Case Report

Exome-First Approach in Fetal Akinesia Reveals Chromosome 1p36 Deletion Syndrome

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1. Introduction

Fetal akinesia is a condition characterized by reduced or absent fetal movements independent of the etiologies [1–3]. A definitive diagnosis of the cause could be helpful for perinatal management, perinatal decision-making within local limits, and genetic counseling for future pregnancies [1]. Although next-generation sequencing (NGS) technologies have identified some underlying genetic mutations associated with fetal akinesia, some cases remain genetically unsolved [2, 3].

Chromosome 1p36 deletion syndrome (MIM# 607872) is the most common subtelomeric terminal deletion syndrome with a prevalence of 1:5000 newborns [4]. The typical clinical features of this syndrome include generalized hypotonia, severe developmental delay, seizure, growth restriction, microcephaly, congenital heart defects, flat nasal bridge, and midface hypoplasia [4]. It is classically diagnosed postnataally from typical craniofacial features, although prenatal characteristic findings have been described [4, 5]. The recurrence risk depends on the mechanism of the deletion, such as de novo deletion or inheritance from one of the parents with balanced translocations [6].

Here, we present a case with fetal akinesia associated with chromosome 1p36 deletion syndrome, which was not suspected from prenatal clinical findings before genetic testing and was diagnosed postnataally by the exome-first approach.

2. Clinical Case

A 28-year-old nulliparous pregnant Japanese woman was referred for prenatal evaluation at 30 weeks of gestation because of abnormal ultrasound findings of fetal congenital...
heart defects. The family history of the parents was unremarkable. Fetal ultrasonography at 30 weeks and 5 days of gestation showed vascular ring, Ebstein's anomaly, ventricular septal defect, and single umbilical artery. The estimated fetal body weight corresponded to the Japanese standard for the gestational age. Fetal ultrasonography showed vertex presentation of the moving fetus and the fetal stomach appeared to be normally dilated (Table 1). The pregnant woman had not felt any fetal movements since 31 weeks of gestation. At 32 weeks and 5 days of gestation, fetal ultrasonography showed absence of fetal movement with breech presentation, polyhydramnios, absent filling of stomach, and fetal growth restriction (FGR) (Table 1). However, abnormal Doppler findings regarding the fetal middle cerebral artery, umbilical cord artery, and ductus venous were not observed. Clinical diagnosis of fetal akinesia was made at this point. At 32 weeks and 6 days of gestation, amniocentesis was performed to assess the possibility of chromosomal aberrations. Interphase fluorescence in situ hybridization (FISH) analysis on uncultured amniocytes for chromosome 13, 18, and 21 revealed two signals, respectively. At 34 weeks and 6 days of gestation, progression of polyhydramnios with maternal respiratory compromise occurred (Table 1) and 2300 mL of amniotic fluid was removed. G-banding analysis on cultured amniocytes revealed a karyotype of 46,XY,22pstk+ (Figure 1). After discussion with the parents about the prognosis of the fetus based on ultrasound findings, including fetal akinesia since 31 weeks of gestation, FGR, congenital heart defects, and left-sided pleural effusion that indicated severe phenotype with prenatal onset of genetic disorders, perinatal palliative care was chosen. At 36 weeks and 3 days of gestation, fetal ultrasonography showed further progression of polyhydramnios with maternal compromise (Table 1), and 2000 mL of amniotic fluid was removed and labor was induced with oxytocin. The breech neonate was delivered vaginally at 36 weeks and 4 days of gestation with an Apgar score of 1 at 1 min and 1 at 5 min. Birth weight was 1839 g, length 45.5 cm, head circumference 31.8 cm, and chest circumference 23.5 cm. External examination revealed marked muscular hypoplasia of upper and lower extremities, extremely thin transverse palmar creases, joint contractures of lower extremities, hypertelorism, and deep-set ears. The neonate died within 2 h after birth due to respiratory failure. Therefore, we could not assess developmental profile after birth. In addition, permission for neonatal autopsy was not obtained from the parents. Clinical features of the neonate were not sufficient to diagnose a specific disease but suggested the possibility of genetic disorders, including diseases caused by either a single gene or a chromosomal defect. After genetic counseling and obtaining written consent from the parents, whole exome sequencing (WES) was performed with genomic DNA extracted from the placenta using the eXome Hidden Markov Model v1.0 (XHMM). Although the causative gene mutations related to the phenotype of the neonate were not identified, a 3 Mb deletion of chromosome 1p was suspected (Figure 2(a)). The suspected deleted region by the exome analysis using XHMM was further validated by chromosomal microarray (CMA). CMA analysis demonstrated monoallelic deletion located from positions 849466 to 3347420 on chromosome
Figure 2: Postnatal molecular cytogenetic analyses. (a) Whole exome sequencing analysis using the eXome Hidden Markov Model (XHMM). XHMM analysis using WES data detected the copy number loss located within 1p36.33–p36.32, suggesting a 3 Mb deletion (black bar). x axis shows the physical position, and y axis shows the Z score of the principal component analysis that was normalized to read depth. Purple circles connected by red lines represent values of the placenta to WES. Gray dots with gray connected lines indicate the results of normalized read depth. Copy number losses (red dots) without gains (green dots) on chromosome 1p36 were detected in the placenta. (b) Chromosomal microarray (CMA) analysis using Cytoscan 750 k Array. CMA analysis results for the copy number log2 ratio of chromosome 1p region for the placenta (purple), father (pink), and mother (blue). CMA analysis demonstrated a 3 Mb heterozygous deletion within 1p36.33–p36.32 in the placenta. The fetus had arr[hg]1p36.33–p36.32 (849466_3347420)x1. There were no copy number variations in the parents detected by CMA.
the first report describing a case with chromosome 1p36 deletion syndrome presenting with fetal akinesia.

Fetal akinesia is a condition characterized by reduced or absent fetal movement [1–3]. Prenatal sonographic findings of fetal akinesia include lack of extremity motions, persistent abnormal posture of the extremities, polyhydramnios due to decreased fetal swallowing, thorax hypoplasia due to absent fetal breathing, and fetal hydrops [1]. However, these prenatal ultrasound findings are nonspecific to identify the cause, and as yet there is no established approach for prenatal diagnosis of fetal akinesia. Fetal akinesia can result from primary defects at any point along the motor system pathway from the central nervous system to the skeletal muscle cell, which cause diseases such as spinal muscular atrophy, congenital myasthenic syndromes, and congenital muscular dystrophies [1–3]. In addition, a differential diagnosis should include a trisomy 18, metabolic dysfunction such as pyruvate dehydrogenase deficiency, maternal antibodies against acetylcholine receptor, and maternal infections such as cytomegalovirus and toxoplasmosis [1, 8]. A family history is helpful because some diseases are inherited [1–3]. Although a definitive diagnosis helps parents with perinatal decision-making, it would not be possible to do so based on sonographic findings alone. Prenatal sonography could provide sufficient information about a severity of the fetus for parents.

In the present case, while fetal movements and a normal fluid-filled stomach were seen until second trimester, these were absent during the third trimester. In addition, multiple fetal abnormalities such as congenital heart defects, FGR, ventriculomegaly, choroid plexus cysts, and single umbilical artery were found through fetal sonography. Based on these fetal sonographic findings, we performed amniocentesis to rule out trisomy 18. Although the amniocytes showed a normal karyotype, perinatal palliative care was performed based on the prenatal sonographic findings alone. Prenatal sonography could provide sufficient information about a severity of the fetus for parents.

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among affected individuals [12]. This phenotypic variation is due, at least in part, to the genetic heterogeneity seen in 1p36 deletions, which include deletions of varying lengths located throughout the 30 Mb of DNA that comprise chromosome 1p36 [6, 12]. In addition, the terminal 1p36 deletion can be missed using conventional G-banding analysis because of the low level of resolution and light staining of the region [5, 11]. In this regard, CMA analysis or subtelomeric FISH may be required to identify chromosome 1p36 deletion [5, 11]. In the present case, a 3 Mb deletion of 1p36 was not seen prenatally in the cultured amniocyte karyotype (Figure 1). Given the WES and CMA findings, the cultured amniocyte karyotype was reanalyzed and the deletion was still not seen. In Japan, the use of sub-telomeric FISH or CMA analysis for prenatal screening are not considered due to legal constraints. However, given the implications for prognosis and higher rate of hypotonia and severe developmental delay, molecular prenatal diagnosis, specifically for deletion of 1p36, should be considered in the setting of a fetal akinesia.

In the present case, the 76 deleted genes included the SKI gene that is responsible for the 1p36 deletion phenotype and is one of the candidate genes involved in hypotonia [4, 7]. However, there are no reports of the association between fetal akinesia and 1p36 deletion within a segment from 849466 to 3347420. Four genes AGRN, B3GALT6, ATAD3A, and PEX10, which were also among the 76 genes, cause recessive syndrome with hypotonia. As no mutations were detected in the four genes in the nondeleted allele, fetal akinesia may not result from the unmasking of recessive diseases. Therefore, we did not speculate that a deletion located from positions 849466 to 3347420 on chromosome 1p36.33–p36.32 resulted in fetal akinesia, which may present a more severe phenotype with prenatal onset of chromosome 1p36 deletion syndrome. Trio whole genome sequencing is helpful to search other causes of fetal akinesia. Further molecular analyses are essential to clarify this point.

In regard to diagnostic approach for the present case, we performed exome-first approach postnatally following prenatal G-banding analysis. Although the CMA-first approach is still widely used to detect CNVs, WES is becoming available to detect CNVs, leading to an appropriate clinical diagnosis [13]. The advantage of using exome sequencing for a combined analysis of not only single nucleotide variants but also CNVs is to increase the analysis resolution and detection rate with one single test. Therefore, in postnatal testing, WES may be advantageous to screen genetic abnormalities as a first-choice diagnostic approach before performing CMA in undiagnosed syndromic individuals suspected of having either single gene defects or CNVs, such as the present case [13]. After WES analysis, CMA analysis with or without FISH is necessary to accurately determine the range of the region of genomic imbalance for accurate cytogenetic diagnosis.

In conclusion, we propose that chromosome 1p36 deletion syndrome, which may be missed using conventional G-banding karyotype with amniocytes, be considered for differential diagnosis in cases of fetal akinesia and prompt molecular cytogenetic analysis if necessary. As a practical approach to the diagnosis of the cause of fetal akinesia, a detailed anomaly scan should be performed with karyotyping and if unremarkable, exome-first approach may be offered postnatally.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References


