Point prevalence study of antibiotic susceptibility of genital group B streptococcus isolated from near-term pregnant women in Calgary, Alberta

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BACKGROUND: Genital group B streptococcus (GBS) may be transmitted from a colonized mother to her infant if appropriate intrapartum antibiotic prophylaxis is not given. A recent case of GBS neonatal sepsis occurred due to an erythromycin-intermediate strain after inappropo""""
TABLE 1
Genital group B streptococcus antibiogram profile for Calgary, Alberta (2011)

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>% Susceptible</th>
<th>% Resistant</th>
<th>Resistance Mechanism</th>
<th>Total, %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Penicillin</td>
<td>100</td>
<td>None</td>
<td>N/A</td>
<td>N/A</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>75</td>
<td>25</td>
<td>Efflux</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>MLSB$_{R}^{-1}$</td>
<td>8.1</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>MLSB$_{C}$</td>
<td>14.2</td>
</tr>
<tr>
<td>Inducible ribosomal methylation (Clindamycin)</td>
<td>78</td>
<td>22</td>
<td>MLSB$_{R}^{-1}$</td>
<td>8.1</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>MLSB$_{C}$</td>
<td>14.2</td>
</tr>
</tbody>
</table>

*Based on the phenotypic antibiotic susceptibility profile. Efflux msrA (erythromycin resistant [R], clindamycin sensitive [D-test negative]); MLSB$_{R}^{-1}$ (Ribosomal methylation constitutive) (erythromycin R, clindamycin R); MLSB$_{C}$ (Ribosomal methylation inducible) (erythromycin R, clindamycin R [D-test positive]); N/A Not applicable

Our clinical microbiology laboratory performs routine isolation of GBS from vaginal/rectal swabs, but antibiotic susceptibility testing is not routinely performed unless the physician indicates that the patient has a penicillin allergy or specifically requests this testing be performed. This practice is consistent with current guidelines for antibiotic susceptibility testing of genital GBS isolates that is followed by clinical microbiology laboratories in Canada and the United States (13), based on the premise that genital GBS remain almost universally susceptible to penicillin and other β-lactam drugs that are prescribed to women without allergy. Canadian guidelines have not been recently updated and continue to recommend the use of erythromycin and clindamycin as alternative antibiotics in women colonized with genital GBS who have a penicillin allergy (12).

However, our experience with a recent case of early onset neonatal GBS bacteremia, and the subsequent results of a regional point prevalence study of genital GBS isolate antibiotic susceptibility profiles, highlights the need for changes to laboratory and clinical practice in this regard.

CASE PRESENTATION
A near-term pregnant woman had a vaginal/rectal swab collected for GBS culture. The patient was confirmed to be colonized with GBS, but antibiotic susceptibility testing was not performed because the physician had not made this request nor was there an indication of penicillin allergy on the laboratory requisition. A second vaginal/rectal swab was collected two weeks later, and the physician requested that antibiotic susceptibility testing be performed due to the patient’s penicillin allergy. The patient went into labour and delivered before the testing was complete. Following the Canadian guideline for penicillin-allergic patients (12), she was prescribed prophylaxis with intravenous erythromycin in the recommended dose of 500 mg. The infant developed GBS bacteremia three days after delivery. Both the maternal genital and neonatal blood GBS isolates had the same antibiogram profile, with a penicillin Errest (bioMérieux, France) minimum inhibition concentration of 0.047 μg/mL, which is sensitive, a Kirby-Bauer zone size of 25 mm for clindamycin, which is sensitive, but a zone size of 17 mm for erythromycin, which is intermediate. Both the maternal genital and neonatal blood GBS isolates did not produce a D-zone, indicating that these strains had a phenotype consistent with an efflux mechanism of resistance (ie, erythromycin [intermediate or resistant], clindamycin [sensitive] and a negative D-zone). Subsequently, a clinical and laboratory incident review of this case was performed to implement measures that would prevent a repeat of this serious occurrence.

METHODS
Laboratory confirmation of GBS colonization requires collection of a vaginal/rectal swab. Calgary Laboratory Services performs approximately 15,000 vaginal/rectal swab cultures per year using culture-based detection for genital group B streptococcus that includes the use of a selective broth as previously described (14). Vaginal/rectal swabs are physician collected using Copan swabs (Copan Italia SPA, Italy) in liquid Amies transport tubes, and transported within 4 h to 6 h to the laboratory. Swabs are inoculated into StrepB Carrot Broth (SCB, Hardy Diagnostics Inc, USA) on receipt and incubated for 18 h to 24 h at 35°C. All SCB cultures were read as positive if a visible colour change, from colourless to orange or red, occurred. Samples negative by SCB culture were subsequently subcultured onto 5% sheep blood agar, incubated for an additional 18 h to 24 h at 35°C and analyzed for the presence of GBS using standard biochemical methods. Genital GBS isolates recovered from pregnant women with a reported penicillin allergy are routinely tested for antibiotic susceptibility to penicillin by Errest, and the Kirby-Bauer disk method for clindamycin and erythromycin. All isolates that demonstrate intermediate or full resistance to erythromycin have a D-test performed to determine inducible resistance to clindamycin (13,15).

Antibiotic susceptibility testing was performed on a total of 309 genital GBS isolates that were prospectively isolated and randomly selected from vaginal/rectal swabs collected between March and May 2011 to determine the regional antibiogram profile. The sample size (ie, 300 isolates tested) was based on the total number of genital GBS isolates per year (2250 [15%]) (ie, recovered from approximately 15,000 vaginal/rectal swabs), a margin of error of 5% and a CI of 95%. Errest strips were used for the surveillance study to determine the minimum inhibitory concentrations to penicillin, erythromycin and clindamycin according to standard methods (13). All isolates that demonstrated either intermediate or full resistance to erythromycin had a D-test performed to determine inducible resistance to clindamycin (16). The resistance mechanism was inferred from the isolate’s individual antibiogram profile. Molecular testing to detect resistance genes was not performed.

RESULTS
The overall antibiogram pattern derived from our point prevalence study is outlined in Table 1. Although all genital GBS isolates were susceptible to penicillin, there were high rates of resistance to both erythromycin (25%) and clindamycin (22%). Most genital GBS isolates that were resistant to erythromycin and clindamycin had a resistance phenotype (MLSB$_{R}^{-1}$; 8.1%) (Table 1) (16). Erythromycin resistance due to an efflux mechanism (msrA) (3%) was much less common.

DISCUSSION
The present study is the first point prevalence study of genital GBS antibiogram profile to be reported from a Canadian health care region. Our data highlight the high rates of resistance to erythromycin (25%) and clindamycin (22%) of genital GBS isolates from near-term pregnant women, as has been previously reported recently in other jurisdictions (17-24). In these studies, genital GBS resistance rates for erythromycin ranged from 15% to 54% of the strains tested, and slightly lower rates were found for clindamycin resistance. Our results confirm the increasing resistance to both erythromycin and clindamycin worldwide, mainly due to the acquisition and spread of erm genes conveying the MLSB$_{R}^{-1}$ phenotype (16). Most commonly, erm genes are constitutively expressed rather than being inducible (positive D-test for clindamycin), similar to the resistance phenotypes recently found in our region (15,16). Erythromycin resistance due to an efflux mechanism (msrA) has been reported much less commonly among genital GBS isolates (16).

Our study was prompted by a recent incident of intrapartum prophylaxis failure of erythromycin in a case where both the maternal genital and neonatal blood culture GBS isolates were not susceptible to this drug. Although inadequate prophylaxis may have contributed to the subsequent development of early onset neonatal sepsis in this case, future changes to laboratory and clinical practice in this regard are necessary, as well as continued surveillance for resistance patterns among genital GBS isolates in our region.


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case, inadequate maternal peak serum and fetal tissue levels of erythromycin may have also been a factor. The mean transplacental transfer of macrolide antibiotics, including erythromycin, has been studied by perfusing term placental tissue after delivery (25). In this study of 21 placentas, erythromycin (2 µg/mL) was given as a 2 h, nonrecirculating perfusion of a single placental cotyledon, and drug concentrations were measured by high-performance liquid chromatography. The mean transplacental transfer of erythromycin was only 3% when calculated as the ratio between the steady state concentrations in fetal venous and maternal arterial sides, but similar results were obtained for calculations of the absolute amount of drug transferred across the placenta during the 2 h perfusion (25). Although this was a small study, it suggests that the placenta is an effective barrier that limits transplacental transfer of macrolides, which may compromise the efficacy of intrapartum prophylaxis with this compound. No clinical trials of intrapartum prophylaxis have been performed using either erythromycin or clindamycin as alternate agents when a β-lactam cannot be used.

The clinical significance of increasing rates of genital erythromycin-resistant GBS isolates is also shown by more recent assessments of the rate of early onset neonatal sepsis. Chen et al (26) performed a time-trend study at a tertiary care centre in the United States over several periods of time including no protocol for group B streptococcus prophylaxis (1990 to 1992), risk-based protocol (1993 to 1996) and a screening-based protocol (1997 to 2002). A total of 120,952 neonates were born during this time with 118 cases of GBS early onset neonatal sepsis. Ninety percent of the GBS isolates were resistant to ampicillin, penicillin, cefazolin or vancomycin. Although the rate of GBS early-onset neonatal sepsis decreased significantly across these periods (from 2.0 to 1.1 to 0.4 per 1000 live births, respectively; P<0.0001), and the overall rate of erythromycin-resistant GBS neonatal sepsis did not change (from 0.14 to 0.03 to 0.08 per 1000 live births, respectively; P=0.60), an increasing proportion of cases were due to erythromycin-resistant GBS strains (from 7% to 2.6% to 23.8%, respectively; P=0.07) (26).

The high rates of genital GBS resistance to both erythromycin and clindamycin in our region makes it necessary to modify both current clinical and laboratory practice. Routine antibiotic susceptibility testing must be performed for all GBS isolates recovered from near-term pregnant women who have a reported or documented history of penicillin allergy. Currently, the onus is on the physician to provide the laboratory with a clear clinical history of penicillin allergy for antibiotic susceptibility testing to be performed. However, even with a well-designed laboratory requisition, laboratory orders will sometimes be missed (27). Online order entry with a requirement for penicillin allergy history or a modified laboratory requisition (eg, checkbox should be included) could improve physician ordering and minimize laboratory errors.

The safest way to ensure availability of appropriate antibiotic susceptibility at delivery would be to institute routine antibiotic testing on all genital GBS isolates. Cost estimates, however, indicate this would add $16.58 per specimen (approximately $81,476.84 per year in our laboratory), mainly due to an increase of a 0.72 full-time equivalent of additional medical laboratory assistant/technologist labour.

Canadian guidelines for the prescription of alternative antibiotic agents for women with penicillin allergy also need to be updated to align with those recently published by the Centers for Disease Control (Atlanta, Georgia, USA) in the United States (1). Penicillin or an equivalent β-lactam antibiotic (ie, cefazolin, third generation cephalosporin) should remain the drug of choice for prophylaxis of women who do not have a drug allergy. Erythromycin should no longer be used as an alternative antimicrobial agent for intrapartum prophylaxis of pregnant women colonized with GBS, as highlighted by our case. Clindamycin may still be used if the isolate has been confirmed to be susceptible to this agent. However, if the isolate is clindamycin resistant or antibiotic susceptibility testing has not been performed before delivery, then vancomycin should be used instead as the primary alternative agent of choice in penicillin-allergic women. Although vancomycin requires prolonged infusion (2 h), genital GBS remain universally susceptible, and the drug's transplacental passage occurs in a predictable manner in concentrations that exceed the usual GBS breakpoint provided an appropriate dose is given based on body weight and renal function (28,29).

A multicentre prevalence study of genital GBS isolate antibiotic susceptibility profiles also needs to be performed to confirm the levels of resistance to erythromycin and clindamycin in other Canadian health care regions. The derived antibiogram profile for genital GBS isolates should then be used to publish new Canadian guidelines for the prevention of neonatal GBS infection that discourage the empirical use of either erythromycin or clindamycin in near-term pregnant women with laboratory-confirmed genital GBS colonization.

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