Synchronous occurrence of collagenous colitis and pseudomembranous colitis

Z Vesoulis MD1, G Lozanski MD1, T Loiudice MD2

More than 100 cases of collagenous colitis have been described in the pathology and gastroenterology literature since the original report by Lindstrom in 1976 (1-8). The generally accepted histological definition originally required microscopic evidence of subepithelial collagen deposition in excess of 10 µm in mean thickness (versus a normal mean thickness of 2 to 7 µm). Thickened subepithelial collagen alone is not diagnostic of collagenous colitis because normal rectal mucosa may demonstrate subepithelial collagen that approaches 10 µm in thickness. More recently, the
Pseudomembranous colitis, also a chronic diarrheal illness with an often acute explosive onset, has been closely associated with intestinal overgrowth of *Clostridium difficile* and elaboration of bacterial endotoxins. The etiology, pathological attributes and pathogenetic mechanisms have evolved since its first documentation in 1893, particularly since the inception of widespread antibiotic use in the 1950s (9-11). Collagenous colitis has been reported in association with lymphocytic and/or microscopic colitis, lymphocytic gastritis, Crohn’s disease, ulcerative colitis and rheumatic arthritis (12-17). Pseudomembranous colitis has been associated with autoimmune diseases, particularly arthritis (18-20). To our knowledge, there have been no reports of the synchronous occurrence of collagenous and pseudomembranous colitis. One case of resolved pseudomembranous colitis followed by biopsy-proven collagenous colitis has been documented (21).

**CASE PRESENTATION**

A 73-year-old white woman presented with a chief complaint of chronic watery diarrhea and lower abdominal cramping without fever. She reported a 6.8 kg weight loss and eight to 10 bowel movements per day, six weeks in duration. The diarrhea was not alleviated by loperamide. Some episodes of diarrhea were nocturnal. The patient had no previous medical history of protracted diarrhea and no history of arthritis, arthralgia or collagen vascular disease. Pertinent history included gastritis (clinical diagnosis), coronary artery disease and hypertension treated with a daily regimen of cilnidipine, nitroglycerin and tenormin, respectively. The patient had taken ampicillin several weeks before the onset of the diarrhea for an unspecified infection. There was no history of nursing home or institutional admissions or stays. The patient required four days of hospitalization with intravenous hydration due to the severity of her presenting diarrheal illness. Stool cultures and examination for ova and parasites were negative, as were duodenal washings for *Giardia* and the CLOtest (Delta West Ltd, Bentley, Australia) for *H pylori*. Stool was positive for the presence of leukocytes and red blood cells. Endoscopic examinations taken during hospitalization revealed discrete ulcerated plaques of the left colon and an endoscopically normal right colon. Stool C difficile toxin A assay by cell culture cytotoxin method and enzyme immunonassay were positive. These results, together with the endoscopy and biopsy findings, prompted treatment with metronidazole (500 mg tid). This treatment was continued for three weeks, with a significant decrease in diarrhea and a follow-up negative C difficile toxin study. Subsequently, steroid therapy (methylprednisolone 32 mg dose pack) was added; the dose was tapered for two additional months, and symptoms resolved completely. The patient has remained symptom-free for more than a year on mesalamine (400 mg bid). A follow-up examination at four months demonstrated an endoscopically normal colon. Follow-up biopsies of the left and right colon were obtained.

**MATERIALS AND METHODS**

Eight endoscopic biopsies from the left colon and rectosigmoid colon were received in 10% buffered formalin and processed for histological evaluation in the typical manner. Sections were stained with hematoxylin and eosin, and Mason’s trichrome. Morphometric measurements of subepithelial collagen in the hematoxylin and eosin-stained sections were carried out using the CAS 200 image analyzer (Becton Dickinson Co, San Jose, California). Mean measurements for each biopsy fragment were determined using Ocular Micrometer version 1.0 software (Cell Analysis Systems, Elmhurst, Illinois). Biopsy fragments with an intact surface epithelium, proper orientation and a continuous uninterrupted zone (at least 200 µm in length) of subepithelial collagen were used for measurements. Fragments sectioned tangentially, those with crush or other artifact and those with extensive inflammatory cell infiltrates obliterating the collagenous layer were excluded from measurement. Each of the eight qualified fragments was measured in at least three separate, optimally oriented areas representing the thickest visually determined zones of subepithelial collagen. Each measurement consisted of 10 independent readings at 10 to 20 µm intervals. Deposition of thick collagen at each point of measurement was confirmed by correlation with Mason’s trichrome staining of parallel sections. An identical technique was used for measurements of subepithelial collagen in the four-month follow-up biopsies.

**PATHOLOGY**

Biopsies of the right colon revealed markedly thickened subepithelial collagen in a patchy but widespread distribution in most of the biopsy pieces. This thickened collagen table
ranged from 6.7 to 44.4 µm. The mean thickness of subepithelial collagen in the measured zones was 23.4 µm. Only well oriented biopsy fragments with at least a 200 µm length of uninterrupted collagen were used to maintain objectivity in measurements. The thickened subepithelial collagen was associated with a prominent increase in the inflammatory cell content of the lamina propria, predominantly lymphocytes and plasma cells and including focal eosinophilia and scattered neutrophils. Intraepithelial lymphocytes were conspicuous (Figure 1). The superficial colonic epithelium showed extensive degenerative and regenerative changes (Figure 2). Biopsies from the left colon revealed similar markedly thickened subepithelial collagen material, entrapped capillaries with fibrin thrombi and an inflammatory component containing a higher content of neutrophils than that seen in the right colon biopsies (Figure 3). Intraepithelial lymphocytes and epithelial degeneration were also identified. In addition, most biopsy fragments from the left colon demonstrated the presence of inflammatory pseudomembranes superimposed on the thickened subepithelial collagen (Figure 4). The pseudomembranes were comprised of fibrin with linear streams of neutrophils and karyorrhectic debris ‘erupting’ from a degenerated surface epithelium (Figure 5).

Follow-up biopsies of the left and right colon at four months revealed persistence of the thickened subepithelial collagen (range from 6.2 to 42.6 µm, mean 20.3 µm**), fewer intraepithelial lymphocytes but no objective decrease in the chronic inflammatory cell content of the lamina propria – persistent collagenous colitis changes. Active inflammatory cells and pseudomembranes were conspicuously absent in the follow-up biopsy (Figure 6).

**DISCUSSION**

Pseudomembranous colitis has been used descriptively to encompass not only the diarrheal syndrome following antibiotic use, but also any endoscopically or histologically recognized mucosal exudative process. Inflammatory pseudomembranes have been described in ischemic bowel disease, uremia, after irradiation or chemotherapy, and secondary to some bacterial infections, most notably verocytotoxin-producing *Escherichia coli* O157-H7. It was not until the late 1970s that Price and Davies (22) found that a toxin elaborated by *C difficile* was the etiological factor of the antibiotic-associated cases of pseudomembranous colitis. At least four enterotoxins are elaborated by *C difficile*, two of which (toxin A and B) induce mucosal injury. These toxins cause marked permeability in the mucosal epithelial cells and small blood vessels, with resultant fluid and cell loss, causing symptoms of cramping, abdominal pain, watery diarrhea, and findings of blood, fecal blood and leukocytosis. Fe-

Figure 2) Right colon biopsy showing thick irregular subepithelial collagen (large arrowhead) with vacuolar change and surface epithelial degeneration including vacuolar change (small arrowhead). (hematoxylin and eosin stain, original magnification ×400)

Figure 3) Left colon biopsy with collagenous colitis changes including deposition of subepithelial collagen (large arrowheads), entrapped capillaries with fibrin thrombi (small arrowheads). There is a relative increase in the number of polymorphonuclear cells compared with that seen in right colon biopsies (hematoxylin and eosin stain, original magnification ×400)

Figure 4) Left colon biopsy showing an inflammatory pseudomembrane comprised of fibrin and neutrophils (arrow) superimposed on irregularly thickened subepithelial collagen (arrowheads) (hematoxylin and eosin stain, original magnification ×200)
cal leukocytes, commonly seen in C difficile and other forms of infectious colitis, have also been described in collagenous colitis (23,24).

Collagenous colitis is a disease with an unknown etiology, poorly understood pathogenesis, irregular colonic distribution, arguable progression and variable, sometimes spontaneous resolution. Lindstrom (6), in his initial report, considered the collagen deposition to be a phenomenon secondary to the chronic inflammatory process that accompanied it. He considered the collagen material to be an anatomical barrier to fluid absorption – thus the watery diarrhea syndrome (6). Others consider the chronic inflammatory process that often coexists with the collagen to be the etiology of the diarrheal disease (25). Direct correlation of lamina propria cellularity and stool weight has been offered as evidence of the importance of the inflammatory component in the pathological process (25).

The relationship of ‘microscopic’ and lymphocytic colitis with collagenous colitis is not well understood. Some consider these processes to be a continuum, with lymphocytic colitis being the early manifestation and collagenous colitis being the late manifestation of the same disease. Similar clinical and endoscopic parameters as well as overlapping microscopic features are cited as evidence of a disease continuum (17,24,26,27). As with other forms of inflammatory bowel disease, there is an association of collagenous colitis with autoimmune diseases, thyroid disease, arthritis, celiac disease and idiopathic inflammatory bowel disease. Serum antinuclear antibody is found in up to 50% of collagenous colitis, as is rheumatoid factor, complement C3 and C4, and perinuclear antineutrophilic cytoplasmic antibody (28-32). However, the significance of autoantibodies or elevated serum immunoglobulin levels as an etiological factor in collagenous colitis has not been validated (28). No deposition of immunoglobulin or complement has ever been demonstrated in colonic tissue (1,26,33). Collagenous colitis has been found in association with inflammatory bowel diseases that ostensibly have more convincing evidence of an immunological pathogenesis, including celiac sprue, collagenous sprue, Crohn’s disease and ulcerative colitis (13,16,24,34-38). Other proposed etiological factors in the development of collagenous colitis have included bacterial cytotoxins and local toxic factors. Cytotoxic factors have been promulgated in studies of fecal cytotoxic activity after intestinal bypass where collagenous colitis resolved in the bypassed intestinal segment and recurred when the ostomy was closed (26,39). An undesignated bacterial cytotoxin has been demonstrated in collagenous colitis with profound reactivity against McCoy cells in culture but lacking neutralization by C difficile or Clostridium sordellii antitoxins (26). Cholestyramine, shown to induce resolution of symptoms and histological changes in several described cases of collagenous colitis, is also effective in pseudomembranous colitis. The bile sequestrating resin has a recognized cytoxin-binding capability (40,41). Likewise, the success of metronidazole and sulphasalazine as antibacterial agents, and their success in the treatment of collagenous colitis suggests a possible infectious etiology (4). Fecal leukocytes have been described in colla-

Figure 5) Left colon biopsy inflammatory pseudomembrane with linear streams of neutrophils and karyorrhectic debris in fibrin (curved arrow). Note the thickened subepithelial collagen (arrow) with entrapped capillaries and fibrin thrombi (arrowhead) (hematoxylin and eosin stain, original magnification ×400)

Figure 6) Follow-up biopsy of the left colon at four months reveals persistent subepithelial collagen (arrowhead), chronic inflammation of the lamina propria and complete absence of surface inflammatory pseudomembranes (hematoxylin and eosin stain, original magnification ×400)
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...and are commonly seen in pseudomembranous as well as other forms of infectious colitis.

Nonsteroidal anti-inflammatory drugs (NSAIDs) have also been implicated in the pathogenesis of collagenous colitis. Two large studies of patients with collagenous colitis reported NSAID use in 61% and 71% of patients, respectively, and resolution of diarrhea in some patients when the drug was discontinued (3,43). NSAIDs have been implicated in a wide range of intestinal pathology, including ischemic lesions, ulcers, bleeding, perforation and collagenous colitis. NSAID injury is believed to be a multistage process, with subcellular organelle damage and nonspecific tissue injury (44).

There is no standard, uniformly effective therapy for collagenous colitis; however, steroids, particularly prednisone, are widely believed to be the most effective therapy. Other drugs used with success include 5-aminosalicylate, sulphasalazine, azathioprine and methotrexate; locally active steroids such as budesonide have been advocated in patients refractory to prednisone (3,45-47). Efficacy of these therapies is difficult to verify because collagenous colitis often undergoes spontaneous regression of symptoms and histopathological lesions.

A direct relationship between pseudomembranous and collagenous colitis cannot be extrapolated from the findings in this report; their simultaneous occurrence in this patient may be coincidental. However, as described, intriguing overlapping clinical and pathological features may imply an infectious or cytotoxic basis for collagenous colitis as well as pseudomembranous colitis. The superimposition of historical and clinical findings of collagenous and pseudomembranous colitis has not been previously described, but as with other previously unrelated colitides and diarrheal syndromes, further understanding of the pathogenesis and related clinical syndromes may eventually demonstrate some association.

REFERENCES


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