CASE REPORT

Persistent urethritis and prostatitis due to *Trichomonas vaginalis*: A case report

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The present report describes a case of persistent urethritis accompanied by prostatitis due to *Trichomonas vaginalis* in a young male patient. The importance of the laboratory diagnosis of trichomoniasis in persistent or recurrent urethritis (ie, testing samples from multiple sites) is highlighted, with the aim of improving the clinical recognition of this pathogen.

Key Words: Persistent urethritis; Prostatitis; *Trichomonas vaginalis*

Trichomoniasis is highly prevalent worldwide and is associated with urethritis (1-3) and prostatitis (4,5) in men. Moreover, infection with *Trichomonas vaginalis* (TV) may increase the risk of HIV acquisition; some studies have associated TV infection with elevated seminal HIV RNA levels in men with HIV infection and symptomatic urethritis (2,6,7). However, the importance of trichomoniasis in men is poorly understood, in part, due to difficulties in the diagnosis of the infection.

Detection of TV in men has traditionally been based on wet mount microscopy or culture of a single specimen, through either a urethral swab or a urine sediment. Although microscopic examination of the saline wet mount or culture has poor sensitivity (ranging from 60% to 90%), they are still used as routine diagnostic tests for TV in many laboratories (3). A case of persistent urethritis with prostatitis due to TV, which was diagnosed by direct microscopy and cultivation of samples from multiple sites, is reported.

CASE PRESENTATION

A 26-year-old heterosexual man presented with a six-month history of persistent urethritis. He complained of urethral discharge, dysuria and a recent onset of perineal and ejaculatory pain. He had already been treated with multiple antibiotics, including ciprofloxacin, doxycycline, tetracycline and erythromycin, during his four previous visits to different clinicians. He had not received any metronidazole due to all previous negative investigations for TV on urethral and urine samples. He denied any extramarital sexual partners since his marriage one month ago, but acknowledged three casual female partners before that. He was using condoms with his wife for contraception. No information was available concerning clinical or laboratory follow-ups on any of his female contacts, including his wife.

A physical examination revealed scanty mucoid urethral discharge. There was no evidence of inguinal adenopathy, genital lesions or intrascrotal abnormalities. Three urethral specimens were obtained using calcium alginate swabs. The first swab was used to make a Gram stain preparation and to inoculate Chocolate agar and New York City medium for the isolation of *Neisseria gonorrhoeae*. The second swab was used for detection of *Chlamydia trachomatis* by direct immunofluorescence assay. Wet mount microscopy and inoculation of modified Diamond's medium were performed on the third urethral swab to detect TV.

After the physical examination, the patient was requested to provide 10 mL to 20 mL of first-voided and midstream urine as part of the four-glass localization test previously described by Meares and Stamey (8). The patient then underwent prostatic massage and his expressed prostatic secretions were collected. Immediately after his prostatic massage, the patient collected 10 mL to 15 mL of urine. For all urine samples, 10 mL of urine was centrifuged at 1500 g for 10 min and 50 μL of urine sediment was used to inoculate the TV culture bottles. Mycoplasma and ureaplasma culture media were inoculated using 50 μL of first-voided urine sediment separately. Finally, the patient was asked to provide a semen specimen for TV culture. The semen was allowed to liquefy at room temperature for approximately 1 h before processing. The liquefied semen was centrifuged at 2000 g for 10 min, and approximately 50 μL of semen was used to
inoculate the last TV culture bottle as previously described by Kaydos-Danniels et al (9).

The Gram-stained urethral exudate showed more than 15 polymorphonuclear leukocytes (PMNLs) per oil immersion field and there was no evidence of intracellular Gram-negative diplococci. The direct immunofluorescence test for C trachomatis was negative. After 48 h of incubation at 37°C in 5% atmospheric CO2, no N gonorrhoeae strains were isolated. Cultures for Mycoplasma genitalium and Ureaplasma urealyticum were negative after five days. Direct examination of expressed prostatic secretions revealed 10 PMNLs to 15 PMNLs per high power field (original magnification ×400) with few clumps, indicating prostate infection and inflammation. Wet-mount microscopy of all collected samples was negative for mobile trichomonal trophozoites, except for urine sediment after prostate massage. The first two Diamond’s bottles that were inoculated by urethral swab and first-catch urine sediment, respectively, yielded negative results. TV trophozoites were observed in the third, fourth and fifth bottles, which were inoculated by prostate secretions, postprostate massage urine sediment and semen sample, respectively. All TV cultures were checked on days 2, 5 and 7 after inoculation. Interestingly, all culture bottles were negative until day 5, when the first positive culture was found. Table 1 compares the results of direct microscopic examination of wet mounts and cultures based on different sampling sites.

The patient was diagnosed with TV infection and treated with 500 mg metronidazole twice daily for seven days. He returned to the clinic for a follow-up three weeks after his first visit. The urethral discharge disappeared by day 3 of treatment and he acknowledged that his dysuria, perineal and ejaculatory pain had all completely resolved. Although follow-up cultures were suggested for test of cure, the patient rejected any additional investigations and did not return for further medical care. We did not have the opportunity to establish any TV investigations on vaginal samples from the patient’s wife or his previous sexual partners.

DISCUSSION

Persistent or recurrent urethritis develops in a small proportion of men who suffer from nongonococcal urethritis (NGU); TV probably accounts for only a minority of these cases (9,10). Clinical and laboratory diagnosis of trichomaniasis is more complicated in male patients (11). Although urethral swabs are usually considered to be the best specimen to diagnose TV infections in men, addition of other samples, such as urine or semen, increases the sensitivity of results. Addition of metronidazole to the syndromic management of male urethritis, in places where trichomoniasis is common, can not only eliminate the infection but may also help to reduce the transmission of HIV.

There is no consensus of opinion in the management of prostatitis caused by TV. In one study (16), two weeks of metronidazole (250 mg twice daily) was used to treat chronic TV prostatitis. Skerk et al (17) demonstrated that TV eradication and clinical cure in patients with chronic prostatitis did not significantly differ with regard to the administered dose of metronidazole (three doses of 500 mg, for either seven or 14 days). We assumed that in our patient, the concurrent treatment of his partner(s) was unlikely. Our patient was treated with 500 mg metronidazole twice daily for seven days because there is some evidence to suggest that the failure rate of a single dose is higher if partners are not treated simultaneously (18). Moreover, due to the recent onset of prostatitis in our patient, we did not put him under a ‘chronic TV prostatitis’ category and, therefore, neither a higher dose of metronidazole nor prolonged regimens were prescribed for him.

Young men with persistent or recurrent NGU apparently represent a high-risk group for TV infection. These patients are subjected to repeated diagnostic evaluation and, although

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**TABLE 1**

Comparison of the results of wet-mount microscopy, culture and the number of polymorphonuclear leukocytes (PMNLs) from different sample sources

<table>
<thead>
<tr>
<th>Test results</th>
<th>Urethral swab</th>
<th>First-catch urine</th>
<th>Mid-stream urine</th>
<th>Expresssed prostate secretions</th>
<th>Urine after prostate massage</th>
<th>Semen</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wet mount microscopy</td>
<td>–ve</td>
<td>–ve</td>
<td>NA</td>
<td>–ve</td>
<td>–ve</td>
<td>–ve</td>
</tr>
<tr>
<td>Culture (PMNLs/hpf)</td>
<td>&gt;20</td>
<td>15–20</td>
<td>1–3</td>
<td>10–15</td>
<td>15–20</td>
<td>1–3</td>
</tr>
</tbody>
</table>

*The midstream urine sample was only tested for the presence of PMNLs to exclude the possibility of utrinary tract infection. The number of PMNLs/high power field (hpf) (original magnification ×400) was counted from the sediments of first-catch, midstream and postprostate massage urine samples, while urethral swab expressed prostate secretions and semen samples were examined directly without concentration. –ve Negative; +ve Positive; NA Not applicable.
no infectious etiology has been found, they are often empirically treated with multiple courses of antibiotics. The present case highlights the importance and necessity of performing accurate diagnostic procedures in the case of persistent or recurrent urethritis. Collection of multiple specimens from different sites may increase the sensitivity of testing for TV in male patients with negative results by routine investigative methods.

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