Relationship of the 1793G-A and 677C-T polymorphisms of the 5,10-methylenetetrahydrofolate reductase gene to coronary artery disease

Cory B. Kebert^a, June E. Eichner^b, William E. Moore^b, Eliot Schechter^c, Takuro Yaoi^d, Steve Vogel^a, Richard A. Allen^a and S. Terence Dunn^{a,*}

^aDepartment of Pathology, University of Oklahoma Health Sciences Center, Oklahoma City, OK, 73104, USA

^bUniversity of Oklahoma Prevention Research Center, University of Oklahoma Health Sciences Center, Oklahoma City, OK 73104, USA

^cDepartment of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, OK 73104, USA

^dFreshGene, Inc., 200 Mason Circle, Suite C, Concord, CA 94520, USA

Abstract. Numerous studies have investigated the relationship between polymorphisms, in particular 677C-T and 1298A-C, of the methylene-tetrahydrofolate reductase (*MTHFR*) gene and coronary artery disease (CAD) with conflicting results. This study investigates the potential association of two point mutations in *MTHFR*, 677C-T and 1793G-A, along with other risk factors, with CAD. This is the first hospital-based study to investigate 1793G-A in this context. Genotype analysis was performed on 729 Caucasians and 66 African Americans undergoing coronary angiography using a novel PCR-based assay involving formation of Holliday junctions. Allelic frequencies for 677C-T were 66.2% C and 33.8% T for Caucasians and 90.9% C and 9.1% T for African Americans. With respect to the 1793G-A polymorphism, allelic frequencies were 94.7% G and 5.3% A for Caucasians and 99.2% G and 0.8% A for African Americans. Disease associations were examined in the Caucasian patients due to their greater genotype variability and larger number in the patient cohort. Results suggest that neither 677CT heterozygotes (OR-1.36; 95% CI 0.95 to 1.96) nor mutant homozygotes (OR-0.73; 95% CI 0.44 to 1.20) have either an increased or decreased risk for CAD compared to the 677CC genotype. Likewise, the 1793GA genotype did not demonstrate a statistically significant association with CAD compared to 1793GG patients (OR-0.79; 95% CI 0.47 to 1.33). Mean homocysteine levels ($\mu\text{mol/L}$) increased from normal to mutant for 677C-T (677CC: 10.2; 677CT: 11.0; 677TT: 11.6) and normal to heterozygous in 1793G-A (1793GG: 10.7; 1793GA: 11.5). These *MTHFR* polymorphisms did not contribute to the prediction of clinically defined CAD in Caucasians.

Keywords: Methylene-tetrahydrofolate reductase, *MTHFR*, CAD, C677T, G1793A, atherosclerosis, homocysteine, Holliday junction

1. Introduction

Methylene-tetrahydrofolate reductase (*MTHFR*) has the important role in folate metabolism of catalyz-

ing the conversion of 5,10-methylene-tetrahydrofolate to 5-methyltetrahydrofolate which is required for the remethylation of homocysteine to methionine. Two common polymorphisms, 677C-T and 1298A-C, have been described as causing thermolability and reduced enzyme activity [9,15,16,41]. Homozygosity for 677C-T, homozygosity for 1298A-C and compound heterozygosity have been shown to reduce *MTHFR* enzyme activity *in vitro* to 45%, 68%, and 41%, respectively [42].

*Corresponding author: Department of Pathology, University of Oklahoma Health Sciences Center, 940 Stanton L. Young Blvd., Oklahoma City, OK 73104, USA. Tel.: +1 405 2715249; Fax: +1 405 2713881; E-mail: terry-dunn@ouhsc.edu.

With a decrease in proper functioning, hyperhomocysteinemia can occur which has been recognized as an independent risk factor for arteriosclerotic vascular disease unrelated to hyperlipidemia, hypertension, diabetes, and smoking [2,26]. Currently, 677C-T and 1298A-C polymorphisms have been linked to coronary artery disease (CAD) [8,20,21,24,25,29,36]; while a third, 1793G-A, has yet to be studied in this context.

Studies relating *MTHFR* polymorphisms to CAD have produced conflicting results. While several studies have found positive associations [8,20,21,24,25,29,36], others have not [1,4,18,27,37,46]. Some studies have even suggested that hyperhomocysteinemia may actually be a consequence of cardiovascular disease rather than a causal factor [3,28]. Folate levels have also been extensively studied, and moderate to high plasma folate levels have been shown to reduce the incidence of acute coronary events [40]. Also, the 677C-T mutant genotype along with low folate levels has been associated with increases in plasma homocysteine levels [11,13]. Klerk et al. [19] reported in their meta-analysis that individuals with 677C-T and low folate status had a significantly higher risk of CAD. In combined analysis of European studies ($n = 6207$), CAD risk for the TT mutant genotype was increased 14% over the CC wild-type genotype. By comparison, analysis of the North American studies ($n = 3146$) did not show such a relationship and even suggested a protective association with CAD (OR 0.87). One of the speculations of this study was that, with higher dietary intakes of folate, there might be no adverse effect on plasma homocysteine levels or subsequent risk of coronary heart disease. In addition to an association with CAD, *MTHFR* polymorphisms have been associated with arterial and venous thromboembolism [10,17], diabetes [12,35], neural tube defects [5,39], breast cancer [32,33], and lung cancer [31]. Possible protective effects of 677C-T have been shown against colon cancer [22,23].

A novel polymorphism of *MTHFR*, 1793G-A, was first described by Rady et al. [30] who reported polymorphism frequencies in US Caucasians, African-Americans, Hispanics, and Ashkenazi Jews. This polymorphism was found to have a lower allelic frequency in comparison to 677C-T and 1298A-C. It is unknown whether 1793G-A polymorphism causes a decrease in *MTHFR* enzyme activity, however, in a recent study of kidney transplant recipients, it was suggested that 1793G-A may have a stabilizing, rather than destabilizing, effect on *MTHFR* activity [43]. To the best of our knowledge, there have been no studies that have

looked at a possible association of this polymorphism with CAD. In the present study, we investigate the role of two *MTHFR* polymorphisms, 677C-T and 1793G-A in a population of 795 patients receiving coronary angiography in an attempt to examine a possible role in CAD.

2. Materials and methods

2.1. Study population

Consenting consecutive men and women ($n = 795$) undergoing coronary angiography at Veterans Affairs Medical Center (VAMC) and the University Hospital (Oklahoma City, OK) between November 1992 and March 1994, were enrolled in the study, as approved by the Institutional Review Board at The University of Oklahoma Health Sciences Center. The 795 patients included 604 Caucasian men, 125 Caucasian women, 51 African American men, and 15 African American women. Most patients were undergoing catheterization for angina; however, other indications included previous MI, aortic stenosis, aortic or mitral regurgitation, and atypical chest pain. Coronary angiography was performed using standard techniques. Measurements and views taken during the procedure have been described previously [7]. Severity of vessel obstruction was graded visually by two independent cardiologists in the view in which it was most severe. Other CAD risk factors were obtained by review of angiography reports and medical records.

2.2. Laboratory methods

Fifty-five milliliters of blood were collected at the time of coronary angiography through the arterial sheath prior to administration of heparin. Total cholesterol, HDL cholesterol, and triglycerides were measured on EDTA plasma samples using the Centers for Disease Control standardized protocols. Genomic DNA was isolated from blood by a standard proteinase K-phenol-chloroform extraction method. The presence of single nucleotide polymorphisms (SNP) in the 5,10-methylenetetrahydrofolate reductase (*MTHFR*) gene was determined at positions 677 (C-T) and 1793 (G-A) using a novel one-step Holliday junction-based allele-specific genotyping technology (*MTHFR* Polymorphism Genotyping Sample Kits, Cat. No. 100001 and 100005; FreshGene Inc., Concord, CA) described previously [45]. Briefly, each 10 μ L reaction contained

AmpliTaQ Gold™ (Perkin Elmer, Wellesley, MA), dNTPs, primers flanking the substitution sites, approximately 1 ng of DNA and the following amplification parameters performed in a Biometra T-Gradient Thermoblock (Whatman, Göttingen, Germany): 94°C initial denaturation then 45 cycles of 94°C denaturation for 15 sec, 58°C primer annealing for 23 sec, and 72°C extension for 45 sec, as per the manufacturers instructions. Two reactions were run per SNP for each patient sample (i.e., a C tube and a T tube for the 677C-T analysis and a G tube and an A tube for the 1793G-A analysis). For each SNP, a total of five primers were used: three short forward primers and two reverse primers. One forward primer is used to produce “target amplicons” in conjunction with the pair of reverse primers. The other two forward primers are designed to be complementary for either allele 1 or allele 2 of each SNP and to produce “reference amplicons” in conjunction with the reverse primers. Either a “normal” or “mutant” reference primer is placed in the two reactions. The forward primers and the two reverse primers are designed with complementary tails in order to encourage the formation of 4-stranded cruciform structures, or Holliday junctions (HJ) in subsequent steps. Following thermal cycling, PCR products were subject to denaturation at 95°C for 2 mins then a branch migration step for HJ formation at 62°C for 30 mins. During this process the allele-specific reference amplicons hybridize with the target amplicons to give HJ products depending on the presence of a complementary target (i.e., normal or mutant allele). The reference primers were constructed with 3' ends that introduced a single base mismatch before each SNP in order to impede branch migration and cause the formation of stable HJ products. In addition, reference primers had GC-clamps at the 5' ends to aid in HJ formation. HJ products were then analyzed by electrophoretic separation on 3% Super Fine Resolution (Amresco, Solon, Ohio) agarose or 6% pre-cast TBE PAGE (Invitrogen Inc., Carlsbad, CA) gels stained with SYBR-Gold (Molecular Probes, Eugene, OR). Each PCR run included a no-DNA control to monitor for carry-over contamination and a known SNP heterozygote to verify optimal performance of each reaction set. “Blind” processing of 24 duplicate samples for each polymorphism analyzed served as an internal control of genotype specificity and assay reproducibility. In addition, genotypes of 20 samples were verified independently by separate PCR-RFLP analysis, as previously described [9,30].

2.3. Statistical methods

For the purposes of description and analysis, we created two separate data sets; one containing all genotyped subjects (729 Caucasians and 66 African Americans) and the other with African Americans omitted to be used for logistic regression modeling. We performed chi-square tests for independence on the complete data set to determine the relationship between race and *MTHFR* polymorphism. For logistic regression analysis, the sample was further restricted due to incomplete data on some patients. Since African Americans represented a small portion of our overall patient group and moreover, because *MTHFR* allelic frequencies in this ethnic group were divergent from Caucasians, the final data set used for CAD analysis comprised 706 Caucasians. We also calculated mean homocysteine levels by *MTHFR* polymorphism on a reduced subset with available data ($n = 393$).

We generated crude and adjusted odds ratios to determine the risk for CAD associated with each *MTHFR* polymorphism. The adjusted logistic regression model included age, sex, diabetes, fibrinogen, triglycerides, non-HDLc, and smoking status as potential confounders of the relationship between *MTHFR* polymorphisms and CAD. Patients were dichotomized into those without disease (no stenosis equal or greater than 50%) and those with disease (at least one stenosis greater or equal to 50%). Lastly, the amount of vessel disease was also quantified for each patient by summing the values of all the identified stenoses; a term we refer to as the “atherosclerosis score”.

3. Results

3.1. Prevalence of 677C-T and 1793G-A polymorphisms

Of the 795 patients genotyped, there were 381 (47.9%) 677CC wild-types, 323 (40.6%) 677CT heterozygotes, and 91 (11.4%) homozygous 677TT mutants (Table 1). The 1793G-A polymorphism was less frequent than the 677C-T (Table 2); allelic frequency was 78/1590 (4.9%) for all patients. There were 717 patients (90.2%) with the 1793GG wild-type genotype and 78 (9.8%) 1793GA heterozygotes. There were no homozygous 1793AA mutant individuals detected in the cohort analyzed. There were 29 677CT/1793GA compound heterozygotes. The distribution of compound genotypes within the total patient data set is

Table 1
Frequency of 677C-T genotypes in coronary angiography patients by race and gender

	Caucasian n (column %)	African American n (column %)	All races n (column %)
Males and females			
Normal	326 (44.7%)	55 (83.3%)	381 (47.9%)
Heterozygotes	313 (42.9%)	10 (15.2%)	323 (40.6%)
Homozygotes	90 (12.3%)	1 (1.5%)	91 (11.4%)
Total (row%)	729 (91.7%)	66 (8.3%)	795
Chi-square test for independence, 36.5 _{2df} , $p < 0.001$			
Males			
Normal	262 (43.4%)	42 (82.3%)	304 (46.4%)
Heterozygotes	270 (44.7%)	8 (15.7%)	278 (42.4%)
Homozygotes	72 (11.9%)	1 (2.0%)	73 (11.1%)
Total (row%)	604 (92.2%)	51 (7.8%)	655
Chi-square test for independence, 28.9 _{2df} , $p < 0.001$			
Females			
Normal	64 (51.2%)	13 (86.7%)	77 (55%)
Heterozygotes	43 (34.4%)	2 (13.3%)	45 (32.1%)
Homozygotes	18 (14.4%)	0 (0.0%)	18 (12.9%)
Total (row%)	125 (89.3%)	15 (10.7%)	140
Chi-square test for independence, 7.1 _{2df} , $p = 0.029$			

Table 2
Frequency of 1793G-A genotypes in coronary angiography patients by race and gender

Patient	Caucasian n (column %)	African American n (column %)	All races n (column %)
Males and females			
Normal	652 (89.4%)	65 (98.5%)	717 (90.2%)
Heterozygotes	77 (10.6%)	1 (1.5%)	78 (9.8%)
Homozygotes	0 (0.0%)	0 (0.0%)	0 (0.0%)
Total (row %)	729 (91.7%)	66 (8.3%)	795
Chi-square test for independence, 5.6 _{1df} , $p = 0.018$			
Males			
Normal	542 (89.7%)	50 (98.0%)	592 (90.4%)
Heterozygotes	62 (10.3%)	1 (2.0%)	63 (9.6%)
Homozygotes	0	0	0
Total (row %)	604 (92.2%)	51 (7.8%)	655
Chi-square test for independence, 3.7 _{1df} , $p = 0.053$			
Female			
Normal	110 (88.0%)	15 (100%)	125 (89.3%)
Heterozygotes	15 (12%)	0	15 (10.3%)
Homozygotes	0	0	0
Total (row %)	125 (89.3%)	15 (10.7%)	140
Chi-square test for independence, 2.0 _{1df} , $p < 0.156$			

presented in Table 3. There were significantly fewer African Americans with either mutation as compared to Caucasians (Tables 1 and 2). Allelic frequencies for 677C-T were 66.2% C and 33.8% T for Caucasians and 90.9% C and 9.1% T for African Americans. With respect to the 1793G-A polymorphism, allelic frequencies were 94.7% G and 5.3% A for Caucasians and 99.2% G and 0.8% A for African Americans. The differences in 677C-T and 1793G-A frequencies between Caucasians and African Americans were statistically significant when considering the overall popula-

tion (Tables 1 and 2), but there were no significant differences in the frequencies of the 1793G-A polymorphism between these groups when considering males or females only.

3.2. 677C-T and 1793G-A polymorphisms and CAD

Logistic regression analysis of Caucasian patients produced an odds ratio indicating that 677CT heterozygotes do not have an increased risk of disease compared to 677CC individuals (Crude OR-1.36; 95% CI

Table 3

Distribution of compound genotypes in coronary angiography patients

n	677CC	677CT	677TT	Totals
1793 GG	332	294	91	717
1793 GA	49	29	0	78
1793 AA	0	0	0	0
Totals	381	323	91	795

0.95 to 1.96) (Table 4). Likewise, 677TT homozygous Caucasians do not have a reduced risk for disease when compared to normal 677CC individuals (Crude OR-0.73; 95% CI 0.44 to 1.20). Although point estimates diverged from 1.00, the 95% confidence intervals included 1.00 in both cases. Logistic regression analysis of the less frequent 1793G-A polymorphism produced an odds ratio for CAD that was not statistically significant as compared to the normal genotype (Crude OR-0.79; 95% CI 0.47 to 1.33). The results for both polymorphisms were consistent for crude odds ratios and those generated from a model including potential confounders: age, sex, diabetic status, smoking status, fibrinogen, lipid profile and triglycerides. Overall, *MTHFR* genotype did not demonstrate any statistically significant association with the presence of atherosclerosis using the clinical definition of one or more stenosis $\geq 50\%$ in any of the 3 major coronary arteries. Lastly, a descriptive table of possible confounding variables in this population was generated for completeness and future investigators (Table 5).

3.3. 677C-T and 1793G-A polymorphisms, CAD and homocysteine

Homocysteine levels were obtained on 393 patients. The mean Hcy levels ($\mu\text{mol/L}$) in our population included 677CC: 10.2; 677CT: 11.0; 677TT: 11.6; 1793GG: 10.7; 1793GA: 11.5 (Table 6). These levels increase in a mutation dose-dependent manner for both the 677C-T and the 1793G-A mutations.

4. Discussion

4.1. Prevalence of 677C-T and 1793G-A polymorphisms

The 677C-T genotype (CC-47.9%, CT-40.6%, TT-11.4%) and allelic frequencies (31.8%) for all races found in our predominately Caucasian population are similar to those reported for other populations [20, 37,38], including a recent meta-analysis [19]. The

allelic frequency amongst African Americans in our population (9.1%) was similar to those described for other African American populations [6,30,34]. Few studies have investigated the population prevalence of the 1793G-A polymorphism. The 1793G-A polymorphism had lower frequencies in our Caucasian patient population (GG-89.4%, GA-10.6%, AA-0%; allelic frequency- 5.3%) as compared to the 677C-T polymorphism. A recent Austrian study [44] of 1793G-A, with a similar patient sample size ($n = 733$) and likely Caucasian-enriched, had similar findings (GG-89.9%, GA-9.9%, AA-0.3%; allelic frequency- 5.2%). Rady et al. [30] studied the distribution of the 1793G-A polymorphism amongst several ethnic groups and found that allelic frequencies were highest amongst Caucasians (6.9%), lowest in Ashkenazi Jews (1.3%) and intermediate in Hispanics (5.8%) and African-Americans (3.1%). Allelic frequencies for the 1793G-A polymorphism in our population were slightly lower for Caucasians (5.3%) and substantially lower for African Americans (0.8%). Although other studies have documented homozygous 1793AA mutants (1/507 [30]; 2/733 [43]), this genotype was absent from our patient group despite the larger cohort size.

Compound 677CT/1793GA heterozygotes were present in 3.6% of our cohort (Table 3). It is unclear whether these mutated alleles existed in *cis* or *trans*; however, of the ninety-one 677TT homozygous mutants, none had a 1793G-A polymorphism, which suggests that mutated alleles generally reside in *trans* conformation. In addition, other studies have failed to document 677CT/1793AA genotype combinations [30]. Isotalo et al. [14] postulated that 677CT/1298CC and 677TT/1298CC genotypes, which contain three and four mutated alleles respectively, may compromise fetal viability especially during times of folate insufficiency and therefore would be very rare in the population. Clearly, evaluation of any potential synergistic effect of 677C-T, 1298A-C, 1793G-A in *cis* and *trans* will require study of a large cohort considering the low prevalence of compound heterozygotes and homozygous mutants in the general population.

4.2. 677C-T and 1793G-A polymorphisms and CAD

Numerous studies have been completed relating *MTHFR* polymorphisms to CAD with 677C-T being studied in most detail followed by 1298A-C. While many have shown risk associations [8,20,21,24,25,29, 36], others have not [1,4,18,27,37,46]. Unfortunately, we were unable to complete analysis of the 1298A-C

Table 4
Crude and adjusted* Odds Ratios for CAD by *MTHFR* genotype among Caucasian coronary angiography patients

n = 706	Adjusted odds ratio (95% CI)	Crude odds ratio (95% CI)
677 CC	1.0	1.0
677 CT	1.41 (0.93 to 2.12)	1.36 (0.95 to 1.96)
677 TT	0.69 (0.39 to 1.23)	0.73 (0.44 to 1.20)
Age (1 Year)	1.07 (1.04 to 1.09)	
Diabetes (Yes)	2.03 (1.25 to 3.29)	
Fibrinogen (1 mg/dL)	1.003 (1.002 to 1.005)	
NonHDLc (1 mg/dL)	1.004 (1.0001 to 1.008)	
Sex (Male)	4.44 (2.72 to 7.23)	
Smoker (Current v. never)	1.90 (1.02 to 3.53)	
Triglycerides (1 mg/dL)	1.003 (1.001 to 1.005)	
n = 706	Adjusted odds ratio (95% CI)	Crude odds ratio (95% CI)
1793 GG	1.0	1.0
1793 GA	0.80 (0.44 to 1.44)	0.79 (0.47 to 1.33)
Age (1 Year)	1.07 (1.04 to 1.09)	
Diabetes (Yes)	2.03 (1.25 to 3.29)	
Fibrinogen (1 mg/dL)	1.003 (1.002 to 1.005)	
NonHDLc (1 mg/dL)	1.004 (1.0005 to 1.008)	
Sex (Male)	4.45 (2.80 to 7.41)	
Smoker (Current v. never)	1.92 (1.03 to 3.56)	
Triglycerides (1 mg/dL)	1.003 (1.001 to 1.005)	

* Adjusted for age, diabetes, fibrinogen, nonHDLc, gender, smoking, and triglycerides using logistic regression.

Table 5

Descriptive statistics for possible continuous confounding factors (mean and standard error) and possible categorical confounding factors (%) for CAD and *MTHFR* genotype in Caucasian coronary angiography patients (n = 706)

Continuous variables	677CC		677CT		677TT		1793GG		1793GA	
mean (s.e.)	CAD	No CAD	CAD	No CAD	CAD	No CAD	CAD	No CAD	CAD	No CAD
Age (years)	60.56 (0.65)	54.39 (1.04)	59.74 (0.64)	53.13 (1.18)	58.43 (1.30)	57.40 (1.81)	59.92 (0.45)	54.70 (0.77)	60.31 (1.38)	52.13 (2.07)
Number of stenoses (Count)	6.64 (0.18)	1.39 (0.29)	6.71 (0.18)	1.34 (0.33)	5.74 (0.36)	1.83 (0.51)	6.47 (0.13)	1.41 (0.21)	7.56 (0.38)	1.65 (0.58)
Atherosclerosis score*	405.09 (12.51)	34.00 (20.12)	404.58 (12.32)	33.73 (22.67)	350.09 (25.07)	46.67 (34.86)	393.13 (8.71)	34.61 (14.71)	451.54 (26.44)	45.22 (39.75)
Total cholesterol (mg/dL)	225.75 (3.82)	213.21 (6.15)	222.28 (3.77)	209.27 (6.93)	226.81 (7.67)	217.77 (10.66)	224.85 (2.66)	211.96 (4.50)	219.15 (8.09)	216.13 (12.16)
NonHDLc (mg/dL)	194.47 (3.82)	179.47 (6.15)	190.68 (3.77)	174.66 (6.92)	195.38 (7.66)	183.17 (10.65)	193.39 (2.66)	178.48 (4.50)	187.92 (8.08)	176.70 (12.15)
Triglycerides (mg/dL)	202.07 (9.94)	179.06 (15.99)	198.08 (9.79)	155.61 (18.00)	241.71 (19.91)	182.83 (27.69)	203.91 (6.94)	173.38 (11.72)	210.92 (21.07)	153.09 (31.68)
Fibrinogen (mg/dL)	385.34 (8.72)	333.02 (14.04)	376.15 (8.64)	331.10 (15.95)	366.59 (17.70)	340.33 (24.18)	375.87 (6.08)	339.48 (10.26)	409.23 (18.28)	290.43 (27.49)
Categorical variables %										
Diabetes	31.6	22.2	29.6	8.7	34.5	26.7	31.0	18.1	30.8	17.4
Current smokers	48.9	48.9	56.3	54.9	44.8	56.7	50.1	52.4	67.3	52.2

* Atherosclerosis score = Sum of blockages in coronary arteries.

polymorphism for this cohort due to technical problems encountered during the design of primers for this specific Holliday junction assay. Moreover, due to the constraints imposed on analysis by the low numbers of African Americans included in the cohort, the present study is limited to investigation of possible associations between CAD and two polymorphisms of *MTHFR*, 677C-T and 1793G-A, in Caucasians only. To the au-

thors' knowledge, this is the first study to look at the association between 1793G-A and CAD.

Surprisingly, although our results suggest that 677CT heterozygotes have a slightly increased (albeit not significant) risk of disease compared to 677CC individuals (Crude OR-1.36; 95% CI 0.95 to 1.96), the mutant TT population did not (Crude OR-0.73; 95% CI 0.44 to 1.20, respectively). Clearly, one would expect

Table 6

Homocysteine levels associated with 677C-T and 1793G-A genotypes in Caucasian coronary angiography patients (n = 393)

mean (s.e.)	Homocysteine $\mu\text{mol/L}$
677 CC	10.2 (0.4)
677 CT	11.0 (0.4)
677 TT	11.6 (0.7)
1793 GG	10.7 (0.3)
1793 GA	11.5 (0.7)

to see a mutation dose-related response if indeed the 677C-T polymorphism plays a significant role in predisposition to CAD. The 1793GA population demonstrated a statistically non-significant protective association with CAD compared to the 1793GG patients (Crude OR-0.79; 95% CI 0.47 to 1.33).

A recent meta-analysis has provided convincing evidence that low folate status, resulting in high homocysteine levels, causes an increased risk for CAD in 677C-T mutants [19]. In this study, European patients with the mutant 677TT genotype had an OR of 1.14. Conversely, North American subjects, who were expected to be well nourished and with higher dietary folate and vitamin usage than that of the Europeans, were slightly protected by the mutation and had an OR of 0.87. In 1996, the US Food and Drug Administration permitted voluntary fortification of enriched cereal and grain products, and in January 1998 required that all enriched grain products contain 140 mcg of folic acid per 100 g. Therefore, our patient population may not have benefited from dietary folate supplementation since they underwent coronary angiography between November 1992 and March 1994 and had their blood samples drawn at that time for the laboratory values indicated in this paper. In this sense, our cohort probably has a folate status resembling that of European patients. However, the mutant 677TT genotype was associated with a crude OR of 0.73 in our population and an OR of 1.14 in the meta-analysis of Europeans [19]. Unfortunately, the initial design of the study of our patient population in relation to CAD did not anticipate investigation of markers related to *MTHFR* metabolism, so we do not have folate levels gathered on these patients. However, considering the fact that these patients were studied at a time prior to folate supplementation, it is likely that their dietary folate intake was sub-optimal and their homocysteine levels accordingly elevated. Indeed, we found an increasing trend of homocysteine levels: 10.2, 11.0, 11.6 $\mu\text{mol/L}$ in 677CC, 677CT and 677TT individuals, respectively (Table 6). An increasing trend was also seen in our 1793G-A population: 10.7 $\mu\text{mol/L}$ for 1793GG and 11.5 $\mu\text{mol/L}$ for 1793GA.

While functional studies have confirmed the effects of 677C-T and 1298A-C polymorphisms on *MTHFR* thermolability, similar studies have not been performed on *MTHFR* containing the 1793G-A mutation. If the 1793G-A polymorphism were to cause enzyme instability and decreased activity, we would expect homocysteine levels to be higher, especially in those with low folate status. Although homocysteine levels demonstrated a dose-response for both mutations, these values are probably too low to have independently contributed to any significant cardiovascular effect.

Our findings confirm previous studies that the 677C-T polymorphism probably contributes only minimally, if at all, to an increased risk for CAD in Caucasians. The *MTHFR* 1793G-A polymorphism likely contributes no risk for CAD in Caucasians.

Acknowledgements

Patient enrollment and collection and processing of specimens were supported by Oklahoma Center for the Advancement of Science and Technology HS2-025 and American Heart Association Oklahoma Affiliate 93078550 (JEE). *MTHFR* genotyping was made possible by funds from the Department of Pathology Research Studentship (CK and SV) and by the Clarence and Matilda Bower Memorial Fund.

References

- [1] K.K. Abu-Amro, C.A. Wyngaard and N. Dzimiri, Prevalence and role of methylenetetrahydrofolate reductase 677 C→T and 1298 A→C polymorphisms in coronary artery disease in Arabs, *Archives of Pathology & Laboratory Medicine* **127** (2003), 1349–1352.
- [2] C.J. Boushey, S.A. Beresford, G.S. Omenn and A.G. Motulsky, A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes, *Journal of the American Medical Association* **274** (1995), 1049–1057.
- [3] L. Brattstrom and D.E. Wilcken, Homocysteine and cardiovascular disease: cause or effect? *The American Journal of Clinical Nutrition* **72** (2000), 315–323.
- [4] E.S. Brilakis, P.B. Berger, K.V. Ballman and R. Rozen, Methylenetetrahydrofolate reductase (*MTHFR*) 677C>T and methionine synthase reductase (*MTRR*) 66A>G polymorphisms: association with serum homocysteine and angiographic coronary artery disease in the era of flour products fortified with folic acid, *Atherosclerosis* **168** (2003), 315–322.
- [5] B. Christensen, L. Arbour, P. Tran, D. Leclerc, N. Sabbaghian, R. Platt, B.M. Gilfix, D.S. Rosenblatt, R.A. Gravel, P. Forbes and R. Rozen, Genetic polymorphisms in methylenetetrahydrofolate reductase and methionine synthase, folate levels in red blood cells, and risk of neural tube defects, *American Journal of Human Genetics* **84** (1999), 151–157.

- [6] A. Dilley, W.C. Hooper, M. El-Jamil, M. Renshaw, N.K. Wenger and B.L. Evatt, Mutations in the genes regulating methylene tetrahydrofolate reductase (*MTHFR* C→T677) and cystathione beta-synthase (CBS G→A919, CBS T→C833) are not associated with myocardial infarction in African Americans, *Thrombosis Research* **103** (2001), 109–115.
- [7] J.E. Eichner, V.J. Christiansen, W.E. Moore, S.T. Dunn and E. Schechter, Angiotensin-converting enzyme gene polymorphism in a cohort of coronary angiography patients, *Atherosclerosis* **154** (2001), 673–679.
- [8] O. Fletcher and A.M. Kessling, *MTHFR* association with arteriosclerotic vascular disease? *Human Genetics* **103** (1998), 11–21.
- [9] P. Frosst, H.J. Blom, R. Milos, P. Goyette, C.A. Sheppard, R.G. Matthews, G.J.H. Boers, M. den Heijer, L.A.J. Kluijtmans, L.P. van den Heuvel and R. Rozen, A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase, *Nature Genetics* **10** (1995), 111–113.
- [10] D. Gemmati, M.L. Serino, C. Trivellato, S. Fiorini and G.L. Scapoli, C677T substitution in the methylenetetrahydrofolate reductase gene as a risk factor for venous thrombosis and arterial disease in selected patients, *Haematologica* **84** (1999), 824–828.
- [11] D. Girelli, N. Martinelli, F. Pizzolo, S. Friso, O. Olivieri, C. Stranieri, E. Trabetti, G. Faccini, E. Tinazzi, P.F. Pignatti and R. Corrocher, The interaction between *MTHFR* 677 C→T genotype and folate status is a determinant of coronary atherosclerosis risk, *The Journal of Nutrition* **133** (2003), 1281–1285.
- [12] G. Hasegawa, H. Obayashi, K. Kamiuchi, M. Nakai, T. Kanatsuna, M. Yamaguchi, T. Tanaka, H. Shigeta, M. Fujii, T. Yoshikawa and N. Nakamura, The association between end-stage diabetic nephropathy and methylenetetrahydrofolate reductase genotype with macroangiopathy in type 2 diabetes mellitus, *Experimental and Clinical Endocrinology & Diabetes* **111** (2003), 132–138.
- [13] H.J. Huh, H.S. Chi, E.H. Shim, S. Jang and C.J. Park, Gene-nutrition interactions in coronary artery disease: Correlation between the *MTHFR* C677T polymorphism and folate and homocysteine status in a Korean population, *Thrombosis Research* (2005), Epub ahead of print.
- [14] P.A. Isotalo, G.A. Wells and J.G. Donnelly, Neonatal and fetal methylenetetrahydrofolate reductase genetic polymorphisms: an examination of C677T and A1298C mutations, *American Journal of Human Genetics* **67** (2000), 986–990.
- [15] S.-S. Kang, J. Zhou, P.W.K. Wong, J. Kowalisyn and G. Strokosch, Intermediate homocysteinemia: a thermolabile variant of methylenetetrahydrofolate reductase, *American Journal of Human Genetics* **43** (1988), 414–421.
- [16] S.-S. Kang, P.W.K. Wong, A. Susmano, J. Sora, M. Norusis and N. Ruggie, Thermolabile methylenetetrahydrofolate reductase: an inherited risk factor for coronary artery disease, *American Journal of Human Genetics* **48** (1991), 536–545.
- [17] P.J. Kelly, J. Rosand, J.P. Kistler, V.E. Shih, S. Silveira, A. Plo-maritoglou and K.L. Furie, Homocysteine, *MTHFR* 677C→T polymorphism, and risk of ischemic stroke: results of a meta-analysis, *Neurology* **59** (2002), 529–536.
- [18] C.H. Kim, K.Y. Hwang, T.M. Choi, W.Y. Shin and S.Y. Hong, The methylenetetrahydrofolate reductase gene polymorphism in Koreans with coronary artery disease, *International Journal of Cardiology* **78** (2001), 13–17.
- [19] M. Klerk, P. Verhoef, R. Clarke, H.J. Blom, F.J. Kok and E.G. Schouten, *MTHFR* Studies Collaboration Group. *MTHFR* 677C→T polymorphism and risk of coronary heart disease: a meta-analysis, *Journal of the American Medical Association* **288** (2002), 2023–2031.
- [20] L.A. Kluijtmans, J.J. Kastelein, J. Lindemans, G.H. Boers, S.G. Heil, A.V. Bruschke, J.W. Jukema, L.P. van den Heuvel, F.J. Trijbels, G.J. Boerma, F.W. Verheugt, F. Willems and H.J. Blom, Thermolabile methylenetetrahydrofolate reductase in coronary artery disease, *Circulation* **96** (1997), 2573–2577.
- [21] L.A. Kluijtmans, L.P. van den Heuvel, G.H. Boers, P. Frosst, E.M. Stevens, B.A. van Oost, M. den Heijer, F.J. Trijbels, R. Rozen and H.J. Blom, Molecular genetic analysis in mild hyperhomocysteinemia: a common mutation in the methylenetetrahydrofolate reductase gene is a genetic risk factor for cardiovascular disease, *American Journal of Human Genetics* **58** (1996), 35–41.
- [22] L. Le Marchand, L.R. Wilkens, L.N. Kolonel and B.E. Henderson, The *MTHFR* C677T polymorphism and colorectal cancer: the multiethnic cohort study, *Cancer Epidemiology, Biomarkers, & Prevention* **14** (2005), 1198–1203.
- [23] J. Ma, M.J. Stampfer, E. Giovannucci, C. Artigas, D.J. Hunter, C. Fuchs, W.C. Willet, J. Selhub, C.H. Hennekens and R. Rozen, Methylenetetrahydrofolate reductase polymorphism, dietary interactions, and risk of colorectal cancer, *Cancer Research* **57** (1997), 1098–1102.
- [24] A. Mager, S. Lalezari, T. Shohat, Y. Birnbaum, Y. Adler, N. Magal and M. Shohat, Methylenetetrahydrofolate reductase genotypes and early-onset coronary artery disease, *Circulation* **100** (1999), 2406–2410.
- [25] H. Morita, J. Taguchi, H. Kurihara, M. Kitaoka, H. Kaneda, Y. Kurihara, K. Maemura, T. Shindo, T. Minamino, M. Ohno, K. Yamaoki, K. Ogasawara, T. Aizawa, S. Suzuki and Y. Yazaki, Genetic polymorphism of 5,10-methylenetetrahydrofolate reductase (*MTHFR*) as a risk factor for coronary artery disease, *Circulation* **95** (1997), 2032–2036.
- [26] A.G. Motulsky, Nutritional ecogenetics: homocysteine-related arteriosclerotic vascular disease, neural tube defects, and folic acid, *American Journal of Human Genetics* **58** (1996), 17–20.
- [27] K. Nakai, C. Itoh, K. Nakai, W. Habano and D. Gurwitz, Correlation between C677T *MTHFR* gene polymorphism, plasma homocysteine levels and the incidence of CAD, *American Journal of Cardiovascular Drugs: Drugs, Devices, and Other Interventions* **1** (2001), 353–361.
- [28] O. Nygard, S.E. Vollset, H. Refsum, L. Brattstrom and P.M. Ueland, Total homocysteine and cardiovascular disease, *Journal of Internal Medicine* **246** (1999), 425–454.
- [29] D.A. Payne, A.J. Chamoun, S.I. Seifert and G.A. Stouffer, *MTHFR* 677C→T mutation: a predictor of early-onset coronary artery disease, *Thrombosis Research* **103** (2001), 275–279.
- [30] P.L. Rady, S. Szucs, J. Grady, S.D. Hudnall, L.H. Kellner, H. Nitowsky, S.K. Tying and R.K. Matalon, Genetic polymorphisms of methylenetetrahydrofolate reductase (*MTHFR*) and methionine synthase reductase (MTRR) in ethnic populations in Texas; a report of a novel *MTHFR* polymorphic site, G1793A, *American Journal of Human Genetics* **107** (2002), 162–168.
- [31] Q. Shi, Z. Zhang, G. Li, P.C. Pillow, L.M. Hernandez, M.R. Spitz and Q. Wei, Sex differences in risk of lung cancer associated with methylene-tetrahydrofolate reductase polymorphisms, *Cancer Epidemiology, Biomarkers & Prevention* **14** (2005), 1477–1484.
- [32] M.J. Shrubsole, X.O. Shu, Z.X. Ruan, Q. Cai, H. Cai, Q. Niu, Y.T. Gao and W. Zheng, *MTHFR* genotypes and breast cancer

- survival after surgery and chemotherapy: a report from the Shanghai Breast Cancer Study, *Breast Cancer Research and Treatment* **91** (2005), 73–79.
- [33] M.J. Shrubsole, Y.T. Gao, Q. Cai, X.O. Shu, Q. Dai, J.R. Hebert, F. Jin and W. Zheng, *MTHFR* polymorphisms, dietary folate intake, and breast cancer risk: results from the Shanghai Breast Cancer Study, *Cancer Epidemiology, Biomarkers & Prevention* **13** (2004), 190–196.
- [34] R.E. Stevenson, C.E. Schwartz, Y.Z. Du and M.J. Adams Jr, Differences in methylenetetrahydrofolate reductase genotype frequencies, between Whites and Blacks, *American Journal of Human Genetics* **60** (1997), 229–230.
- [35] J. Sun, Y. Xu, Y. Zhu and H. Lu, Genetic polymorphism of methylenetetrahydrofolate reductase as a risk factor for diabetic nephropathy in Chinese type 2 diabetic patients, *Diabetes Research and Clinical Practice* **64** (2004), 185–190.
- [36] A. Szczeklik, M. Sanak, M. Jankowski, J. Dropinski, R. Czachor, J. Musial, I. Axenti, M. Twardowska, T. Brzostek and M. Tendera, Mutation A1298C of methylenetetrahydrofolate reductase: risk for early coronary disease not associated with hyperhomocysteinemia, *American Journal of Human Genetics* **101** (2001), 36–39.
- [37] F.M. van Bockxmeer, C.D. Mamotte, S.D. Vasikaran and R.R. Taylor, Methylenetetrahydrofolate reductase gene and coronary artery disease, *Circulation* **95** (1997), 21–23.
- [38] N.M. van der Put, F. Gabreels, E.M. Stevens, J.A. Smeitink, F.J. Trijbels, T.K. Eskes, L.P. van den Heuvel and H.J. Blom, A second common mutation in the methylenetetrahydrofolate reductase gene: an additional risk factor for neural-tube defects? *American Journal of Human Genetics* **62** (1998), 1044–1051.
- [39] N.M.J. van der Put, T.K.A.B. Eskes and H.J. Blom, Is the common 677C→T mutation in the methylenetetrahydrofolate reductase gene a risk factor for neural tube defects? A meta-analysis, *The Quarterly Journal of Medicine* **90** (1997), 111–115.
- [40] S. Voutilainen, T.H. Rissanen, J. Virtanen, T.A. Lakka and J.T. Salonen, Kuopio Ischemic Heart Disease Risk Factor Study. Low dietary folate intake is associated with an excess incidence of acute coronary events: The Kuopio Ischemic Heart Disease Risk Factor Study, *Circulation* **103** (2001), 2674–2680.
- [41] I. Weisberg, P. Tan, B. Christensen, S. Sibani and R. Rozen, A second genetic polymorphism in methylenetetrahydrofolate reductase (*MTHFR*) associated with decrease enzyme activity, *Molecular Genetics and Metabolism* **64** (1998), 169–172.
- [42] I.S. Weisberg, P.F. Jacques, J. Selhub, A.G. Bostom, Z. Chen, R. Curtis Ellison, J.H. Eckfeldt and R. Rozen, The 1298A→C polymorphism in methylenetetrahydrofolate reductase (*MTHFR*): *in vitro* expression and association with homocysteine, *Atherosclerosis* **156** (2001), 409–415.
- [43] W.C. Winkelmayr, A. Huber, O.F. Wagner, W.H. Horl, G. Sunder-Plassmann and M. Fodinger, Associations between *MTHFR* 1793G>A and plasma total homocysteine, folate, and vitamin B in kidney transplant recipients, *Kidney International* **67** (2005), 1980–1985.
- [44] W.C. Winkelmayr, G. Sunder-Plassmann, A. Huber and M. Fodinger, Patterns of co-occurrence of three single nucleotide polymorphisms of the 5,10-methylenetetrahydrofolate reductase gene in kidney transplant recipients, *European Journal of Clinical Investigation* **34** (2004), 613–618.
- [45] Q. Yang, A. Lishanski, W. Yang, S. Hatcher, H. Seet and J. Gregg, Allele-specific Holliday junction formation – a new mechanism of allelic discrimination for SNP scoring, *Genome Research* **13** (2003), 1754–1764.
- [46] H. Yilmaz, S. Isbir, B. Agachan, A. Ergen, B. Farsak and T. Isbir, C677T mutation of methylenetetrahydrofolate reductase gene and serum homocysteine levels in Turkish patients with coronary artery disease, *Cell Biochemistry and Function* (2005), Epub ahead of print.