

# Analysis of polymorphisms in genes (*AGT*, *MTHFR*, *GPIIIa*, and *GSTP1*) associated with hypertension, thrombophilia and oxidative stress in Mestizo and Amerindian populations of México

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**Abstract.** Several polymorphisms related to hypertension, thrombophilia, and oxidative stress has been associated with the development of cardiovascular disease. We analyzed the frequency of M235T *angiotensinogen* (*AGT*), A222V *5,10 methylenetetrahydrofolate reductase* (*MTHFR*), L33P *glycoprotein IIIa* (*GPIIIa*), and I105V *glutathione S-transferase P1* (*GSTP1*) polymor-

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phisms in 285 individuals belonging to Mexican-Mestizo and five Amerindian population from México, by real time PCR allelic discrimination. Allele and genotype frequencies were compared using  $\chi^2$  tests.

All populations followed the Hardy Weinberg equilibrium for assay markers with the exception of the Triki, whose were in Hardy Weinberg disequilibrium for the *glutathione S-transferase P1* polymorphism.

Interestingly, according to all the analyzed single nucleotide polymorphisms (SNPs), the Triki population was the most differentiated and homogeneous group of the six populations analyzed. A comparison of our data with those previously published for some Caucasian, Asian and Black populations showed quite significant differences. These differences were remarkable with all the Mexican populations having a lower frequency of the 105V allele of the *glutathione S-transferase P1* and reduced occurrence of the 222A allele of the *5,10 methylenetetrahydrofolate reductase*. Our results show the genetic diversity among different Mexican populations and with other racial groups.

Keywords: Gene polymorphisms, *AGT*, *GPIIIa*, *GSTP1*, *MTHFR*, Mexican populations

## 1. Introduction

Several epidemiological and clinical studies have reported associations between polymorphisms of various genes related to hypertension, thrombophilia, and oxidative stress with development of cardiovascular and cerebrovascular diseases. Among these genes are *angiotensinogen (AGT)* (MIM 106150) [1], *glycoprotein IIIa (GPIIIa)* (MIM 173470) [2], *5,10 methylenetetrahydrofolate reductase (MTHFR)* (MIM 607093) [3], and *glutathione S-transferase P1 (GSTP1)* (MIM 134660) [4]. However, other studies have shown no evidence of association between these polymorphisms and risk of these diseases [1,5,6].

Approximately 30% to 40% of the population variability in blood pressure is genetically determined [7]. The importance of the renin-angiotensin system for maintenance of normal cardiovascular homeostasis is well established [8]. It has been suggested that individuals with a p.M235T (c.704C>T) polymorphism in the *AGT* gene in the homozygous TT state have increased plasma angiotensinogen levels and a corresponding increase in risk of hypertension [9]. Likewise, it was demonstrated that moderate hyperhomocysteinemia is considered as an independent risk factor for ischemic cardiovascular disease [10]. The genetic basis of this disease may be due to a polymorphism in the *MTHFR* gene (p.A222V, c.677C>T) where homozygosity for the C-677T substitution results in reduced MTHFR enzyme activity and subsequently elevated homocysteine concentrations of ~20% [11].

*GPIIIa* is a thrombophilic gene involved in the regulation of vascular thrombosis. The GPIIb/GPIIIa complex mediates platelet aggregation by acting as a receptor for fibrinogen. This complex also acts as a receptor for von Willebrand factor and fibronectin [12]. The polymorphism c.98C>T in this gene causes a p.L33P substitution and the existence of two antigeni-

cally distinct forms of the mature GPIIb/IIIa antigen on platelets [13]. This variant itself has been associated with risk of premature acute coronary syndromes and stroke in young Caucasian women [14]. In addition, oxidative stress is thought to play an important role in the pathophysiology of hypertension, although this statement lies within the context of the development of preeclampsia [15]. It has been hypothesized that reduced levels of GSTP1 due to the lower activity of the *GSTP1* (p.I105V, c.313A>G) allele in this syndrome may be an indicator of decreased capacity of the GST detoxification system and may cause a prolonged exposure to reactive by-products, which may contribute to maternal endothelial dysfunction [15,16].

It has been observed that diverse genetic polymorphisms associated with cardiovascular diseases are usually found in most human populations but often with variations in the allele frequencies [17]. The Mexican-Mestizo population is constituted by a mixture of Europeans and Africans with native Indian subjects [18]. These individuals have a proportion of 56% Amerindian genes, 40% Caucasian genes, and 4% African genes [19]. According to the National Institute of Anthropology, a Mexican-Mestizo is defined as a person who was born in Mexico, has a Spanish-derived last name, and has a family of Mexican ancestors back to the third generation [20]. Moreover, in Mexico ~7% of the total population corresponds to an ethnic group, using the language as a classification criterion [21]. Interestingly, several of these groups have maintained a limited admixture level with the Mestizo population as a consequence of the geographical isolation and/or cultural barriers.

Because the Mestizos and Amerindian populations are genetically heterogeneous and considering the putative role of polymorphisms in genetic susceptibility to cardiovascular diseases, the principal aim of this study was to analyze the frequency of p.M235T an-

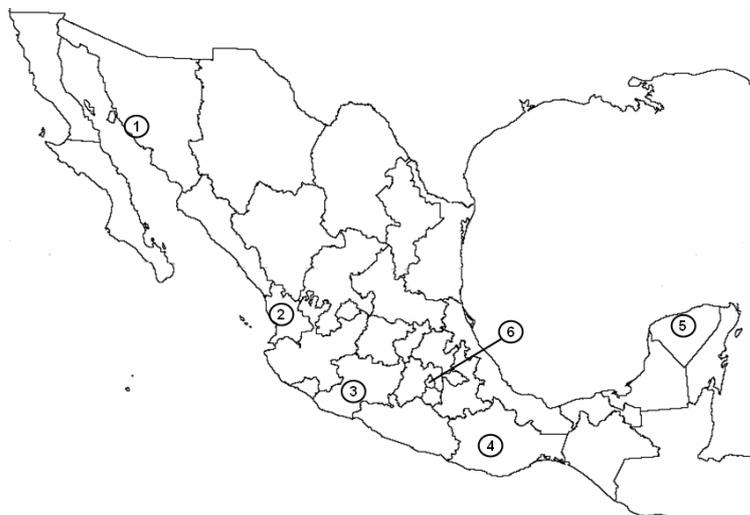


Fig. 1. Geographical location of the six Mexican populations analyzed in the study. 1. Yaqui, 2. Huichol, 3. Puépecha, 4. Triki, 5. Maya, 6. Mestizo.

*giotensinogen*, p.A222V *5,10 methylenetetrahydrofolate reductase*, p.L33P *glycoprotein IIIa*, and p.I105V *glutathione S-transferase P1* polymorphisms in the Mestizo and five Amerindian populations from México.

## 2. Subjects

The study was approved by the Institute's Human Research Committee. Informed consent was obtained from all subjects before participating in the study. The local authorities of the Amerindian population gave their approval to participate in the study, and a translator was used as needed. From the 285 subjects, DNA from blood samples was obtained from apparently normal male individuals with no phenotypic abnormalities. Individuals ranged in age from 18–60 years ages. The study was comprised of subjects from six Mexican populations from six different geographical regions (Fig. 1): 71 Mexican-Mestizos living in Mexico City or its surroundings; 29 Purépechas from the state of Michoacán in western Mexico; 27 Yaquis from the state of Sonora located in northern Mexico; 15 Huicholes from Nayarit in northwestern Mexico; 89 Trikis from the state of Oaxaca and 54 Mayas from the state of Yucatán, both located in southeastern México.

The Mexican-Mestizo group resulted from the admixture between Native American and European (Spanish) populations with a much smaller contribution of African groups. Because México City has been a site of massive immigration during the last century receiving inhabitants from all around the country, this

group can be considered representative of the overall Mexican population. Only individuals born in México whose parents and grandparents were born in México were considered Mexican-Mestizo. All Amerindian individuals and their ancestors throughout three generations were born in the same community and spoke their own native language.

## 3. Methods

### 3.1. Genotyping

The study was performed to determine the frequencies of the polymorphisms in *AGT* (p.M235T), *MTHFR* (p.A222V), *GPIIIa* (p.L33P), and *GSTP1* (p.I105V) genes. Peripheral blood samples were obtained from all individuals, and genomic DNA was purified by standard techniques [22]. Single nucleotide polymorphism (SNP) analysis was performed using real-time PCR allelic discrimination TaqMan assays (AB) with minor modifications. All PCR reactions contained 20 ng of DNA, 2.5  $\mu$ l TaqMan Universal Master Mix (AB) (2X), 0.25  $\mu$ l primers and probes (10X) and water for a final volume of 5  $\mu$ l, including the appropriate negative controls in all assays. Real-time PCR was performed on an ABI Prism 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA, USA) under the following conditions: 50°C for 2 min, 95°C for 10 min, and 40 cycles of amplification (95°C for 15 s and 62°C for 1 min). For each cycle, the software determined the fluorescent signal from the VIC or FAM-labeled probe

Table 1  
Primers and probes used for the PCR allelic discrimination TaqMan assay

Gene	Primers	Probe
<i>AGT</i>	5'-GCTGTGACAGGATGGAAGACT-3' (sense)	VIC 5'-CTGGCTCCCATCAGG-3' wt
	5'-AGTGGACGTAGGTGTTGAAAGC-3' (antisense)	FAM 5'-CTGGCTCCCGTCAGG-3' mt
<i>MTHFR</i>	5'-GCACTGAAGGAGAAGGTGTCT-3' (sense)	VIC 5'-CTGGCTCCCATCAGG -3' wt
	5'-CCTCAAAGAAAAGCTGCGTGATG-3' (antisense)	FAM 5'-CTGGCTCCCGTCAGG-3' mt
<i>GPIIIa</i>	5'-TCTCTTTGGGCTCCTGTCTTACA-3' (sense)	FAM 5'-TGAGCCCAGAGGCA-3' wt
	5'-CAGATTCTCCTCCGGTCACA-3' (antisense)	VIC 5'-TGAGCCCAGAGGCA-3' mt
<i>GSTP1</i>	5'-CCTGGTGGACATGGTGAATGAC-3' (sense)	VIC 5'-CTGCAAATACATCTCC-3' wt
	5'-CAGATGCTCACATAGTTGGTGTAGA-3' (antisense)	FAM 5'-CTGCAAATACGCTCTCC-3' mt

Table 2  
Allele frequencies of the *MTHFR* c.677C>T, *AGT* c.704C>T, *GPIIIa* c.98C>T and *GSTP1* c.313A>G polymorphisms in several populations

SNP	Allele	Huichol	Maya	Purépecha	Trikis	Yaquis	Mestizo	Caucasian	Asian	Black
<i>MTHFR</i>	C	0.467 <sup>a</sup>	0.389 <sup>b</sup>	0.293 <sup>a</sup>	0.078 <sup>a</sup>	0.537 <sup>a</sup>	0.472 <sup>a</sup>	0.68 <sup>26</sup>	0.55 <sup>27</sup>	0.9 <sup>28</sup>
	C→T	0.533	0.611	0.707	0.922	0.463	0.528	0.32	0.45	0.1
<i>AGT</i>	C	0.733 <sup>c</sup>	0.88 <sup>c</sup>	0.879 <sup>c</sup>	0.983 <sup>d</sup>	0.759 <sup>c</sup>	0.775 <sup>c</sup>	0.41 <sup>40</sup>	0.81 <sup>38</sup>	0.83 <sup>39</sup>
	C→T	0.267	0.12	0.121	0.017	0.241	0.225	0.59	0.19	0.17
<i>GPIIIa</i>	C	1.0 <sup>c</sup>	0.973 <sup>e</sup>	0.949 <sup>c,f</sup>	1.0 <sup>e</sup>	0.907 <sup>f</sup>	0.958 <sup>e,f</sup>	0.83 <sup>14</sup>	0.99 <sup>41</sup>	0.89 <sup>42</sup>
	C→T	0.0	0.027	0.051	0.0	0.093	0.042	0.17	0.01	0.11
<i>GSTP1</i>	A	0.143 <sup>g</sup>	0.38 <sup>g</sup>	0.413 <sup>g</sup>	0.298 <sup>g</sup>	0.352 <sup>g</sup>	0.415 <sup>g</sup>	0.64 <sup>43</sup>	0.83 <sup>44</sup>	0.58 <sup>45</sup>
	A→G	0.857	0.62	0.587	0.702	0.648	0.585	0.36	0.17	0.42
								(2n = 820)	(2n = 936)	(2n = 658)
								(2n = 200)	(2n = 1028)	(2n = 806)
								(2n = 432)	(2n = 632)	(2n = 92)
								(2n = 236)	(2n = 626)	(2n = 274)

**Note:**The distributions of allele frequencies obtained in our populations (Amerindian and Mexican Mestizos) were compared to those previously described in other populations (Caucasian, Asian and Black population) using the chi-square test ( $p$  significant values were in the range of  $p < 0.05$  to  $p < 0.0001$ ).

<sup>a</sup>Decreased frequency when compared to Caucasian and Black populations

<sup>b</sup>Decreased frequency when compared to Caucasian Asian and Black populations.

<sup>c</sup>Increased frequency when compared to Caucasian population

<sup>d</sup>Increased frequency when compared to Caucasian, Asian, and Black populations.

<sup>e</sup>Increased frequency when compared to Caucasian, and Black populations.

<sup>f</sup>Decreased frequency when compared to Asian population.

<sup>g</sup>Decreased frequency when compared to Caucasian, Asian, and Black population.

(Applied Biosystems). Allelic discrimination was performed using specific primers and probes for each allele. The sequences of the primers and probes for each polymorphism are shown in Table 1.

### 3.2. Statistical analysis

Results are expressed as mean  $\pm$  standard deviation (SD). Statistical analyses were performed using SPSS v.10 (SPSS, Chicago, IL, USA). Continuous variables were compared by unpaired Student's test. Genotype distributions were tested for deviation from Hardy-Weinberg equilibrium in all groups. Allele and genotype frequencies were tested using the  $\chi^2$  test (<http://ihg.gsf.de/cgi-bin/hw/hwal.pl>);  $p$  value  $< 0.05$  was accepted as statistically significant. Genetic distances were estimated by calculating the coancestor coefficient of Reynolds et al. [23].

## 4. Results

The frequencies of the *AGT* c.704T>C, *MTHFR* c.677C>T, *GPIIIa* c.98C>T and *GSTP1* c.313A>G polymorphisms were analyzed in different Indian and Mestizo-Mexican populations. Distribution of allelic frequencies and comparison of these with other populations are shown in Table 2. The most distinguishing group among the six Mexican populations was the Trikis, given that this group presented a significant genetic differentiation in most of the studied SNPs ( $p < 0.0002$ ) when compared to the other studied populations; the only exception was *GSTP1*.

Among the most remarkable differences was that observed for the *MTHFR* c.677C>T polymorphism where the Trikis had a very low frequency of the allele C (7.8%), whereas the rank was 29.3% and 53.7% for Purépechas and Yaquis, respectively. Considering the

Table 3  
Genotype frequency, heterozygosity (H) and Hardy-Weinberg equilibrium test<sup>a</sup> of four SNPs from six Mexican populations

Population		<i>MTHFR</i>			<i>AGT</i>			<i>GPIIIa</i>			<i>GSTP1</i>		
		CC	CT	TT	CC	CT	TT	CC	CT	TT	AA	AG	GG
Mestizo	n	17	33	21	42	26	3	65	6	0	13	33	25
(n = 71)	H (%)	He = 49.8; Ho = 46.5			He = 35.0; Ho = 36.6			He = 8.1; Ho = 8.4			He = 48.6; Ho = 46.5		
Maya	n	7	28	19	42	11	1	51	3	0	6	29	19
(n = 54)	H (%)	He = 47.6; Ho = 51.8			He = 21.2; Ho = 20.4			He = 5.4; Ho = 5.5			He = 47.1; Ho = 53.7		
Trikis	n	0	14	75	86	3	0	89	0	0	14	25	50
(n = 89)	H (%)	He = 14.6; Ho = 15.7			He = 3.3; Ho = 3.4			He = 0; Ho = 0			He = 42.0; Ho = 28.1 <sup>b</sup>		
Purépechas	n	2	13	14	22	7	0	26	3	0	4	16	9
(n = 29)	H (%)	He = 41.5; Ho = 44.8			He = 21.3; Ho = 24.1			He = 9.8; Ho = 10.3			He = 48.5; Ho = 55.2		
Yaquis	n	9	11	7	16	9	2	22	5	0	3	13	11
(n = 27)	H (%)	He = 49.7; Ho = 40.7			He = 37.0; Ho = 33.3			He = 17.0; Ho = 18.5			He = 45.6; Ho = 48.1		
Huicholes	n	3	8	4	7	8	0	15	0	0	1	2	12
(n = 15)	H (%)	He = 49.8; Ho = 53.3			He = 40.0; Ho = 53.3			He = 0; Ho = 0			He = 23.1; Ho = 13.3		

<sup>a</sup>Heterozygosity expected (He), Heterozygosity observed (Ho) and Hardy-Weinberg equilibrium test.

<sup>b</sup>Significant Hardy-Weinberg disequilibrium by Pearson's goodness-of-fit  $\chi^2$  test (1 df).  
SNP, single nucleotide polymorphism.

Table 4  
Coancestry coefficient (above diagonal) and combined probability of pairwise comparison<sup>a</sup> (below diagonal) between Mexican populations and three worldwide racial groups

	Mestizo	Maya	Triki	Huichol	Purépecha	Yaqui	Caucasian	Asian	Blacks
Mestizo	*****	0.0064	0.19114	0.0358	0.0158	-0.0088	0.0780	0.1380	0.1661
Maya	0.1814	*****	0.1194	0.0476	-0.0081	0.0131	0.1436	0.1807	0.2388
Triki	0.0000	0.0000	*****	0.2748	0.0740	0.2694	0.4056	0.4110	0.6021
Huichol	0.1123	0.0556	0.0000	*****	0.0568	-0.0066	0.1639	0.3478	0.2881
Purépecha	0.1257	0.7341	0.0000	0.0133	*****	0.0227	0.1534	0.1834	0.2879
Yaqui	0.5707	0.0637	0.0000	0.1750	0.0510	*****	0.0597	0.1743	0.1304
Caucasian	0.0000	0.0000	0.0000	0.0000	0.0000	0.0001	*****	0.0910	0.0723
Asian	0.0000	0.0000	0.0000	0.0029	0.0000	0.0000	0.0000	*****	0.1672
Blacks	0.0000	0.0000	0.0000	0.0000	0.0002	0.0000	0.0000	0.0000	*****

<sup>a</sup>Exact test: 10,000 permutations.

Bonferroni correction factor, to establish pair comparison significance (Table 2;  $p < 0.003$ ), the other genetically distinctive group was the Mestizo, which was differentiated from the Mayas, Purépechas and Yaquis.

Comparing allelic distribution in the six Mexican ethnic groups with those reported previously in other populations (Table 2), we observed in the case of the *MTHFR* a significant reduction in the frequency of allele C in all studied populations when compared with Caucasian, Asian and Black populations. Moreover, the frequency of *AGT* 704C allele in the Mestizo, Maya, Purépecha, Huichol and Yaqui groups was high when compared to the Caucasian population. Concerning the Trikis, the same allele showed an increased frequency in relation not only to the Caucasian population but also to the Asian and Black populations.

Frequency of *GPIIIa* 98T allele was slightly increased in Mestizo and indigenous Mexican groups when compared with Caucasian and Black populations. On the other hand, the frequency of the allele was similar between the Mexican population and an Asian population.

In regard to the *GSTP1* 313G allele, all Mexican populations (ranking from 14% to 41%) had a reduction in frequency when compared with Caucasian (64%), Asian (83%) and Black (58%) populations.

Distribution of genotypes for all analyzed genetic markers was in Hardy-Weinberg equilibrium (Table 3) with the exception of the distribution observed for *GSTP1* polymorphism in the Trikis. In this ethnic group there was an increment in the homozygous proportion, suggesting a significant endogamous process, which is supported by comparison of the heterozygosity in all the studied Mexican populations. In this respect, the Trikis presented the lowest frequency of heterozygosity in most of the genetic markers as compared with the other groups, with the exception of the *GPIIIa* polymorphism.

Genetic distances were estimated by calculating the coancestry coefficient of Reynolds et al. [23] among the different populations (Table 4). The neighbor-joining tree (Fig. 2) was obtained by using the genetic distances of the Mexican populations and three other racial

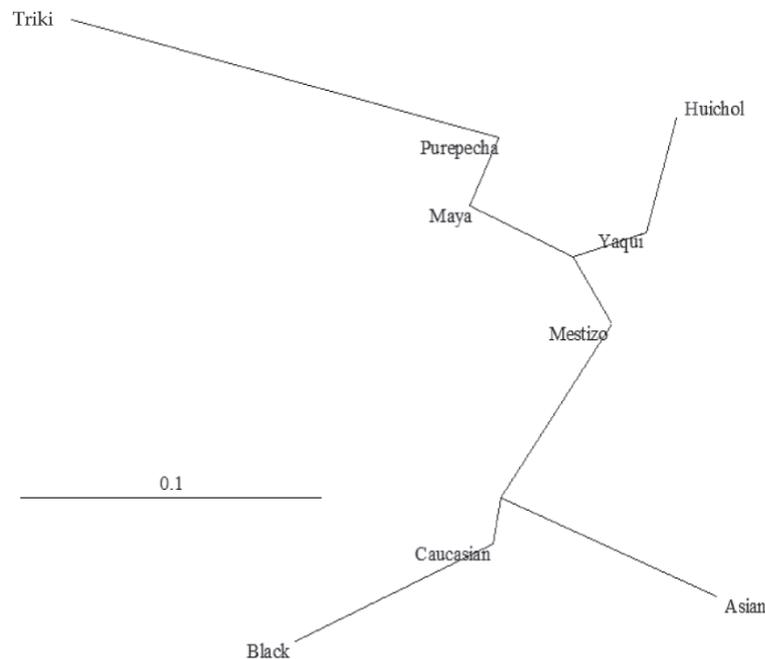


Fig. 2. Neighbor-joining tree using concenstry coefficient distances between Mexican populations and three racial groups based on four SNPs (*MTHFR*, *AGT*, *GPIIIa* and *GSTP1*).

groups (Caucasian, Asian and Black) and graphically demonstrates the relationship among the populations. As suggested for the preceding analysis, Trikis were the most genetically differentiated group.

## 5. Discussion

The Mexican population has a historical distinction for its ethnic diversity, and the country presents one of the highest concentrations of indigenous populations on the entire American continent (obtained from the Instituto Nacional Indigenista) [24]. According to the most recent population census, the indigenous population of Mexico is 8.7 million (obtained from the Instituto Nacional de Estadística Geografía e Informática) [21]. These indigenous subjects are descendants of Amerindian populations with an ancient Asian origin [25]. In addition, the Mexican Mestizos population appears to be the result of the genetic admixture among Amerindians, Caucasian and Black genes. The general pattern has a high Indian ancestry followed by Caucasian and Black ancestry [18]. Because it has been determined that genetic polymorphisms have an ancient origin and are usually found in most human populations but often with different allele frequencies [17], it would be very probable that Mexican populations may

have diverse types and frequencies of genetic polymorphisms of different genes that predispose to cardiovascular diseases.

We analyzed the allelic and genotype frequencies in polymorphisms of hypertension, thrombophilic and oxidative stress-related genes *MTHFR*, *AGT*, *GPIIIa*, and *GSTP1* in different Mexican populations. Regarding the *MTHFR* c.677C>T polymorphism, we observed an allelic frequency ranking from 46% to 92%. In general, these frequencies were higher than that found in the Caucasian [26], Asian [27] and Black [28] populations. The elevated frequency of the 677T allele in Mexican populations has also been observed in earlier studies [29,30]. It is noteworthy that our results determined that the Triki population has the highest frequency (92%) of the mutant allele with no homozygous individuals for the 677C allele. In a previous study [30], the frequency of the 677T allele in the Purépecha population was lower (57%) than observed in this study (70%). This difference may be caused by some genetic admixture of the former indigenous population with Caucasian genes.

It has been observed that homozygosity for the 677T allele results in reduced MTHFR enzyme activity and, subsequently, elevated homocysteine concentrations of ~20% [11], which represents a risk factor for ischemic vascular disease [10]. Likewise, several studies de-

scribed the association between the *MTHFR* 677TT homozygous genotype and an increased risk of hypertension [31], macrovascular abnormalities in systemic sclerosis patients [32], intima-media thickening in patients with cognitive impairment [33], and preeclampsia [34]. However, in a study carried out by our group, it was found that the 677TT genotype confers a reduced risk of preeclampsia in Maya-Mestizo women under a recessive model for the 677T allele [35]. The inconsistency of the results in different populations may be caused by the differential phenotypic expression of specific genotypes as a consequence of diverse factors such as genetic background, age, gender, physiological and pathological conditions, intake of food and drugs, and physical activity [36,37]. All of these findings demonstrate the importance of studying the allelic and genotype frequency of different polymorphisms in diverse human groups.

The *AGT* 704TT genotype increases plasma angiotensinogen levels that, in turn, augment the risk of hypertension [9]. In this study, *AGT* 704C allele and genotype frequencies distribution were similar among Mexicans. In all populations the 704C allele was predominant, and Trikis bear the highest frequencies of this (98%). In contrast, the 704TT genotype was absent in Trikis, Purépechas and Huicholes and was very low in Mestizo, Maya and Yaquis. Comparison of the allele frequencies of the Mexican populations with other previously described allele frequencies in different human groups resulted in being similar between Mexicans and Asians [38] as well as Blacks [39]. In contrast, the 704T allele was lower than that observed in the Caucasian group [40].

Concerning the c.98C>T polymorphism of the *GPIIIa* gene, it seems to predispose premature acute coronary syndrome and stroke in young Caucasian women [14]. The frequency of the mutant allele (98T) was very low in all the studied Mexican populations and was similar to that reported for an Asian [41] and Black population [42]. In contrast, a Caucasian group has the highest presence of the mutant allele with a frequency of 17% [14]. To the best of our knowledge, this is the first report of this polymorphism in a Mexican population.

In regard to the *GSTP1* gene, the 313G allele was present in a higher frequency in Mexicans as compared with Caucasians [43], Asians [44], and Blacks [45]. Currently there are no studies of this genetic marker with other Amerindian populations. In contrast, Hatagima et al. [46] studied the same polymorphism in a Brazilian Mestizo population, finding that the frequen-

cy of the 313G allele was 38%, similar to that observed in Caucasian and Black populations. This may be attributed to the fact that the genetic constitution of Brazilians has a high influence over these human groups.

Interestingly, the neighbor-joining tree using coancestry coefficient distances between Mexican populations and three racial groups showed that Trikis were the most genetically differentiated group. Consequently, this population presented the highest relative genetic homogeneity, a characteristic that may be a consequence of the conservation of its endogamous reproductive cultural patterns [47].

In summary, according to the study of polymorphisms involved in hypertension, thrombophilia and oxidative stress (*AGT*, *GPIIIa*, *MTHFR* and *GSTP1*), we determined their allele and genotype frequencies in different Mexican populations with diverse geographical, anthropological, genetic, and cultural antecedents. Because these polymorphisms have been associated with distinct cardiovascular pathologies, we are currently studying whether the presence of some alleles may be a risk factor associated with these types of diseases in the studied Mexican populations.

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## References

- [1] T.V. Pereira, A.C. Nunes, M. Rudnicki, Y. Yamada, A.C. Pereira and J.E. Krieger, Meta-analysis of the association of 4 angiotensinogen polymorphisms with essential hypertension: a role beyond M235T? *Hypertension* **51** (2008), 778–783.
- [2] J. Mikkelsen, M. Perola, A. Penttilä, P.J. Goldschmidt-Clermont and P.J. Karhunen, The GPIIIa (beta3 integrin) PLA polymorphism in the early development of coronary atherosclerosis, *Atherosclerosis* **154** (2001), 721–727.
- [3] J.P. Casas, L.E. Bautista, L. Smeeth, P. Sharma and A.D. Hingorani, Homocysteine and stroke: evidence on a causal link from mendelian randomization, *Lancet* **365** (2005), 224–232.
- [4] C.N. Palmer, V. Young, M. Ho, A. Doney and J.J. Belch, Association of common variation in glutathione S-transferase genes with premature development of cardiovascular disease in patients with systemic sclerosis, *Arthritis Rheum* **48** (2003), 854–855.

- [5] G. Benze, J. Heinrich, H. Schulte, S. Rust, U. Nowak-Göttl, M.C. Tataru, E. Köhler, G. Assmann and R. Junker, Association of the GPIa C807T and GPIIIa PIA1/A2 polymorphisms with premature myocardial infarction in men, *Eur Hear J* **23** (2003), 325–330.
- [6] J. Frederiksen, K. Juul, P. Grande, G.B. Jensen, T.V. Schroeder, A. Tybjaerg-Hansen and B.G. Nordestgaard, Methylenetetrahydrofolate reductase polymorphism (C677T), hyperhomocysteinemia, and risk of ischemic cardiovascular disease and venous thromboembolism: prospective and case-control studies from the Copenhagen City Heart Study, *Blood* **104** (2004), 3046–3051.
- [7] R. Ward, Familial aggregation and genetic epidemiology of blood pressure, in: *Hypertension: Pathophysiology, Diagnosis and Management*, J.H. Laragh and D.M. Brenner, eds, Raven Press, New York, 1990, pp. 81–100.
- [8] G.A. MacGregor, N.D. Markandu, J.E. Roulston, J.C. Jones and J.J. Morton, Maintenance of blood pressure by the renin-angiotensin system in normal man, *Nature* **291** (1981), 329–331.
- [9] F. Paillard, D. Chansel, E. Brand, A. Benetos, F. Thomas, S. Czekalski, R. Ardaillou and F. Soubrier, Genotype-phenotype relationships for the renin-angiotensin-aldosterone system in a normal population, *Hypertension* **34** (1999), 423–429.
- [10] D.S. Wald, M. Law and J.K. Morris, Homocysteine and cardiovascular disease: evidence on causality from a meta-analysis, *BMJ* **325** (2002), 1202.
- [11] P. Frosst, H.J. Blom, R. Milos, P. Goyette, C.A. Sheppard, R.G. Matthews, G.J. Boers, M. den Heijer, L.A. Kluijtmans and L.P. van den Heuvel, A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase, *Nat Genet* **10** (1995), 111–113.
- [12] M.H. Prandini, E. Denarier, P. Frachet, G. Uzan and G. Marguerie, Isolation of the human platelet glycoprotein IIb gene and characterization of the 5-prime flanking region, *Biochem Biophys Res Commun* **156** (1988), 595–601.
- [13] P.J. Newman, R.S. Derbes and R.H. Aster, The human platelet alloantigens, PIA1 and PIA2, are associated with a leucine33/proline33 amino acid polymorphism in membrane glycoprotein IIIa, and are distinguishable by DNA typing, *J Clin Invest* **83** (1989), 1778–1781.
- [14] A.M. Carter, A.J. Catto, J.M. Bamford and P.J. Grant, Platelet GP IIIa PIA and GP Ib variable number tandem repeat polymorphisms and markers of platelet activation in acute stroke, *Arterioscler Throm Vasc Biol* **18** (1998), 1124–1131.
- [15] P.L. Zusterzeel, W. Visser, W.H. Peters, H.W. Merkus, W.L. Nelen and E.A. Steegers, Polymorphism in the glutathione S-transferase P1 gene and risk for preeclampsia, *Obstet Gynecol* **96** (2000), 50–54.
- [16] P.L. Zusterzeel, W.H. Peters, M.A. De Bruyn, M.F. Knapen, H.M. Merkus, and E.A. Steegers, Glutathione S-transferase isoenzymes in decidua and placenta of preeclamptic pregnancies, *Obstet Gynecol* **94** (1999), 1033–1038.
- [17] F. Cambien and L. Tire, Genetics of cardiovascular diseases: from single mutations to the whole genome, *Circulation* **116** (2007), 1714–1724.
- [18] R. Lisker, E. Ramirez, R. Perez-Briceño, J. Granados and V. Babinsky, Gene frequencies and admixture estimates in four Mexican urban centers, *Hum Biol* **62** (1990), 791–801.
- [19] C. Bekker-Mendez, J.K. Yamamoto-Furusho, G. Vargas-Alarcón, D. Ize-Ludlow, J. Alcocer-Varela and J. Granados, Haplotype distribution of class II MHC genes in Mexican patients with systemic lupus erythematosus, *Scand J Rheumatol* **27** (1998), 373–376.
- [20] C. Gorodezky, C. Alaez, M.N. Vázquez-García, G. de la Rosa, E. Infante, S. Balladares, R. Toribio, E. Pérez-Luque and L. Muñoz, The genetic structure of Mexican Mestizos of different locations: tracking back their origins through MHC genes, blood group systems, and microsatellites, *Hum Immunol* **62** (2001), 979–991.
- [21] Instituto Nacional de Estadística Geografía e Informática (INEGI), La población indígena en México, www.inegi.org.mx (2004).
- [22] J. Sambrook and D. Russell, Preparation and analysis of eukaryotic genomic DNA, in: *Molecular Cloning: A Laboratory Manual*, C. Nolan, ed., Cold Spring Harbor Laboratory Press, New York, 2001, pp. 6.1–6.31.
- [23] J. Reynolds, B.S. Weir and C.C. Cockerham, Estimation of the coancestry coefficient: basis for a short-term genetic distance, *Genetics* **105** (1983), 767–779.
- [24] Instituto Nacional Indigenista, Información básica sobre los pueblos indígenas de México, México, Instituto Nacional Indigenista, www.sedesol.gob.mx (1999).
- [25] L. Cavalli-Sforza, Genes, peoples, and languages, *Proc Natl Acad Sci USA* **94** (1997), 7719–7724.
- [26] J. Lin, M.R. Spitz, Y. Wang, M.B. Schabath, I.P. Gorlov, L.M. Hernandez, P.C. Pillow, H.B. Grossman and X. Wu, Polymorphisms of folate metabolic genes and susceptibility to bladder cancer: a case-control study, *Carcinogenesis* **25** (2004), 1639–1647.
- [27] X. Miao, D. Xing, W. Tan, J. Qi, W. Lu and D. Lin, Susceptibility to gastric cardia adenocarcinoma and genetic polymorphisms in methylenetetrahydrofolate reductase in an at-risk Chinese population, *Cancer Epidemiol Biomarkers Prev* **11** (2002), 1454–1458.
- [28] T. Keku, R. Millikan, K. Worley, S. Winkel, A. Eaton, L. Biscocho, C. Martin and R. Sandler, 5,10-Methylenetetrahydrofolate reductase codon 677 and 1298 polymorphisms and colon cancer in African Americans and whites, *Cancer Epidemiol Biomarkers Prev* **11** (2002), 1611–1621.
- [29] O.M. Mutchinick, M.A. Lopez, L. Luna, J. Waxman and V.E. Babinsky, High prevalence of the thermolabile methylenetetrahydrofolate reductase variant in Mexico: a country with a very high prevalence of neural tube defects, *Mol Genet Metab* **68** (1999), 461–467.
- [30] I.P. Dávalos, N. Olivares, M.T. Castillo, J.M. Cantú, B. Ibarra, L. Sandoval, M.C. Morán, M.P. Gallegos, R. Chakraborty and F. Rivas, The C677T polymorphism of the methylenetetrahydrofolate reductase gene in Mexican mestizo neural-tube defect parents, control mestizo and native populations, *Ann Genet* **43** (2000), 89–92.
- [31] X. Qian, Z. Lu, M. Tan, H. Liu and D. Lu, A meta-analysis of association between C677T polymorphism in the methylenetetrahydrofolate reductase gene and hypertension, *Eur J Hum Genet* **15** (2007), 1239–1245.
- [32] S. Szamosi, Z. Csiki, E. Szomják, E. Szolnoki, G. Szoke, Z. Szekanecz, G. Szegedi, Y. Shoenfeld and G. Szucs, Plasma homocysteine levels, the prevalence of methylenetetrahydrofolate reductase gene C677T polymorphism and macrovascular disorders in systemic sclerosis: risk factors for accelerated macrovascular damage? *Clin Rev Allergy Immunol* **36** (2009), 145–149.
- [33] G. Gorgone, F. Ursini, C. Altamura, F. Bressi, M. Tombini, G. Curcio, P. Chioyenda, R. Squitti, M. Silvestrini, R. Ientile, F. Pisani, P.M. Rossini and F. Vernieri, Hyperhomocysteinemia, intima-media thickness and C677T MTHFR gene

- polymorphism: a correlation study in patients with cognitive impairment, *Atherosclerosis* **206** (2009), 309–313.
- [34] I.P. Kosmas, A. Tatsioni and J.P. Ioannidis, Association of C677T polymorphism in the methylenetetrahydrofolate reductase gene with hypertension in pregnancy and preeclampsia: a meta-analysis, *J Hypertens* **22** (2004), 1655–1662.
- [35] P. Canto, T. Canto-Cetina, R. Juárez-Velázquez, S. Canizales-Quinteros, H. Rosas-Vargas, H. Rangel-Villalobos, A.C. Velázquez-Wong, G. Fernández, M.T. Villarreal-Molina and R. Coral-Vázquez, Methylenetetrahydrofolate reductase C677T and glutathione S-transferase P1 A313G are associated with a reduced risk of preeclampsia in Maya-Mestizo women, *Hypertens Res* **31** (2008), 1015–1019.
- [36] L. Tiret, Gene-environment interaction: a central concept in multifactorial diseases, *Proc Nutr Soc* **61** (2002), 457–463.
- [37] K. Hemminki, J. Lorenzo Bermejo and A. Forsti, The balance between heritable and environmental aetiology of human disease, *Nat Rev Genet* **7** (2006), 958–965.
- [38] H. Akasaka, T. Katsuya, S. Saitoh, K. Sugimoto, Y. Fu, S. Takagi, H. Ohnishi, H. Rakugi, N. Ura, K. Shimamoto and T. Ogihara, Effects of angiotensin II type 1 receptor gene polymorphisms on insulin resistance in a Japanese general population: The Tanno-Sobetsu Study, *Hypertens Res* **29** (2006), 961–967.
- [39] L.J. Rasmussen-Torvik, K.E. North, C.C. Gu, C.E. Lewis, J.B. Wilk, A. Chakravarti, Y.-P. C. Chang, M.B. Miller, N. Li, R.B. Devereux and D.K. Arnett, A population association study of angiotensinogen polymorphisms and haplotypes with left ventricular phenotypes, *Hypertension* **46** (2005), 1294–1299.
- [40] Y. Pei, J. Scholey, K. Thai, M. Suzuki and D. Cattran, Association of angiotensinogen gene T235 variant with progression of immunoglobulin nephropathy in Caucasian patients, *J Clin Invest* **100** (1997), 814–820.
- [41] J. Lim, S. La, K.C. Ng, K.-S. Ng, N. Saha and C.-K. Heng, Variation of the platelet glycoprotein IIIa  $PI^{A1/A2}$  allele frequencies in the three ethnic groups of Singapore, *Int J Cardiol* **90** (2003), 269–273.
- [42] K.R. Wagner, W.H. Giles, C.J. Johnson, C.Y. Ou, P.F. Bray, P.J. Goldschmidt-Clermont, J.B. Croft, V.K. Brown, B.J. Stern, B.R. Feaser, D.W. Buchholz, C.J. Earley, R.F. Macko, R.J. McCarter, M.A. Sloan, P.D. Stolley, R.J. Wityk, M.A. Wozniak, T.R. Price and S.J. Kittner, Platelet glycoprotein receptor IIIa polymorphism P1A2 and ischemic stroke risk: the Stroke Prevention in Young Women Study, *Stroke* **29** (1998), 581–585.
- [43] M.A. Carless, R.A. Lea, J.E. Curran, B. Appleyard, P. Gaffney, A. Green and L.R. Griffiths, The GSTM1 null genotype confers an increased risk for solar keratosis development in an Australian Caucasian population, *J Invest Dermatol* **119** (2002), 1373–1378.
- [44] J.H. Fowke, X.O. Shu, Q. Dai, A. Shintani, C.C. Conaway, F.L. Chung, Q. Cai, Y.T. Gao and W. Zheng, Urinary isothiocyanate excretion, brassica consumption, and gene polymorphisms among women living in Shanghai, China, *Cancer Epidemiol Biomarkers Prev* **12** (2003), 1536–1539.
- [45] M.A. Watson, R.K. Stewart, G.B. Smith, T.E. Massey and D.A. Bell, Human glutathione S-transferase P1 polymorphisms: relationship to lung tissue enzyme activity and population frequency distribution, *Carcinogenesis* **19** (1998), 275–280.
- [46] A. Hatagima, E.C. Costa, C.F. Marques, R.J. Koifman, P. Boffetta and S. Koifman, Glutathione S-transferase polymorphisms and oral cancer: a case-control study in Rio de Janeiro, Brazil, *Oral Oncol* **44** (2008), 200–207.
- [47] E.E. Hollenbach, El parentesco entre los triques de Copala, Oaxaca, *América Indígena* **33** (1973), 167–186.



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