**Helicobacter pylori** infection and markers of gastric cancer risk in Alaska Native persons: A retrospective case-control study

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**BACKGROUND:** Alaska Native persons experience gastric cancer incidence and mortality rates that are three to four times higher than in the general United States population.

**OBJECTIVE:** To evaluate pepsinogen I, pepsinogen I/II ratio, anti-*Helicobacter pylori* and cytotoxin-associated gene A (CagA) antibody levels, and blood group for their associations with gastric cancer development in Alaska Native people.

**METHODS:** The present analysis was a retrospective case-control study that matched gastric cancers reported to the Alaska Native Tumor Registry from 1969 to 2008 to three controls on known demographic risk factors for *H pylori* infection, using sera from the Alaska Area Specimen Bank. Conditional logistic regression evaluated associations between serum markers and gastric cancer.

**RESULTS:** A total of 122 gastric cancer cases were included, with sera predicting cancer diagnosis (mean = 13 years) and 346 matched controls. One hundred twelve cases (91.8%) and 285 controls (82.4%) had evidence of previous or ongoing *H pylori* infection as measured by anti-*H pylori* antibody levels. Gastric cancer cases had a 2.63-fold increased odds of having positive anti-*H pylori* antibodies compared with their matched controls (P=0.01). In a multivariate model, non-cardia gastric cancer (n=94) was associated with anti-*H pylori* antibodies (adjusted OR 3.92; P=0.004) and low pepsinogen I level (adjusted OR 6.04; P=0.04). No association between gastric cancer and blood group, anti-CagA antibodies or pepsinogen I/II ratio was found.

**CONCLUSION:** Alaska Native people with gastric cancer had increased odds of previous *H pylori* infection. Low pepsinogen I level may function as a precursor marker for noncardia cancer.

**Key Words:** Alaska Native; cagA+; Gastric cancer; Helicobacter pylori; Pepsinogen I

**ORIGINIAL ARTICLE**

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impractical. Therefore, we sought associations between gastric cancer and serological markers that could form the basis of screening efforts to more efficiently identify individuals at higher risk for cancer so they may be targeted for early detection and treatment.

Studies involving other populations have investigated serum markers and H pylori virulence factors for their association with gastric cancer. Researchers have found associations between exposure to H pylori strains expressing the virulence factor cytotoxin-associated gene A (CagA) and gastric cancer (11,12). Low serum pepsinogen I levels and a low pepsinogen I/I ratio, indicative of chronic gastritis (a precursor of gastric cancer) (13), have shown an association with gastric cancer in some studies but not in others (14,15). Finally, some studies have suggested a possible association between blood group A and gastric adenocarcinoma (16,17), although other studies did not demonstrate this association (18,19). No studies have investigated these potential gastric cancer risk markers in Alaska Native people; furthermore, the aforementioned studies examined the association between the markers and patients at the time of their gastric cancer diagnosis. In the present study, our objective was to measure the association between gastric cancer development in Alaska Native people and potential serological cancer markers from samples obtained years before the cancer diagnosis.

METHODS

Study design
A retrospective matched case-control study was designed to investigate the association between gastric cancer and various serological and serum markers. Cases included Alaska Native individuals diagnosed with gastric adenocarcinoma in adulthood (≥18 years of age) residing in Alaska at the time of diagnosis. Alaska Native people belong to a diverse group of populations indigenous to Alaska. Patients with pathology-confirmed gastric cancer, who had at least one serum specimen in the Alaska Area Specimen Bank collected before their gastric cancer diagnosis, were identified from the Alaska Native Tumor Registry from 1969 through 2008. The Alaska Area Specimen Bank is a collection of >300,000 residual biological specimens from 92,000 people participating in various research studies, public health investigations and clinical testing conducted in Alaska since 1963.

Controls were Alaska Native people without known gastric adenocarcinoma (confirmed by review of the Alaska Native Tumor Registry) who resided in Northwest, Southeast, Southwest or Western Alaska, and had at least one serum specimen available from the Alaska Area Specimen Bank during the time period 1969 to 2008. To control for the known demographic risk factors for H pylori infection within the Alaska Native population (5), controls were matched to cases (3:1) according to region of residence in Alaska (southwest, southeast, west, northwest), age group (10-year age groupings), sex and date of serum specimen collection (±10 years). For cases in which multiple serum samples were available, samples collected >10 years before gastric cancer diagnosis were selected because H pylori serological titres have been reported to decline up to 10 years before cancer diagnosis (20). The study protocol received approval from the Centers for Disease Control and Prevention Institutional Review Board and the Alaska Area Institutional Review Board, including a waiver of informed consent because of the use of deidentified, previously collected medical information from the Alaska Native Tumor Registry. Study approval was received from the Bristol Bay Area and the Yukon-Kuskokwim Health Corporations (southwest), Maniilaq Association (northwest), Norton Sound Health Corporation (west) and the Southeast Alaska Regional Health Consortium (southeast).

Data abstraction
The Alaska Native Tumor Registry (www.anthc.org/clu/epicenter/), established in 1973, was used to obtain information about the gastric cancers. The Alaska Native Tumor Registry is a full member of the National Cancer Institute Surveillance, Epidemiology, and End Results (SEER) program, and provides comprehensive cancer surveillance of approximately 127,000 Alaska Native people residing in Alaska (21). Additional patient data were obtained from death certificates and RPMS, the health information system used by the Alaska Tribal Health System. The national SEER database (22), a collection of 17 regional registries that provide population-based surveillance for approximately 28% of the US population (http://seer.cancer.gov/), was accessed to obtain national gastric cancer data. SEER*Stat version 6.6.2 (National Cancer Institute, USA) was used to calculate frequencies from the national database (excluding cases from the Alaska Native Tumor Registry) for reported gastric cancers that occurred from 1973 to 2007.

Laboratory testing
Commercial kits were used to test samples for anti-H pylori antibodies (Helicobacter pylori IgA/IgG ELISA; Biohit, Finland), anti-CagA antibodies (Helicobacter pylori p120 [CagA] ELISA; ravo Diagnostika, Germany), pepsinogen I (Pepsinogen I ELISA, Biohit), pepsinogen II (Pepsinogen II ELISA, Biohit) and blood grouping (Affirmagen pooled reagent red blood cells; Ortho Clinical Diagnostics, USA). The manufacturer’s test procedures and analysis instructions were followed for all tests. The manufacturer’s cut-off values were used for normal versus abnormal levels of pepsinogen I (25 µg/L) and the pepsinogen I/I ratio (2.5). An abnormally low value for either indicates advanced corpus atrophy. Also followed was manufacturer guidance to determine anti-H pylori antibody positivity (≥50 enzyme immune units [EUI]) and anti-CagA antibody positivity (>7.5 units; 5 to 7.5 units indeterminate).

Sample size calculation and statistical analysis
Using previously reported estimates for H pylori and CagA seropositivity in Alaska Native people, sample sizes were calculated under a variety of assumptions. To detect an OR ≥2 at a 95% significance level (P<0.05) with 80% power and 1:3 case-control matching, the calculations produced sample sizes of 107 to 163 cases and 321 to 489 controls (data not shown). Descriptive analyses of case, control and gastric cancer characteristics were undertaken, which are reported as frequencies and percentages; z, χ² and paired t tests were used to evaluate differences in proportions and their distributions and paired serological data, as appropriate. The analysis excluded pepsinogen values from five grossly hemolyzed serum samples and anti-H pylori antibody enzyme immunoassay values >1500 (but not the positive test result) due to the instability of the test kit in that range (14 samples). To preserve matching, associations between serological markers and gastric cancer development were checked using univariate and multivariate conditional logistic regression. When more than one serum sample per case or control was available, the earlier-collected sample was used for modeling. Purposeful backward stepwise regression was used for multivariate models and initially included variables with P≤0.2. In addition, subgroup analyses were performed and restricted to: gastric cancers not located in the cardia region of the stomach (noncardia) because of the reported lack of association between H pylori infection and gastric cancers arising from the cardia of the stomach (3,4,23); cases with serum samples collected ≥10 years before gastric cancer diagnosis due to the reported decline of H pylori antibodies leading up to gastric cancer diagnosis (20); gastric cancer cases diagnosed before 50 years of age (approximately the first age quartile in the sample) because of genetic differences in early onset cancers (24,25); and anti-H pylori antibody-positive cases and controls. All reported P values are two-sided; P<0.05 was considered to be statistically significant. Stata version 10 (StataCorp, USA) was used to perform statistical analyses.

RESULTS

Participant characteristics
Of the 206 reported cases of gastric adenocarcinoma with at least one available serum sample over the 40-year study period, permission from the regional tribal health organizations was granted to include 129 (62.6%) of these cases and initially matched to 377 controls. From this group were
excluded: three cases for lack of matched controls; one case and 21 controls for sex mismatch; three cases with non-adenocarcinoma histology (two epithelial and one squamous cell) and their nine matched controls; and one control who identified as non-Native. One hundred twenty-two individuals with gastric cancer and 346 matched controls were retained for analysis (Figure 1). Two serum samples were available for 38 (31.1%) cases; the other cases and all of the controls had one sample. Samples obtained from cases predated gastric cancer diagnosis by a mean of 13 years (interquartile range nine to 18 years). Of the gastric cancer group, 73.0% (89 of 122) were male, the group's mean (± SD) age at the time of serum sample collection was 45.2±16.1 years, mean age at time of diagnosis was 58.6±15.7 years and the proportion <50 years of age was 29.5% (36 of 122). The control group was 72.8% (252 of 346) male and had a mean age of 41.2±17.6 years at serum collection. Almost all cases and controls, 92.6% and 92.8%, respectively, identified as Eskimo, and 93.4% and 94.2%, respectively, lived in rural western Alaska communities.

Serum markers and gastric cancer

It was found that 91.8% (112 of 122) of the gastric cancer patients and 82.4% (285 of 346) of the control group had evidence of previous or ongoing H pylori exposure indicated by elevated anti-H pylori antibody levels (Table 1). A greater percentage of cases (95.1% [n=116]) and controls (93.1% [n=322]) had evidence of H pylori exposure as measured by anti-CagA immunoglobulin G (IgG). All of the cases (n=122) and 342 (98.8%) of the controls demonstrated evidence of previous or ongoing H pylori exposure when combining these two serological markers of H pylori infection. Low pepsinogen I level was uncommon, with 4.1% (five of 121) of cases and 2.1% (seven of 342) of controls recording serum pepsinogen I levels <25 µg/L. Equally uncommon was a low pepsinogen I/II ratio: 5.0% (six of 121) of case and 2.9% (10 of 342) of control samples had a calculated ratio <2.5.

Individuals who developed gastric cancer had a 2.59-fold higher odds of positive H pylori serology than their matched controls (P=0.013; 95% CI 1.22 to 5.50), the only significant univariate association (Table 2). Also calculated were matched ORs using a pepsinogen threshold of 75 µg/L and a pepsinogen I/II ratio of 10; however these analyses did not result in any additional statistically significant associations (data not shown). In the multivariate analysis, which retained variables with P<0.25 (positive H pylori serology and pepsinogen I), only the presence of H pylori antibody was significantly associated with gastric cancer (OR 2.63 [95% CI 1.21 to 5.62], P=0.01) (Table 2).

Analysis of gastric cancer patient subgroups revealed associations between gastric cancer and H pylori exposure and low pepsinogen I levels, as shown in Table 2. A multivariate model of noncardia gastric cancer cases (n=94) showed associations with anti-H pylori antibodies (adjusted OR [aOR] 3.92; P=0.004) and low pepsinogen I levels (aOR 6.04; P=0.04). Individuals diagnosed with gastric cancer before 50 years of age (n=36) had a stronger association with anti-H pylori antibodies (aOR 7.96; P=0.047) than cases diagnosed in individuals ≥50 years of age (n=86; aOR 1.88; P=0.14). Gastric cancer patients with serum specimens collected ≥10 years before their cancer diagnosis (n=86) had an association with anti-H pylori antibodies (aOR 3.20; P=0.013), while cases with specimens from <10 years before diagnosis did not (n=36; aOR 0.13; P=0.86). Also investigated were cases and controls positive for anti-H pylori antibodies, which found a nonsignificant association between gastric cancer and antecedent low pepsinogen I level (aOR 4.48; P=0.08). To assess for temporal changes in specimen values and gastric cancer associations, serum specimens collected before and after 1980 were grouped and analyzed. The association of gastric cancer with anti-H pylori and anti-CagA antibodies, pepsinogen I, and the pepsinogen I/II ratio between the two time periods were similar and not significantly different (data not shown). Additionally, the mean values for these serum markers were not statistically different across the two time periods (data not shown).

**Pair ed sera**

For 38 of the gastric cancer cases, two separate prediagnosis serum samples were collected a mean of 7.2 years apart (interquartile range four to nine years) (Table 3). Of the measured serum markers, only anti-H pylori antibody levels changed significantly between the earlier and later samples (mean increase = 31.4±90.0 EIU; P=0.04), increasing an average of 5.9 EIU per year. Paired specimen antibody levels...
TABLE 2
Gastric cancer predictors according to case group

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>All cases (n=122)</th>
<th>Cases with specimens ≥10 years before diagnosis (n=86)</th>
<th>Noncardia cases (n=94)</th>
<th>Helicobacter pylori-positive cases and controls (n=112)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Univariate analysis</td>
<td>Multivariate analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>OR</td>
<td>P</td>
<td>OR</td>
<td>P</td>
</tr>
<tr>
<td>H pylori positive</td>
<td>2.59</td>
<td>0.01</td>
<td>3.39</td>
<td>0.01</td>
</tr>
<tr>
<td>CagA intermediate and positive†</td>
<td>0.98</td>
<td>0.97</td>
<td>1.32</td>
<td>0.63</td>
</tr>
<tr>
<td>CagA positive†</td>
<td>1.40</td>
<td>0.47</td>
<td>1.95</td>
<td>0.23</td>
</tr>
<tr>
<td>Pepsinogen I &lt;25 µg/L</td>
<td>1.97</td>
<td>0.27</td>
<td>2.50</td>
<td>0.20</td>
</tr>
<tr>
<td>Pepsinogen I/II ratio &lt;2.5</td>
<td>1.72</td>
<td>0.33</td>
<td>2.33</td>
<td>0.19</td>
</tr>
<tr>
<td>Blood group A versus others</td>
<td>1.11</td>
<td>0.62</td>
<td>1.27</td>
<td>0.36</td>
</tr>
<tr>
<td>H pylori positive</td>
<td>2.63</td>
<td>0.01</td>
<td>3.32</td>
<td>0.01</td>
</tr>
<tr>
<td>Pepsinogen I, low (&lt;25 µg/L)</td>
<td>2.56</td>
<td>0.15</td>
<td>2.98</td>
<td>0.15</td>
</tr>
</tbody>
</table>

Bolded values indicate statistical significance. *Anti-H pylori immunoglobulin (Ig) G/IgA ≥30 enzyme immune units; †Cytotoxin-associated gene A (CagA) IgG: <5 units (negative), 5 to 7.5 units (indeterminate), >7.5 units (positive); ‡Only variables significant at P<0.2 were retained in the multivariate model. na Not applicable

TABLE 3
Characteristics of paired sera taken before gastric cancer diagnosis

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Specimen</th>
<th>Earlier (n=38)</th>
<th>Later (n=38)</th>
<th>Paired, change (Δ)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at serum collection, years</td>
<td>109.6±77.5</td>
<td>141.7±121.7</td>
<td>31.4^†</td>
<td>5.9</td>
</tr>
<tr>
<td>Serum collection before cancer diagnosis, years</td>
<td>29.3±20.6</td>
<td>28.3±19.7</td>
<td>-0.9</td>
<td>-0.4</td>
</tr>
<tr>
<td>Pepsinogen I, µg/L</td>
<td>138.6±65.9</td>
<td>140.8±91.2</td>
<td>2.2</td>
<td>0.5</td>
</tr>
<tr>
<td>Pepsinogen I/II ratio</td>
<td>8.9±3.9</td>
<td>8.9±5.4</td>
<td>0.01</td>
<td>0.1</td>
</tr>
<tr>
<td>Blood group A versus others</td>
<td>49.3±15.9</td>
<td>56.6±15.3</td>
<td>7.2</td>
<td></td>
</tr>
<tr>
<td>Serum pepsinogen I, µg/L</td>
<td>12.1±6.5</td>
<td>4.9±5.0</td>
<td>7.2</td>
<td></td>
</tr>
</tbody>
</table>

Data presented as mean ± SD unless otherwise indicated. *One sample censored from each case group due to instability of laboratory test at values >1500 enzyme immune units. †P<0.05 for paired t test between earlier and later specimens

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DISCUSSION

For the first time, we report an association between the presence of serum anti-H pylori antibodies and the development of gastric cancer in Alaska Native people. This association was larger among people with noncardia gastric cancer and individuals <50 years of age. The larger OR found in younger individuals likely arises from control H pylori seropositivity increasing with age, which diminishes the association apparent in the older age group. Unlike results reported by most previous studies (11,12), exposure to CagA-positive H pylori strains, as measured by anti-CagA antibody levels, did not appear to increase an individual’s risk for gastric cancer in our study population. The near-ubiquitous presence of anti-CagA antibodies (95.1% of each of cases and controls), which was significantly higher than reported in other studies (28,29), may have masked any true association between this virulence factor and gastric carcinogenesis. However, other studies have also found no association between infection with CagA-positive H pylori strains and gastric cancer (30), particularly in populations in which H pylori prevalence more closely mirrors that of rural Alaska (31). The high proportion of participants seropositive for CagA is likely due to prevalent H pylori infection and the relative antigenicity of the CagA protein, resulting in persistent CagA antibodies even after clearance of H pylori infection and H pylori IgG antibodies (32).

Few studies have evaluated pepsinogen levels and ratios before gastric cancer diagnosis to determine the predictive or screening value of these markers for gastric cancer. Most studies examined these serum markers in individuals at the time of their gastric cancer diagnosis. However, other studies have also found no association between infection with CagA-positive H pylori strains and gastric cancer (30), particularly in populations in which H pylori infection and the relative antigenicity of the CagA protein, resulting in persistent CagA antibodies even after clearance of H pylori infection and H pylori IgG antibodies (32).

The gastric cancers included in this study were moderately differentiated (n=35 [28.7%]) or poorly differentiated (n=55 [43.1%]) histologically, which was similar to national data (12). Most gastric cancers in the study were moderately differentiated (n=35 [28.7%]) or poorly differentiated (n=55 [43.1%]) histologically, which was similar to national data. This distribution of reported gastric cancer sites was different from national reports (12). Most gastric cancers occurring in this sample were in a variety of locations in the stomach: 28% in the cardia; 69% in the body (42% greater and lesser curvatures, and antrum); six (4.9%) in the pylorus; four (3.3%) in overlapping regions; and 15 (12.3%) at unspecified locations. This distribution of reported gastric cancer sites was different from national reports (12). The proportion of Alaska Native cancers occurring in the greater and lesser curvatures, and fewer occurring in the body and overlapping regions, was similar to national data. Summary staging information was available for 101 (82.8%) of the cancers, and 21 (20.8%) of the staged cancers were localized, 44 (43.6%) were regional and 36 (35.6%) were distant, which was similar to the national data (12).
and other published studies may relate to variations in laboratory methods, particularly duration of sample storage, and is unlikely a reflection of differing gastric cancer pathophysiology given the similarity of our sample cases to the SEER national sample.

The descriptive pathology of our study sample of Alaska Native gastric cancers was similar to that reported by SEER registries from other US populations. Summary staging information from our study population and from a recent report on cancer in Alaska Natives (36) suggests that gastric cancers in Alaska Native people were diagnosed at a stage similar to those reported in the national SEER database. This finding implies that delayed diagnosis due to access to healthcare (availability and utilization) is not the main explanation for increased Alaska Native gastric cancer mortality. Histology and histological grade were also similar between the two groups, indicating that Alaska Native people did not experience different or more aggressive tumours than other US populations. The main difference between the two groups was in the recorded sites of the gastric adenocarcinomas, although the differences were mainly in the specific location within the body of the stomach (body, lesser and greater curvatures), which is likely due to variance in surgical reporting and not cancer location. The similarity in descriptive gastric cancer pathologies and stage at diagnosis between the national and study samples suggests that the reason for a higher gastric cancer mortality rate in Alaska Native people is due to elevated cancer incidence in this population.

In the 38 individuals with gastric cancer and available paired serum specimens, we saw a significant increase in anti-H pylori EU1 levels from earlier to later specimens, although antibody level changes were heterogeneous, with some individuals experiencing a decline in antibody levels. Our finding confirms the report by Tulinius et al (20) describing a decline in H pylori antibody levels approaching cancer diagnosis. The increasing antibody levels apparent in our study may have resulted from the high prevalence of H pylori infection in this population, leading to re-exposure to H pylori and priming of the immune system. Focused exploration of anti-H pylori antibody levels and their temporal relation to gastric cancer diagnosis may further clarify the association.

We aimed to sera obtained before a diagnosis of gastric cancer. However, because of the retrospective nature of the study, we did not perform endoscopy on individuals to confirm their lack of gastric cancer at the time of serum collection, meaning that we may have included serum samples from individuals (cases) with undiagnosed gastric cancer. However, this scenario is unlikely because 70% of samples predated diagnosis by 10 years and >90% predated diagnosis by at least five years, during which time the likelihood of identifying gastric cancer would have been high. Additionally, our subanalysis was restricted to cases with samples predated diagnosis by at least 10 years and similar findings to the primary analysis (Table 2). Another challenge was the multiple changes made to summary staging and ICD-O-3 code definitions during the 40 years of study data. To correct for this, SEER provides updated coding and staging manuals on a regular basis and recodes data within SEER*Stat to provide uniformity across the multiple years of cancer registry data. Finally, the sample of Alaska Native people with gastric adenocarcinomas may not represent all Alaska Native people because the study population resided primarily in rural areas.

Evidence of H pylori infection was widespread in our study population. Although we demonstrated an association between previous infection and subsequent gastric cancer, the utility of H pylori seropositivity screening to predict gastric cancer in a population with such elevated rates of infection and reinfection is less. While noncardia gastric cancer cases showed an association with previous low pepsinogen I levels, the extremely low sensitivity of this potential screening test for predicting gastric cancer minimizes its clinical utility in this population. Because of the disproportionate burden of gastric cancer in the Alaska Native population, we will continue to examine potential markers of gastric cancer risk and to evaluate potential screening strategies to identify individuals at risk. As a next step, we will characterize H pylori strains circulating in the Alaska Native population to search for genotypes associated with gastric cancer and study how host characteristics predispose individuals to gastric cancer.

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REFERENCES


