CASE REPORT

Granulomatous *Pneumocystis jiroveci* pneumonia associated with immune reconstituted HIV

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*Pneumocystis jiroveci* pneumonia uncommonly presents with pulmonary nodules and granulomatous inflammation. An unusual case of granulomatous *P. jiroveci* pneumonia in an HIV patient with a CD4⁺ lymphocyte count of greater than 200 cells/mm³, occurring in the context of immune reconstitution with highly active antiretroviral therapy, is described. The case highlights the importance of establishing this diagnosis to institute appropriate therapy.

Key Words: HIV/AIDS; Immune reconstitution disease; Multiple pulmonary nodules; *Pneumocystis jiroveci*

CASE PRESENTATION

A 40-year-old man presented with progressive cough, dyspnea and 20 kg weight loss over several weeks. Chest imaging revealed bilateral ground-glass infiltrates, and *Pneumocystis jiroveci* pneumonia (PCP) was confirmed by bronchoalveolar lavage (BAL) analysis. The patient was subsequently found to be positive for HIV, with a CD4⁺ lymphocyte count of 16 cells/mm³, an HIV viral load >5×10⁶ copies/mL and evidence of HIV dementia. He responded to a 21-day course of trimethoprim-sulfamethoxazole and prednisone, and was started on highly active antiretroviral therapy (HAART) shortly before completing treatment for PCP. Two months after initiating HAART, his CD4⁺ count had increased to 359 cells/mm³ and his HIV viral load was 3.1×10⁴ copies/mL. Due to an allergic reaction to sulfamethoxazole, PCP prophylaxis was discontinued. A routine chest radiograph performed four months later revealed multiple bilateral pulmonary nodules (Figure 1). Subsequent chest computed tomography (CT) scans confirmed the presence of multiple homogeneous and noncalcified pulmonary nodules, several of which were cavitary. These were most prominent in the lung apices (Figure 2). The patient was asymptomatic, had been compliant with therapy and had a CD4⁺ count of 266 cells/mm³.

During the following year, the patient developed malaise, cough and wheeze. His CD4⁺ count had been consistently >250 cells/mm³ for the previous seven months, with undetectable HIV viral loads. Chest CT revealed progression of the pulmonary nodules. Bronchoscopy with BAL and transbronchial biopsy was performed, in addition to transthoracic needle biopsy. Direct fluorescent antibody testing for pneumocystis was negative on all specimens, and bacterial culture, transthoracic needle biopsy. Direct fluorescent antibody testing for pneumocystis was negative on all specimens, and bacterial culture, cytology and acid-fast bacilli (AFB) smears were consistently negative. Thoracoscopic lung biopsy was ultimately performed, and confirmed the presence of necrotizing granulomas containing *Pneumocystis* organisms surrounded by a hyalinized capsule (Figure 3). Treatment was initially deferred because the patient’s symptoms were mild and not clearly attributable to PCP infection. However, the patient’s symptoms progressed to include drenching night sweats, worsening cough and progressive dyspnea. He was subsequently treated for PCP with atovaquone, despite a CD4⁺ count of 374 cells/mm³. Within two weeks, his symptoms resolved, his weight increased and his diffusing capacity for carbon monoxide increased from 62% predicted to 87% predicted. A repeat chest CT scan three months later showed no significant change in the pulmonary nodules.

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DISCUSSION

Granulomatous PCP is an uncommon form of pneumocystis infection, occurring in only 3% to 5% of patients (1). Cases have most commonly been described in HIV patients with CD4⁺ counts <200 cells/mm³, and rarely in other immunocompromised patients (2). An insidious presentation is the rule, with minimal systemic or respiratory symptoms. Risk factors for the development of granulomatous PCP include PCP prophylaxis with aerosolized pentamidine, active malignancy, recent corticosteroid use and immune reconstitution disease (1,2). BAL and transbronchial biopsy often fail to reveal organisms, and the diagnosis...
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is often made on open lung biopsy or postmortem. Biopsy specimens typically reveal *Pneumocystis* organisms within hyalinized granulomas.

Immune reconstitution disease (IRD) is a paradoxical worsening of a pre-existing opportunistic infection in HIV-positive individuals after the initiation of HAART, despite improvement in markers of HIV infection. It occurs in 10% to 25% of patients receiving HAART, often within eight weeks of therapy, and is most commonly seen with mycobacterial infections (3). IRD with PCP has been rarely reported, occurring two weeks to seven months after initiation of HAART (4,5). Risk factors for IRD include a pretreatment CD4+ count <50 cells/mm³, a brisk CD4+ recovery of two- to four-fold in the first 12 months of therapy or a rapid decrease in HIV viral load after initiation of HAART (6). Pathogenic mechanisms include immune recognition of antigens associated with ongoing infection or an inflammatory response to nonreplicating antigens from previous infection (6,7). Granuloma formation is characteristic due to T lymphocyte activation (8). The inflammatory reaction is responsive to a combination of glucocorticoids and anti-inflammatories, whereas IRD occurring as a result of active infection requires treatment with antimicrobials. HAART should be discontinued if the inflammatory response is life threatening (6).

PCP colonization is defined by the presence of *Pneumocystis* DNA in the respiratory specimens of patients without clinical PCP infection. Colonization is detected with polymerase chain reaction assays targeting the large subunit of mitochondrial ribosomal RNA (9). PCP colonization is seen in 20% of healthy adults (10), 44% of non-HIV immunocompromised individuals and 46% of HIV-infected hosts without active PCP (11). Risk factors for PCP colonization include HIV infection, corticosteroid use and chronic lung disease (9). It is unclear whether PCP colonization represents the presence of active organisms and, as such, predisposes patients to subsequent PCP infection.

We hypothesize that our patient developed an immune response to pneumocystis antigens remaining in the lung after treatment of primary pneumocystosis. The lysis of nodules during surgery likely resulted in the release of PCP antigens, intensifying the immune response and causing a progression of symptoms. Clinical improvement with directed antimicrobial therapy is evidence of active infection and not merely PCP colonization – either alone or in association with IRD. The patient’s presentation is unique in that there was a significant delay in symptoms after the radiographic demonstration of pulmonary nodules, without a clear inciting factor for the development of symptoms and in the setting of an adequate CD4+ count. The present case highlights the need to consider PCP in all HIV patients, even those with an unusual presentation; although establishing a diagnosis can be challenging, it is critical to provide appropriate therapy at the onset of infectious or immune complications.

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REFERENCES