Childhood Ménétrier’s disease: A rare cause of exudative enteral protein loss

MARK R OLIVER MBBS, CYNTHIA TREVENEN MD, R BRENT SCOTT MDCM

MÉNÉTRIÉR’S DISEASE IS A RARE cause of gut-related protein loss in childhood. It was first described by Ménétrier in 1888 (1), and since its initial description there have been only 47 pediatric cases reported (2). The disorder is characterized by an enlargement of gastric rugal folds and characteristic histological changes in full thickness biopsy specimens, and is often associated with hypochlorhydria, hypoproteinemia and edema (2-10). Compared with the adult form of Ménétrier’s disease, which follows a chronic pattern often requiring active treatment, the childhood form is usually self-limited. Although the etiology of childhood Ménétrier’s disease is unclear, mainly allergic phenomena and infection have been implicated in the pathogenesis. Epidemiological evidence mainly supports cytomegalovirus (CMV), but other microorganisms including Helicobacter pylori and Mycoplasma pneumoniae have been implicated (11-17). This report describes a young child with Ménétrier’s disease and provides additional evidence for an association between this disorder and CMV infection in childhood.

CASE PRESENTATION

A six-year-old Canadian Aboriginal presented to Alberta Children’s Hospital following a two-week illness characterized by colicky upper abdominal pain, increasing edema, intermittent nonbilious vomiting and self-limiting nonbloody diarrhea. There were no symptoms to suggest an underlying cardiac, renal, hepatic or lymphatic disorder that might have explained the edema. In addition, there was a negative history for atopy and immunosuppressive disorders. Examination showed him to be a well nourished child with mild pharyngitis and no lymphadenopathy. He had impressive periorbital edema, and pitting edema of his legs up to the knees, small bilateral pleural effusions and abdominal distension with ascites. He also complained of epigastric tenderness on deep palpation. Rectal examination was normal.

Laboratory findings on admission to hospital showed hyponatremia (serum sodium 127 mmol/L), hypokalemia (se-
rum potassium 2.8 mmol/L) and hypo-chloremia (serum chloride 94 mmol/L). Serum albumin was 14 g/L (normal range 30 to 50 g/L) and total protein 26 g/L (normal range 55 to 75 g/L) with normal hepatocellular enzymes. Urine analysis was consistently negative for both protein and blood. A complete blood count demonstrated normal hemoglobin, white blood cell and platelet counts. The differential showed no lymphopenia, but peripheral eosinophilia with an absolute count of 1.2 \times 10^9/L (normal range 0.1 to 0.7 \times 10^9). Immunoglobulin G levels were low at 3.16 g/L (normal range 6.33 to 12.8 g/L), but other immunoglobulin levels were normal. Plain x-rays of his abdomen and chest, and an abdominal ultrasound demonstrated bilateral pleural effusions and ascitic fluid without other significant abnormalities.

Stool cultures were positive for the trophozoites of *Entamoeba histolytica* and *Entamoeba nana* without any pus or red blood cells. A fluoroscopy guided duodenal aspirate and small bowel biopsy performed using a Carey capsule (Precise Products Corporation, Minnesota) excluded *Giardia lamblia* enteritis and bacterial overgrowth and demonstrated normal duodenal histology. A 72 h fecal \alpha-1 antitrypsin clearance was increased well above the normal range at 0.278 g/day (normal less than 0.06 g/day), suggesting a protein losing enteropathy. A limited colonoscopy showed mild confluent inflammation of the distal colon that was histologically confirmed as a mild acute nonspecific colitis. He was treated with metronidazole for seven days and indomethacin for 21 days.

Despite rapid resolution of the vomiting and diarrhea, the patient remained hypoproteinemnic with ongoing epigastric pain. A barium upper gastrointestinal series demonstrated diffuse thickening of the gastric folds with free flow of barium into the structurally normal small intestine (Figure 1). An esophagogastroduodenoscopy confirmed marked thickening of the rugal folds in the body of the stomach with sparing of the antrum. The thickened folds appeared gelatinous and hemorrhagic. The esophagus and duodenum had a normal macroscopic appearance that was confirmed histologically. Biopsies of gastric tissue were sent for culture of *H pylori* and for viral isolation. Superficial gastric biopsies from the fundus demonstrated chronic gastritis with conserved foveolar epithelium and nonatrophic glands. The lamina propria was expanded by an inflammatory infiltrate including eosinophils (Figure 2). There was no evidence of *H pylori*-like organisms or viral inclusion bodies on light microscopy of gastric fundal or antral biopsies.

Cultures of gastric tissue were negative for *H pylori*. However, both fundal biopsy specimens and urine were subsequently shown to be positive for CMV by an early antigen method that depends on a fluorescein-labelled monoclonal antibody that is specific for CMV and recognizes the immediate early protein of CMV (18). Serology done at six and 12 weeks was positive for complement fixing CMV antibodies. Immunohistochemical studies conducted on both gastric and rectal tissues were negative. Within two and a half months of the onset of symptoms, and without additional therapy, the patient recovered both clinically and biochemically. A follow-up esophagogastroduodenoscopy performed three months after presentation showed both macroscopic and microscopic resolution of the gastric disease. Repeat evaluation of gastric biopsies

**Figure 1** Left Barium meal demonstrating hypertrophic gastric folds

**Figure 2** Gastric mucosa focally infiltrated by lymphocytes, plasma cells and eosinophils (original magnification x118)
for CMV by the early antigen method were negative. Similarly, a repeat limited colonoscopy showed macroscopic resolution of distal colonic changes with only mild residual microscopic colitis. Stool cultures were now negative for Entamoeba histolytica. Eighteen months following initial presentation he remains completely well.

**DISCUSSION**

The diagnosis of Ménétriér’s disease was based on the presence of giant hypertrophy of gastric folds seen both radiologically and at endoscopy, hypoproteinemia with associated enteral protein loss and a histological gastritis that is often associated with hypochlorhydria. As has been previously reported in children with Ménétriér’s disease, this patient’s clinical course was self-limited and benign (2-8,10-13). Although the microscopic appearance of his fundal biopsy was not classical of this disorder (there was no evidence of foveolar hyperplasia and atrophied glands), there was tissue eosinophilia and edema which is seen histologically in this disease. This variation may have been secondary to either sampling error or the superficial nature of the biopsies (2,9). Or it could be that, as in adults, the clinical features of childhood Ménétriér’s disease can be divided into at least two distinct histopathological entities, one representing massive foveolar hyperplasia and the other hypertrophic lymphoctic gastritis (19).

It is interesting to note that our patient also presented with diarrhea and mild colitis, and excreted E histolytica trophozoites in his stools. While it is possible that amoebic colitis may have contributed to the enteric protein loss, severe hypoproteinemia is not reported as a presenting factor of this infection (20,21). The marked hypoproteinemia in this patient persisted after the treatment of amoebiasis and after resolution of the diarrhea. In addition, the colitis was macroscopically and histologically mild. It is likely that the massive gut-related protein loss observed in this patient was secondary to Ménétriér’s disease, which is associated not only with widening of the gastric mucosal tight junctions (10,22) but also with hypochlorhydria. The latter results in significantly less degradation of α-1 antitrypsin activity and hence measurable gastric clearance in this disease.

At least four possible factors can contribute to an increased gastric mucosal mass, in health or in response to disease: intraluminal substances such as nutrients or growth factors from the more proximal gut; trophic factors produced locally or by infectious agents; neural stimulation; and immunological phenomena secondary to an autoimmune process or allergy (9). It is likely that in Ménétriér’s disease the irritant agent initiates a significant inflammatory reaction (as shown by the presence of gastritis) which then directly or indirectly causes the observed hypertrophic changes through the action of unidentified mediators. It has recently been shown that transforming growth factor-α, an epithelial cell mitogen that inhibits gastric acid secretion and increases mucin production, may be involved in the pathogenesis of this disorder (23,24).

The strongest evidence for an etiological factor in the pediatric population is for CMV infection (2-13), although it is difficult to prove such a causative role because of the ubiquitous nature of this organism. The virus could be directly toxic to the gastric mucosa and allow increased macromolecular uptake that, in turn, initiates an allergic response with the typical peripheral eosinophilia observed in this disease. Infection might also suppress specific regulatory T cell subsets and allow an allergic type of response to be manifested (7), or stimulate a local inflammatory response with the release of mediators, eg, tumour necrosis factor-α, that provoke a hypertrophic response lasting only as long as the infection is active (2). Of the 47 pediatric patients previously reported as having Ménétriér’s disease, only 27 were investigated for CMV infection (2). Nine of these patients (33%) had evidence of gastric CMV infection by characteristic intranuclear inclusions, early antigen and/or positive gastric CMV culture. However, only two previously reported cases had evidence of current CMV infection on the basis of viral culture (urine and gastric tissue) and serology (2). In our case, there is not only supportive evidence of current CMV infection (fundal biopsy and urine positive for CMV early antigen), but also a clinical, endoscopic and histological resolution of Ménétriér’s disease that was temporally related to clearance of infection. This suggests that CMV had a role in the pathogenesis of the disorder in our patient. A direct causal relationship between gastric CMV infection and Ménétriér’s disease in childhood will require a thorough examination for CMV in subsequent patients with this disorder, and further study of both host defence mechanisms and the manner in which they can be altered in CMV infection.

ACKNOWLEDGEMENTS: The authors thank Roberta Funk for typing the manuscript. Mark Oliver is a recipient of an Alberta Children’s Hospital Foundation Fellowship.

**REFERENCES**

and review of the literature.