A comparison between the Strep A Rapid Test Device and conventional culture for the diagnosis of streptococcal pharyngitis

Kevin R Forward MD FRCP1,2, David Haldane MB FRCP1,2, Duncan Webster MD FRCP1, Carolyn Mills MLT1, Cheryl Brine MLT1, Diane Aylward MLT1

BACKGROUND: Rapid antigen detection tests are frequently used to diagnose pharyngitis due to Streptococcus pyogenes. Because a large number of kits are available commercially, performance characteristics may vary considerably. The present study evaluated one such kit currently in use in Canadian laboratories for which published evaluations are not available.

OBJECTIVE: To evaluate the performance characteristics of the Strep A Rapid Test Device (SARTD) (Nova Century Scientific Inc., Canada).

METHODS: Pharyngeal swabs from 818 patients with suspected streptococcal pharyngitis were tested. Swabs were initially inoculated onto the surface of a blood agar plate and then used to perform the rapid antigen test. The test was performed in accordance with the product monograph. Beta-haemolytic colonies were identified as S pyogenes using conventional means.

RESULTS: Four hundred ninety specimens were obtained from children and 328 from adults. S pyogenes was recovered from 171 (21%) patients. The SARTD detected S pyogenes antigens in 123 of 171 specimens from which S pyogenes was isolated on culture; the screen was negative in 610 of 647 specimens from which cultures were negative. The positive and negative predictive values of the SARTD were 76.9% and 92.7%, respectively.

CONCLUSIONS: The SARTD was much less sensitive (72%) than was suggested in the product monograph (90%). Laboratories should vigorously evaluate such products in-house, optimize specimen collection and transport, and choose more sensitive kits for use.

Key Words: Antigen detection; Pharyngitis; Streptococcus pyogenes

Streptococcal pharyngitis remains an important cause of morbidity and is one of the leading reasons for physician visits (1). Although there are clinical algorithms to assess the probability that pharyngitis is due to Streptococcus pyogenes, the diagnosis of streptococcal pharyngitis cannot be made on clinical grounds alone (2,3). Physicians are encouraged to submit throat swabs for antigen detection, culture or both (4,5). Same-day testing by antigen detection can be an important strategy to reduce unnecessary antibiotic use. In many settings, patient care may be improved, and fewer laboratory resources may be used when streptococcal antigen testing is performed at the point of care rather than sent to a clinical laboratory.

The performance characteristics of rapid antigen tests vary in both sensitivity and specificity (6-10). When tests are highly sensitive and the pretest probability is low, supplemental culture may not be necessary if the screening test is negative (4). False-positive tests may lead to unnecessary antibiotic use.

We compared the results of a commercially available screening device with conventional culture to evaluate its performance characteristics.
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TABLE 1

Performance characteristics of the Strep A Rapid Test Device* compared with conventional culture

<table>
<thead>
<tr>
<th></th>
<th>Culture-positive (%)</th>
<th>Culture-positive, screen-positive (n)</th>
<th>Culture-negative, screen-positive (n)</th>
<th>Culture-positive, screen-negative (n)</th>
<th>Culture-negative, screen-negative (n)</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>PPV (%)</th>
<th>NPV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Children (n=490)</td>
<td>24.1</td>
<td>80</td>
<td>23</td>
<td>38</td>
<td>349</td>
<td>67.8</td>
<td>93.8</td>
<td>77.7</td>
<td>90.2</td>
</tr>
<tr>
<td>Adults (n=328)</td>
<td>16.2</td>
<td>43</td>
<td>14</td>
<td>10</td>
<td>261</td>
<td>81.1</td>
<td>94.9</td>
<td>75.4</td>
<td>96.3</td>
</tr>
<tr>
<td>Overall (n=818)</td>
<td>21.0</td>
<td>123</td>
<td>37</td>
<td>48</td>
<td>610</td>
<td>71.9</td>
<td>94.3</td>
<td>76.9</td>
<td>92.7</td>
</tr>
</tbody>
</table>

*Nova Century Scientific Inc, Canada. NPV Negative predictive value; PPV Positive predictive value

METHODOLOGY

Study design

In February and March 2006, pharyngeal swabs from children (16 years of age or younger) and adults received at the Queen Elizabeth II Health Sciences Clinical Microbiology Laboratory (Halifax, Nova Scotia) were subjected to both conventional culture and the Strep A Rapid Test Device (SARTD) (distributed in Canada by Nova Century Scientific Inc, Canada), the latter being a chromatographic immunoassay used to detect group A streptococcus. Swabs were received in clear Amies Transport Medium (M40 Transystem, Copan Innovation, USA). The swab was used to inoculate a 5% sheep blood agar plate (Oxoid, Canada) and then screened with the study device.

Culture and identification

Plates were incubated anaerobically at 35°C (11) and examined at 24 h and 48 h; typical colonies that were catalase-negative and exhibited typical Gram stain morphology were tested for the presence of pyrrolidonyl peptidase by using the PYR test (Remel, USA) as per the manufacturer's instructions (12). The screening test and the colony work-up were performed by different technologists who had no knowledge of prior results.

Rapid antigen detection test

The SARTD was used according to the manufacturer's recommendations. Briefly, kits were stored at 4°C and brought to room temperature before testing. To extract the group A streptococcal carbohydrate antigen, five drops of reagent A and five drops of reagent B were added to the test kit chamber, and the swab was then inserted. The swab was agitated and left to sit for 1 min to 5 min, removed halfway, and rotated against the chamber ribs to release liquid into the chamber. The chamber valve was subsequently opened, allowing liquid to flow from the chamber along a test strip to an indicator coated with antibody to the group A streptococcal antigen. At 5 min, the strip was observed for the presence of a red line in the control and test regions. The appearance of two distinct red lines was interpreted as a positive result, while a single red line in the control region was considered to be a negative test result. If the positive control line did not appear, then the test was considered invalid.

Statistical analysis

Sensitivity, specificity and predictive values were calculated on the basis of the definition and formulas presented in the study by Lilstrup (13).

RESULTS

Four hundred ninety specimens originated from children and 328 from adults (Table 1). S pyogenes was recovered from 171 (21%) patients. The SARTD detected group A streptococcal antigens in 123 of 171 specimens from which S pyogenes was isolated on culture; the screen was negative in 610 of 647 specimens from which cultures were negative. The rapid screen was positive on 37 occasions when cultures were negative. Overall, the positive and negative predictive values of the SARTD were 76.9% and 92.7%, respectively (positive predictive values were 77.7% for children and 75.4% for adults; negative predictive values were 90.2% for children and 96.3% for adults). The test detected a higher proportion of adults with positive cultures than did conventional culture (81.1% versus 67.8%), but this did not reach statistical significance. There were no equivocal test results or instances where the control failed to turn positive.

DISCUSSION

A number of kits have been marketed for the detection of group A streptococcal antigens on throat swabs (6-10). Many of these products have not been subjected to peer-reviewed study. Typically, those in wider use have reported sensitivities of approximately 90% and specificities of approximately 95% (7). When tests are less sensitive or specific, the utility of antigen testing is reduced and their use is less easily justified. When rapid tests are highly sensitive, subsequent cultures may be unnecessary. As new tests are marketed, their performance characteristics should be carefully evaluated, especially when the results of the prior evaluations undertaken for licensing purposes are not published in peer-reviewed journals.

In all, we examined 818 specimens submitted in Amies Transport Medium without charcoal. Overall, the positivity rate was 22% in children and 15.6% in adults. These rates, particularly the adult positivity rates, were somewhat higher than we had anticipated. This may reflect the time of the year that the study was conducted (mid-winter). The SARTD detected S pyogenes antigens in 123 of 171 (71.9%) specimens. The test was more often correctly positive in adults than in children (81% versus 68%). It is possible that this may be due to a spectrum bias (ie, the proportion of adults with true pharyngitis, rather than colonization, was greater than that of children), although this seems unlikely. Perhaps adults were more effectively swabbed than children. The overall specificity when clear Amies Transport Medium was used was 94.9%. Because it can be assumed that all patients with a positive antigen test would receive an antibiotic, and because the predictive value of a positive antigen test was 78%, it means that a significant proportion of patients with positive antigen tests would receive unnecessary antibiotics.

The SARTD product monograph indicated an overall sensitivity of 90% and a specificity of 94%. We considered this test for use in our laboratory because it was of low complexity and was suitable for use with the Amies transport swabs that we use. It is unclear why, in our hands, the test sensitivity differed so markedly from that stated in the product monograph. In the
manufacturer's unpublished study, 758 swabs were examined by both culture and the SARTD. In all, they detected 240 of 265 swabs from which cultures grew \textit{S} \textit{pyogenes} (sensitivity 90\%). The SARTD was negative in 464 of 494 cases in which \textit{S} \textit{pyogenes} was not recovered from culture (specificity 94\%). The sensitivity of the SARTD was higher for adults than for children (81\% versus 68\%) and is in line with that stated in the product monograph. The manufacturer's evaluations were carried out in the same manner as ours, that is, with a single swab that was first rolled onto the agar plate and then subjected to antigen extraction. It is our understanding that this evaluation was performed on dry swabs, whereas we used Amies Transport Medium in the present study. The company advises that transport swabs containing modified Stuart's or Amies medium may also be used with this product. It is also suggested that specimens may be held for up to 8 h at room temperature or refrigerated for 72 h before testing. The testing we performed was, therefore, entirely in compliance with the product monograph.

We evaluated the SARTD in a manner that reflected our usual laboratory practice (ie, we used single swabs submitted by physicians in the Capital District Health Authority, Halifax, Nova Scotia). The vast majority of swabs are processed on the same day they are collected; those received in the late evening were refrigerated and tested the following morning. Swabs were first used to inoculate the surface of a blood agar plate. Studies have suggested that this has a negligible effect on the recovery of organisms on the swab, and this strategy has not been shown to significantly reduce the proportion of antigen tests that are positive (14,15). The test was simple to use (Clinical Laboratory Improvement Amendments waived) and was performed by laboratory technologists experienced with the use of antigen detection tests. Although the test sometimes produced faint bands, these were interpreted as being positive. We examined the possibility that our identification scheme for beta-hemolytic streptococci may have resulted in false-positive cultures. To exclude this possibility, we examined 25 consecutive beta-hemolytic streptococci identified as \textit{S} \textit{pyogenes} using Gram stain and pyrrolidonyl-beta-naphthylamide tests (data not shown). All were subsequently confirmed using a commercially available agglutination grouping kit (Prolex Streptococcal Grouping Latex Kit, Pro-Lab Diagnostics, Canada).

We used only a single blood agar plate and did not perform a broth enrichment step. It may therefore be argued that the true proportion of throat swabs containing \textit{S} \textit{pyogenes} may have been underestimated (10). However, it has also been argued that very small numbers of organisms that can be detected using a culture 'gold standard' may be more likely to represent colonization rather than acute infection.

Kits used for rapid streptococcal antigen testing need to have adequate sensitivity and specificity to inform patient management and to justify the additional expense. Only test kits with very high sensitivities in low prevalence populations (ie, adults) may have a negative predictive value that justifies antigen testing alone without backup culture of initial negative screens. In our setting and using our manner of testing, we did not feel that this kit was adequately sensitive to fulfill that requirement. Numerous streptococcal antigen kits are commercially available, many in the absence of published evaluations of their performance. In many laboratories, the procedures used may differ from the way in which manufacturer-sponsored evaluations were conducted. Laboratories introducing new kits should perform thorough in-house evaluations before placing them into routine use.

ACKNOWLEDGEMENTS: The authors acknowledge the contribution of Dorothy Gillie, who prepared the manuscript.

**REFERENCES**
