

Research Article

Diagnostic Value of Combined Intravoxel Incoherent Motion Diffusion-Weighted Magnetic Resonance Imaging with Diffusion Tensor Imaging in Predicting Parametrial Infiltration in Cervical Cancer

Ting-Ting Lin ^{1,2}, Xin-Xiang Li ², Wei-Fu Lv ³, Jiang-Ning Dong ², Chao Wei ²,
Ting-Ting Wang ², Chuan-Bin Wang ², and Ping Zhang ²

¹Anhui Provincial Hospital, Cheeloo College of Medicine, Shandong University, Jinan, Shandong 250012, China

²Department of Radiology, The First Affiliated Hospital of USTC, Division of Life Sciences and Medicine, University of Science and Technology of China, Hefei, Anhui 230031, China

³Anhui Provincial Hospital, Department of Radiology, The First Affiliated Hospital of USTC, Division of Life Sciences and Medicine, University of Science and Technology of China, Hefei, Anhui 230001, China

Correspondence should be addressed to Wei-Fu Lv; lwf09@163.com and Jiang-Ning Dong; dongjn@163.com

Received 8 December 2020; Revised 19 April 2021; Accepted 22 April 2021; Published 11 May 2021

Academic Editor: Guillermina Ferro Flores

Copyright © 2021 Ting-Ting Lin et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Objective. This study sought to determine the diagnostic value of combined intravoxel incoherent motion (IVIM) diffusion-weighted magnetic resonance imaging (MRI) with diffusion tensor imaging (DTI) in predicting parametrial infiltration (PMI) in patients with cervical cancer. **Materials and Methods.** We enrolled 65 patients with cervical cancer confirmed by radical hysterectomy (25 PMI-negative and 40 PMI-positive) who underwent IVIM and DTI pretreatment. The parameters of IVIM (ADC, D, D*, and f) and DTI (average diffusion coefficient (DCavg) and fractional anisotropy (FA)) were recorded by two observers. All parameter differences were tested, and the receiver operating characteristic (ROC) curves were generated to estimate the diagnostic performance of significant metrics and their combinations. **Results.** Compared to the PMI-negative group, the PMI-positive group had significantly lower D (0.632 ± 0.017 vs. 0.773 ± 0.024 , $p < 0.001$) and lower FA (0.073 ± 0.002 vs. 0.085 ± 0.003 , $p = 0.003$). The area under the ROC curve (AUC) of D and FA was 0.801 and 0.726, respectively, and the combination of D and FA improved the AUC to 0.931, with a sensitivity and specificity of 80.0% and 97.5%, respectively. **Conclusion.** D and FA values could be used to help diagnose PMI in patients with cervical cancer. The combination of IVIM and DTI was more valuable than either option alone.

1. Introduction

Cervical cancer is the third most prevalent type of malignant tumor in females worldwide. Parametrial infiltration (PMI) is a key predictor of a poor prognosis and is associated with increased rates of tumor metastasis, tumor recurrence, and mortality [1–4]. Accurate pretreatment evaluation of PMI is needed to determine the International Federation of Gynecology and Obstetrics (FIGO) stage of the cervical cancer and select individualized treatment strategies [5]. Magnetic resonance imaging (MRI), as an accurate and noninvasive

pretreatment method, has immense clinical value in evaluating PMI.

MRI techniques have been used in several studies to assess PMI in patients with cervical cancer. When considering tumor volume combined with findings that are suggestive of PMI, preoperative MRI is useful in ruling out PMI diagnosis [6]. As shown by Park et al., the apparent diffusion coefficient (ADC) of cervical cancer is lower in PMI-positive than PMI-negative cases [4, 7]. Woo et al. showed that 3.0 T MRI scanners and diffusion-weighted imaging (DWI) can be used to improve the diagnostic efficiency of detecting

cervical cancer with PMI [8]. Intravoxel incoherent motion (IVIM) is the stretched modality of conventional DWI sequence based on a bi-exponential model and the derived parameters include standard ADC (ADC), slow ADC (D), fast ADC (D*), and perfusion fraction (f). The ADC value quantifies the pure water molecular diffusion (represented by D) and microcirculation perfusion (represented by D*). All parameters can be measured simultaneously, which can better reveal the microstructure of tumors and biological issues and has been used for diagnosing, differentiating pathological subtype and histological grade, and evaluating treatment efficacy of cervical cancer [9–11].

In addition, diffusion tensor imaging (DTI) is widely used in the study of uterine diseases. In 2006, Weiss et al. investigated the use of 3-dimensional (3D) imaging of normal fiber architecture of an *ex vivo* human uterus by DTI [12]. In another *ex vivo* study, Toba et al. found that DTI is a beneficial and complementary technique in detecting myometrial invasion in patients with endometrial cancer [13]. In an *in vivo* investigation, He et al. showed that the dynamic changes of fractional anisotropy (FA) are correlated with menstrual cycles [14]. Another *in vivo* study by Fujimoto et al. compared DTI parameter variations in different uterine layers during the luteal phase of menstrual cycles [15]. DCavg and FA are the main quantitative parameters obtained from DTI data [16]. DCavg value reflected water diffusion in all directions and FA value represented the proportion of the anisotropic components of the water molecule to the overall diffusion tensor. The quantitative parameters have been shown to be useful in detecting, diagnosing, and predicting pathological grade of tumors [17, 18].

To the best of our knowledge, the method of combining IVIM and DTI to detect PMI in cervical cancer cases has not been published previously. The purpose of the current study was, therefore, to investigate the diagnostic value of the combination of IVIM and DTI for predicting PMI in cervical cancer.

2. Materials and Methods

2.1. Patients. This study was supported by our institutional review board, and all patients provided written informed consent to participate. Between February 2019 and April 2020, information from 90 consecutive patients with clinically suspected or pathologically proven cervical cancer was retrieved from our hospital. This study's inclusion criteria were as follows: (1) patients had cervical cancer diagnosed by pathology, (2) patients had not undergone biopsy, surgery, or other treatment, before MRI evaluation, (3) conventional MRI was performed with uniform IVIM, with DTI protocols performed simultaneously before treatment, and (4) the image quality was adequate for subsequent analysis (no obvious artifacts and serious imaging distortion; the maximum diameter of tumor was more than 1 cm and displayed clearly). The exclusion criteria were as follows: (1) the lesion diameter was too small to measure (<1 cm), (2) the MRI scan sequences were incomplete, or (3) the image quality could not meet the needs of diagnosis and measurement (imaging with obvious motion artifacts or distortion).

2.2. MRI Evaluation. MRI data were acquired using a 3.0T scanner (Signa HDxt; GE Healthcare, Milwaukee, Wisconsin, USA) with an 8-channel phased array body coil. All patients received an intramuscular injection of 15 mg of scopolamine butyl bromide 30 minutes before the examination to reduce artifacts caused by intestinal movement. Pelvic imaging included routine sequences (axial T1-weighted imaging (T1WI), axial/sagittal T2-weighted imaging (T2WI), and axial T2WI with fat suppression), functional imaging (IVIM and DTI), and contrast-enhanced (CE) protocols. CE-MRI was performed after an intravenous injection of gadodiamide (0.1 mmol/kg, 3 ml/s; Omniscan, GE Healthcare, Cork, Ireland). All scanning processes were performed with free breathing.

IVIM with $10b$ values (0, 10, 20, 50, 100, 200, 400, 800, 1000, and 1200 s/mm²) was obtained with single-shot spin-echo echo planar imaging (repetition time (TR) = 4000 ms, echo time (TE) = 85 ms, slice thickness = 4 mm, number of excitations (NEX) = 2, matrix size = 130 × 128, and field of view (FOV) = 40 × 22 cm). The DTI parameters were TR = 7000 ms, TE = 89.6 ms, slice thickness = 4 mm, diffusion direction = 15, number of excitations (NEX) = 1, matrix size = 96 × 130, and FOV = 32 cm. The acquisition time for IVIM and DTI was 5 min, 44 s, and 1 min, 59 s, respectively.

2.3. Radiological Evaluation. The original IVIM and DTI data were transferred to the Advantage Windows 4.5 workstation and processed using the in-house Function Tool (GE Healthcare, Milwaukee, Wisconsin, USA). The signal-to-noise ratio (SNR) of the images was evaluated before analysis. The MADC or Diffusion Tensor interface was chosen for IVIM or DTI data set respectively after entered the Function Tool software. Subsequently, the IVIM parameters (ADC, D, D*, and f) and DTI metric maps (DCavg and FA) were generated automatically. All data were evaluated by 2 radiologists with 21 (observer 1) and 15 (observer 2) years of experience in gynecological imaging. The two radiologists were blinded to measure with each other's findings. The IVIM dataset was processed using the bi-exponential model fitting formula [19]: $Sb/S0 = (1 - f) * \exp(-b * D) + f * \exp[b * (D * + D)]$, where b was the b -value represents the diffusion sensitizing factors, D indicated the slow component of diffusion, D* stood for the incoherent microcirculation, f was the fractional perfusion linked to the intravascular component, Sb reflects the mean signal intensity with the diffusion gradient b , and $S0$ reflects a 0 of b value. Referencing the T2WI and CE T1WI images, the maximum solid area of the tumor with the most restricted diffusion and the most obvious display of IVIM images and original DTI images was outlined as the regions of interest (ROIs). ROIs were drawn manually as large as possible along the tumor margin of the tumor's largest area. Macroscopic cystic degeneration, necrotic regions, hemorrhage, surrounding vessels, and artifacts induced by air-water interface were avoided during the tumor contour. Each tumor was measured 3 times in the selected ROI regions on IVIM and DTI parametric maps. Measurements were performed by observer 1 twice, the interval between the 2 measurements

being at least 1 week. The third measurement was performed by observer 2. The arithmetic mean of the 3 measurements by the 2 observers was recorded as the final result and was used for the following statistical analysis.

2.4. Statistical Analysis. Statistical analyses were conducted with SPSS software version 23.0 (Chicago, IL, USA) and GraphPad Prism 8.0 (San Diego, CA, USA). A p value <0.05 was considered to indicate statistical significance. An intraclass correlation coefficient (ICC) was calculated to test intraobserver (using the 2 results obtained by observer 1) and interobserver (using the first result from observer 1 and the single result by observer 2) reproducibility (an ICC value >0.8 representing an almost perfect agreement). Quantitative data were described as mean \pm standard deviation (SD). A one-way analysis of variance was used to estimate the data with a normal distribution; the comparison of parameters between two groups was performed using least significant difference (LSD) test. Mann–Whitney test in the nonparametric test was used to compare the data with a nonnormal distribution. Receiver operating characteristic (ROC) curves were calculated to evaluate the diagnostic performance of the significant parameters and their combinations. The optimal cut-off value of each parameter was determined by the maximum of Youden index (the maximum of Youden index = sensitivity + specificity – 1).

3. Results

3.1. Patients. A total of 65 patients between the ages of 27 and 88 years (median age: 53.0 years) with pathologically confirmed cervical cancer were enrolled in the study. The staging was made according to the FIGO staging criteria of 2018 [20]. The final diagnosis revealed that 40/65 (61.5%) of the patients had low-FIGO-stage cervical cancer (IB to IIA) without PMI and 25/65 (38.5%) had high-FIGO-stage cervical cancer (IIB to IVA) with PMI. The clinical 2018 FIGO stage breakdown was IB, $n = 27$; IIA, $n = 13$; IIB, $n = 11$; IIIA, $n = 7$; IIIB, $n = 4$; and IIIC, $n = 3$.

3.2. Reproducibility of IVIM and DTI Measurements. The intraobserver ICC, obtained using the 2 measurements taken by observer 1, ranged from 0.867 to 0.925 (Table 1). The interobserver ICC, obtained using observer 1's first result and observer 2's single result, ranged from 0.836 to 0.934 (Table 1). Both the intra- and interobserver assessments indicated excellent reproducibility.

3.3. Diagnostic Efficiency of IVIM Parameters, DTI Metrics, and Their Combination. IVIM-, DWI-, and DTI-derived parameters between the PMI-positive and PMI-negative groups were compared and are summarized in Table 1 and Figures 1 and 2. Compared to the PMI-negative group, the PMI-positive group demonstrated significantly lower D (0.632 ± 0.017 vs. 0.773 ± 0.024 , $p < 0.001$) and lower FA (0.073 ± 0.002 vs. 0.085 ± 0.003 , $p < 0.05$); however, there was no significant difference for ADC , D^* , f , and $DCavg$.

ROC analyses revealed that setting the cut-off value of D to $0.741 \times 10^{-3} \text{ mm}^2/\text{s}$ resulted in the diagnostic efficiency for differentiating between PMI-positive and PMI-negative lesions, with a sensitivity of 60.0% and a specificity of 85.0% ($AUC = 0.801$) (Table 2). When setting the cut-off value of FA to 0.078, differentiation between PMI-positive and PMI-negative lesions achieved a sensitivity of 72.0% and a specificity of 70.0% ($AUC = 0.726$). The combination of D and FA improved the diagnostic performance obviously, with the highest AUC (0.931), sensitivity (80.0%), and specificity (97.5%) (Figure 3). Representative IVIM-DWI and DTI images of the patient are shown in Figure 4.

4. Discussion

This study investigated IVIM and DTI in cervical cancer with PMI in 65 patients. To the best of our knowledge, this is the first study to use these technologies to evaluate PMI in patients with cervical cancer. Moreover, we found the D and FA value showed a significant difference between the PMI-positive and PMI-negative groups, especially the combination of D and FA improving the diagnosis of PMI in cervical cancer.

IVIM-DWI with multiple b values using a double exponential model reflects the microcirculatory and diffusivity information of tumors [21, 22]. There were no differences between the PMI-positive and PMI-negative groups in terms of ADC , D^* , and f values in this study, which is reflective of differences in tumor heterogeneity, tissue cellularity density, and tissue perfusion. The ADC value quantifies tissue water molecule diffusion and reflects the capillary microcirculation perfusion [23]. The D^* value represents the microcirculation perfusion and tissue microstructure in cervical cancers, and the f value represents the volume ratio of the local microcirculation perfusion and the overall effect of diffusion [11, 24]. The lack of significant differences between the PMI-positive and PMI-negative groups indicate that they were the similar in terms of water molecule diffusion and angiogenesis. In our study, there was a significant difference in the D value of the PMI-positive and PMI-negative groups. Perucho et al. [25] showed that high FIGO stages had higher D^* values than low stages, but D values had no significance in distinguishing between high and low stages. The D value, excluding the low b value perfusion, represents pure water molecule diffusion of the tumor. The diffusion restriction of the tumor is related to cell gap. In this study, the D value of the PMI-positive group was smaller than that of the PMI-negative group, which means that the tumor cell gap in the PMI-positive group was smaller than that in the PMI-negative group. This feature reflects that the tumor cells with indefinite direction growth in the PMI-positive group were more obvious than those of the PMI-negative group, which indirectly reflects the higher degree of malignancy of the PMI-positive group.

$DCavg$ values are affected by cell density, cell organization, and microcirculation [15, 26]. In our study, there was no significant difference between the PMI-positive and PMI-negative groups, indicating that the $DCavg$ value could not be used to diagnose PMI in cervical cancer cases. The FA

TABLE 1: Comparison of IVIM-, MRI-, and DTI-derived parameters between the PMI-positive and PMI-negative groups.

Parameters	PMI-negative ($n = 40$)	PMI-positive ($n = 25$)	P value	ICC	
				Inter (95% CI)	Intra (95% CI)
IVIM-MRI					
ADC	0.918 ± 0.045	0.904 ± 0.016	0.732	0.932 (0.865, 0.973)	0.867 (0.789, 0.943)
D	0.773 ± 0.024	0.632 ± 0.017	< 0.001	0.934 (0.869, 0.958)	0.902 (0.833, 0.972)
D^*	46.22 ± 13.46	33.90 ± 3.63	0.290	0.865 (0.796, 0.924)	0.906 (0.858, 0.965)
f	0.273 ± 0.028	0.225 ± 0.076	0.080	0.932 (0.862, 0.974)	0.867 (0.784, 0.921)
DTI					
DCavg	1.080 ± 0.058	1.062 ± 0.033	0.765	0.925 (0.834, 0.946)	0.912 (0.872, 0.978)
FA	0.085 ± 0.003	0.073 ± 0.002	0.003	0.836 (0.778, 0.895)	0.925 (0.876, 0.954)

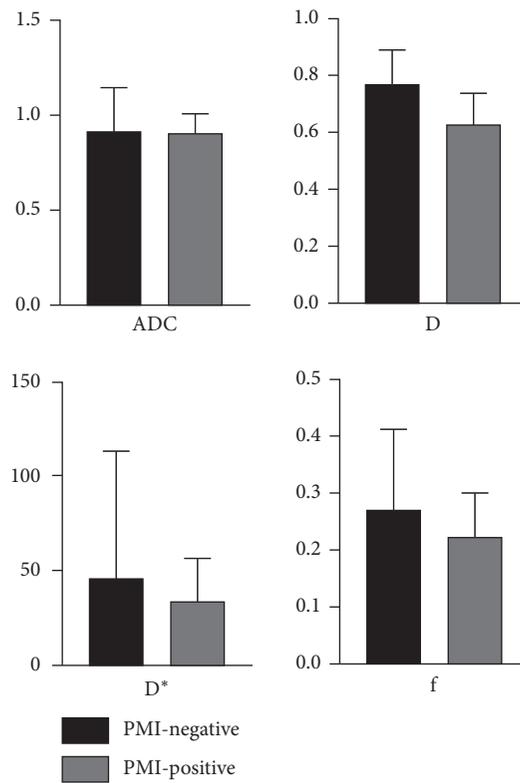
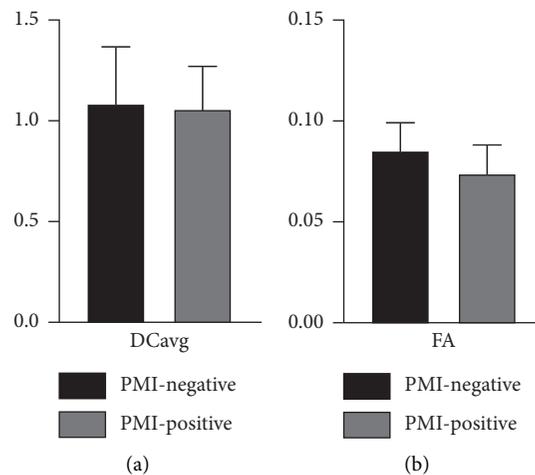
FIGURE 1: ADC, D, D^* , and f in the PMI-positive and PMI-negative groups.

FIGURE 2: DCavg and FA in the PMI-positive and PMI-negative groups.

TABLE 2: Diagnostic efficiency of each metric and their combined parameters.

Parameters	Cut-off value	AUC	Sensitivity	Specificity
D	0.741	0.801 (0.695–0.907)	0.600	0.850
FA	0.078	0.726 (0.600–0.852)	0.720	0.700
D + FA	—	0.931 (0.865–0.997)	0.800	0.975

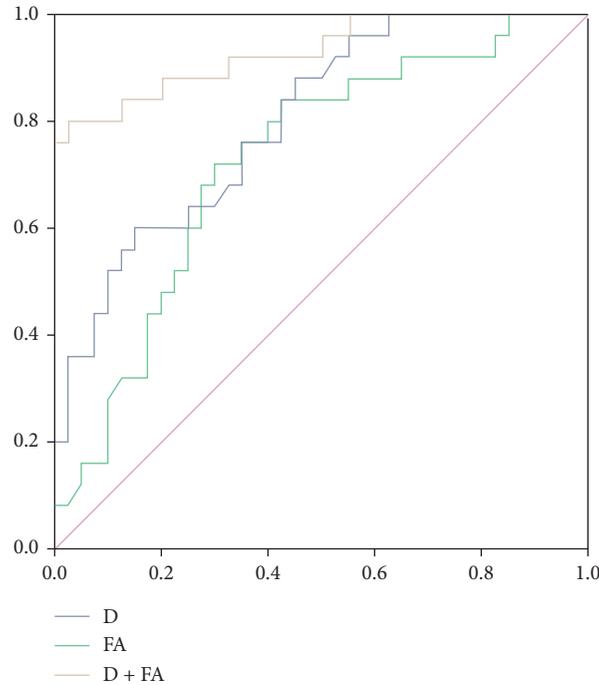


FIGURE 3: ROC curve showing D (blue line), FA (green line), and the combination of D and FA (yellow line) in the diagnosis of cervical cancer with parametrial invasion.

value is mainly related to cell density, the form of nucleus division, and the arrangement of muscle fibers [27]. The present study showed that compared with the PMI-negative group, the PMI-positive group had a lower FA value. This result corresponded with a previous report that the FA values of endometrial cancer (0.21 ± 0.05) were lower than those of the inner myometrium (0.44 ± 0.01) and outer myometrium (0.32 ± 0.08), while the DCavg values of endometrial cancer could not be used to evaluate myometrial invasion [13]. The tumor cells infiltrated into the space of muscle fiber bundles and disrupted a varying degree of muscle fibers, resulting in a decreased amount of muscle fiber bundles in the unit voxel. After that, the water diffusive limitation in all directions increased. In our study, the PMI-positive group showed a higher degree of malignancy than the PMI-negative group, reflecting higher damage to the muscle fibers. Meanwhile, the space of the muscle fibers decreased.

Using a combination of D and FA resulted in the best diagnostic performance in terms of diagnosing PMI, and use of the combination was significantly better than the use of D

or FA alone. Moreover, the combination of D and FA had higher sensitivity and specificity (80.0% and 97.5%, respectively) than either alone. Therefore, a combination of IVIM and DTI could be used to diagnose PMI with improved efficacy. This improvement might considerably benefit clinical decision-making, facilitate the management of individual patients, and improve patients' clinical prognosis.

Several limitations of the present study should be mentioned. First, the number of patients with cervical cancer was relatively small. Second, a wide range of clinical stages (early and advanced cervical cancer) were included in the study, and different pathological subtypes were not analyzed. Third, certain biases related to this study were inevitable because none of our team members were involved in the initial diagnostic processes or documentation of treatment. In addition, the ROIs were drawn manually by radiologists in the study, so certain subjective factors of inter- and intraobserver variability and slice-selection bias may exist. Future research with a larger sample size will be carried out.

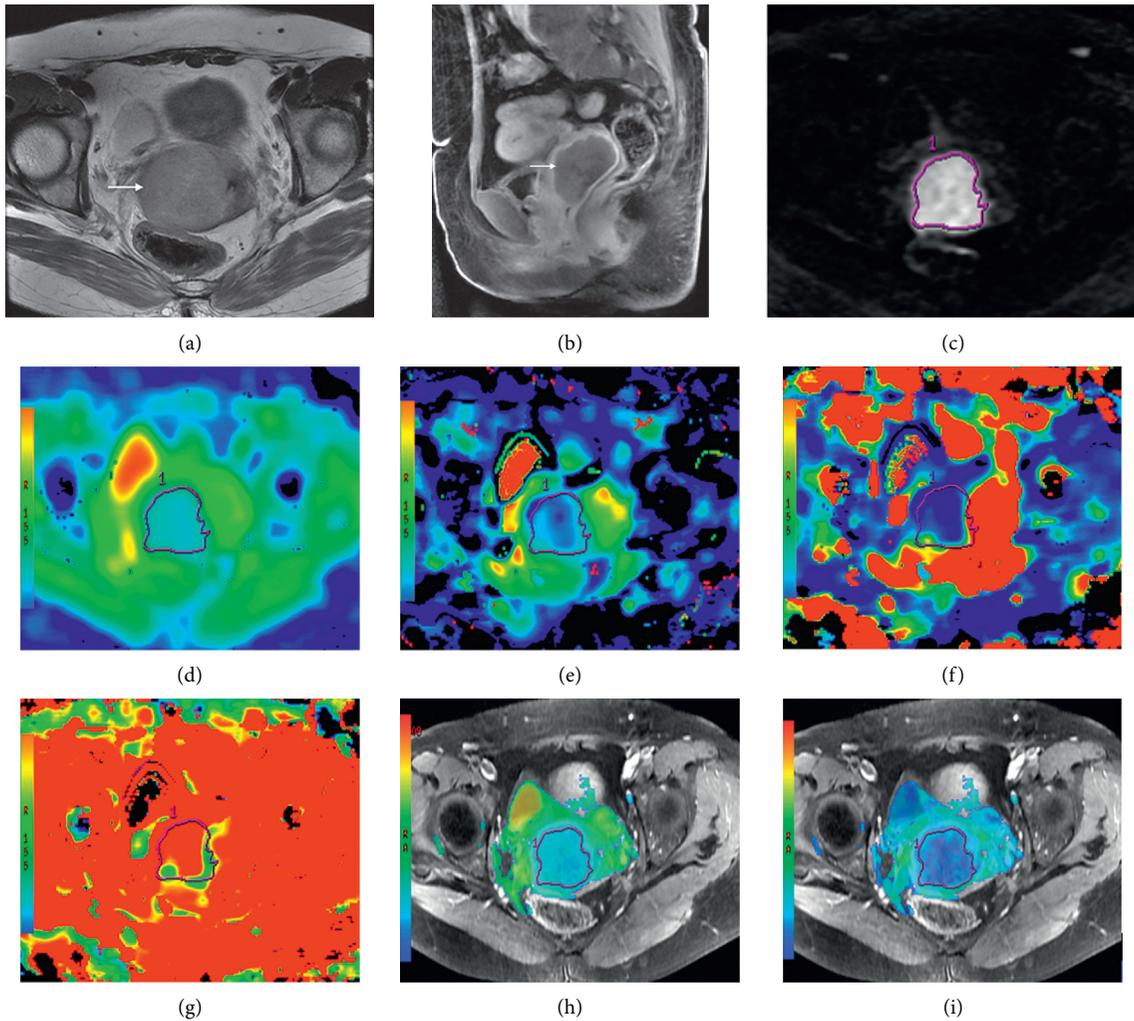


FIGURE 4: A 52-year-old woman with stage IIA cervical cancer. (a) is T2-weighted image (T2WI) shows a slightly hyperintense cervical mass, which was slightly enhanced on the contrast-enhanced magnetic resonance image (b). (c) shows the tumor with hypertension on the IVIM-DWI image with the b value of 800 s/mm^2 . (d–g) display the ADC, D , D^* , and f parametric maps are derived from IVIM-DWI. (h–i) show the D_{CAvg} and FA maps derived from DTI.

5. Conclusions

The present study showed that PMI-positive lesions had significantly lower D and lower FA values than PMI-negative lesions. Both IVIM and DTI were promising methods for diagnosing PMI in cervical cancer cases. However, the combination of IVIM and DTI is more valuable than either option alone, as the combination obviously improved the diagnostic performance, resulting in higher AUC and higher diagnostic sensitivity and specificity than either option alone.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

This work was supported by the Anhui Provincial Key Research and Development Program Project, China (1804h08020294), Anhui Provincial Natural Science Foundation for Youths, China (1908085QH364), and Fundamental Research Funds for the Central Universities (WK9110000061).

References

- [1] H. U. Je, S. Han, Y. S. Kim et al., "A nomogram predicting the risks of distant metastasis following postoperative radiotherapy for uterine cervical carcinoma: a Korean radiation

- oncology group study (KROG 12-08),” *Radiotherapy and Oncology*, vol. 111, no. 3, pp. 437–441, 2014.
- [2] G. Zhou, X. Chen, F. Tang, J. Zhou, Y. Wang, and Z. Wang, “The value of diffusion-weighted imaging in predicting the prognosis of stage IB-IIA cervical squamous cell carcinoma after radical hysterectomy,” *International Journal of Gynecological Cancer*, vol. 26, no. 2, pp. 361–366, 2016.
 - [3] Y. Ikeda, A. Furusawa, R. Kitagawa et al., “Practice patterns of adjuvant therapy for intermediate/high recurrence risk cervical cancer patients in Japan,” *Journal of Gynecologic Oncology*, vol. 27, no. 3, 2016.
 - [4] K. Nakamura, I. Joja, T. Nagasaka et al., “The mean apparent diffusion coefficient value (ADC_{mean}) on primary cervical cancer is a predictive marker for disease recurrence,” *Gynecologic Oncology*, vol. 127, no. 3, pp. 478–483, 2012.
 - [5] X. Tang, J. Qiu, C. Guo, J. Guo, and K. Hua, “A novel prognostic nomogram utilizing the 2018 FIGO staging system for cervical cancer: a large multicenter study,” *International Journal of Gynecology & Obstetrics*, 2021.
 - [6] M. Kim, D. H. Suh, K. Kim, H. J. Lee, Y. B. Kim, and J. H. No, “Magnetic resonance imaging as a valuable Tool for predicting parametrial invasion in stage IB1 to IIA2 cervical cancer,” *International Journal of Gynecologic Cancer*, vol. 27, no. 2, pp. 332–338, 2017.
 - [7] J. J. Park, C. K. Kim, S. Y. Park, B. K. Park, and B. Kim, “Value of diffusion-weighted imaging in predicting parametrial invasion in stage IA2-IIA cervical cancer,” *European Radiology*, vol. 24, no. 5, pp. 1081–1088, 2014.
 - [8] S. Woo, C. H. Suh, S. Y. Kim, J. Y. Cho, and S. H. Kim, “Magnetic resonance imaging for detection of parametrial invasion in cervical cancer: an updated systematic review and meta-analysis of the literature between 2012 and 2016,” *European Radiology*, vol. 28, no. 2, pp. 530–541, 2017.
 - [9] D. L. Bihan, E. Breton, D. Lallemand et al., “Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging,” *Radiology*, vol. 168, no. 2, pp. 497–505, 1988.
 - [10] X. Zheng, W. Guo, J. Dong, and L. Qian, “Prediction of early response to concurrent chemoradiotherapy in cervical cancer: value of multi-parameter MRI combined with clinical prognostic factors,” *Magnetic Resonance Imaging*, vol. 72, no. 6, pp. 159–166, 2020.
 - [11] M. Lin, X. Yu, Y. Chen et al., “Contribution of mono-exponential, bi-exponential and stretched exponential model-based diffusion-weighted MR imaging in the diagnosis and differentiation of uterine cervical carcinoma,” *European Radiology*, vol. 27, no. 6, pp. 2400–2410, 2017.
 - [12] S. Weiss, T. Jaermann, P. Schmid et al., “Three-dimensional fiber architecture of the nonpregnant human uterus determined ex vivo using magnetic resonance diffusion tensor imaging,” *The Anatomical Record Part A: Discoveries in Molecular, Cellular, and Evolutionary Biology*, vol. 288A, no. 1, pp. 84–90, 2006.
 - [13] M. Toba, N. Miyasaka, U. Sakurai, I. Yamada, Y. Eishi, and T. Kubota, “Diagnostic possibility of diffusion tensor imaging for the evaluation of myometrial invasion in endometrial cancer: an ex vivo study,” *Journal of Magnetic Resonance Imaging*, vol. 34, no. 3, pp. 616–622, 2011.
 - [14] Y. He, N. Ding, Y. Li et al., “3-T diffusion tensor imaging (DTI) of normal uterus in young and middle-aged females during the menstrual cycle: evaluation of the cyclic changes of fractional anisotropy (FA) and apparent diffusion coefficient (ADC) values,” *The British Journal of Radiology*, vol. 88, no. 1049, Article ID 20150043, 2015.
 - [15] K. Fujimoto, A. Kido, T. Okada, M. Uchikoshi, and K. Togashi, “Diffusion tensor imaging (DTI) of the normal human uterus in vivo at 3 tesla: comparison of DTI parameters in the different uterine layers,” *Journal of Magnetic Resonance Imaging*, vol. 38, no. 6, pp. 1494–1500, 2013.
 - [16] M. L. White, Y. Zhang, F. Yu, and S. A. Jaffar Kazmi, “Diffusion tensor MR imaging of cerebral gliomas: evaluating fractional anisotropy characteristics,” *American Journal of Neuroradiology*, vol. 32, no. 2, pp. 374–381, 2011.
 - [17] R. Jiang, Z. Ma, H. Dong, S. Sun, X. Zeng, and X. Li, “Diffusion tensor imaging of breast lesions: evaluation of apparent diffusion coefficient and fractional anisotropy and tissue cellularity,” *The British Journal of Radiology*, vol. 89, no. 1064, Article ID 20160076, 2016.
 - [18] K. Yamaguchi, T. Nakazono, R. Egashira et al., “Diagnostic performance of diffusion tensor imaging with readout-segmented echo-planar imaging for invasive breast cancer: correlation of ADC and FA with pathological prognostic markers,” *Magnetic Resonance in Medical Sciences*, vol. 16, no. 3, pp. 245–252, 2017.
 - [19] D. L. Bihan, R. Turner, and J. R. Macfall, “Effects of intravoxel incoherent motions (IVIM) in steady-state free precession (SSFP) imaging: application to molecular diffusion imaging,” *Magnetic Resonance in Medicine*, vol. 10, no. 3, pp. 324–337, 1989.
 - [20] S. I. Lee and M. Atri, “FIGO staging system for uterine cervical cancer: enter cross-sectional imaging,” *Radiology*, vol. 292, no. 1, pp. 15–24, 2018.
 - [21] G. Y. Cho, L. Moy, S. G. Kim et al., “Evaluation of breast cancer using intravoxel incoherent motion (IVIM) histogram analysis: comparison with malignant status, histological subtype, and molecular prognostic factors,” *European Radiology*, vol. 26, no. 8, pp. 2547–2558, 2016.
 - [22] W. Ma, G. Zhang, J. Ren et al., “Quantitative parameters of intravoxel incoherent motion diffusion weighted imaging (IVIM-DWI): potential application in predicting pathological grades of pancreatic ductal adenocarcinoma,” *Quantitative Imaging in Medicine and Surgery*, vol. 8, no. 3, pp. 301–310, 2018.
 - [23] E. Y. P. Lee, X. Yu, M. M. Y. Chu et al., “Perfusion and diffusion characteristics of cervical cancer based on intravoxel incoherent motion MR imaging—a pilot study,” *European Radiology*, vol. 24, no. 7, pp. 1506–1513, 2014.
 - [24] Q. Wu, Y. Wang, L. Shi et al., “Intravoxel incoherent motion diffusion-weighted magnetic resonance imaging of cervical cancer with different *b*-values,” *Journal of Computer Assisted Tomography*, vol. 41, no. 4, pp. 592–598, 2017.
 - [25] J. A. U. Perucho, H. C. C. Chang, V. Vardhanabhuti et al., “*B*-value optimization in the estimation of intravoxel incoherent motion parameters in patients with cervical cancer,” *Korean Journal of Radiology*, vol. 21, no. 2, p. 218, 2020.
 - [26] H. Y. Lee, D. G. Na, I.-C. Song et al., “Diffusion-tensor imaging for glioma grading at 3-T magnetic resonance imaging: analysis of fractional anisotropy and mean diffusivity,” *Journal of Computer Assisted Tomography*, vol. 32, no. 2, pp. 298–303, 2008.
 - [27] M. Kinoshita, N. Hashimoto, T. Goto et al., “Fractional anisotropy and tumor cell density of the tumor core show positive correlation in diffusion tensor magnetic resonance imaging of malignant brain tumors,” *Neuroimage*, vol. 43, no. 1, pp. 29–35, 2008.