Bronchiectasis exacerbations: The role of atypical bacteria and respiratory syncytial virus

Eugenios I Metaxas MD MSc¹, Evangelos Balis MD PhD¹, Joseph Papaparaskevas MD PhD², Nicholas E Spanakis MD PhD², Georgios Tatsis MD PhD¹, Athanasios Tsakris MD PhD²

BACKGROUND: Aside from the known role of common bacteria, there is a paucity of data regarding the possible role of atypical bacteria and viruses in exacerbations of non-cystic fibrosis bronchiectasis.

OBJECTIVE: To explore the possible role of atypical bacteria (namely, Mycoplasma pneumoniae and Chlamydophila pneumoniae) and respiratory syncytial virus (RSV) as causative agents of bronchiectasis exacerbations.

METHODS: A cohort of 33 patients was studied over a two-year period (one year follow-up for each patient). Polymerase chain reaction for the detection of M pneumoniae, C pneumoniae and RSV in bronchoalveolar lavage samples were performed during all visits. Antibody titres (immunoglobulin [Ig]M and IgG) against the aforementioned pathogens were also measured. In addition, cultures for common bacteria and mycobacteria were performed from the bronchoalveolar lavage samples.

RESULTS: Fifteen patients experienced a total of 19 exacerbations during the study period. Although RSV was detected by polymerase chain reaction during stable visits in four patients, it was never detected during an exacerbation. M pneumoniae and C pneumoniae were never detected at stable visits or during exacerbations. IgM antibody titres for these three pathogens were negative in all patient visits.

CONCLUSIONS: Atypical pathogens and RSV did not appear to be causative agents of bronchiectasis exacerbations.

Key Words: Atypical bacteria; Bronchiectasis; Exacerbations; Respiratory syncytial virus

Les exacerbations de la bronchectasie : le rôle des bactéries atypiques et du virus respiratoire syncytial

HISTORIQUE : On connaît le rôle des bactéries courantes dans les exacerbations des bronchectasies non attribuables à la fibrose kystique, mais on possède peu de données sur celui des bactéries et des virus atypiques.

OBJECTIF : Explorer la possibilité que des bactéries atypiques (le Mycoplasma pneumoniae et la Chlamydophila pneumoniae) et le virus respiratoire syncytial (RSV) soient responsables d’exacerbations de la bronchectasie.

MÉTHODOLOGIE : Une cohorte de 33 patients a fait l’objet d’une étude de deux ans (suivi de chaque patient pendant un an). À chaque rendez-vous, les chercheurs ont effectué une amplification en chaîne de la polymérase (PCR) de prélèvements de lavage broncho-alvéolaire pour déceler le M pneumoniae, la C pneumoniae et le VRS. Ils ont mesuré les titres d’anticorps (immunoglobuline [Ig]M et IgG) contre ces pathogènes. Enfin, ils ont procédé à des cultures de bactéries et mycobactéries courantes dans les prélèvements des lavages broncho-alvéolaires.

RÉSULTATS : Quinze patients ont subi un total de 19 exacerbations pendant la période de l’étude. Même si le VRS a été décelé par PCR lors du rendez-vous de quatre patients stables, il ne l’a jamais été pendant une exacerbation. On n’a jamais décelé le M pneumoniae ou la C pneumoniae chez les patients stables ou en exacerbation. Les titres d’anticorps de l’IgM de ces trois pathogènes étaient négatifs lors de tous les rendez-vous.

CONCLUSIONS : Les pathogènes atypiques et le VRS ne semblaient pas responsables des exacerbations de la bronchectasie.

METHODS

A two-year, single-centre, observational prospective cohort study was designed and performed in a 900-bed tertiary care general hospital. The study was approved by the Ethics Committees of Evangelismos General Hospital and the Medical School of National and Kapodistrian University of Athens (Athens, Greece). Eligibility criteria were adults with known or newly diagnosed bronchiectasis. To confirm the presence and the extent of bronchiectasis, high-resolution computed tomography (HRCT) was required for all patients. Exclusion criteria comprised inability to undergo bronchoscopy due to severe heart problems or respiratory failure, and the inability to maintain a personal calendar. Cystic fibrosis (CF) patients were also excluded because they have different epidemiological characteristics (4, 5). Candidates were referred to the study team, who informed the patients about the aims and purposes of the study. All patients who were suitable and willing to participate were asked to provide informed written consent.

Definitions

Exacerbation was defined as the deterioration of at least three respiratory symptoms (cough, increased sputum volume or change in viscosity, sputum purulence with or without increasing wheeze, increased sputum

¹Pulmonology Department, Evangelismos General Hospital; ²Microbiology Department, Medical School, National and Kapodistrian University of Athens, Athens, Greece

Correspondence: Dr Eugenios I Metaxas, Ethnikis Antistaseos 29, Limassol, Cyprus. Telephone 357-99315080, fax 357-25108148, e-mail eumetaxas@yahoo.gr

Can Respir J Vol 22 No 3 May/June 2015 ©2015 Pulsus Group Inc. All rights reserved
Subjects initially screened to enter the study (n=59)

Subjects initially rejected from the study team
• 7 were considered to be high risk for bronchoscopy
• 6 could not use a peak-flow meter and/or could not keep a personal calendar

Subjects further tested to enter the study (n=46)

Subjects that entered the study (n=41)

Subjects excluded from the study
• 5 did not sign the inform consent form

Subjects excluded from the statistical analysis
• 4 withdrew consent
• 4 had missing values

Subjects included in the statistical analysis (n=33)

Figure 1) Flow diagram of the bronchiectasis cohort study

Production, dyspnea, hemoptysis and chest pain) for >24 h and/or systemic complaints, such as fever, and alterations in chest radiograph (3,6). Stable condition was defined as the simultaneous absence of clinical symptoms and elevated inflammation markers. An acute or presumed acute infection from atypical pathogens or RSV was defined as the presence of a positive polymerase chain reaction (PCR) test from bronchoalveolar lavage (BAL) samples with or without immunoglobulin (Ig)M antibodies above cut-off values, or a fourfold increase in IgG antibody titre combined with clinical features (7-9).

Management and follow up

Subjects were enrolled during a two-year period and were followed for 12 months from enrollment day. The first baseline visit occurred during a stable condition period and, if the patients were in a stable condition, they were followed up every four months. If an exacerbation occurred, patients were monitored during the exacerbation and the next planned visit was omitted. If an exacerbation occurred during the last planned visit, a fifth visit was performed six weeks after the exacerbation. Finally, if a patient did not experience an exacerbation, the last visit was omitted. During the first visit, medical history and epidemiological characteristics were recorded. Patients were given a personal calendar to record their bronchiectasis-related symptoms (eg, dyspnea, sputum production) and a peak-flow meter. On every visit – stable condition or exacerbation – blood samples were obtained and patients were submitted to bronchoscopy and pulmonary function tests.

Collection and process of samples

During visits, blood samples were obtained to measure inflammation parameters such as white blood cell count, C-reactive protein level, and antibodies (IgM and IgG) against C pneumoniae, M pneumoniae and RSV. Subsequent to the initial baseline measurement of antibodies, a second baseline measurement, taken three weeks later, was performed to identify any latent or ongoing infections. Antibodies against M pneumoniae, C pneumoniae and RSV were detected using immunofluorescence assay (Focus, USA), microimmunofluorescence assay (MRL Diagnostics, USA) and ELISA (Serion, Germany), respectively. BAL samples were obtained during bronchoscopy from the most affected lobe (according to HRCT). Cultures for mycobacteria and quantitative cultures for common pathogens were performed. Bacterial species were classified as potential pathogens or not, as described previously (10). In BAL samples, conventional PCR for M pneumoniae and C pneumoniae was performed as described previously (11,12). Real-time PCR was performed using the M pneumoniae, C pneumoniae and the RSV Real-Time RT-PCR Kits (Obelis SA, 1040, Belgium). Before the study’s PCRs tests, positive control tests were performed to confirm test accuracy.

Statistical analysis

Nonparametric tests and t tests were used to identify differences in measurable parameters between subgroups of the cohort. To identify factors associated with exacerbations, univariate logistic regression models and cross-tabulations were used. Univariate logistic regression and cross-tabulations for nonrepeated measurements were applied including each patient once. All reported P values are two tailed; the level of significance (ie, alpha value) was 0.05 with a 95% level of confidence.

RESULTS

A total of 59 patients with bronchiectasis were screened during recruitment period, of whom 41 were enrolled. Only 33 patients completed the follow-up procedure and were included in the final analysis (Figure 1). During the study, 116 patient visits were occurred. Ninety-seven (84%) visits were performed during periods of stable condition, baseline or follow-up and 19 (16%) during exacerbations. Fifteen patients (45%) experienced at least one exacerbation and four experienced two exacerbations during follow-up; three of these were hospitalized but were eventually discharged. Bronchoscopies were generally well tolerated because high-risk patients were excluded. Only minor adverse events, such as cough and throat irritation, occurred. No deaths were recorded during the study. A summary of the mean values of the inflammation markers are presented in Table 1.

Serology and PCR results

IgM-specific antibodies against C pneumoniae and M pneumoniae were negative in all cases. There was only one patient who had a fourfold increase of the IgG antibodies against C pneumoniae during the baseline visit. This patient did not exhibit any signs of infection and did not fulfill exacerbation criteria. Regarding RSV, there were no positive IgM samples or any increase in IgG titres. Molecular detection of atypical pathogens and RSV was performed in all 116 samples collected. There was no detection of C pneumoniae or M pneumoniae DNA during baseline periods. In contrast, RSV RNA was detected four times during baseline periods (Table 2). All four patients had positive IgG antibodies against RSV. Three of them had a change in IgG titres during the second measurement three weeks later but it was less than fourfold. The same three patients experienced some baseline respiratory symptoms, according to their personal calendars, but did not fulfill the criteria for an exacerbation. All PCR samples during exacerbations were negative for both atypical bacteria and RSV, including two of the patients from whom RSV RNA was isolated during the baseline periods.

Common bacterial pathogens

Isolation of a microorganism from BAL cultures was achieved in 14 (42%) different patients in 29 visits (24%). Eighteen visits were during stable condition and 11 during exacerbations. In some cultures, more than one bacterial species was isolated. The only non-potential pathogenic microorganism isolated was Candida albicans (Table 3). Eleven patients who presented during an exacerbation were treated successfully with antibiotics. Nontuberculous mycobacteria or multiresistant bacteria were not isolated. With regard to exacerbations, nine different patients had a total of 11 positive cultures. Three of them were colonized from the same bacterium that was isolated during the exacerbation: one with Staphylococcus aureus and two with P aeruginosa. Attempts to eradicate these bacteria from these three patients were unsuccessful. They experienced more than one exacerbation during the study period and exhibited increased production of purulent sputum. Patients with an exacerbation and negative cultures were treated empirically with antibiotics according to the results of previous studies (13). Long-term antibiotic regimens were prescribed in the present study (14).
TABLE 1
Summary and comparison of inflammation markers recorded during the bronchiectasis study

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Total</th>
<th>Stable condition</th>
<th>Exacerbation</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Men</td>
<td>Women</td>
<td>Men</td>
</tr>
<tr>
<td>CRP, mg/L</td>
<td>6.2 (0.0–24.0)</td>
<td>5.0 (0.0–18.0)</td>
<td>7.3 (0.0–24.0)</td>
</tr>
</tbody>
</table>

Comparison of CRP and WBC between exacerbation and stable condition periods

<table>
<thead>
<tr>
<th>Parameter</th>
<th>P</th>
<th>Mean</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>CRP, mg/L</td>
<td>&lt;0.0001</td>
<td>5.62</td>
<td>42.32–70.08</td>
</tr>
<tr>
<td>WBC, ×10⁹/L</td>
<td>0.098</td>
<td>1.165</td>
<td>−0.227–2.558</td>
</tr>
</tbody>
</table>

Data presented as mean (range). CRP = C-reactive protein; WBC = White blood cells

TABLE 2
Characteristics of the four patients from whom respiratory syncytial virus (RSV) RNA was detected in bronchoalveolar lavage (BAL) samples during the baseline visit

<table>
<thead>
<tr>
<th>Patient</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Visit category</td>
<td>Baseline</td>
<td>Baseline</td>
<td>Baseline</td>
<td>Baseline</td>
</tr>
<tr>
<td>BAL RSV PCR</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>Serum RSV IgM</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Serum RSV IgG*</td>
<td>Twofold increase</td>
<td>Twofold Stable</td>
<td>Twofold decrease</td>
<td></td>
</tr>
<tr>
<td>Month of visit</td>
<td>September</td>
<td>June</td>
<td>August</td>
<td>November</td>
</tr>
<tr>
<td>Baseline FEV₁, L (%)*</td>
<td>1.37 (33)</td>
<td>1.16 (62)</td>
<td>1.5 (63)</td>
<td>3.84 (82)</td>
</tr>
<tr>
<td>Baseline symptoms</td>
<td>Cough</td>
<td>Cough</td>
<td>None</td>
<td>Cough</td>
</tr>
<tr>
<td>Dyspnea</td>
<td>Dyspnea</td>
<td>None</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>White blood cells, ×10⁹/L</td>
<td>7.050</td>
<td>6.420</td>
<td>5.640</td>
<td>8.690</td>
</tr>
<tr>
<td>C-reactive protein, mg/L</td>
<td>5</td>
<td>10</td>
<td>4</td>
<td>2</td>
</tr>
</tbody>
</table>

*Three weeks after initial measurement; *Percentage of the mean predicted value. FEV₁ = Forced expiratory volume in 1 s; Ig = Immunoglobulin; PCR = Polymerase chain reaction

DISCUSSION

To our knowledge, the present study was the first to investigate the role of viruses and atypical bacteria in non-CF bronchiectasis using PCR in BAL samples. These data are missing from large collaborative networks and guidelines regarding bronchiectasis.

The small size of the cohort and the low exacerbation rate were the main limitations. This reduced the probability of isolating culprit microorganisms. The small size of the cohort reduced the power of the study and, thus, the ability to confirm the results. Small cohort size is one of the limitations of single-centre studies. The cohort size was further reduced because of the strict protocol. Some patients with clinical suspicion of bronchiectasis were not referred to the study team because their diagnosis was not confirmed using HRCT. The protocol also required the ability to undergo bronchoscopy and to maintain a personal calendar – requirements that were not met by 13 patients who were initially screened for study entry. Furthermore, the results from four patients (almost 10% of the cohort) were not included in the final analysis because of missing data. All of these limitations derived from the strict protocol reduced the cohort size by almost one-half. In addition to the small cohort size, the exacerbation rate per patient per year in our study was only 0.58, one of the lowest reported in the literature (14). This could be partly attributed to the continuous monitoring of the patients in the study group. As a final point, some patients with mild exacerbation may have been misclassified in stable condition because the clinical criteria of an exacerbation are subjective.

The microbiological status of the patients in our study did not differ from previous studies investigating non-CF bronchiectasis during baseline (2,10,15) and exacerbation periods (13). This is also an important result because the present study was performed in a country with increased antibiotic resistance. Again, we verified that a positive culture was strongly correlated with an exacerbation.

The attribution of M pneumoniae as a causative agent of bronchiectasis has been previously reported (16,17). We did not find any data regarding baseline colonization from atypical bacteria or their role in exacerbations. However, there are data from other obstructive lung diseases. The role of viruses and atypical bacteria has been investigated in chronic obstructive pulmonary disease (COPD) exacerbations (18). Atypical bacteria have also been investigated among CF patients (19,20), although few studies used molecular techniques (21,22). Noteworthy, the investigators who studied the role of viruses using molecular techniques used nasal secretions – either exclusively or in conjunction – with sputum samples (18,22). Samples from the upper respiratory tract were also used by investigators who studied atypical bacteria in studies not related to bronchiectasis (9,18). Consequently, the use of BAL samples in our study, and not nasal or sputum samples, combined with the fact that we were not looking for upper respiratory viruses, may have contributed to these low isolation figures.

Nevertheless, our results regarding atypical bacteria appear to be consistent with the aforementioned studies, although performed in different disease groups; however, extrapolation of these results should be done with caution (23). For example, in 66 exacerbations recorded from 83 COPD patients, Seemungal et al (18) reported only one positive sample for C pneumoniae and none for M pneumoniae. This was also the case for Emre et al (20), who reported only negative results when investigating the role of M pneumoniae in CF patients.

The role of RSV has previously been investigated in COPD and CF, but we could not find any studies examining non-CF bronchiectasis. Isolation of RSV RNA in COPD patients is usually associated with symptomatic infections, but it was also detected in stable patients (8,18). This was characterized as a low-grade asymptomatic infection and has been associated with disease severity in stable COPD (18).
TABLE 4
Parameters associated with bronchiectasis exacerbations during an observational study in patients with non-cystic fibrosis bronchiectasis

<table>
<thead>
<tr>
<th>Parameter</th>
<th>P (Fisher’s exact test)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bacterial colonization</td>
<td>0.071</td>
</tr>
</tbody>
</table>

Logistic regression analysis

<table>
<thead>
<tr>
<th>Parameter</th>
<th>P OR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive culture (any bacteria)</td>
<td>0.012 7.78 (1.56–38.75)</td>
</tr>
</tbody>
</table>

Possible explanations given for the detection of RSV RNA in stable COPD patients were laboratory contamination, aborted infection and low-grade asymptomatic infection (8). In our study, the four positive isolations of RSV RNA were in patients in a clinically stable situation and negative for IgM antibodies during their first baseline visit. The nondiagnostic change in RSV IgG titres, in three of these four patients, may indicate a recent past RSV infection (24). As mentioned, two of these patients were checked again in an exacerbation and, again, RSV RNA was not isolated. Future studies investigating viruses in bronchiectasis should examine both the upper and lower respiratory tract. Additionally, the sampling method (eg, nasal swab or BAL) should be standardized because sampling method appears to be important (25).

CONCLUSION

We did not confirm an association between bronchiectasis exacerbations and atypical bacteria or RSV, although our results were derived from a small cohort. If our results are validated in larger studies, treatment of bronchiectasis exacerbations should be focused solely on already known culprits, the common bacteria.

DISCLOSURES: The authors have no financial disclosures or conflicts of interest to declare.

REFERENCES


AUTHOR CONTRIBUTIONS: Dr Metaxas: contributed to patient enrollment, design of the study, conception of the experiments, analysis and interpretation of the data, drafting of the manuscript and approval of the final version. Dr Balis: contributed to the conception and design of the study, patient enrollment, conception of the experiments, analysis and interpretation of the data, review of the report and approval of the final version. Dr Papapanakeas: contributed to conception of the experiments, analysis and interpretation of the data, review of the report and approval of the final version. Dr Spanakos: contributed to the conception of the experiments, review of the report and approval of the final version. Dr Tatsis: contributed to patient enrollment, review of the report and approval of the final version. Dr Tsakris: contributed to the conception of the experiments, design of the study, interpretation of the data, review of the report and approval of the final version.

OTHER CONTRIBUTIONS: The authors are grateful to all patients agreeing to participate in their study. They gratefully acknowledge Dr D Chryssovergi for her invaluable help in patient enrollment.

Submit your manuscripts at
http://www.hindawi.com