

Review Article

Diagnostic Value of Salivary Markers in Neuropsychiatric Disorders

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A growing interest in the usability of saliva has been observed recently. Using saliva as a diagnostic material is possible because it contains a varied range of composites, organic and inorganic like proteins, carbohydrates, and lipids, which are secreted into saliva. Moreover, this applies to drugs and their metabolites. Saliva collection is noninvasive, and self-collection is possible. There is a lack of risk of injuries related to injection with needle, and it is generally safe. Human saliva has been successfully used, for example, in the diagnosis of many systemic diseases like cancers, autoimmune diseases, infectious diseases (HIV, hepatitis, and malaria), and endocrinological diseases, as well as diseases of the gastrointestinal tract. Also, it is used in toxicological diagnostics, drug monitoring, and forensic medicine. The usefulness of saliva as a biological marker has also been extended to psychiatry. The specificity of mental illness and patients limits or prevents cooperation and diagnosis. In many cases, the use of saliva as a marker seems to be the most sensible choice.

1. Introduction

At present, growing interest in the usability of saliva has been observed [1–4]. Human saliva takes part in the protection against different pathogens of oral tissues and upper respiratory and digestive systems [1, 2].

One of the most important roles of saliva is to provide the right environment for oral mucosa and teeth. It protects against the variability of destructive biological or chemical substances and mechanical damage. Also, saliva plays a significant part in the primary phase of digestion and participates in the perception of different kinds of tastes. Moreover, saliva has antibacterial, antifungal, and antiviral properties due to the presence of immunoglobulins, lactoferrin, and lysozyme [4–6].

Using saliva as a diagnostic material is possible because it contains a varied range of composites, organic and inorganic

like proteins, carbohydrates, and lipids, which are secreted into saliva. This also applies to drugs and their metabolites [6–10]. Its components are very sensitive, and they have a great response to toxic substances. They also correlate to the real-time level of these markers. Moreover, saliva collection is noninvasive, and self-collection is possible. There are no risk of injuries related to injection with needle, and it is generally safe [2, 11, 12].

Hence, many studies recommended saliva as the model of noninvasive diagnostic material. Nowadays, human saliva might be used in the monitoring and the early diagnosis of different systemic diseases, such as infectious cardiovascular disorders and cancers [6, 13]. Analysis of the concentrations of various salivary components is becoming increasingly important in laboratory medicine and the monitoring of the therapeutic range of drugs [6, 14–19]. Currently, saliva is used in toxicological diagnostics, e.g., detection of drug

dependence and alcohol abuse [2, 5, 6, 11, 20–22], neurology, psychiatry [6, 23–25], and forensic medicine (DNA) [26] (Figure 1).

In recent years, the usefulness of saliva as a biological marker has also been extended to psychiatry. The specificity of mental illness and patients limits or prevents cooperation and diagnosis. In many cases, the use of saliva as a marker seems to be the most sensible choice (Figure 2).

2. Drug Monitoring

It was proved that the concentrations of drugs in saliva correlate with the level of the drug in the blood [6, 27–31]. Therapeutic drug monitoring is used to optimize the management of patients receiving drug therapy. It encompasses the quantity of drug concentrations in biologic fluids. It also correlates with the patient's clinical condition and helps recognize the need to change the dosage, for example. Saliva use in drug monitoring is valuable and results from reflecting the free non-protein-bound pharmacologically active component in the serum [13, 32].

One example is valproic acid, used not only in the treatment of epilepsy but also in psychiatry. It is used in schizophrenia along with other medications and as a second-line treatment for bipolar disorder. Drug determination in saliva can be a simple test checking whether the patient is taking the drugs systematically as well as drug toxicity. It also makes it possible to determine the approximate level in the serum without blood sampling [33]. Dwivedi et al. [34] showed that the mean ratio of saliva to serum-free valproic acid concentration indicates that the saliva levels can predict the free drug concentrations in serum, and it also shows the protein binding of valproic acid in both. Carbamazepine, methadone, nicotine, cocaine, amphetamines, or buprenorphine has also been measured in oral fluid [13, 32, 35].

3. Dementia

Recent studies showed that saliva might be a valuable marker of neurodegenerative diseases [36–39].

An example is dementia, which is characterized by progressive cognitive impairment and behavioral changes. There are five types of dementia, for now, namely, Alzheimer's disease, vascular dementia, Lewy body dementia, frontotemporal dementia, and mixed dementias [36, 38]. It is estimated that about 50% of all dementia instances are Alzheimer's disease [36, 39], in which amyloid β and tau protein accumulate in the central nervous system.

Amyloid β is one of the most significant sources of reactive oxygen species in patients with dementia. It is deposited in the brain and also in the peripheral regions like the nasal mucosa, lacrimal glands, or lingual glands (salivary gland epithelium cells) [24, 36].

It is proved that oligomer forms of amyloid β activate nicotinamide adenine dinucleotide phosphate-oxidase (NADPH), increase the formation of hydrogen peroxide, and increase reactive oxygen species production in the mitochondria. This happens through modulation of alcohol dehydrogenase activity, which binds α -ketoglutarate dehydrogenase and

amyloid β . Accumulation of amyloid β in the secretory epithelium of salivary glands in patients with dementia disrupts the local redox balance and is responsible for damage to the structure and function of salivary glands [24, 36]. Changes in the composition of saliva can involve worsening in the quality of life of patients with dementia. These changes may cause problems with swallowing, inflammatory and fungal lesions, and worse cavital digestion [24, 36, 40, 41].

It is possible that oxidative stress is a significant factor that might cause dysfunction of the salivary glands. Scientists compare this to the mechanism observed in metabolic syndromes, such as insulin resistance [36, 42], obesity [36, 43], and diabetes [36, 44, 45], or autoimmune diseases, such as Sjögren syndrome and rheumatoid arthritis [36, 46]. The newest studies show that saliva might be an alternative diagnostic material to blood plasma or serum. In cases of dementia, it is used as an indicator of redox homeostasis biomarkers [24, 36, 40]. Choromańska et al. [36] proved decreased antioxidant properties of saliva and increased levels of DNA products in dementia patients. Moreover, they showed oxidative damage of protein and lipid, with simultaneously reduced secretion of nonstimulated and stimulated saliva. They suggested that changes in salivary redox homeostasis are independent of systemic changes in the progression of dementia [36].

4. Alcohol Dependence

Alcohol consumption is a serious public health problem and has been associated with high mortality rates. The world's population of adults suffering from alcohol abuse is estimated at about 4.9%. More than 2% of the world's population is alcohol dependent, while in Europe, it is estimated at 4% and in America 3.4% [47]. The World Health Organization assessed that the problem of binge drinking concerns more than 7% of the world's population (over 16% in Europe and 13% in America). In the last years, binge drinking has become the dominant pattern of alcohol consumption among adults [47].

So far, some chronic alcohol markers have been found in saliva, namely, aminotransferases and gamma-glutamyl-transferase, ethanol, sialic acid, hexosaminidase A, and glucuronidase. Waszkiewicz et al. [11, 47, 48] suggested that alcohol such as methanol, diethylene, ethylene, and glycol and salivary glycoproteins like oral peroxidase, α -amylase, clusterin, haptoglobin, heavy and light chains of immunoglobulins, and transferrin may be possible alcohol markers. In addition, chronic drinking leads to disturbances in adaptive and innate immunities, like immunoglobulin A, peroxidase, and lactoferrin [11, 48].

Waszkiewicz et al. [1, 49] found increased activity or concentration of β -hexosaminidase and immunoglobulin A in binge drinking [1, 49]. They also showed specific changes in salivary immunity in binge drinkers and alcohol-dependent patients. Furthermore, it was showed that even a single high dose of alcohol (2 g/kg) increases the level of salivary immunoglobulin A [2, 50]. Binge drinking caused disturbances in innate salivary immunity (lysozyme). They found possible applicability of raised immunoglobulin A

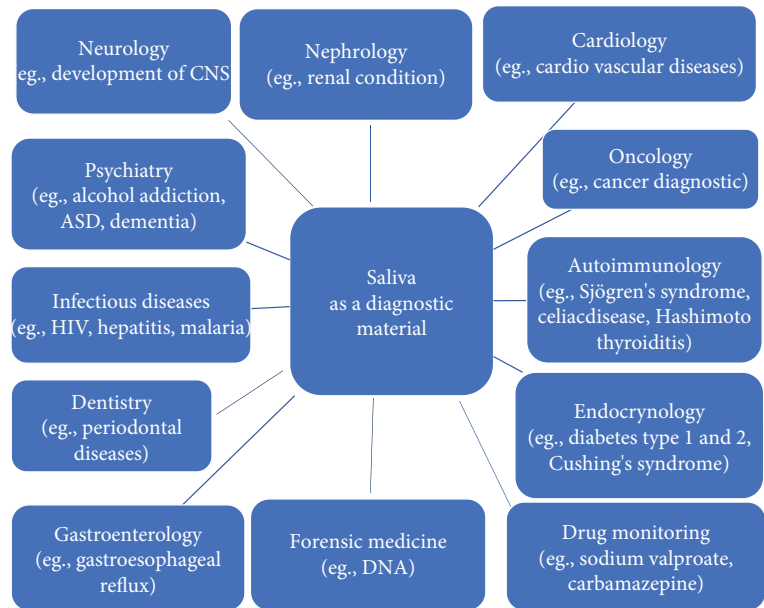


FIGURE 1: Saliva as a diagnostic material in medicine.

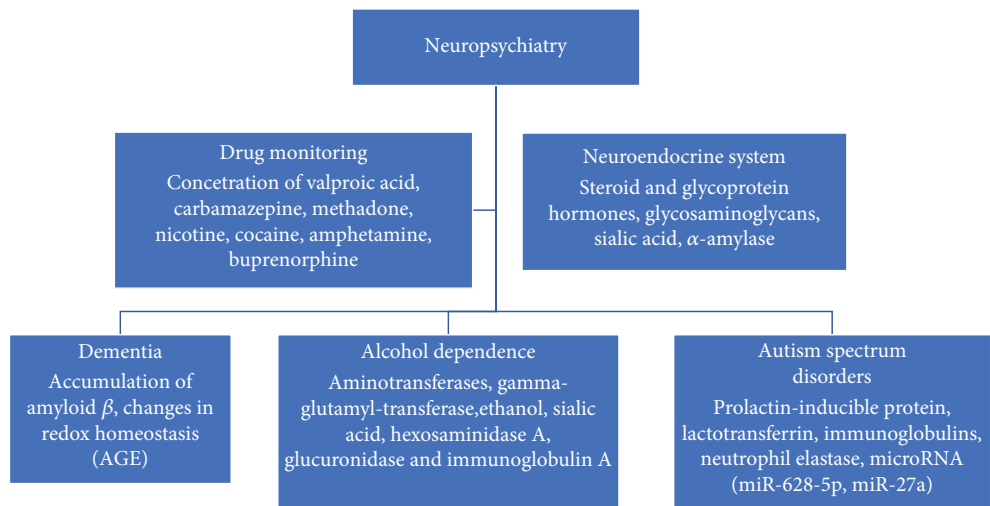


FIGURE 2: Saliva as a diagnostic material in neuropsychiatry.

concentration and oral peroxidase activity in binge and chronic drinking differentiation [2, 50].

5. Autism Spectrum Disorders

Autism spectrum disorder is a neurological and developmental disorder that affects communication and behavior [51]. It is included in the group of developmental disorders because symptoms begin early in childhood, mostly appearing in the first three years of life [52]. Scientists estimate the prevalence of autism spectrum disorders as 6 per 1,000. However, the frequency rates vary for each of the developmental disorders in the spectrum [52]. Early diagnosis and intervention might improve functional outcomes in children with autism spectrum disorder. Diagnosis, prognosis, and monitoring of

symptoms of autism spectrum disorder can also be helped with biomarkers [53].

Ngounou Wetie et al. [53] tried to optimize salivary proteomic biomarker methods and to identify initial biomarkers in children with autism spectrum disorders. They assumed that mass spectrometry-based proteomics could help expose biomarkers for autism spectrum disorder. Scientists have analyzed the salivary proteome in individuals with autism spectrum disorders compared to control subjects. They found statistically significant differences in several salivary proteins, e.g., the elevation of prolactin-inducible protein, lactotransferrin, Ig kappa chain C region, Ig gamma-1 chain C region, Ig lambda-2 chain C regions, neutrophil elastase, and polymeric immunoglobulin receptor and deletion in malignant brain tumors 1. Their achievement supports the concept that immune system and gastrointestinal

disturbances may be present in individuals with autism spectrum disorders [53].

Bhandary and Hari [54] studied the role of saliva as a biomarker and oral health status of children with autism spectrum disorders. They observed that salivary pH and buffering capacity were lower in children with autism spectrum disorders than their healthy siblings [54].

In another study, the authors measured salivary microRNA. They assumed that epigenetic mechanisms including microRNAs might contribute to the autism spectrum disorder phenotype by changing the neurodevelopmental gene networks. They showed the presence of the differential expression of 14 microRNAs (e.g., miR-628-5p, miR-27a), which are expressed in the developing brain. Furthermore, the impact of microRNAs on brain development and its correlates with neurodevelopmental behaviors were shown. MicroRNAs found in saliva showed high specificity and cross-validated utility. MicroRNAs seem to be a potential screening tool for autism spectrum disorders [55].

6. Neuroendocrine System

The use of saliva for monitoring steroid hormone levels has received increasing attention in recent years. The monitoring of steroid hormone levels is currently commercially available. There is nothing unusual in that, since levels of salivary steroid hormones reflect the free and thus the active level of these hormones in the blood [56]. The levels of cortisol, dehydroepiandrosterone, estradiol, estriol, progesterone, testosterone, etc. can be accurately assessed in saliva, being useful in evaluations of mood and cognitive-emotional behavior, in the diagnosis of premenstrual depression, to assess ovarian function, to evaluate risk for preterm labor and delivery, in full-term and preterm neonate monitoring, to study child health and development, as well as to predict sexual activity in adolescent males, or in Cushing's syndrome screening.

Protein hormones are too large to reach saliva through passive diffusion and can reach saliva through contamination from serum as a result of the outflow of gingival crevicular fluid or from oral wounds [14]. Protein hormones are therefore not useful in routine salivary analyses. Archunan et al. [57] presented that cyclic variations in salivary levels of glycosaminoglycans (GAGs) and sialic acid (SA) as well as in steroid (estrogens, progesterone) and glycoprotein (luteinizing hormone, LH) hormones can be helpful in predicting ovulation. SA and GAG content showed a distinct peak at ovulation during a normal menstrual cycle. Such hormonal changes in estrogen levels and a peak in LH might be the reason for proteoglycan degradation. Estrogen can inhibit the synthesis of the extracellular matrix, shifting normal proteoglycan turnover toward degradation processes. Identification of the period of ovulation in humans is critical in the treatment of infertility, which may result in mental disorders [21, 57, 58]. An easy, new, and noninvasive method of ovulation detection may help in the infertility treatment. Besides the salivary hormonal changes, changes in salivary GAGs and SA seem to show promise in the identification of the period of ovulation as well as the assessment of endocrine function.

Cortisol plays an important role as a marker of psychiatric disorders, such as anxiety and depression. Changes in cortisol levels appear in response to stress as well as emotional support. Chronic stress may lead to disease by activating the hypothalamic-pituitary-adrenocortical (HPA) axis. The correlation of cortisol levels in blood and saliva is extremely strong, and the noninvasive quantification of this hormone in saliva meets the detection criteria in biomedical research, both scientific and diagnostic [59–61].

Another parameter that is very helpful in assessing a neurotic disorder is alpha-amylase, which reflects catecholamines in the blood. Therefore, it reflects stress levels, reacting even faster than cortisol [62, 63].

Thus, further studies focusing on changes in salivary components during different physiological and pathophysiological states seem to be warranted.

7. Conclusions

Based on these properties, human saliva has successfully been used in the diagnosis of many systemic diseases, like cancers (ovarian, lung, breast, and pancreatic), autoimmune diseases (Sjögren's syndrome, celiac disease, and Hashimoto's thyroiditis), infectious diseases (HIV, hepatitis, and malaria), and endocrinological diseases (types 1 and 2 diabetes, Cushing's syndrome) as well as diseases of the gastrointestinal tract (gastroesophageal reflux disease). Also, it is used in toxicological diagnostics, drug monitoring, and forensic medicine. The usefulness of saliva as a biological marker has also been extended to psychiatry. Saliva is recommended as an excellent material for biochemical, toxicological, and immunological diagnostics of not only oral cavity or systemic diseases but also in the still unexplored field of neuropsychiatry.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

- [1] N. Waszkiewicz, S. D. Szajda, A. Jankowska et al., "The effect of acute ethanol intoxication on salivary proteins of innate and adaptive immunity," *Alcoholism, Clinical and Experimental Research*, vol. 32, no. 4, pp. 652–656, 2008.
- [2] N. Waszkiewicz, B. Galińska-Skok, A. Zalewska et al., "Salivary immune proteins monitoring can help detection of binge and chronic alcohol drinkers: preliminary findings," *Drug and Alcohol Dependence*, vol. 183, pp. 13–18, 2018.
- [3] N. Waszkiewicz, S. D. Szajda, A. Jankowska et al., "The effect of the binge drinking session on the activity of salivary, serum and urinary beta-hexosaminidase: preliminary data," *Alcohol and Alcoholism*, vol. 43, no. 4, pp. 446–450, 2008.
- [4] N. Waszkiewicz, B. Zalewska-Szajda, A. Zalewska et al., "Salivary lysozyme in smoking alcohol-dependent persons," *Folia Histochemica et Cytobiologica*, vol. 50, no. 4, pp. 609–612, 2012.
- [5] N. Waszkiewicz, B. Zalewska-Szajda, A. Zalewska et al., "Decrease in salivary lactoferrin output in chronically intoxicated alcohol-dependent patients," *Folia Histochemica et Cytobiologica*, vol. 50, no. 2, pp. 248–254, 2012.

- [6] S. Chojnowska, T. Baran, I. Wilińska, P. Sienicka, I. Cabaj-Wiater, and M. Knaś, "Human saliva as a diagnostic material," *Advances in Medical Sciences*, vol. 63, no. 1, pp. 185–191, 2018.
- [7] F. Amado, M. J. C. Lobo, P. Domingues, J. A. Duarte, and R. Vitorino, "Salivary peptidomics," *Expert Review of Proteomics*, vol. 7, no. 5, pp. 709–721, 2014.
- [8] E. Ganowicz, "Salivary diagnostics diseases of the oral cavity," *Dental and Medical Problems*, vol. 48, no. 3, pp. 421–430, 2011.
- [9] E. V. Kochurova and S. V. Kozlov, "The diagnostic possibilities of saliva," *Klinicheskaia Laboratornaia Diagnostika*, vol. 1, pp. 13–15, 2014.
- [10] S. Marti-Alamo, A. Mancheno-Franch, C. Marzal-Gamarra, and L. Carlos-Fabuel, "Saliva as a diagnostic fluid. Literature review," *Journal of Clinical and Experimental Dentistry*, vol. 4, no. 4, pp. e237–e243, 2012.
- [11] N. Waszkiewicz, S. Chojnowska, A. Zalewska, K. Zwierz, A. Szulc, and S. D. Szajda, "Salivary exoglycosidases as markers of alcohol dependence," *Alcohol and Alcoholism*, vol. 49, no. 4, pp. 409–416, 2014.
- [12] N. Waszkiewicz, E. M. Kratz, S. Chojnowska et al., "Long-term changes of salivary exoglycosidases and their applicability as chronic alcohol-drinking and dependence markers," *The World Journal of Biological Psychiatry*, vol. 20, no. 1, pp. 64–75, 2019.
- [13] L. A. Soares Nunes, S. Mussavira, and O. Sukumaran Bindhu, "Clinical and diagnostic utility of saliva as a non-invasive diagnostic fluid: a systematic review," *Biochemical Medicine*, vol. 25, no. 2, pp. 177–192, 2015.
- [14] E. Kaufman and I. B. Lamster, "The diagnostic applications of saliva- a review," *Critical Reviews in Oral Biology and Medicine*, vol. 13, no. 2, pp. 197–212, 2016.
- [15] T. D. Rees, "Drugs and oral disorders," *Periodontology 2000*, vol. 18, no. 1, pp. 21–36, 1998.
- [16] S. Al Kawas, Z. H. A. Rahim, and D. B. Ferguson, "Potential uses of human salivary protein and peptide analysis in the diagnosis of disease," *Archives of Oral Biology*, vol. 57, no. 1, pp. 1–9, 2012.
- [17] M. Klichowska-Palonna and T. Bachanek, "Possible use of saliva in the diagnostics and treatment review of the literature," *Przegląd Lekarski*, vol. 68, no. 2, pp. 114–117, 2011.
- [18] S. A. Kolesov and L. V. Korkotashvili, "The proteome of saliva and its diagnostic possibilities," *Klinicheskaia Laboratornaia Diagnostika*, vol. 60, no. 5, pp. 54–58, 2015.
- [19] D. T. Wong, "Saliva the body's mirror," *Dimensions of Dental Hygiene*, vol. 4, pp. 14–17, 2006.
- [20] N. Waszkiewicz, S. Chojnowska, A. Zalewska, K. Zwierz, A. Szulc, and S. D. Szajda, "Salivary hexosaminidase in smoking alcoholics with bad periodontal and dental states," *Drug and Alcohol Dependence*, vol. 129, no. 1–2, pp. 33–40, 2013.
- [21] N. Waszkiewicz, S. D. Szajda, A. Jankowska et al., "Catabolism of salivary glycoconjugates in acute ethanol intoxication," *Medical Science Monitor*, vol. 15, no. 8, pp. CR413–CR417, 2009.
- [22] N. Waszkiewicz, S. D. Szajda, A. Zalewska et al., "Alcohol abuse and glycoconjugate metabolism," *Folia Histochemica et Cytobiologica*, vol. 50, no. 1, pp. 1–11, 2012.
- [23] D. Gazzolo and F. Michetti, "Perinatal S100B protein assessment in human unconventional biological fluids: a minireview and new perspectives," *Cardiovascular Psychiatry and Neurology*, vol. 2010, Article ID 703563, 5 pages, 2010.
- [24] F. Bermejo-Pareja, D. Antequera, T. Vargas, J. A. Molina, and E. Carro, "Saliva levels of Abeta1–42 as potential biomarker of Alzheimer's disease: a pilot study," *BMC Neurology*, vol. 10, no. 1, p. 108, 2010.
- [25] T. L. Sletten, S. Vincenzi, J. R. Redman, S. W. Lockley, and S. M. W. Rajaratnam, "Timing of sleep and its relationship with the endogenous melatonin rhythm," *Frontiers in Neurology*, vol. 1, p. 137, 2010.
- [26] K. Ackermann, K. N. Ballantyne, and M. Kayser, "Estimating trace deposition time with circadian biomarkers: a prospective and versatile tool for crime scene reconstruction," *International Journal of Legal Medicine*, vol. 124, no. 5, pp. 387–395, 2010.
- [27] E. J. Cone, J. Clarke, and L. Tsanacis, "Prevalence and disposition of drugs of abuse and opioid treatment drugs in oral fluid," *Journal of Analytical Toxicology*, vol. 31, no. 8, pp. 424–433, 2007.
- [28] O. H. Drummer, "Drug testing in oral fluid," *Clinical Biochemistry Reviews*, vol. 27, no. 3, pp. 147–159, 2006.
- [29] A. J. Jenkins, J. M. Oyler, and E. J. Cone, "Comparison of heroin and cocaine concentrations in saliva with concentrations in blood and plasma," *Journal of Analytical Toxicology*, vol. 19, no. 6, pp. 359–374, 1995.
- [30] K. Langel, H. Gjerde, D. Favretto et al., "Comparison of drug concentrations between whole blood and oral fluid," *Drug Testing and Analysis*, vol. 6, no. 5, pp. 461–471, 2014.
- [31] D. Malamud, "Salivary diagnostics: the future is now," *Journal of the American Dental Association (1939)*, vol. 137, no. 3, pp. 284–286, 2006.
- [32] P. N. Patsalos and D. J. Berry, "Therapeutic drug monitoring of antiepileptic drugs by use of saliva," *Therapeutic Drug Monitoring*, vol. 35, no. 1, pp. 4–29, 2013.
- [33] W. Sobaniec, B. Wroński, K. Czerwińska-Ciechan, and L. Szczepaniak, "Monitoring sodium valproate treatment of epilepsy in the development years," *Neurologia i Neurochirurgia Polska*, vol. 21, no. 1, pp. 1–5, 1987.
- [34] R. Dwivedi, Y. K. Gupta, M. Singh et al., "Correlation of saliva and serum free valproic acid concentrations in persons with epilepsy," *Seizure*, vol. 25, pp. 187–190, 2015.
- [35] O. Beckonert, H. C. Keun, T. M. D. Ebbels et al., "Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts," *Nature Protocols*, vol. 2, no. 11, pp. 2692–2703, 2007.
- [36] M. Choromańska, A. Klimiuk, P. Kostecka-Sochoń et al., "Antioxidant defence, oxidative stress and oxidative damage in saliva, plasma and erythrocytes of dementia patients. Can salivary AGE be a marker of dementia?," *International Journal of Molecular Sciences*, vol. 18, no. 10, 2017.
- [37] M. Mousavi, P. Jonsson, H. Antti et al., "Serum metabolomic biomarkers of dementia," *Dementia and Geriatric Cognitive Disorders Extra*, vol. 4, no. 2, pp. 252–262, 2014.
- [38] M. Ragusa, P. Bosco, L. Tamburello et al., "miRNAs plasma profiles in vascular dementia: biomolecular data and biomedical implications," *Frontiers in Cellular Neuroscience*, vol. 10, p. 51, 2016.
- [39] E. Altunoglu, G. Guntas, F. Erdenen et al., "Ischemia-modified albumin and advanced oxidation protein products as potential biomarkers of protein oxidation in Alzheimer's disease," *Geriatrics & Gerontology International*, vol. 15, no. 7, pp. 872–880, 2015.

- [40] J. Figueira, P. Jonsson, A. Nordin Adolfsson, R. Adolfsson, L. Nyberg, and A. Öhman, "NMR analysis of the human saliva metabolome distinguishes dementia patients from matched controls," *Molecular BioSystems*, vol. 12, no. 8, pp. 2562–2571, 2016.
- [41] J. A. Ship, C. DeCarli, R. P. Friedland, and B. J. Baum, "Diminished submandibular salivary flow in dementia of the Alzheimer type," *Journal of Gerontology*, vol. 45, no. 2, pp. M61–M66, 1990.
- [42] U. Kołodziej, M. Maciejczyk, A. Miąsko et al., "Oxidative modification in the salivary glands of high fat-diet induced insulin resistant rats," *Frontiers in Physiology*, vol. 8, 2017.
- [43] M. Knaś, M. Maciejczyk, K. Sawicka et al., "Impact of morbid obesity and bariatric surgery on antioxidant/oxidant balance of the unstimulated and stimulated human saliva," *Journal of Oral Pathology & Medicine*, vol. 45, no. 6, pp. 455–464, 2016.
- [44] A. Y. Al-Maskari, M. Y. Al-Maskari, and S. Al-Sudairy, "Oral manifestations and complications of diabetes mellitus: a review," *Sultan Qaboos University Medical Journal*, vol. 11, no. 2, pp. 179–186, 2011.
- [45] A. Zalewska, M. Knaś, M. Maciejczyk et al., "Antioxidant profile, carbonyl and lipid oxidation markers in the parotid and submandibular glands of rats in different periods of streptozotocin-induced diabetes," *Archives of Oral Biology*, vol. 60, no. 9, pp. 1375–1386, 2015.
- [46] A. Zalewska, M. Knaś, N. Waszkiewicz, D. Waszkiel, S. Sierakowski, and K. Zwierz, "Rheumatoid arthritis patients with xerostomia have reduced production of key salivary constituents," *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology*, vol. 115, no. 4, pp. 483–490, 2013.
- [47] N. Waszkiewicz, B. Galińska-Skok, A. Nestsiarovich et al., "Neurobiological effects of binge drinking help in its detection and differential diagnosis from alcohol dependence," *Disease Markers*, vol. 2018, Article ID 5623683, 9 pages, 2018.
- [48] N. Waszkiewicz, W. Jelski, A. Zalewska et al., "Salivary alcohol dehydrogenase in non-smoking and smoking alcohol-dependent persons," *Alcohol*, vol. 48, no. 6, pp. 611–616, 2014.
- [49] N. Waszkiewicz, S. D. Szajda, A. Jankowska et al., "The effect of the binge drinking session on the activity of salivary, serum and urinary β -hexosaminidase: preliminary data," *Alcohol Alcohol*, vol. 43, no. 4, pp. 446–450, 2008.
- [50] I. Jastrzębska, A. Zwolak, M. Szczurek, A. Wawryniuk, B. Skrzydło-Radomańska, and J. Daniluk, "Biomarkers of alcohol misuse: recent advances and future prospects," *Gastroenterology Review*, vol. 2, pp. 78–89, 2016.
- [51] American Psychiatric Association, *Diagnostic and Statistical Manual of Mental Disorders*, American Psychiatric Association, Arlington, VA, USA, 5th edition, 2013.
- [52] C. J. Newschaffer, L. A. Croen, J. Daniels et al., "The epidemiology of autism spectrum disorders," *Annual Review of Public Health*, vol. 28, no. 1, pp. 235–258, 2007.
- [53] A. G. Ngounou Wetie, K. L. Wormwood, S. Russell, J. P. Ryan, C. C. Darie, and A. G. Woods, "A pilot proteomic analysis of salivary biomarkers in autism spectrum disorder," *Autism Research*, vol. 8, no. 3, pp. 338–350, 2015.
- [54] S. Bhandary and N. Hari, "Salivary biomarker levels and oral health status of children with autistic spectrum disorders: a comparative study," *European Archives of Paediatric Dentistry*, vol. 18, no. 2, pp. 91–96, 2017.
- [55] S. D. Hicks, C. Ignacio, K. Gentile, and F. A. Middleton, "Salivary miRNA profiles identify children with autism spectrum disorder, correlate with adaptive behavior, and implicate ASD candidate genes involved in neurodevelopment," *BMC Pediatrics*, vol. 16, no. 1, p. 52, 2016.
- [56] C. F. Streckfus and L. R. Bigler, "Saliva as a diagnostic fluid," *Oral Diseases*, vol. 8, no. 2, pp. 69–76, 2002.
- [57] G. Archunan, V. S. Prabhu, E. A. Orozco B, R. G. Guzman, and S. Alagendran, "Biochemical evaluation in human saliva with special reference to ovulation detection," *Indian Journal of Dental Research*, vol. 21, no. 2, pp. 165–168, 2010.
- [58] J. A. S. Richards, D. L. Russell, S. Ochsner, and L. L. Espey, "Ovulation: new dimensions and new regulators of the inflammatory-like response," *Annual Review of Physiology*, vol. 64, no. 1, pp. 69–92, 2002.
- [59] G. E. Miller, E. Chen, and E. S. Zhou, "If it goes up, must it come down? Chronic stress and the hypothalamic-pituitary-adrenocortical axis in humans," *Psychological Bulletin*, vol. 133, no. 1, pp. 25–45, 2007.
- [60] G. A. Carrasco and L. D. Van de Kar, "Neuroendocrine pharmacology of stress," *European Journal of Pharmacology*, vol. 463, no. 1-3, pp. 235–272, 2003.
- [61] P. La Marca-Ghaemmaghami, R. La Marca, S. M. Dainese, M. Haller, R. Zimmermann, and U. Ehlert, "The association between perceived emotional support, maternal mood, salivary cortisol, salivary cortisone, and the ratio between the two compounds in response to acute stress in second-trimester pregnant women," *Journal of Psychosomatic Research*, vol. 75, no. 4, pp. 314–320, 2013.
- [62] M. R. Rashkova, L. S. Ribagin, and N. G. Toneva, "Correlation between salivary alpha-amylase and stress-related anxiety," *Folia Medica*, vol. 54, no. 2, pp. 46–51, 2012.
- [63] I. S. Lim, "Correlation between salivary alpha-amylase, anxiety, and game records in the archery competition," *Journal of Exercise Nutrition & Biochemistry*, vol. 20, no. 4, pp. 44–47, 2016.

