Thompson, "Quantifying heterogeneity in a meta-analysis," *Statistics in Medicine*, vol. 21, no. 11, pp. 1539–1558, 2002.
- [15] M. B. Mathur and T. J. VanderWeele, "Sensitivity analysis for publication bias in meta-analyses," *Journal of the Royal Statistical Society. Series C, Applied Statistics*, vol. 69, no. 5, pp. 1091–1119, 2020.
- [16] N. Karunasinghe, D. Y. Han, M. Goudie et al., "Prostate disease risk factors among a New Zealand cohort," *Journal of Nutrigenetics and Nutrigenomics*, vol. 5, no. 6, pp. 339–351, 2012.
- [17] M. S. Geybels, C. M. Hutter, E. M. Kwon et al., "Variation in selenoenzyme genes and prostate cancer risk and survival," *The Prostate*, vol. 73, no. 7, pp. 734–742, 2013.
- [18] C. Méplan, L. O. Dragsted, G. Ravn-Haren, A. Tjønneland, U. Vogel, and J. Hesketh, et al., "Association between polymorphisms in glutathione peroxidase and selenoprotein P genes, glutathione peroxidase Activity, HRT Use and Breast Cancer Risk," *PLoS ONE*, vol. 8, no. 9, article e73316, 2013.
- [19] A. Steinbrecher, C. Méplan, J. Hesketh et al., "Effects of selenium status and polymorphisms in selenoprotein genes on prostate cancer risk in a prospective study of European men," *Cancer Epidemiology and Prevention Biomarkers*, vol. 19, no. 11, pp. 2958–2968, 2010.
- [20] C. Meplan, D. J. Hughes, B. Pardini et al., "Genetic variants in selenoprotein genes increase risk of colorectal cancer," *Carcinogenesis*, vol. 31, no. 6, pp. 1074–1079, 2010.
- [21] M. L. Cooper, H. O. Adami, H. Gronberg, F. Wiklund, F. R. Green, and M. P. Rayman, "Interaction between single nucleotide polymorphisms in selenoprotein P and mitochondrial superoxide dismutase determines prostate cancer risk," *Cancer Research*, vol. 68, no. 24, pp. 10171–10177, 2008.
- [22] E. Jablonska, J. Gromadzinska, B. Peplonska et al., "Lipid peroxidation and glutathione peroxidase activity relationship in breast cancer depends on functional polymorphism of GPX1," *BMC Cancer*, vol. 15, no. 1, 2015.
- [23] A. Sutherland, D. Kim, C. Relton, Y. Ahn, and J. Hesketh, "Polymorphisms in the selenoprotein S and 15-kDa selenoprotein genes are associated with altered susceptibility to colorectal cancer," *Genes & Nutrition*, vol. 5, no. 3, pp. 215–223, 2010.
- [24] R. F. Burk and K. E. Hill, "Selenoprotein P-Expression, functions, and roles in mammals," *Biochimica et Biophysica Acta* (*BBA*) - *General Subjects*, vol. 1790, no. 11, pp. 1441–1447, 2009.
- [25] C. W. Barrett, S. P. Short, and C. S. Williams, "Selenoproteins and oxidative stress-induced inflammatory tumorigenesis in the gut," *Cellular and Molecular Life Sciences*, vol. 74, no. 4, pp. 607–616, 2017.

- [26] C. Méplan, L. K. Crosley, F. Nicol et al., "Genetic polymorphisms in the human selenoprotein P gene determine the response of selenoprotein markers to selenium supplementation in a gender-specific manner (the SELGEN study)," *The FASEB Journal*, vol. 21, no. 12, pp. 3063–3074, 2007.
- [27] C. Méplan, F. Nicol, B. T. Burtle et al., "Relative abundance of selenoprotein P isoforms in human plasma depends on genotype, se intake, and cancer status," *Antioxid Redox Signal*, vol. 11, no. 11, pp. 2631–2640, 2009.