Cutaneous Tuberculosis: Clinicopathologic Arrays and Diagnostic Challenges

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1. Background

Cutaneous tuberculosis is a relatively uncommon, comprising 1-1.5% of all extrapulmonary tuberculosis manifestations, which manifests only in 8.4-13.7% of all tuberculosis cases [1]. Although rare, given its global prevalence, it is imperative for the clinicians to distinguish the many clinical variants of cutaneous tuberculosis and the masquerading infections—granulomatous syphilis, discoid lupus erythematosus, psoriasis, tuberculosis leprosy, sarcoidosis, actinomycosis, mycetoma, bacterial abscesses, and other skin infections—to preclude missed or delayed diagnosis [2, 3]. Most of the diagnostic methods for cutaneous tuberculosis confer lower sensitivity and specificities. Therefore, the physicians must resort to every possible test along with broad clinical consideration; hence the summation of positive rudiments would be auxiliary in precise diagnosis.

2. Epidemiology

Tuberculosis represents a major public health problem in Southeast Asia, since a larger proportion (45%) of total estimated 10.4 million infective cases were listed in the region [4]. Compiling the toll death rate, Southeast Region and African Regions accounted for 85% of total death due to tuberculosis [4]. TB ranks the 6th leading cause of death in Nepal [5]. The prevalence study was not done in Nepal due to impassiveness of government participation in the health sector; however, annually, 34,122 cases of tuberculosis were reported to NTP [6].

Tuberculosis is endemic in Nepal; limited cases of cutaneous tuberculosis were reported, however. The incidence of cutaneous tuberculosis in Central Nepal was reported as 0.1%; nonetheless, the exact incidence is still anonymous over the country. The clincioepidemiological study done in Nepal by Dwari et al. 2010 revealed tuberculosis verrucous (48%) as predominant clinical type [7]; however, on referencing to earlier studies, Lupus vulgaris was the most common (64%), followed by tuberculosis verrucosa cutis (19%) and papulonecrotic tuberculid (4%) [8]. Ironically, cases of cutaneous multidrug resistant tuberculosis (MDR-TB)—resistant with at least two of the most potent first-line anti-TB medications, isoniazid and rifampicin—and XDR-TB—MDR strains that are resistant to fluoroquinolones plus one of the injectables...
such as kanamycin, amikacin, and capreomycin—have also been reported from India and China abutting Nepal [9–11]. Nevertheless, the exact epidemiological entity of perchance MDR/XDR cutaneous tuberculosis cases is still unbeknownst or unreported from Nepal.

3. Etiological Agent

The main etiological agent of the Cutaneous tuberculosis is *Mycobacterium tuberculosis*—occasionally *M. bovis* or BCG vaccine (an attenuated strain of *M. bovis*) [12, 13].

*Mycobacterium tuberculosis* is a straight or slightly bent (rod-shaped), nonmotile, nonsporulated, bacillus, being 1 to 10 μm long and 0.2 to 0.6 μm wide; its most important feature is acid-fastness due to high lipid content in the cell wall. Approximately there are 4000 genes with most of them involved in the mechanism of immune system invasion and 200 of them for lipid metabolism; consequently, the pathogen is able to survive both inside and outside the phagocytic cells [14]. Meanwhile, as lipids are the main energy source of *Mycobacterium tuberculosis*, the pathogen is directly responsible for multiplying in host tissue and forming cellular walls [14, 15].

4. Route of Infection

Cutaneous tuberculosis can be acquired from hematogenous or lymphatic dissemination of a pulmonary focus or by direct inoculation. The pivotal factor for the clinical presentations prior to contact with bacilli is the host natural immune response, however.

Exogenous infection occurs with direct inoculation of bacilli into the skin of predisposed individuals (tuberculous chancre, tuberculosis verrucosa cutis) [1].

Endogenous infection is secondary to a preexisting primary focus and may result from contiguous (orificial tuberculosis, scrofuloderma), hematogenous (acute miliary tuberculosis, tuberculous gumma, and lupus vulgaris), or lymphatic dissemination (lupus vulgaris) [2, 16].

5. Classifications of Cutaneous Tuberculosis

Based on a Load of Pathogens

Based on a load of the pathogens on skin, the tuberculosis variant can be classified into two broad categories.

Multibacillary forms (easily detected in cutaneous tissue) include tuberculous chancre, scrofuloderma, orificial tuberculosis, acute miliary tuberculosis, and tuberculosis gumma [17, 18].

Paucibacillary forms (bacilli being sparse) include TB verrucosa cutis, tuberculoid, and lupus vulgaris [17, 18].

6. Clinical Manifestations of Cutaneous Tuberculosis

Cutaneous tuberculosis exhibits diverse clinical manifestations: inflammatory papules, verrucous plaques, suppurative nodules, chronic ulcers, and other atypical lesions [19].

7. Exogenous Cutaneous Tuberculosis

7.1. Tuberculosis Chancre. The direct inoculations of *Mtb* in the skin from the traumatic injuries or surgical procedures performed with unsterilized materials and even after tattoos or body piercing lead to acquired tuberculosis chancre. Progressing from a firm, painless, reddish-brown, slow-growing papule, or nodule, after 2 to 4 weeks it develops into the friable ulcers—tendency to bleed with a granular surface [20]. Furthermore, the bacilli disseminate to regional lymph nodes via lymph.

Presumptive identification can be done with histopathological examinations, where the acute neutrophilic inflammatory reaction prolific in AFB and necrotic areas are usually noticed [16]. Sequentially, the lesion acquires a granulomatous form with enlarged giant cells after 3 to 6 weeks with the reduced number of bacilli [20].

7.2. Tuberculosis Verrucosa Cutis. Tuberculosis verrucosa cutis, the usual exogenous form of tuberculosis, is more common in an anatomist, physicians, and bare-footed children of tropical zones, since the infection proceeds with an injured dermal layer [1]. The lesions—solitary, painless, and without adenopathy—are more seen commonly in the extremities prone to trauma [16]. The lesions jerk as erythematous papules to verrucous plaques with peripheral extension.

8. Endogenous Tuberculosis

8.1. Scrofuloderma. Scrofuloderma, also called colliquative cutis, is a common form of cutaneous tuberculosis; it results from direct extension from an underlying tuberculosis lesion in lymph node, bone, joints, or testicles [1, 2]. The neck, axillae, and groin are often involved, with the cervical lymph nodes as a common source of infection [1]. Early lesions appear as firm, painless, subcutaneous, and red-brown nodules which advanced to ulcers and discharging sinus [21]. Spontaneous healing may occur, leaving keloid scars, retractions, and the atrophic sequel [21].

8.2. Orificial Tuberculosis. Orificial tuberculosis—a very rare form of cutaneous tuberculosis—is clinically characterized by ulcerations at mucocutaneous orifices including mouth, nose, perianal region, and genitalia and adjacent skin, usually advanced form of lungs, intestinal, or genitourinary tuberculosis [22]. The lesions, about 1 to 3 cm in diameter, appear as friable, painful erythematous-to-yellowish papules and nodules, which may advance to painful ulcers [16]. Edema and inflammation are obvious in perilesional tissue.

8.3. Lupus Vulgaris. Lupus vulgaris is the most common form of cutaneous tuberculosis in Europe, India, and Nepal [8, 13, 16]. It is a chronic, progressive, paucibacillary form of cutaneous tuberculosis which occurs primarily in the previously sensitized individual [23, 24]. The infection occurs endogenously via lymphohematogenous route and occasionally via exogenous route—with drainage scar of scrofuloderma [25].

The most typical clinical feature of lupus vulgaris is a papulotubercular lesions commonly on the legs and buttocks,
which eventually coalesce into a plaque (Figures 1, 2(a), and 2(b)) [12]. The plaques grow peripherally, with serpiginous or verrucous borders, accompanied by central discoloration and atrophy [25]. Besides, the classic appearance is described as “apple jelly nodules” observed on diascopy [24, 26].

8.4. Tuberculous Gumma. Tuberculous gumma, also known as metastatic tuberculosis abscess, is an outcome of hematogenous dissemination of mycobacteria from primary focus especially in an immunocompromised host, scarcely in an immunocompetent host too [17, 27]. Clinically it may bear a semblance to scrofuloderma; few lesions affecting trunks and extremities with inconsistent subcutaneous nodules having tendency to ulcerate and drain caseous secretion are seen in tuberculous gumma [23].

8.5. Acute Miliary Tuberculosis. It is a rare presentation of cutaneous tuberculosis predominantly in severely immunocompromised host, demonstrating anergy. The bulk of cases have been increasing primarily due to coinfection with HIV with declining CD4 count below 100 cells/μL [28]. Clinically, diverse cutaneous lesions—erythema and erythematous-whitish or erythematous-purplish papules—may be noticed which later on break to form umbilication and crust formation leaving hypochromic scars [17].

9. Tuberculids

Tuberculids are acute or chronic cutaneous forms of tuberculosis, appearing with diverse clinical forms, having a propensity of hyperergic expressions, active TB, or disseminated forms [20]. The discrete relationship between tuberculids and TB continues to be debated because the clinical forms usually have a symmetrical distribution, tuberculous involvement (usually inactive) of viscera or lymph nodes, and the absence of AFB (low positivity to culture and PCR) in the lesions [16, 26].

9.1. Papulonecrotic Tuberculids. Papulonecrotic tuberculids are the commonly observed form of cutaneous in children and young people [29]. They appear as painless, symmetrical erythematous, or violaceous papulonodular lesions noted particularly around the face, ears, extensor areas of the trunk, extremities, and buttocks, leaving a depressed scar [26].

9.2. Lichen Scrofulosorum. Lichen scrofulosorum is an eruption of multiple, small, grouped, asymptomatic, firm, perifollicular, lichenoid papules or plaques often affecting children and adults with underlying diseases of bone and lymph nodes [16, 26]. The dermatosis leaves no scar after months or years. The onset of this tuberculid was speculated, after BCG vaccinations and in the patient infected with M. avium-intracellulare [30].

9.3. Erythema Induratum of Bazin. Erythema induratum of Bazin is a granulomatous lobular panniculitis, which appears as erythematous-purplish subcutaneous nodules usually in legs and thighs [26]. The nodules advance few centimeters in diameter forming deep ulcers with caseous discharges and leave pigmented scar without or after successful treatment.

Figure 1: Erythematous plaque (2 cm) of lupus vulgaris on right forearm of a 17-year-old female with a history of trauma forming a linear scar (4 cm), visiting TUTH.

The relapse, however, may occur in flares every 3-4 months with similar clinical presentations [1]. Besides, the tendency of coinfectivity with systemic diseases like sarcoidosis is the differential clinical diagnosis of erythema nodosum [16, 21].

10. Diagnosis of Cutaneous Tuberculosis

10.1. Differential Diagnosis. The precise diagnosis is often significantly deferred and delayed, as cutaneous TB is not routinely considered in the differential diagnosis due to the relative paucity of pathogens in lesions and varied clinical manifestations (Table 1) [2, 16, 19, 31–33]. Hence, differential diagnosis is obligatory for the successful clinical management and treatment.

10.2. Laboratory Diagnosis

10.2.1. Tuberculin Skin Test. This technique involves an injection of 0.1 ml tuberculin, purified protein derivatives (PPD) derived from the attenuated strain of M. tuberculosis, intradermally, and read after 48 to 72 hours; on positive interpretation, the induration diameter exceeds the measuring of 10mm. The reaction is the classic example of delayed hypersensitivity reaction, where sensitized T-cells by prior infection are recruited thereby releasing the lymphokine [34]. These lymphokines induce inductions through local vasodilation, edema, fibrin deposition, and recruitment of other inflammatory cells to the area [34, 35]. TST has the sensitivity between 33% and 96% and specificity of 62.5% with cutoff 10mm for cutaneous tuberculosis; the sensitivity, however, exceeds 97% in an unvaccinated population [36, 37]. Furthermore, on analyzing clinical forms of cutaneous tuberculosis separately, positivity, intensity of the tuberculin skin test also diverges (Table 2). Conclusively, neither a positive TST necessarily indicates active infection nor a negative TST rules out the infection persistence.

10.2.2. Immunological Tests (Interferon Gamma-Release-Assay). The FDA approved immunological tests, QuantIFERON and
Table 1: Clinical manifestations of cutaneous tuberculosis and its differential diagnosis.

<table>
<thead>
<tr>
<th>S. N</th>
<th>Classification of cutaneous tuberculosis</th>
<th>Diagnostic considerations</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Exogenous cutaneous Tuberculosis</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Tuberculosis chancre</td>
<td>sporotrichosis, leishmaniasis, atypical mycobacteriosis, syphilis, cat scratch disease and tularemia</td>
</tr>
<tr>
<td></td>
<td>Tuberculosis verrucosa cutis</td>
<td>paracoccidioidomycosis, leishmaniasis, sporotrichosis, tuberculosis verrucosa and chromomycosis. Lobomycosis, atypical mycobacteriosis, hypertrophic lichen planus, verrucous carcinoma, iododerma, bromoderma, verruca vulgaris, keratoacanthoma centrifugum and pyoderma vegetans</td>
</tr>
<tr>
<td>2</td>
<td>Endogenous cutaneous tuberculosis</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Scrofuloderma</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Orificial tuberculosis</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Tuberculids</td>
<td></td>
</tr>
</tbody>
</table>

Table 2: TST result in different forms of cutaneous tuberculosis.

<table>
<thead>
<tr>
<th>Clinical forms of cutaneous tuberculosis</th>
<th>Tuberculin skin test result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tuberculosis chancre</td>
<td>initially negative, but becomes positive during course of disease (usually after 15 days)</td>
</tr>
<tr>
<td>Tuberculosis verrucosa</td>
<td>strongly positive</td>
</tr>
<tr>
<td>Lupus vulgaris</td>
<td>usually positive</td>
</tr>
<tr>
<td>Scrofuloderma</td>
<td>strongly positive</td>
</tr>
<tr>
<td>Orificial tuberculosis</td>
<td>negative</td>
</tr>
<tr>
<td>Acute cutaneous miliary tuberculosis</td>
<td>negative</td>
</tr>
<tr>
<td>Papulonecrotic tuberculid</td>
<td>positive</td>
</tr>
<tr>
<td>Lichen scrofulosorum</td>
<td>positive</td>
</tr>
<tr>
<td>Erythema induratum of Bazin</td>
<td>positive</td>
</tr>
</tbody>
</table>
EliSpot, assess sensitizations to \textit{M. tuberculosis} by measuring the amount of INF gamma released by lymphocytes confronted with \textit{M. tuberculosis} specific antigens [16]. The sensitivity and specificity of QuantiFERON are 89\% and 99\%, respectively, while EliSpot has the sensitivity of 98.8\% and a specificity of 100\% [38]. Unlike tuberculin skin test (TST), it detects disease in patients who have been vaccinated against BCG (latent infection)—and active infection too.

These tests are still not in routine-practice in our midst, because of high cost and laborious cell extract procedure from culture to antigen preparation (particularly in EliSpot).

\textbf{10.2.3. Histopathology.} Histopathology of a skin biopsy shows granulomatous presentations as those of cutaneous diseases with different etiology—cutaneous leishmaniasis, tuberculous leprosy, superficial granulomatous pyoderma, cutaneous sarcoidosis, lupus miliaris disseminatus faciei, and chromomycosis [16, 19, 33]. Meanwhile, the exact elucidation in diagnosis of cutaneous tuberculosis could not be done; however, the characteristic feature (well-formed granulomas with absence of caseous necrosis, granulomas with caseous necrosis, and the presence of poorly formed granulomas with intense caseous necrosis) would be auxiliary to differentiate types of cutaneous tuberculosis (Table 3) [16, 19, 21, 26, 28, 33, 39].

The equivocal manifestation of cutaneous tuberculosis to correlate the histologic with clinical observations in an evidence-based diagnosis is imperfect and lacking pragmatics.

\textbf{10.2.4. Diagnosis by Test: Staining and Culture.} The mycobacterial cell wall is rich in complex lipids which resists the acid and alcohol; hence the pathogen is termed as acid-fast bacilli (AFB). Staining techniques include Ziehl-Neelsen (common in practice), Kinyoun, and fluorochrome-based techniques with auramine-rhodamine. Microscopic observation of AFB in staining of tissue or secretions enables the empiric therapy if there are sufficient clinical suspicions. However, this does not necessarily suggest the cutaneous tuberculosis, since the other pathogens like \textit{Nocardia}, \textit{Corynebacterium}, nontuberculous mycobacteria, and even artifacts may reveal acid-fast characteristics [38, 40].

Furthermore, the lower sensitivities of staining results in extrapulmonary compared to pulmonary tuberculosis limit the applicability of the test [16, 37, 38]. The cultures of the pathogen, \textit{Mycobacterium tuberculosis}, on specific solid media or by automatic detection of its metabolites in liquid media remain the gold standard method, for identifications and their drug sensitivities. However, the long generation time of the pathogens to grow and lower sensitivity of culture results for lesions and tissue samples attribute further challenges in prompt and accurate diagnosis of cutaneous tuberculosis [16, 38].

\textbf{10.2.5. Amplifications of Nucleic Acids (PCR).} The detection of \textit{Mycobacterium} genus using bacterial 16S ribosomal DNA with PCR assays is now termed as a milestone in a diagnosis of pulmonary tuberculosis and several forms of cutaneous tuberculosis. DNA present in a sample of fresh tissues, blood, or a paraffin block even formalin fixed paraffin embedded sections, is amplified and it can then be identified, confirming the presence of mycobacteria [16, 33, 41].

PCR assay has augmented sensitivity and specificity in the diagnosis of cutaneous tuberculosis (Table 4) [42–55]; nevertheless, like other diagnostic approaches it is inconclusive in paucibacillary forms due to unevenly microbial distributions [25, 45].

\textbf{10.2.6. Genotyping.} Genotyping, the recent advance in the diagnosis of cutaneous tuberculosis, has a tendency to separate atypical mycobacteria from Mtbb—and detect mutant if it persists inducing drug resistance in the pathogen. The major molecular typing methods—Spoligotyping, MIRU-VNTR (Mycobacterial Interspersed Repetitive Unit-Variable Number Tandem Repeats), and RFLP—detect \textit{Mycobacterium tuberculosis}, DNA, or RNA in clinical specimens by in vitro nucleic acid amplifications, empowering investigations into epidemiology, transmission, and PTB outbreaks [56]. The
Table 3: Histopathological features of cutaneous tuberculosis.

<table>
<thead>
<tr>
<th>Different forms of cutaneous tuberculosis</th>
<th>Histopathological features</th>
<th>Observation of AFB</th>
</tr>
</thead>
<tbody>
<tr>
<td>Well-formed granulomas with absence of caseous necrosis</td>
<td>epidermis may be atrophic or hypertrophic, featuring acanthosis, papillomatosis and even pseudo-epitheliomatous hyperplasia. Presence of well-formed tuberculous granulomas accompanied more often by Langhans giant cells, or foreign body-like granulomas in the reticular dermis.</td>
<td>infrequent</td>
</tr>
<tr>
<td>Lupus vulgaris</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lichen scrofulosorum</td>
<td>granulomas in upper dermis and around dermal appendages</td>
<td>not seen</td>
</tr>
<tr>
<td>Intermediate forms: granulomas with caseous necrosis</td>
<td>marked pseudoepitheliomatous hyperplasia of the epidermis with hyperkeratosis and dense inflammatory cell infiltrate consisting of neutrophils, lymphocytes, and giant cells. The presence of granulomatous infiltrates is a cardinal sign it varies according to the time of inoculation; in recent lesions there is the presence of necrotizing neutrophilic infiltrate with numerous AFB. At a later stage there is organization of granulomas skin consists of areas of an inflammatory infiltrate composed of lymphocytes, plasma cells, and neutrophils with focal superficial dermal areas of necrosis and abscess formation without true caseating granuloma. The presence of acid-fast bacilli with vascular thrombi is characteristic of these lesions</td>
<td>can be seen decreased number</td>
</tr>
<tr>
<td>Tuberculosis verrucosa cutis</td>
<td></td>
<td>can be seen</td>
</tr>
<tr>
<td>Primary cutaneous tuberculosis</td>
<td></td>
<td>not usually found</td>
</tr>
<tr>
<td>Acute miliary tuberculosis</td>
<td></td>
<td>not usually found</td>
</tr>
<tr>
<td>Tuberculosis orificialis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Papulonecrotic tuberculid</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Poorly formed granulomas with intense caseous necrosis</td>
<td>Massive central necrosis with abscess formation and in many cases, suppuration, traces of granulomas can be observed at periphery of the lesions</td>
<td>may be found</td>
</tr>
<tr>
<td>Scrofuloderma</td>
<td>Central ulceration with abundant caseous necrosis, surrounded by a rim of giant cells and macrophages can be observed</td>
<td>frequently detected</td>
</tr>
<tr>
<td>Metastatic abscesses and gumma</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table 4: Sensitivity and specificity of PCR in the diagnosis of cutaneous tuberculosis (literature review).

<table>
<thead>
<tr>
<th>References and date</th>
<th>No. of samples</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>(Lee et al. 2016)</td>
<td>574</td>
<td>51.1</td>
<td>100</td>
</tr>
<tr>
<td>(Tan et al. 2001)</td>
<td>105</td>
<td>Overall 73</td>
<td>not calculated</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(positivity of 55% in cases of tuberculous verrucosa and 60% in cases of lupus vulgaris; positivity of 54% for cases of erythema induratum)</td>
<td>(pauci-bacillary form)</td>
</tr>
<tr>
<td>(Chawla et al. 2009)</td>
<td>104</td>
<td>74.1</td>
<td>96.1</td>
</tr>
<tr>
<td>(Agarwal et al. 2017)</td>
<td>70</td>
<td>24.5</td>
<td>not calculated</td>
</tr>
<tr>
<td>(Salian et al. 1998)</td>
<td>60</td>
<td>73.6</td>
<td>100</td>
</tr>
<tr>
<td>(Ogusku et al. 2003)</td>
<td>37</td>
<td>43.7</td>
<td>90.4</td>
</tr>
<tr>
<td>(Negi et al. 2005)</td>
<td>37</td>
<td>95.2</td>
<td>100</td>
</tr>
<tr>
<td>(Abdalla et al. 2009)</td>
<td>34</td>
<td>88</td>
<td>83</td>
</tr>
<tr>
<td>(Hsiao et al. 2003)</td>
<td>34</td>
<td>56</td>
<td>not calculated</td>
</tr>
<tr>
<td>(Lall et al. 2017)</td>
<td>31</td>
<td>25.8</td>
<td>not calculated</td>
</tr>
<tr>
<td>(Khosravi et al. 2006)</td>
<td>30</td>
<td>75</td>
<td>not calculated</td>
</tr>
<tr>
<td>(Ramam et al. 2013)</td>
<td>28</td>
<td>25</td>
<td>73.7</td>
</tr>
<tr>
<td>(Khine et al. 2017)</td>
<td>25</td>
<td>52</td>
<td>not calculated</td>
</tr>
<tr>
<td>(Quiros et al. 1996)</td>
<td>20</td>
<td>85</td>
<td>not calculated</td>
</tr>
</tbody>
</table>

Clinical applicability testing of these genotyping techniques was also accessed in the patients with cutaneous tuberculosis in China by Ziang et al., 2017, with augmented sensitivity and specificity [57].

10.2.7. RFLP (Restriction Fragment Length Polymorphism). The gold standard in genotyping, IS6110-based restriction fragment length polymorphism (RFLP), has been for more than an epoch; however, it is laborious and costly and requires a large amount of chromosomal DNA [56].

10.2.8. Spoligotyping. Spoligotyping—commonly used to differentiate *Mycobacterium tuberculosis* complex strain—is based on polymorphisms of the chromosomal direct repeat (DR) locus, which contains a variable number of short DRs interspersed with nonrepetitive spacers [56, 57].

10.2.9. Mycobacterial Interspersed Repetitive Unit-Variable Number Tandem Repeat (MIRU-VNTR). Lately, the International consortium has proposed MIRU-VNTR as a standardized genotyping scheme, with 15- and 24-locus sets proven to have ample discriminatory power for tracing transmission and investigating the phylogenetics of tuberculosis [57].

11. Conclusions

In a limelight, almost all of the investigative methods confer lesser sensitivity and specificities for cutaneous tuberculosis, considering atypical erythema nodosum, nonspecific appearance, insufficiently elucidative radio-imaging approaches, histopathology features, and even microbial culture techniques too. The genotyping techniques, nevertheless, could be an assistant to cope with this diagnostic challenge, paradoxically beyond reach to the third world like ours, due to expensive running cost and wanting equipped laboratory setup. In this perspective, the clinicians must resort to every possible test, so that supporting positive rudiments would be ancillary in the early and precise diagnosis of cutaneous tuberculosis.

### Abbreviations

AFB: Acid-fast bacilli  
MDR-TB: Multiple drug resistant tuberculosis  
MIRU-VNTR: Mycobacterial Interspersed Repetitive Unit-Variable Number Tandem Repeat  
Mtb: *Mycobacterium tuberculosis*  
NTP: National Tuberculosis control Programme  
PCR: Polymerase chain reaction  
TST: Tuberculin skin test  
XDR-TB: Extensively drug resistant tuberculosis.

### Conflicts of Interest

The authors declare that they have no conflicts of interest.

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