Pharmacokinetic properties of clarithromycin: A comparison with erythromycin and azithromycin

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OBJECTIVE: To compare the pharmacokinetic properties of two new macrolide antibiotics, clarithromycin and azithromycin, with those of the prototype macrolide, erythromycin. DATA SOURCES: Primarily peer review journals were searched for papers describing the pharmacokinetics of these new macrolides. STUDY SELECTION: Fifteen in vitro and clinical studies of clarithromycin and azithromycin and one clinical abstract on clarithromycin from the past four years were selected for review. DATA EXTRACTION: Data relevant to the pharmacokinetic characteristics of clarithromycin, azithromycin and, to a lesser extent, erythromycin were selected for presentation in this comparison. DATA SYNTHESIS: By reviewing the available studies, it was possible to construct pharmacokinetic profiles of the new compounds, and to compare them with each other and with erythromycin. CONCLUSIONS: Both clarithromycin and azithromycin have been shown to have an antibacterial spectrum and pharmacokinetic profile superior to that of erythromycin. The differences between the new compounds, however, may not be that significant. Each is likely to become a first-line therapeutic option in specific instances, which will become better delineated as clinical research on these new macrolides continues.

Key Words: Azithromycin, Clarithromycin, Erythromycin, Macrolides, Pharmacokinetics

Propriétés pharmacocinétiques de la clarithromycine : Comparaison avec l'érythromycine et l'azithromycine

OBJECTIF : Comparer les propriétés pharmacocinétiques de deux nouveaux macrolides, soit la clarithromycine et l'azithromycine, avec celles du macrolide prototype érythromycine. SOURCES DES DONNÉES : Des publications scientifiques avec comité de révision scientifique ont principalement été passées en revue et on y a sélectionné des articles sur la pharmacocinétique de ces nouveaux macrolides. SÉLECTION DES ÉTUDES : Quinze études in vitro et cliniques sur la clarithromycine et l'azithromycine et un résumé clinique sur la clarithromycine publiés au cours des quatre dernières années ont été passés en revue. EXTRACTION DES DONNÉES : Des données relatives aux caractéristiques pharmacocinétiques de la clarithromycine, de l'azithromycine et, dans une proportion plus faible, de l'érythromycine, ont été choisies aux fins de cet article comparatif. Synthèse des données : En passant en revue les études disponibles, il a été possible de dresser un profil pharmacocinétique des nouveaux composés et de les comparer entre eux, de même qu'avec l'érythromycine. CONCLUSIONS : La clarithromycine et l'azithromycine se sont toutes deux dotées d'un spectre antibactérien et d'un profil pharmacocinétique supérieur à ceux de l'érythromycine. Les différences entre ces deux nouveaux produits ne sont nécessairement significatives. Chacun est susceptible de devenir un choix thérapeutique de première ligne dans certains cas spécifiques qui seront définis plus précisément à mesure que la recherche clinique se poursuivra.
Erythromycin is commonly used to treat a variety of infections, including those of the respiratory tract. Although the compound is effective against a fairly broad range of organisms (including atypical pathogens), its clinical success may be limited by erratic low blood and tissue levels, and a tid to qid dosing schedule. In addition, patients complain frequently of gastrointestinal adverse effects associated with erythromycin. A number of congeners are now in development (and one, clarithromycin, is available in Canada) that promise to improve upon the antibiotic spectrum, pharmacokinetics, safety, and tolerability of erythromycin.

Clarithromycin is an acid-stable analogue of erythromycin with a methoxy substitution at C-6 of the erythronolide ring. This structural alteration prevents acid-induced conversion of the molecule to inactive spiropental forms in the stomach and improves bioavailability and gastrointestinal tolerance after an oral dose. It therefore increases antibacterial activity (1) compared with erythromycin. Clarithromycin has demonstrated bactericidal activity against both typical and atypical respiratory pathogens.

When clarithromycin is metabolized in humans, a microbiologically active metabolite, 14-hydroxy clarithromycin, is formed. This active metabolite has been shown to contribute an additive or synergistic effect to the activity of the parent compound in vivo against selected pathogens. Results from in vitro and in vivo testing of the combined compounds suggest that routine in vitro susceptibility tests and animal efficacy studies with clarithromycin alone may underestimate its potential efficacy against Haemophilus influenzae (2).

Azithromycin, a 15-membered macrolide that is classified as the first azalide, differs from erythromycin by the insertion of a methyl-substituted nitrogen into position 9a of the large aglycone ring. The insertion of the nitrogen into the ring distinguishes azalides from macrolides (which have only carbon- and oxygen-containing rings), and significantly alters the chemical, microbiological, and pharmacokinetic properties of these compounds (3).

This paper compares the pharmacokinetic properties of these new compounds, clarithromycin and azithromycin, with those of erythromycin.

### TABLE 1
Mean pharmacokinetic parameters of erythromycin, azithromycin and clarithromycin following a single dose in healthy volunteers*

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Erythromycin base (500 mg)</th>
<th>Erythromycin estolate** (500 mg)</th>
<th>Azithromycin (500 mg)</th>
<th>Clarithromycin (500 mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bioavailability (%)</td>
<td>-25</td>
<td>-25†</td>
<td>37</td>
<td>55</td>
</tr>
<tr>
<td>C_{max} (μg/L)</td>
<td>1.9 - 3.8</td>
<td>3.08</td>
<td>0.4</td>
<td>1.6</td>
</tr>
<tr>
<td>T_{max} (h)</td>
<td>1 - 5</td>
<td>2.65</td>
<td>2.5</td>
<td>1.7 - 1.9</td>
</tr>
<tr>
<td>AUC (μg h/L)</td>
<td>5.8 - 11.2</td>
<td>20.39</td>
<td>3.39†</td>
<td>4 - 4.2</td>
</tr>
<tr>
<td>Protein binding (%)</td>
<td>26 - 35</td>
<td>75</td>
<td>7 - 50§</td>
<td>65 - 75</td>
</tr>
<tr>
<td>half-life (h)</td>
<td>1.5 - 3.0</td>
<td>3.0</td>
<td>11 - 14</td>
<td>2.6 - 2.7§</td>
</tr>
</tbody>
</table>

*Results are reported as mean ± SD where available. †AUC_{0-72}. ‡Varies with formulation, salt form, and relation to meals. §May change with serum concentration. †May increase with increasing doses. AUC area under the curve; C_{max} maximum concentration; T_{max} time to maximum concentration. **Erythromycin estolate data from Croteau D, Bergeron MG, LeBel M. Antimicrob Agents Chemother 1988;32:561-8.

### ABSORPTION

Erythromycin base is incompletely but adequately absorbed from the upper part of the small intestine, and it is inactivated by gastric acid. The drug is therefore administered as enteric-coated formulations or as esters stable to gastric acid, eg, erythromycin estolate or ethylsuccinate. Bioavailability, generally about 25% of an oral dose (3), is decreased when erythromycin base or stearate is administered with food (4). As shown in Table 1, maximum concentration of 1.9 to 3.8 μg/mL and 3.08 μg/mL are attained with erythromycin base and estolate 500 mg single dosing.

Both clarithromycin and azithromycin have been shown to have pharmacokinetic profiles superior to that of erythromycin (Table 1). These new compounds have a more predictable pattern of absorption than the prototype. After an oral dose, clarithromycin reaches peak serum concentrations within 2 h, azithromycin within 2.5 h, and both achieve higher concentrations in tissue than in serum. In trials evaluating the effect of food on the disposition of clarithromycin, it was found that food may enhance absorption and bioavailability of the drug from 55 to 70% (1.5-6). Clarithromycin is stable in gastric acid. Steady-state peak serum concentrations are 1.0 to 1.5 mg/L after a 250 mg twice-daily dose, and 2.0 to 3.0 mg/L after a 500 mg twice-daily dose.

The bioavailability of a 500 mg oral dose of azithromycin is only about 37%, and peak serum concentration attained is 0.4 mg/L (7). Serum concentrations with multiple-dose regimens were slightly higher, but always below 1 mg/L. Studies of the influence of food on azithromycin absorption revealed that peak serum concentration decreased by 52% and area under the plasma concentration-time curve (AUC) diminished by 43% (8).

There are apparent discrepancies between bioavailability and peak plasma concentration (C_{max}) values for erythromycins, clarithromycin, and azithromycin because no equivalent intravenous formulations exist for erythromycin. Lactobionate and guceptate intravenous erythromycins are not fully equivalent. Thus, bioavailability values for erythromycin and erythromycin estolate are only approximate.
DISTRIBUTION AND TISSUE PENETRATION

Malcolide antibiotics are known to bind to plasma proteins, particularly alpha1-acid glycoprotein. These compounds are lipophilic, and they penetrate well into tissues (5).

Scaglione and Fraschini (9) evaluated the diffusion of clarithromycin into respiratory tissues, including the nasal mucosa, tonsils, and lungs, in adult patients undergoing surgery. For the three days preceding their surgical procedures, patients received clarithromycin, 250 mg twice daily (nasal mucosa or tonsillar tissue), or 500 mg twice daily (lung parenchyma). For clarithromycin and its active metabolite, terminal disposition half-life (t1/2b) values in sputum were 1.3- to 1.6-fold longer than those in serum. In tonsils, mean 4 h post dose parent and metabolite concentrations were 5.3 and 3.1 mg/kg, respectively, and mean 12 h post dose values were 2.1 and 1.2 mg/kg (Figure 1). Parent and metabolite concentrations in nasal mucosa 4 h post dose were 5.9 and 3.2 mg/kg, respectively, and mean 12 h post dose values were 2.2 and 1.5 mg/kg. In lung tissue, parent and metabolite concentrations 4 h post dose were 13.5 and 7.2 mg/kg, respectively, and 12 h post dose values were 2.8 and 2.0 mg/kg (9). Significantly, these values exceed the 90% minimum inhibitory concentration (MIC90) for many respiratory pathogens (Table 2).

In a recent study conducted in the United Kingdom, pulmonary tissue concentrations of azithromycin were measured in 22 patients (10). Up to 96 h after a single 500 mg dose, the following findings were observed: mean peak concentration of the drug in sputum, 1.56 mg/L; in bronchial mucosa, 3.89 μg/mL; in alveolar macrophages, 23 mg/L. Serum concentrations were significantly lower (0.13 mg/L 12 h post dose), and were generally sub-MIC throughout the study period.

The concentration of azithromycin in most tissues has been shown to exceed serum concentrations by 10- to 100-fold (7). Single oral dose (500 mg) studies of azithromycin have found that the concentration of azithromycin in most tissues 12 to 48 h after dosing ranges from 1 to 9 mg/kg (3). Mean concentrations for a single tissue type are usually greater than 2.0 mg/kg, and all tissue levels are higher than those of concurrently obtained serum samples. The average 'tissue' half-life is two to four days. One may question whether such an extended half-life is completely beneficial.

Although intraphagocytic bioactivity is not a common property of antimicrobial agents (11), the newer macrolide antibiotics achieve high intracellular concentrations. Both clarithromycin and azithromycin have been shown to penetrate macrophages and leukocytes, which makes them particularly effective against intracellular pathogens such as Legionella pneumophila and Chlamydia species (5). In contrast, penicillin and cephalosporin antibiotics are not actively concentrated by phagocytes, and they possess only modest, intracellular activity (11).

Anderson and colleagues (11) observed that erythromycin was rapidly concentrated by neutrophils, with an intracellular to extracellular (I:E) ratio of 7:3. The I:E ratio for clarithromycin was 9:1. These investigators concluded that the superior pharmacokinetic properties of clarithromycin will lead to increased intraphagocytic accumulation and bioactivity in vivo.

Therapeutic concentrations of clarithromycin have also been found to stimulate protein kinase C activity in polymorphonuclear leukocytes (PMNLs). Thus, in addition to its antimicrobial activity, the drug stimulates cellular host defense mechanisms involving the activation of protein kinase C (12).

It was recently shown that, among all macrolides tested so far, azithromycin provides the highest I:E ratio, confirmed both in vitro and in vivo, with values of approximately 160 obtained in vitro for azithromycin.

**Table 2**

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>MIC90(mg/L)</th>
<th>MIC90(mg/L)</th>
<th>MIC90(mg/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Bacteria</strong></td>
<td><strong>Erythromycin</strong></td>
<td><strong>Azithromycin</strong></td>
<td><strong>MIC90(mg/L)</strong></td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>&gt;64.0</td>
<td>1.0</td>
<td>&gt;64.0</td>
</tr>
<tr>
<td>Streptococcus pneumoniae</td>
<td>0.015 - 0.25</td>
<td>0.015 - 0.25</td>
<td>0.015 - 0.25</td>
</tr>
<tr>
<td>Streptococcus pyogenes</td>
<td>0.03 - 4.0</td>
<td>0.03 - 4.0</td>
<td>0.015 - 0.25</td>
</tr>
<tr>
<td>Moraxella catarrhalis</td>
<td>0.25</td>
<td>0.06</td>
<td>0.12 - 0.25</td>
</tr>
<tr>
<td>Haemophilus influenzae</td>
<td>2.0 - 8.0</td>
<td>0.25 - 1.0</td>
<td>0.25 - 8.0*</td>
</tr>
<tr>
<td>Legionella pneumophila</td>
<td>2.0</td>
<td>2.0</td>
<td>0.25</td>
</tr>
</tbody>
</table>

*MIC90 of clarithromycin plus 14-OH metabolite: 0.25 - 2.0 (Adapted from Bahal N, Nahata M, Ann Pharmacother 1992:26:48)
The theory that onsite, intraphagocytic delivery of azithromycin provides a significant amount of bioactive antimicrobial agent has been demonstrated in vitro and in vivo. However, this concept applies to all macrolides, including erythromycin, that are known to accumulate in phagocytic cells.

**METABOLISM AND ELIMINATION**

Although the metabolism of the macrolide antibiotics has not been extensively studied, it is known that a portion of the dose is metabolized in the liver. Macrolide antibiotics are demethylated by the cytochrome P-450-III microsomal enzyme system. Clarithromycin is metabolized to eight metabolites, but only one, the 14-hydroxy metabolite, has been shown to have antibacterial activity. The activity of this metabolite is comparable to or greater than that of the parent compound. The 14-hydroxy metabolite has been shown to act synergistically or additively with the parent compound, thereby extending clarithromycin's antimicrobial spectrum to include *H. influenzae*. It is not known whether any of the metabolites of azithromycin are active.

The pharmacokinetics of clarithromycin appear to be dose-dependent and nonlinear, apparently as a result of capacity-limited saturation of metabolic pathways. However, such nonlinearity is slight at the recommended dosages. Disproportionate increases in C\text{max}, t\text{1/2b}, and AUC have been reported in patients receiving a single high dose (1.2 g) or multiple doses. Similar dose dependency has been observed with the 14-hydroxy metabolite.

Thirty to 40% of an oral dose of clarithromycin is excreted unchanged or as an active metabolite via the kidneys, and the remainder is excreted via the bile. In individuals with normal renal function, the half-lives of clarithromycin and its 14-hydroxy metabolite after a 500 mg dose are 5 and 7 h, respectively. As renal function declines, the serum half-lives of these compounds increase to 7.7 h and 14 h, respectively. At a creatinine clearance of 30 to 80 mL/min, clarithromycin's half-life is 12 h, and this interval increases to 32 h when the creatinine clearance falls below 30 mL/min. For 14-hydroxy clarithromycin at the lower creatinine clearance, the half-life is 47 h. Clearly, regimen alteration would be advisable in patients with severely impaired renal function.

Severe hepatic impairment could theoretically alter the pharmacokinetics of clarithromycin and its metabolite so that less metabolite would be formed, and renal clearance of the parent compound would increase. Steady-state levels of unchanged clarithromycin in hepatically impaired patients are similar to those in normal subjects, so if renal function is normal, the drug can be administered without dose adjustment.

Azithromycin elimination is polyphasic: the initial rapid decline in drug serum levels is followed by multiple-phase distribution and elimination. After a single 500 mg dose, terminal half-life probably exceeds 40 h. When azithromycin metabolism occurs, demethylation is the primary route. The metabolites, which may number as many as 10, are not thought to have any significant antimicrobial activity. Urinary excretion of unchanged azithromycin in humans appears to be a minor elimination route, usually amounting to less than 6% within 24 h after an oral 500 mg dose. About 20% of the drug that reaches the systemic circulation is excreted unchanged in the urine. The feces are an important route of elimination for azithromycin; binary concentrations of the drug far exceed serum concentrations, suggesting binary excretion. Over half the drug-related material in the bile is unchanged. Intestinal excretion may be the primary route of elimination of the unchanged compound.

Only 2 to 5% of an oral dose of erythromycin is excreted in active form in the urine. The antibiotic is metabolized through the liver and excreted in active form in the bile.

**DRUG REGIMENS**

The usual dose of erythromycin for adults ranges from 1 to 2 g/day, given in equally divided and spaced amounts, usually every 6 h. Azithromycin may be given as a single 1 g dose in specific instances, but the more common regimen is a five-day course of therapy, beginning with a 500 mg dose on day 1, followed by daily 250 mg doses on days 2 through 5.

For clarithromycin, the usual adult dose for infections of the respiratory tract and the skin and soft tissues is 250 to 500 mg every 12 h for seven to 14 days. In patients with both hepatic and renal impairment, or in the presence of severe renal impairment, decreased dosage or prolonged dosing intervals may be appropriate.

**FUTURE OUTLOOK**

The pharmacokinetic advantages and superior spectra of activity of clarithromycin and azithromycin over erythromycin (base and esters) are well delineated; however, it is my opinion that differences between clarithromycin and azithromycin are not as dramatic as they may appear to be. Azithromycin's long half-life is not, in itself, an advantage. When evaluating tissue penetration of clarithromycin and azithromycin, one would prefer data from the same investigators under similar study conditions. Comparative studies are currently under way and are likely to provide further insights into understanding these new compounds.

Both clarithromycin and azithromycin offer therapeutic advantages in certain areas, and they are likely to become first-line therapy in a number of situations. The once- or twice-daily dosing regimens of these new macrolides may also improve patient compliance, a key factor in the management of any infection.
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REFERENCES