Serum and red blood cell antioxidant status in patients with bronchial asthma

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Asthma is characterized by reversible airflow obstruction and the presence of chronic inflammation of the airways. Cells involved in the asthmatic inflammatory process have been shown to generate increased amounts of reactive oxygen species (ROS) (1-3), and levels of ROS generation correlate with asthma severity (4). ROS, released from eosinophils, alveolar macrophages and neutrophils, appear to play a key role in asthma. They may directly contract smooth mus-
 plausible in the airways, stimulate histamine release from mast cells, stimulate mucus secretion and interact with alpha1-protease inhibitor (5-7). Neutrophils as well as eosinophils are increased in number after antigen inhalation challenge, and may be activated in patients with asthma (8).

To prevent the damage caused by ROS, multiple defence systems – collectively called antioxidants – are present in serum, as well as in erythrocytes and the lungs (5). These defence systems consist of both enzymatic and nonenzymatic antioxidants. The major enzymatic antioxidants are superoxide dismutase (SOD), which degrades superoxide anion and catalase, glutathione peroxidase and the glutathione oxidation-reduction system, which inactivates hydrogen peroxide and hydroperoxides (9). However, nonenzymatic antioxidants such as ceruloplasmin, an acute phase reactant, transferrin and albumin are considered antioxidants because they bind oxidation-reduction active metals and limit the production of metal-catalyzed free radicals (10-12). Essential nutrients such as vitamins C and E also protect against reactive oxygen metabolite-mediated cellular damage, through their free radical scavenging properties (13). Vitamin C is the major antioxidant present in the airway surface liquid of the lung, and it may protect against endogenous agents, as well as against exogenous agents such as cigarette smoke and environmental air pollutants (5). The aim of the present study was to investigate possible changes in antioxidant levels in the serum and erythrocytes of patients with asthma.

PATIENTS AND METHODS

The study included 40 patients with asthma and 43 healthy volunteers. The asthma group comprised 22 women and 18 men, with ages ranging from 18 to 70 years (37.8±14.2 years). The control group comprised 24 healthy women and 19 healthy men, aged 21 to 68 years (36.2±11.7 years). None of the participants in the study had eczema, hay fever or recurrent respiratory symptoms. Patients with asthma were selected from the Sanliurfa State Hospital, Sanliurfa, Turkey. The diagnosis of asthma was established according to American Thoracic Society criteria (14). All patients had reversible airflow limitation, showing a minimum 12% increase in their forced expiratory volume in 1 s (FEV1) after inhalation of a beta-mimetic. Mean baseline FEV1 was 75.2% predicted. None of the patients had diabetes mellitus, liver or kidney diseases, or thyroid dysfunction. During the study, all patients were taking a beta-mimetic (terbutaline, salbutamol, etc) and an antihistaminic, mast cell-stabilizing agent such as cromolin sodium. None received systemic or inhaled corticosteroid treatment during the study.

Blood samples were obtained from patients in the morning after an overnight fast. Samples were centrifuged at 3000 g for 15 min at room temperature. All measurements were taken immediately after serum preparation. Other blood samples were put into heparinized tubes to separate the erythrocytes from the plasma. Plasma anduffy coats on the top of the erythrocyte pellet were removed by a pipette, and the remaining erythrocyte pellet was washed three times with 0.9% saline and centrifuged at 1500 g for 5 min. The samples were stored at 4°C until analysis later on the same day.

Vitamin C levels were measured by the method of Omaye et al (15). Briefly, vitamin C in serum was oxidized by copper ions to form dehydroascorbic acid, which reacts with acidic 2,4-dinitrophenylhidrazine to form a red bishydrazone, measured as absorbance at 520 nm. Ceruloplasmin levels were assessed by measuring its oxidative activity using p-phenylenediamine as the substrate, according to the method of Sunderland and Nomoto (16). Transferrin and albumin were measured with an automatic biochemistry analyzer (Hitachi 911 Analyzer, Boehringer Mannheim, Germany). Erythrocyte reduced glutathione concentration was determined by titration with 0.1 mmol/L 5,5′-dithiobis (2-nitrobenzoic acid) in a 0.1 mol/L disodium phosphate buffer solution, pH 8. Formation of the reduced product thionitrobenzene was measured spectrophotometrically at 412 nm (17). Erythrocyte GSH levels were expressed as μmol/g hemoglobin. Quality controls were included with the sample analysis (mean coefficients of variation for all analyses ranged from 3.8% to 5.2%); also, commercial two-point control standards were included with each analysis to calibrate instruments.

Mean ± standard deviation values of all variables in serum were calculated for the patient and control groups. The Student’s t-test was used to compare means, and P<0.05 was considered significant. Pulmonary function and other serum and erythrocyte variables were tested by intracorrelational analysis.

RESULTS

Characteristics of patients are shown in Table 1. There were no correlations among FEV1, peak flow rate, and levels of vitamin C, ceruloplasmin, transferrin, albumin and GSH. Mean serum vitamin C, ceruloplasmin, transferrin and albumin levels, and erythrocyte GSH levels of patients and controls, together with Student’s t-test results, are given in Table 2. Large differences in the variables were found between the patient and control groups. Serum vitamin C levels in the patient group were lower than those in the control group (P<0.001), whereas ceruloplasmin levels in the patient group were higher (P<0.001). Albumin levels were significantly higher in the control group than in the patient group (P<0.05). No difference was observed in the transferrin levels between the patient and control groups. As seen in

<table>
<thead>
<tr>
<th>TABLE 1 Characteristics of 40 patients with bronchial asthma</th>
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<tbody>
<tr>
<td><strong>Sex (M/F)</strong></td>
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<tr>
<td><strong>Smoker (yes/no)</strong></td>
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<tr>
<td><strong>Age (years)</strong></td>
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<tr>
<td><strong>Height (cm)</strong></td>
</tr>
<tr>
<td><strong>Weight (kg)</strong></td>
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<tr>
<td><strong>FEV1 (L (% predicted)</strong></td>
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<tr>
<td><strong>Peak expiratory flow (L/s (% variability)</strong></td>
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<td><strong>Beta-agonist use (salbutamol/terbutaline)</strong></td>
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**FEV1 Forced expiratory volume in 1 s**
Table 2, erythrocyte GSH levels in patients were higher than those in controls. There were no differences in the numbers of cigarette smokers and nonsmokers in the patient and control groups.

**DISCUSSION**

Patients with asthma have increased numbers of low density or hypodense eosinophils in blood, bronchoalveolar lavage (BAL) fluid and lung tissue. This association suggests that low density eosinophils play a role in asthma (18,19) and may be highly relevant cells in the asthma process. Sedgwich et al (20) found that when purified suspensions of either eosinophils or neutrophils were activated by 1 ng/mL phorbol myristate acetate, differences were observed in both the rate and the total amount of superoxide generated by the cell suspensions. Eosinophils (both normal and low density) from patients with asthma generated more superoxide than matching neutrophils. Moreover, the generation of superoxide by neutrophils from patients with asthma during a reaction lasting 30 to 60 min was significantly greater than neutrophils from controls. These findings suggest that patients with asthma are subjected to more free oxygen radicals than are those without asthma. Rahman et al (21) found that plasma trolox-equivalent antioxidant capacity was low in patients presenting with acute exacerbations of chronic obstructive pulmonary disease or asthma, with increases in plasma lipid peroxidation products.

In a healthy state, when ROS production is low, lipid peroxidation is inhibited by the combined activities of various antioxidants present in plasma. However, in the event of excessive ROS production, as is the case in asthma, this protection may be inadequate. Vitamin C is highly water soluble and a strong reducing agent (22). It has recently been shown that, in addition to enhanced lipid peroxidation, levels of vitamins C and E are decreased in the BAL fluid of guinea pigs after an asthmatic response (23). An increasing body of data suggests that low consumption of fresh fruit containing vitamin C is associated with decreased lung function in both children and adults (24). In 1990, Schwartz and Weiss (25) first reported an inverse association between bronchitis and wheezing, and both dietary and plasma vitamin C levels in a representative sample of the adult population in the United States. In our study, the level of vitamin C was lower in patients with asthma than in the control group.Kalaycý et al (26) reported that plasma vitamin C concentration was decreased in asthmatic children. Kelly et al (27) found that vitamin C concentration was low in the fluid lining the lungs of patients with asthma. One possible explanation for the low level of vitamin C is that asthmatic patients eat less vitamin-rich foods in their diet. This explanation is supported by recent studies (28-31). The other possible explanation is that vitamin C is metabolized at an increased rate as part of a defense mechanism against the ongoing oxidative burden.

The same property makes vitamin C an excellent antioxidant, capable of scavenging a wide variety of oxidants. For example, ascorbate has been shown to scavenge effectively superoxide, hydrogen peroxide, hypochlorous acid, aqueous peroxyl radicals and singlet oxygen. During its antioxidation activity, ascorbate undergoes a two-electron oxidation to dehydroascorbic acid (the oxidized form of vitamin C), with the intermediate formation of the relatively unreactive ascorbyl radical. Although dehydroascorbic acid is relatively unstable and hydrolyzes readily to 1,2-diketogulonic acid, it can be reduced back to ascorbate by a variety of cells or thiols such as homocysteine and GSH. Plasma ascorbate plays an important role in protecting plasma lipids from ROS attack; however, it is rapidly oxidized when challenged by oxidants released from activated polymorphonuclear cells (22). This rapid depletion may be the reason for the low serum ascorbate values recorded in our study (P<0.001). Under these conditions, endogenously synthesized proteins in the liver, such as ceruloplasmin and transferrin, may play a significant role in scavenging ROS and their harmful lipid peroxidation products. It has been shown that ceruloplasmin can scavenge free radicals as well as transition metals, and thereby protect against lipid peroxidation (10,11). Two research teams (32,33) observed that when malondialdehyde levels increased as the result of oxidative stresses, ceruloplasmin activity increased in parallel to the oxidant stress. In addition, Boljevic et al (34) found that the concentration of plasma malondialdehyde increased in patients with acute asthma. Moreover, the relation between induced and spontaneous luminol-dependent chemiluminescence of plasma suggested that antioxidant plasma activity was reduced; in remission, plasma malondialdehyde decreased but did not reach the normal range.

GSH is present in most mammalian cells. It is an antioxidant, participating in many biological phenomena (9). Few data exist regarding glutathione levels in the erythrocytes of patients with asthma (35). GSH protects cells against oxygen

**TABLE 2**

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<th>Serum Vitamin C (µM)</th>
<th>Ceruloplasmin (µmol/L)</th>
<th>Transferrin (g/L)</th>
<th>Albumin (g/L)</th>
<th>Erythrocyte GSH (mol/mol hemoglobin)</th>
</tr>
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<tr>
<td>Bronchial asthma (n=40)</td>
<td>36.91±12.50</td>
<td>442±73</td>
<td>2.05±0.58</td>
<td>46.2±3.0</td>
<td>0.59±0.11</td>
</tr>
<tr>
<td>Controls (n=43)</td>
<td>53.38±13.06</td>
<td>308±47</td>
<td>2.11±0.55</td>
<td>48.8±2.1</td>
<td>0.49±0.09</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>NS</td>
<td>&lt;0.05</td>
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Results are expressed as mean ± SD. NS Not significant (P>0.05)
radicals and toxic compounds. Both animal and human studies have shown that GSH levels may increase during oxidative stress (36). Increased levels of GSH were found in the BAL fluid of patients with asthma, erythrocytes of patients with chronic occupational lung disorders and erythrocytes of volunteers participating in exercise training (36-38). We observed an approximately 20% increase in GSH levels in the erythrocytes of patients with asthma (P<0.001). This increase may occur in response to the production of ROS because ROS require GSH for detoxification. Penning et al (39) reported that the level of GSH was elevated in the erythrocytes of patients with asthma and decreased after steroid inhalation. This observed elevation in erythrocyte GSH may reflect an increased oxidative burden (39). The GSH concentration in bronchial fluid (but not in red blood cells) was shown to be higher in patients with asthma; a similar trend was seen in the alveolar fluid (36). In another study (27), oxidized GSH concentrations in bronchial washings and BAL fluid were higher in patients with asthma than in the control group, indicating the presence of oxidative stress in the airways. Glutathione peroxidase, an enzyme present in large amounts in erythrocytes, needs GSH to detoxify hydrogen peroxide. In our study, GSH concentration was higher in patients with asthma than in the control group, which may be explained by increased oxidative stress. Indeed, extracellularly oxidized vitamin C is reduced by intracellular GSH and then released into the plasma (40). Thus, the GSH level reflects the antioxidant capability of the erythrocytes in such circumstances.

CONCLUSIONS

The present study shows that a vitamin C deficiency and a high GSH level, due to increased oxidative stress, may be related to asthma. Future studies evaluating antioxidant effects in asthma need to examine the products of increased ROS activity, such as malondialdehyde, oxidized sulfhydryl groups on albumin and lipid hydroperoxides.

REFERENCES
