Antibodies from a patient with type 1 diabetes and celiac disease bind to macrophages that express the scavenger receptor CD163

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Antibodies against the wheat storage globulin Glo-3A from a patient with both type 1 diabetes (T1D) and celiac disease were enriched to identify potential molecular mimicry between wheat antigens and T1D target tissues. Recombinant Glo-3A was used to enrich anti-Glo-3A immunoglobulin G antibodies from plasma by batch affinity chromatography. Rat jejenum and pancreas, as well as human duodenum and monocytes were probed, and binding was evaluated by immunohistochemistry and confocal microscopy. Glo-3A-enriched antibodies bound to a specific subset of cells in the lamina propria of rat jejenum that co-localized mostly with a marker of resident, alternatively activated CD163+ (CD163+*) macrophages. Blood monocytes and macrophage-like cells in human duodenum were also labelled with the enriched antibodies. Blocking studies revealed that binding to CD163+ macrophages was not due to cross-reactivity with anti-Glo-3A antibodies, but rather to non-Glo-3A antibodies co-purified during antibody enrichment. The novel finding of putative autoantibodies against tolerogenic intestinal CD163+ macrophages suggests that regulatory macrophages were targeted in this patient with celiac disease and T1D.

Key Words: Autoantibodies; CD163; Celiac disease; Gut; Glo-3A; Macrophages; Mimicry; Type 1 diabetes

Defective intestinal immunity can lead to impaired oral tolerance, chronic inflammation and gut leakiness. These conditions have been reported to precede the development of type 1 diabetes (T1D) in diabetes-prone BioBreeding (BBDp) rats and humans (1). We previously described a case involving a patient with both T1D and celiac disease who displayed strong antibody and T cell reactivity to wheat peptides, including those from the diabetes-related wheat storage globulin homologue of Glb1 (2) now known as Glo-3A (3). Additional studies revealed abnormal immune reactivity to Glo-3A in a subset of children at high risk for T1D (4) and in young children with celiac auto-immunity (5). To investigate mechanisms by which the immune response elicited by this dietary protein could contribute to inflammation in the gut or pancreas, and possibly T1D development, we probed target tissues with Glo-3A-enriched antibodies from the aforementioned patient to identify potential molecular mimicry.

METHODS

The clinical characteristics of the index patient with both T1D and celiac disease were described previously (2). Briefly, a young woman with a 10-year history of T1D and positive for tissue transglutaminase antibody was adhering to a standard gluten-free diet when she presented with diarrhea, oral ulcerations and severe swelling of the lips. The patient displayed strong T cell proliferative responses to wheat peptides including Glo-3A (2). Only the initiation of a strict, specified carbohydrate, cereal-free diet resolved the diarrhea and resulted in the healing of her mouth ulcerations. Plasma was prepared and frozen for future analyses. Informed consent was obtained and the study was approved by the Ottawa Hospital Research Ethics Board (Ottawa, Ontario).

To purify Glo-3A antibodies from this patient, recombinant Glo-3A protein was prepared. A His-Glo-3A DNA fragment was excised from the pET17b-His-Glo-3A vector using XbaI and XhoI restriction enzymes (Invitrogen, USA) and cloned into the XbaI/Xhol restriction enzyme sites of the pBacPak9 transfer vector (Clontech Laboratories, USA). The tagged gene was integrated into BacPAK6 viral DNA using Bactector (Clontech Laboratories, USA) according to the manufacturer's instructions. For protein expression and purification of His-Glo-3A, SF21 insect cells were infected with recombinant BacPAK6-His-Glo-3A virus. Recombinant His-Glo-3A was purified using Ni-NTA agarose according to the manufacturer's instructions.

To enrich human Glo-3A antibodies using batch affinity chromatography, 500 mg of lyophilized cyanoegen bromide-activated Sepharose-4B (GE Healthcare, USA) were suspended and coupled to 5 mg of recombinant Glo-3A. Excess ligand was removed by a series of washes with...
Antibody reactivity to rat CD163\(^+\) intestinal macrophages, human intestinal macrophage-like cells and a subset of human peripheral monocytes

Pancreatic and jejunal sections from a 45-day-old BB\(dp\) rat were probed with Glo-3A-enriched antibodies. There was strong staining of a specific population of macrophage-like cells in the lamina propria of the small intestinal villi (Figure 1A). These antibodies also bound to lamina propria cells in control BB and Wistar Furth rat small intestine. Confocal microscopy demonstrated that most of the cells labelled with the enriched anti-Glo-3A antibodies (Cy3) co-localized with a subset of macrophages expressing CD163 (Alexa 488) (Figure 1B). No labelling was observed in non-inflamed pancreas (data not shown). Staining of human duodenal sections with the Glo-3A-enriched antibodies revealed a similar labelling of macrophage-like cells in the lamina propria (Figure 1C). Staining of human peripheral blood monocytes revealed a subset of cells that was labelled with the Glo-3A-enriched antibodies (Figure 1D).

To further confirm the specificity of anti-Glo-3A antibody cross-reactivity in the gut, a series of control blocking experiments was performed with recombinant Glo-3A and SF21 insect cell proteins (without recombinant Glo-3A). The labelling was blocked by pre-absorption with Glo-3A or with SF21 insect cell proteins (Figure 1E). Similarly, labelling of human monocytes was blocked by pre-absorption with Glo-3A or with SF21 insect cell proteins (data not shown). Because Glo-3A was prepared in insect cells, Western blots of Glo-3A were probed with the enriched antibody preparation pre-absorbed with Glo-3A (prepared in insect cells) or insect cell proteins alone (data not shown). The present study revealed that Glo-3A antibodies were specifically blocked only by the Glo-3A preparation, and not blocked by insect cell proteins alone. Thus, pre-absorption with insect cell proteins blocks only non-Glo-3A antibodies that bound to gut tissue. These experiments revealed that antibody reactivity to intestinal macrophages was not due to specific cross-reactivity with human anti-Glo-3A antibodies, but rather to non-anti-Glo-3A antibodies that were co-purified during the enrichment process.

**DISCUSSION**

The original objective was to study whether Glo-3A antibodies could bind structures in the gut or pancreas consistent with the concept of molecular mimicry. Antibodies from a Glo-3A-enriched preparation labelled a subset of CD163-positive (CD163\(^+\)) macrophages in rat jejunum sections (Figure 1D). These antibodies also bound to lamina propria cells in control BB and Wistar Furth rat small intestine. Confocal microscopy demonstrated that most of the cells labelled with the enriched anti-Glo-3A antibodies (Cy3) co-localized with a subset of macrophages expressing CD163 (Alexa 488) (Figure 1B). No labelling was observed in non-inflamed pancreas (data not shown). Staining of human duodenal sections with the Glo-3A-enriched antibodies revealed a similar labelling of macrophage-like cells in the lamina propria (Figure 1C). Staining of human peripheral blood monocytes revealed a subset of cells that was labelled with the Glo-3A-enriched antibodies (Figure 1D).

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**RESULTS**

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jejenum lamina propria, a subset of human peripheral monocytes and macrophage-like cells in human duodenum. However, further control antibody-blocking experiments demonstrated that labelling was not due to anti-Glo-3A antibodies. Therefore, these results do not support molecular mimicry as an explanation for the enriched Glo-3A antibody cross-reactivity to macrophages. The serendipitous discovery of putative auto-antibodies against small intestinal macrophages in a patient with both T1D and celiac disease is an intriguing finding, sug- gesting the potential involvement of resident CD163+ macrophages in the pathophysiology of one or both of these diseases. CD163 is a class B scavenger receptor for hemoglobin-haptoglobin complexes that are expressed exclusively on monocytes and macrophages. CD163+ macrophages are mature, tissue-resident macrophages present at high frequency in the gastrointestinal tract of rats. They are associated with homeostatic functions including the suppression and resolution of inflammation, and wound healing (7). Unfortunately, the limited volume of patient plasma made it difficult to identify specific macrophage molecules targeted by these autoantibodies. To our knowledge, autoantibodies against macrophages have not been described in patients with T1D or celiac disease. However, such antibodies have been described in systemic lupus erythematosus, in which auto-antibodies against the class A scavenger receptors on macrophages of the marginal zone of the spleen were identified (8). A recent analysis of T1D HLA susceptibility gene interactions with can- didate proteins (9) identified macrophage scavenger receptor class A receptor molecules targeted by these autoantibodies. Anti- bodies have been described in diabetes-prone rats and mice, and there are fewer CD163+ CD163+ macrophages in BBdp rats compared with controls (10). Our initial investigations revealed significantly reduced numbers of CD163+ macrophages in the small intestine of diabetes-prone animals compared with controls (unpublished data).

Tripathi et al (11) identified human zonulin – a key regulator of tight junctions in the intestinal epithelium – as prehaptoglobin-2. Prehaptoglobin-2 is a known precursor of haptoglobin and, similar to haptoglobin, contains a CD163 binding site, further suggesting the importance of CD163+ macrophages in the modulation of intestinal immunity and, possibly, gut permeability. The identification of putative auto-antibodies against this macrophage population suggests a mechan- ism by which the normal homeostatic functions of these innate immune cells in the gut could be compromised. There is one report that diabetic patients harboured approximately 50% fewer CD163+ blood monocytes than age-matched controls (12). Taken together, these data raise the novel possibility that a deficit in tolerogenic CD163+ cells is implicated in the chronic intestinal inflammation and loss of oral tolerance observed in some T1D and celiac disease patients. CD163 could be a disease marker and a therapeutic target.

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