Markers of angiogenesis in ovarian cancer

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Abstract. Tumor development and progression are inherently dependent on the process of angiogenesis. Recently, anti-angiogenic therapy has started to show promise as an effective treatment strategy in many solid tumors including ovarian carcinoma. Unfortunately, lack of effective biomarkers presents a challenge for oncologists in treatment planning as well as monitoring response of new anti-vascular agents. Previously, quantification of angiogenesis by microvessel density analysis provided useful prognostic information, however, its utility following anti-angiogenic therapy remains to be determined. Moreover, since secreted cytokines play an active part in angiogenesis by mediating neovascularization in tumors, investigations have focused on their potential role to serve as candidate biomarkers of disease detection, prognosis, and treatment response. In this article, we review the role of key angiogenesis markers as potential biomarkers in ovarian carcinoma.

Keywords: Angiogenesis, biomarker, ovarian carcinoma, therapy

1. Introduction

Tumor growth and metastasis are inherently dependent on the development of a blood supply or neovascularization. Angiogenic processes must be activated for tumor growth beyond 1 mm [33]. These processes include a shift in balance toward greater levels of pro-angiogenic compared to anti-angiogenic factors (Table 1). During angiogenesis, tumors utilize the host’s cellular machinery to develop an adequate vascular supply which is dependent upon the presence of activated endothelial cells. Multiple angiogenic activators play a role in initiating endothelial cell proliferation, migration, and survival [32,69,86,87]. Collectively, these components lead to the formation of new vascular channels which deliver oxygen and nutrients to the tumor beds.

The functional and architectural characteristics of tumor blood vessels are quite different in comparison to normal vessels. For example, tumor vessels are tortuous, highly permeable and irregularly shaped compared to normal vasculature [14]. The formation of tumor blood vessels is complex and likely involves multiple pathways. Angiogenesis can occur from "sprouting" or intussusceptive growth from pre-existing vessels [19,100]. Non-sprouting angiogenesis results from enlargement, splitting and fusion of pre-existing vessels. There is growing evidence that the initial events in tumor vascularization likely involve cooption of existing vessels by tumor cells [49] followed by production of factors such as Angiopoietin-2 that destabilize the host vasculature resulting in central tumor necrosis. In this setting, angiogenesis occurs secondarily in the tumor periphery as a result of increased production of angiogenic factors. Additional mechanisms of tumor neovascularization include vasculogenesis, which is the formation of new blood vessels from precursor mesodermal cells mobilized from the bone marrow [76, 97]. Hendrix and colleagues have described the plasticity of tumor cells whereby aggressive tumor cells adopt molecular features that are similar to endothelial cells (i.e., vasculogenic mimicry) [79,105–107]. This intriguing pathway suggests that aggressive tumor cells...
Table 1
Regulators of angiogenesis

<table>
<thead>
<tr>
<th>Activators</th>
<th>Inhibitors</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vascular endothelial growth factor (VEGF)</td>
<td>Thrombospondin</td>
</tr>
<tr>
<td>Fibroblast growth factor, acidic and basic (FGF)</td>
<td>Angiostatin</td>
</tr>
<tr>
<td>Transforming growth factor-beta (TGF-β)</td>
<td>Endostatin</td>
</tr>
<tr>
<td>Epidermal growth factor (EGF)</td>
<td>N-terminal prolactin fragments</td>
</tr>
<tr>
<td>Platelet derived growth factor (PDGF)</td>
<td>Interferon-alpha (INF-α)</td>
</tr>
<tr>
<td>Tumor necrosis factor-α (TNF-α)</td>
<td>Interleukin-12 (IL-12)</td>
</tr>
<tr>
<td>Interleukin-8 (IL-8)</td>
<td>Vasostatin</td>
</tr>
<tr>
<td>Interleukin-6 (IL-6)</td>
<td>Growth hormone</td>
</tr>
<tr>
<td>Angiopoietin 1,2 (Ang1, Ang2)</td>
<td>Dopamine</td>
</tr>
<tr>
<td>Cyclooxygenase-2 (COX-2)</td>
<td></td>
</tr>
<tr>
<td>Catecholamines</td>
<td></td>
</tr>
<tr>
<td>Hypoxia inducible factor-α (HIF-1α)</td>
<td></td>
</tr>
<tr>
<td>Matrix metalloproteinases (MMPs)</td>
<td></td>
</tr>
<tr>
<td>Ephrins/ Eph receptors</td>
<td></td>
</tr>
<tr>
<td>Prolactin (PRL)</td>
<td></td>
</tr>
<tr>
<td>Angiogenin</td>
<td></td>
</tr>
</tbody>
</table>

may have the ability to directly participate in the development of tumor vasculature.

Anti-angiogenic approaches are starting to show promise in pre-clinical and clinical investigations across multiple tumor types including ovarian carcinoma [18,54]. Bevacizumab was the first anti-vascular agent to receive approval from the Food & Drug Administration (FDA) for clinical use when given in combination with chemotherapy based on results from a phase III trial showing a 4.7 month improvement in overall survival in previously untreated, metastatic colorectal cancer patients [52]. We have previously reported the benefits of developing agents that target specific components of the vascular system and their potential role in ovarian cancer therapy [58]. Furthermore, we have shown in pre-clinical models that targeting genes responsible for angiogenesis with novel therapeutic strategies, such as siRNA targeted therapy, has therapeutic efficacy and these approaches are being developed clinically [65,66]. Traditional biomarkers may not be optimal for following patients on anti-angiogenic therapies. Based on the growing portfolio of anti-angiogenic approaches and the role of angiogenesis in affecting the course of malignant disease, we will review the predictive and prognostic relevance of tumor neovascularization and associated biomarkers in ovarian carcinoma.

2. Quantitative angiogenesis

To the extent that angiogenesis is related to tumor growth, it was hypothesized that quantification of angiogenesis may be useful as a predictive factor. Initial studies focused on assessing the density of blood vessels (MVD) as a marker for angiogenesis in human tumors [16,74,118]. This method utilizes immunohistochemical (IHC) staining of endothelial-specific markers such as CD34, CD31, or factor VIII-related antigen. The number of blood vessels is then counted in multiple regions of the tumor as a measure of angiogenesis.

Increased vascularity and MVD are poor prognostic factors in most human malignancies [96]. Specifically in ovarian carcinoma, several studies have examined the utility of MVD as a prognostic factor [4,73,108]. Hollingsworth and colleagues were one of the first investigators that utilized CD34 staining and found an inverse relationship between MVD counts and both disease free and overall survival [50]. In contrast, Abulafia reported no correlation between overall survival and MVD analysis in primary ovarian tumors, however MVD of omental metastases from 19 patients was an independent prognostic factor for patient survival [2]. More recently, larger studies have shown not only an inverse relationship between MVD and overall patient survival, but also that increased vascularity related to higher stage, higher grade, and the lower likelihood of optimal tumor reductive surgery [4,108]. Table 2 summarizes these and additional studies correlating MVD in patients with ovarian carcinoma. Collectively, these results suggest that MVD may be a clinically useful application for prognosis and assist in treatment planning, especially in patients with low-stage ovarian cancer.

In pre-clinical studies, MVD analysis has been an effective tool to assess the effects of investigational drugs and treatment regimens in animal models [65]. IHC staining of endothelial markers allows investigators to analyze the angiogenic effects in tumors fol-
Table 2
Summary of tumor MVD analyses in patients with ovarian carcinoma

<table>
<thead>
<tr>
<th>Author</th>
<th>Year</th>
<th>Number of Patients</th>
<th>Method</th>
<th>Associated Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hollingsworth et al. [50]</td>
<td>1995</td>
<td>45</td>
<td>IHC–CD34</td>
<td>OS and DFS</td>
</tr>
<tr>
<td>Abulafia et al. [2]</td>
<td>1997</td>
<td>49 (primary) 19 (omenta-</td>
<td>IHC – factor VIII related antigen</td>
<td>OS (only in MVD analysis in omen-</td>
</tr>
<tr>
<td>Heimburg et al. [46]</td>
<td>1999</td>
<td>38</td>
<td>IHC–vWF &amp; CD31</td>
<td>OS and stage</td>
</tr>
<tr>
<td>Obermaier et al. [90]</td>
<td>1999</td>
<td>63</td>
<td>IHC–CD34</td>
<td>OS</td>
</tr>
<tr>
<td>Stone et al. [108]</td>
<td>2003</td>
<td>202</td>
<td>IHC–CD31</td>
<td>OS, stage, grade, and potential for level of cytoreduction</td>
</tr>
<tr>
<td>Chan et al. [21]</td>
<td>2004</td>
<td>46</td>
<td>IHC–CD34</td>
<td>Age and OS</td>
</tr>
<tr>
<td>Raspolini et al. [98]</td>
<td>2005</td>
<td>33</td>
<td>IHC – CD34*</td>
<td>OS and response to therapy</td>
</tr>
<tr>
<td>Karavasilis et al. [59]</td>
<td>2006</td>
<td>33</td>
<td>IHC–CD34</td>
<td>No association</td>
</tr>
<tr>
<td>Lin et al. [73]</td>
<td>2006</td>
<td>77</td>
<td>IHC – CD31</td>
<td>Stage, ascites, potential for level of cytoreduction</td>
</tr>
</tbody>
</table>

IHC- immunohistochemistry; OS – overall survival; DFS – disease-free survival; vWF – von Willibrand factor; *IHC analyzed by computer-aided imaging.

However, there is a sustained increase in growth factor levels in tumors resulting in progressive neovascularization and tumor growth. One of the more prominent angiogenic mediators for new vessel growth in tumors is vascular endothelial growth factor (VEGF), also known as vascular permeability factor (VPF). VEGF overexpression results in increased endothelial cell proliferation, capillary tube formation, and vascular permeability [81,123]. Moreover, VEGF is an important survival factor for endothelial cells [36]. For these reasons, VEGF may be both a potential biomarker as well as a target for therapy. Indeed, VEGF targeted therapy has demonstrated promising results as one of the first clinically available anti-angiogenic agents in cancer therapy [52,54].

The VEGF gene encodes for several members, but VEGFA (commonly noted as VEGF) appears to play a dominant role in angiogenesis. There are four different isoforms of VEGF that occur in humans through alternative RNA splicing: VEGF121, VEGF165, VEGF189, and VEGF206. The release of VEGF by normal and tumor cells is controlled by several mechanisms. VEGF secretion, in addition to other growth factors including EGF, platelet derived growth factor (PDGF), transforming growth factor alpha and beta (TGFα & β), and fibroblast growth factor (FGF), are upregulated by stressful conditions, such as hypoxia, a common finding in solid tumors [30]. The presence of the Ras oncogene [40] and tumors with mutations of the p53 tumor suppressor gene [122] also result in overexpression of VEGF. Lastly, inflammatory cytokines, such as interleukin-1 (IL-1) and interleukin-6 (IL-6), released by tumor cells and the host’s immune system stimulate growth factor secretion [84]. In normal tissue, inactivation of VEGF is mediated by the von Hippel Lindau
Advanced stage disease may provide useful information in a subset of patients with ovarian carcinoma [101]. Markers may be a promising marker of disease presence. Ovarian carcinomas secrete CA-125, serum VEGF measurements may be a formidable setting to depend on angiogenesis as a predictor of disease burden, early stage disease may not be a formidable setting to depend on angiogenesis as a screening tool. However, since only 70–80% of ovarian carcinomas secrete CA-125, serum VEGF measurements may be a promising marker of disease presence in a subset of patients with ovarian carcinoma [101].

The overexpression of VEGF in patients with advanced stage disease may provide useful information as a prognostic factor for oncologists. In patients with ovarian cancer, VEGF overexpression correlated with advanced stage disease, ascites, and decreased overall survival [22,114]. Furthermore, Oehler and colleagues reported that serum VEGF levels significantly decreased following cytoreductive surgery and were also decreased in patients with low residual disease [91]. However, in multivariate analysis, they indicated that serum VEGF levels were not predictive of patient survival [91]. Moreover, Alvarez and colleagues reported that serum VEGF levels were not predictive of disease recurrence in a small subset of patients with a positive second look surgery [5]. Interestingly, when combined with MVD analyses or cyclooxygenase-2 (COX-2) expression, VEGF expression was predictive of disease free interval and chemotherapy response [99,121]. Based on these findings, quantifying different combinations of angiogenic factors, including VEGF, could assist in deciding which patients may benefit from more aggressive adjuvant treatment regimens.

Growth factors have become highly attractive targets for anti-angiogenic therapy. Pre-clinical research has demonstrated that inhibition of specific growth factors can effectively reduce tumor growth of human ovarian cell lines in murine models when used alone and in combination with chemotherapy [6,17,112]. Currently there are no effective surrogate markers for anti-vascular therapy. Bevacizumab, a humanized monoclonal antibody against VEGF, is one of the few approved anti-angiogenic therapeutic agents for cancer therapy and is currently being studied in ovarian cancer clinical trials (Fig. 1). Previously, bevacizumab therapy has shown benefit in patient survival in phase III colorectal and breast trials [54]. In these studies, plasma VEGF levels actually increased thereby questioning the value of VEGF as a surrogate marker following anti-VEGF therapy [54,119].

Anti-angiogenic agents that target VEGF activity via receptor inhibition may also help investigators identify new potential biomarkers in cancer therapy. Recently, Ebos and colleagues identified a soluble form of VEGFR-2 (sVEGFR-2) that binds to VEGF [26]. Measurable levels of sVEGFR-2 were found in the plasma of both mice and humans [26]. While it was proposed that ligand binding to sVEGFR-2 may result in anti-angiogenic activity, as seen with the soluble form of VEGFR-1, the actual biological role has not been elucidated. Based on these data, Motzer and colleagues investigated the benefit of using sVEGFR-2 as a biomarker in a recent phase I trial [83]. In that study, patients with metastatic renal cell carcinoma were treated with
Table 3

Summary of VEGF measurements for disease detection in patients with ovarian carcinoma

<table>
<thead>
<tr>
<th>Author</th>
<th>Year</th>
<th>Study size</th>
<th>Sample</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Obermair A, et al.</td>
<td>1998</td>
<td>C: 131</td>
<td>sVEGF</td>
<td>54</td>
<td>77</td>
<td>No difference in sVEGF levels among all groups</td>
</tr>
<tr>
<td>Oehler MK, et al.</td>
<td>1999</td>
<td>C: 20</td>
<td>sVEGF</td>
<td>71</td>
<td>65</td>
<td>sVEGF levels significantly higher from patients with malignancies versus control and benign groups</td>
</tr>
<tr>
<td>Cooper BC, et al.</td>
<td>2002</td>
<td>B: 34</td>
<td>sVEGF</td>
<td>74</td>
<td>71</td>
<td>sVEGF levels significantly higher from patients with malignancies versus benign or LMP groups</td>
</tr>
<tr>
<td>Dehaven K, et al.</td>
<td>2002</td>
<td>C: 125</td>
<td>sVEGF</td>
<td>N/A</td>
<td>N/A</td>
<td>No difference in sVEGF levels among all study groups</td>
</tr>
<tr>
<td>Tanir HM, et al.</td>
<td>2003</td>
<td>B: 50</td>
<td>sVEGF</td>
<td>92</td>
<td>88</td>
<td>sVEGF levels significantly higher in patients with malignant tumors</td>
</tr>
<tr>
<td>Demirkiran F, et al.</td>
<td>2003</td>
<td>B: 45</td>
<td>sVEGF</td>
<td>77</td>
<td>78</td>
<td>sVEGF and cVEGF significantly elevated in patients with malignant tumors</td>
</tr>
<tr>
<td>Harlozsinska A, et al.</td>
<td>2004</td>
<td>B: 53</td>
<td>sVEGF</td>
<td>N/A</td>
<td>N/A</td>
<td>sVEGF and cVEGF significantly elevated in patients with malignant tumors</td>
</tr>
<tr>
<td>Li L, et al.</td>
<td>2004</td>
<td>C: 90</td>
<td>sVEGF</td>
<td>77</td>
<td>87</td>
<td>sVEGF levels significantly higher from patients with malignancies versus control and benign groups</td>
</tr>
<tr>
<td>Gorelik E, et al.</td>
<td>2005</td>
<td>C: 45</td>
<td>sVEGF, IL-8, IL-6, CA-125, sEGF</td>
<td>84</td>
<td>95</td>
<td>Significant elevation in serum cytokine levels from patients with malignant tumors versus control and benign groups</td>
</tr>
</tbody>
</table>

sVEGF- serum vascular endothelial growth factor; cVEGF- cystic vascular endothelial growth factor; C- healthy control patients; B- benign ovarian neoplasms; M- malignant ovarian carcinoma; L- low malignant potential ovarian neoplasms; IL-8 – interleukin 8; IL-6 – interleukin 6; sEGF – serum epidermal growth factor; N/A – not available.

single agent therapy using a small molecule inhibitor of VEGF and PDGF receptors, SU11248 [83]. Interestingly, they reported a decrease in plasma sVEGFR-2 levels following therapy while VEGF levels increased. The latter finding is consistent with previous reports of sVEGFR-2 modulation following anti-VEGF therapy, however, the mechanism by which SU11248 affects sVEGFR-2 is not fully known.

As investigators begin to understand the molecular pathways involved in tumor angiogenesis, new agents are being developed that target upstream regulators of VEGF expression (Fig. 1). For example, Src, a non-receptor tyrosine kinase, has been reported to mediate angiogenesis by upregulating pro-angiogenic factors such as VEGF and interleukin-8 (IL-8) [109]. Recently, we have demonstrated in ovarian cancer models that Src inhibition decreased tumor growth and significantly decreased serum VEGF and IL-8 levels. Similarly, siRNA based therapy against FAK, a non-receptor kinase known to regulate VEGF, reduced circulating VEGF levels in response to treatment [41]. These findings suggest a unique opportunity to further explore the role of VEGF as a surrogate marker of response to new agents that mediate angiogenic activity.

Development and validation of circulating VEGF levels as a biomarker may also depend on the type of study design and sample collection obtained by investigators. For example, VEGF levels can differ among serum and plasma samples taken from the same patient [3]. This is partly due to the secretion of VEGF from components of the circulatory system including platelets, neutrophils, monocytes, and lymphocytes [34,116]. In addition, anti-coagulants often found in blood collection tubes can falsely elevate VEGF levels due to platelet-derived secretion in non-clotted samples [117]. The significance of platelet-derived VEGF remains controversial due to suggestions that platelets may mediate release of angiogenic molecules in the presence of tumor cells and therefore reflect the true disease process [31,94]. Although, these differences in circulating levels have been demonstrated in several studies from patients with malignant disease [3]
3.2. Interleukin-8

Interleukins are important members of the cytokine family and are known to modulate normal defense systems in the human body. Stressful environments, such as hypoxia and surgical stress, activate release of interleukins from inflammatory cells, peritoneal mesothelial cells, fibroblasts, and endothelial cells into the systemic circulation and in turn initiate protective pathways [63]. Ovarian carcinoma, once labeled a “cytokine propelled disease”, secretes large amounts of interleukins into the circulation, which in turn mediate tumor growth, metastasis, and angiogenesis [77]. Moreover, increased circulating levels of interleukins have been demonstrated in several malignancies including ovarian carcinoma and are associated with poor patient survival [61,75]. For these reasons, interleukins involved in angiogenesis remain of particular interest as biomarkers in ovarian carcinoma.

Interleukin-8 is well known for its role in tumor invasion, metastatic spread, and angiogenesis. IL-8 is a small (8 kDa) chemotactic cytokine that belongs to the CXC cytokine family known for activating and attracting neutrophils [53]. IL-8 binds to the seven-transmembrane spanning G-protein coupled receptors CXCR1 and CXCR2 with high affinity and in turn activates members of the MAPK kinase pathway including ERK 1/2 [72]. IL-8 was initially reported as a prominent mediator of angiogenesis by Koch and colleagues in 1992 [64]. They demonstrated that recombinant IL-8 induced neovascularization in a rat corneal model [64]. Subsequently, Li and colleagues demonstrated the direct effect of IL-8 on human endothelial cell migration, capillary tube formation and survival [69,70].

IL-8 is secreted by multiple sources including monocytes, neutrophils and mesothelial cells. Tumor cells also secrete IL-8, which in turn can act as an autocrine inducer of tumor growth or paracrine modulator of host endothelial cells in angiogenesis. In several small studies, IL-8 levels were elevated in the serum and ovarian cystic fluid in patients with ovarian carcinoma [28,53,75,88]. Furthermore, Lokshin and colleagues demonstrated that IL-8 and anti-IL-8 antibody levels were increased in ovarian cancer patients and more specifically, that anti-IL-8 antibody levels correlated with early stage disease [75]. In addition, they reported a specificity of 98% for both IL-8 and anti-IL-8 antibody levels and sensitivities of 63% and 66%, respectively, in disease detection [75]. Furthermore, the specificity and sensitivity increased to 98% and 88%, respectively in combination with CA-125 [75]. To this end, IL-8 and anti-IL-8 antibodies may be possible screen-
ing biomarkers for patients with ovarian tumors, especially when combined with traditional applications and markers such as pelvic ultrasound and CA-125.

Due to the role of IL-8 in mediating tumor angiogenesis, quantifying circulating IL-8 levels may assist oncologists in treatment surveillance as a biomarker of response. In most circumstances, ovarian cancer patients are treated with platinum and taxane chemotherapy following cytoreductive surgery. Mayerhofer and colleagues reported that IL-8 levels decreased with chemotherapy in 31 patients [80]. In their study, IL-8 levels demonstrated a decreasing trend midway and following six cycles of combination chemotherapy [80]. Conversely, Uslu reported that IL-8 levels actually increased immediately following the initiation of chemotherapy in ovarian cancer patients, specifically in those with residual disease [115]. However, it has been shown that chemotherapy can transiently induce IL-8 secretion from tumor cells [68] and therefore may explain the differences in these two studies, especially those patients with residual disease.

Although anti-VEGF targeted therapy has demonstrated improvement in patient survival, few studies have reported the benefit of targeting IL-8 in cancer therapy. In pre-clinical murine models, Bar-Eli and colleagues demonstrated that therapy with fully humanized anti-IL-8 antibodies decreased tumor growth and MVD [51]. To the best of our knowledge, no studies report the use of IL-8 as an anti-vascular target in ovarian cancer. However, we recently demonstrated in pre-clinical models that circulating IL-8 levels decreased secondary to Src inhibition [42] suggesting that IL-8 may be a useful marker for response to specific therapies. Clearly, with the emergence of new small molecule inhibitors and now effective applications for delivering gene-specific siRNA molecule inhibitors and now effective applications for delivering gene-specific siRNA in vivo [65], IL-8 may be an attractive target for patients with ovarian carcinoma.

3.3. Interleukin-6

IL-6 was originally reported as a mediator in B cell maturation. Recently, Nilson and colleagues demonstrated that IL-6 mediated tumor growth and angiogenesis in ovarian cancer models [85]. In that study, IL-6 receptors were detected on ovarian and endothelial cells and were found to actively participate in the development of tumor angiogenesis [85]. Since IL-6 is secreted into circulation, it was suggested that IL-6 may be a potential marker for disease detection and surveillance in patients with ovarian tumors. Berek and colleagues were the first to report elevated serum IL-6 levels in ovarian cancer patients [13]. They found a direct correlation with IL-6 overexpression and decreased overall survival, increased tumor burden, and disease status [13]. In a study of 73 ovarian cancer patients, Tempfer and colleagues reported that increased IL-6 levels prior to therapy correlated with both decreased disease free and overall survival [111]. However, these findings have not been consistent in the literature [95]. In a more recent study, IL-6 demonstrated no added benefit as a disease biomarker when compared to traditional markers; however, when evaluated within a panel of cytokines, IL-6 was considered useful for disease detection [39]. To date, the benefit of measuring IL-6 as a marker of angiogenesis remains to be determined in ovarian cancer.

4. Circulating endothelial cells

The development of new vasculature requires activation and migration of endothelial cells. In most normal tissues, endothelial cells remain quiescent and divide approximately every 3–5 years. However, rapid proliferation of endothelial cells is crucial for the process of angiogenesis in growing tumors. As new vasculature matures, often endothelial cells can become dislodged into the systemic circulation. Recent studies have shown that levels of circulating endothelial cells (CEC) are elevated in cancer patients 3.6–5 fold compared to healthy controls and may be a reflection of ongoing angiogenesis [12,35,78]. In addition, tumor-derived VEGF has also been shown to mobilize CECs in murine models and in humans [9,11,55,56]. Based on these findings, monitoring CEC levels may provide useful information regarding disease status and treatment efficacy in cancer patients.

Within the systemic circulation, two populations of CECs have been identified. Mature endothelial cells (CEC) are thought to derive from mature vasculature and circulating endothelial progenitor cells (CEP) are mobilized from the bone marrow. CEPs may contribute to the angiogenic process by differentiating into mature endothelial cells, however, their direct role has yet to be determined [7,8]. CECs and CEPs can be identified based on their expression of specific endothelial antigens using flow cytometry [11]. This distinction is critical for determining the effects of CECs and CEPs in response to current cancer treatments. For example, Beaudry and colleagues demonstrated in a murine model that CECs increased in response to ZD6474 (VEG-
FR inhibitor), although CEPs decreased due to the inhibition of VEGF induced mobilization from the bone marrow [11].

CEC and CEP levels may be particularly useful for following response to anti-angiogenic therapy. Preclinical models have demonstrated that CEPs increase following treatment with cyclophosphamide, however treatment with endostatin (an inhibitor of angiogenesis) correlated with an increase in CECs [15,82]. Moreover, following treatment with anti-vascular agents, CEC and CEP levels have been reflective of vascular changes within the tumor and may provide more useful information in treatment response [11]. In support of this theory and based on the growing evidence that the frequency of chemotherapy administration may have anti-angiogenic effects [37,43], we demonstrated a decrease in CEPs using metronomic chemotherapy scheduling, however CEC levels remained unchanged from baseline [15,57]. Moreover, anti-vascular therapy in combination with metronomic chemotherapy demonstrated a 79% decrease in CEP levels [57]. While preclinical models of ovarian cancer have demonstrated that CEP and CEP levels may be beneficial as a surrogate marker to anti-vascular therapy [57] little is known about their role in patients with ovarian carcinoma. Only a few studies have followed CEC levels in patients with malignancies. For example, in breast and lymphoma patients, CEC levels were initially increased and correlated with tumor burden, however, upon remission became normalized [35,78]. As newer anti-vascular agents become available, quantifying CEC and CEP levels may be helpful in monitoring tumor growth and treatment response in patients with ovarian carcinoma.

5. Concluding remarks

Ovarian cancer remains the most deadly disease in women among gynecologic malignancies [1]. The majority of these patients initially respond to surgical debulking and chemotherapeutic regimens, but most eventually develop recurrent cancer and die from this disease. One of the difficulties in managing ovarian cancer patients is the lack of an effective biomarker for disease detection. Moreover, with the emergence of new anti-vascular therapies in ovarian cancer, there is a need for novel biomarkers for treatment response in patients. It is hoped that global analysis with gene microarrays may identify potential candidates to test as screening markers, therapeutic targets, and/or surrogate biomarkers of treatment response. Furthermore, new treatment approaches have allowed investigators to focus on specific targets that not only demonstrate anti-tumor effects, but also anti-angiogenic properties. This progress in cancer treatment supports the current focus for development and validation of new biomarkers of angiogenesis in ovarian cancer. In this review, we have outlined some of the key components involved in angiogenesis in ovarian carcinoma. In addition, several of the major angiogenic factors were reviewed as potential markers for disease detection and treatment surveillance in ovarian cancer patients. Although some markers appear to be useful in initial studies, validation in larger prospective trials is required.

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