Clinical, pathological and molecular genetic findings in small intestinal follicle centre cell lymphoma

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Observations génétiques, cliniques, pathologiques et moléculaires dans le lymphome B du grêle

Lymphoma may involve any site within the gastrointestinal tract and may be focal, multifocal or diffuse. Diagnosis is sometimes difficult, and classification is complicated by schema based largely on lymphomatous involvement of lymph nodes. In 1982, a proposed working formulation based purely on morphological findings was published (1). It closely paralleled the Rappaport classification because it was modified before 1982 (2,3) and it appeared to be clinically useful since it seemed to predict survival and curability. It was not clear, however, whether this classification applied to localized gastrointestinal disease.

Over the next decade, development of increasing numbers of monoclonal antibodies and molecular genetic markers permitted subdivision of lymphoid cells and their tumours into various subtypes, levels of differentiation and proliferation characteristics. Diagnosis of mucosa-associated lymphoid tissue (MALT) lymphomas, including even those with a nodular appearance, for example, were made more precise because MALT lymphomas do not demonstrate bcl-2 or bcl-1 gene rearrangements (4). As a result, another group (4) proposed a new classification and nomenclature to separate the different types of lymphoid neoplasms more...
accurately and facilitate classification of extranodal lymphomas.

A localized lymphoma of the small bowel detected in a man with iron deficiency anemia is presented in this report. Biopsies of the proximal duodenum revealed a small B cell lymphoma, and additional studies, including polymerase chain reaction (PCR) of fresh small intestinal biopsies, permitted a specific diagnosis of a follicular lymphoma, a B cell lymphoma that usually predominates in lymph nodes and spleen. Detection of the bcl-2 gene rearrangement—the molecular equivalent of the t(14;18) chromosomal abnormality—together with histological and immunophenotypic data, permitted a precise classification for clinical management.

CASE PRESENTATION

A 44-year-old male was referred for evaluation of anemia and iron deficiency. He was followed and investigated in his community hospital for 14 months after initial presentation, at which time his hemoglobin was 46 g/L. Physical examinations over the course of his evaluations were normal. Fecal occult blood tests (total of 12) were negative, as were investigations to exclude a hematological cause for anemia, such as studies to exclude hemolysis. Despite negative fecal occult blood tests studies were also done to exclude a gastrointestinal cause for blood loss. These included repeated barium radiological and fiberoptic endoscopic studies of the upper and lower gastrointestinal tracts. Nuclear medicine imaging studies were also performed, including a tagged red blood cell scan and a Meckel's scan. All investigations were normal, including a rectal biopsy; however, biopsies of the upper gastrointestinal tract were not done. Repeated hospitalizations were required and transfusions were administered from March 1994 until May 1995. He was referred for further evaluation due to persistent anemia.

Because a proximal small intestinal mucosal disorder, eg, occult celiac disease, can present as iron deficiency anemia, an upper endoscopic examination was repeated. This revealed an abnormal proximal duodenum. The mucosal surface was erythematous and swollen, with deep linear and serpiginous ulceration that extended from the distal portion of the duodenal bulb to the transverse portion of the distal duodenum. These changes appeared to be reminiscent of those seen in Crohn’s disease involving the upper intestinal tract (5,6) and to be typical of changes previously detailed for duodenal lymphoma (7,8). The gastric mucosa appeared normal, and gastric biopsies from the body and antrum showed no evidence of a lymphoproliferative disorder. Routine hematoxylin-eosin and silver stains of these gastric biopsies for detection of Helicobacter pylori were also negative. Multiple biopsies of the duodenum (Figures 1,2) showed features of a lymphoma: follicular small cleaved cell type. Biopsies also demonstrated a nodular architecture composed of a monomorphic infiltrate of small cleaved lymphocytes. The cells were small with irregular nuclei, indistinct nucleoli and scant cytoplasm. Immunoperoxidase labelling of lymphoid cells was positive for the B cell antigen marker L-26 (CD-20). Kappa and lambda stains failed to disclose light chain restriction while BCL-2 protein labelling of lymphoid cells was strongly positive.

Further studies were done to aid in the treatment evaluation of this patient. A computerized tomographic scan of
the abdomen and pelvis showed uniform thickening of the proximal duodenum but no lymphadenopathy (Figure 3). Another barium radiographic study of the upper gastrointestinal tract revealed mucosal fold thickening in the proximal duodenum, but normal-appearing distal duodenum, jejunum and ileum (Figures 4, 5). Other studies, including lymphangiography, bone marrow aspiration and biopsy, as well as quantitative immunoglobulin studies, were normal.

**Figure 4**) Barium radiological study of the upper gastrointestinal tract revealed thickened duodenal folds; the rest of the small intestine was normal.

**Figure 5**) Barium radiological study of the duodenal C loop showing narrowing of the lumen with thickened mucosal folds.

During diagnostic evaluation of the lymphoma, duodenal mucosal biopsies were also collected for gene rearrangement studies. B cell clonality was assessed using a consensus primer strategy incorporating V region and J region primers for the immunoglobulin heavy chain region (VJ-PCR) (9). Rearrangements of the bcl-2 gene were detected by PCR as previously described (10). These studies confirmed a monoclonal B cell population by VJ-PCR (Figure 6), as well as PCR evidence of a bcl-2 gene rearrangement at the major break-point region (Figure 7).

**DISCUSSION**

Follicular lymphoma is one of the most common human B cell lymphoid neoplasms. Prior studies have demonstrated that the lymphoma cells have a translocation between chromosomes 14 and 18, juxtaposing the immunoglobulin heavy
chain gene with the bcl-2 proto-oncogene (11,12). Most patients with this type of lymphoma have involvement of lymph nodes, spleen, bone marrow and, occasionally, peripheral blood (4). Our patient had a lymphoid neoplasm localized to the proximal duodenum. Based on immunophenotyping of the small intestinal biopsies, the lymphoma was characterized as a low grade B cell lymphoma with BCL-2 protein expression (4). Using the technique of PCR to classify this lymphoma more specifically, a follicular or follicle centre cell lymphoma with a bcl-2 gene rearrangement was defined (4). This lymphoma is typically composed of cleaved follicle centre cells or centrocytes (11). In up to 90% of cases (10,13,14) the t(14;18) chromosomal abnormality can be detected at a molecular level with rearrangement of the bcl-2 gene (10,13,14).

It is believed that the bcl-2 or 'anti-apoptosis' gene is 'switched off' at the translational level in the normal germinal centre cells. Expression of BCL-2 protein permits accumulation of long-lived centrocytes (11-14). In addition, expression of the BCL-2 protein is a useful marker to distinguish reactive follicular hyperplasia (folicles are BCL-2-negative) from follicular lymphoma (folicles usually BCL-2-positive). Moreover, it has been suggested that when a resting B cell carrying the bcl-2 gene translocation undergoes blast transformation in response to antigen, there is a failure to 'switch off' the bcl-2 gene, and this genetic alteration may contribute to lymphoma development (4).

Although histology remains the gold standard for diagnosis and classification of lymphoid neoplasms, other studies, including immunophenotyping, and cytogenetic and molecular genetic analyses may provide useful information for further subclassification of these disorders. In addition to routine histology, both frozen-section immunophenotyping and molecular genetic studies, particularly PCR, can be performed on endoscopic biopsy specimens (15). These techniques can provide important information about both lineage and clonality, as well as provide specific details useful for the subclassification of malignant lymphomas. Rapid freezing in liquid nitrogen results in biopsy specimens that can be used for complex immunophenotyping, followed by extraction of DNA for gene rearrangement studies. This approach maximizes the use of small endoscopic biopsies.

Rearrangements of the bcl-2 gene characterize most follicular lymphomas, as well as 15% to 20% of de novo diffuse large B cell lymphomas (4). Rearrangements of the bcl-1 gene are typically found in mantle cell lymphomas, the histological hallmark of multiple lymphomatous polyposis of the gastrointestinal tract. Neither of these chromosomal translocations is detected in low grade B cell lymphomas of MALT. Therefore this molecular genetic information is helpful in assigning a specific diagnosis once a lymphoma is documented. Additionally, use of molecular techniques to establish a clonal B cell neoplasm helps to distinguish malignant lymphomas from benign infiltrates and has led to the virtual disappearance of the word 'pseudolymphoma', a term of unquestionable confusion for clinicians and pathologists alike.

CONCLUSIONS
Endoscopic biopsies of the gastrointestinal tract provide sufficient material for routine histology, as well as frozen section material for immunophenotyping and molecular genetic studies. A systematic diagnostic strategy using these techniques will greatly facilitate the specific classification of lymphoid neoplasms, including those localized in the small intestine. A more precise classification of lymphomas in the gastrointestinal tract using modern diagnostic methods should discourage the use of the term pseudolymphoma, improve our understanding of the pathogenesis of lymphomas and reduce the number of repeated endoscopic biopsies and more invasive surgical procedures used to establish a diagnosis. All of these factors should permit the development of optimum treatment regimens for these increasingly recognized extranodal lymphomas.

REFERENCES
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