**Introduction**

Biological stress is a response to physical, chemical, biological and emotional changes, consisting of a pattern of metabolic and behavioral reactions that helps to strengthen the organism (1). During stressful situations, the energy requirement of the organism is increased, resulting in enhanced generation of free radicals (2–4). Free radicals cause oxidation of nucleic acids and proteins. Free radicals also damage biomembranes, reflected by increased lipid peroxidation, thereby compromising cell integrity and function. During this process, the ability of the body’s defense system to combat the oxidative stress may diminish due to reduced anti-oxidants. If the stress level increases beyond the threshold limit of an individual, it results in decreased performance and stress-induced disorders. The management of unusual stress therefore has acquired enormous significance in day-to-day life. Such a management does not endeavor to eliminate stress but rather to raise the threshold level of the organism beyond which stress would start injuring and disturbing life processes. It is possible to support the body’s adaptation by using food supplements, dietary elements, herbs and minerals for increasing physical and mental performance, described in various oriental systems of medicine including the ancient Indian medical system Ayurveda. Such substances have been described as **Keywords:** hypoxia – cold – rectal temperature – oxidative stress

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**Anti-stress and Adaptogenic Activity of L-Arginine Supplementation**

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In the present study, oral supplementation of L-arginine in rats was evaluated for its anti-stress and adaptogenic activity using the cold (5°C)–hypoxia (428 mmHg)–restraint (C-H-R) animal model. A dose-dependent study of L-arginine was carried out at doses of 12.5, 25.0, 50.0, 100.0, 200.0 and 500.0 mg/kg body weight, administered orally 30 min prior to C-H-R exposure. The time taken by the rat to attain a rectal temperature of 23°C (\(T_{\text{rec}}\) 23°C) during C-H-R exposure and its recovery to \(T_{\text{rec}}\) 37°C at normal atmospheric pressure and 32 ± 1°C were used as biomarkers of anti-stress and adaptogenic activity. Biochemical parameters related to lipid peroxidation, anti-oxidants, cell membrane permeability, nitric oxide and stress, with and without administration of the least effective L-arginine dose, were measured in rats on attaining \(T_{\text{rec}}\) 23°C and \(T_{\text{rec}}\) 37°C. The least effective adaptogenic dose of L-arginine was 100.0 mg/kg body weight. The C-H-R exposure of control rats, on attaining \(T_{\text{rec}}\) 23°C, resulted in a significant increase in plasma malondialdehyde (MDA), blood lactate dehydrogenase (LDH) and a decrease in blood catalase (CAT) and plasma testosterone levels. On recovery (\(T_{\text{rec}}\) 37°C) of control rats, there was a further decrease in CAT and plasma testosterone, and an increase in LDH. L-Arginine supplementation resulted in a significant decrease in plasma MDA, an increase in blood superoxide dismutase (SOD), CAT levels maintained at control values and a lower increase in LDH compared with controls (45.3 versus 58.5% and 21.5 versus 105.2%) on attaining \(T_{\text{rec}}\) 23°C during C-H-R exposure and on recovery to \(T_{\text{rec}}\) 37°C. The results suggested that L-arginine possesses potent anti-stress activity during C-H-R exposure and recovery from C-H-R-induced hypothermia.

**Keywords:** hypoxia – cold – rectal temperature – oxidative stress
In strenuous conditions, the physical performance of the organism is dependent on the availability of appropriate macro- and micronutrients required in excess on account of their increased utilization during stressful situations (6). Supplementation with various macro- and micronutrient and herbal preparations has been evaluated for their adaptogenic activity during exposure to a stressful environment (7–10).

It has been suggested that amino acid supplementation might be able to increase human performance to a limited extent (11). An amino acid mixture supplementation was observed to enhance adrenocortical hormone, luteinizing hormone and follicle-stimulating hormone response to corticotropin-releasing hormone in athletes (12). Under certain metabolic, developmental or pathophysiological conditions, some of the non-essential amino acids become essential and are designated as ‘conditionally essential’. Arginine and glutamine are known to be conditionally essential amino acids (13). L-Arginine plays important roles in the urea cycle, protein synthesis, as a precursor of polyamines and creatine, and as a substrate for synthesis of nitric oxide (NO). NO was shown to be an endothelial-derived relaxation factor, a vasodilator, which acted as a modulator of vascular tone to regulate blood flow and blood pressure (14). NO is also involved in enhancement of the thermogenic function of brown adipose tissue in rats (15).

Panax ginseng has been shown to contain large amounts of arginine (16).

It was shown that endogenous plasma arginine levels decreased significantly after 30 min immobilization stress and remain suppressed during a 3.5 h post-stress period (17). In burn patients, there was a higher rate of arginine loss from the body and supplementation of arginine was required to maintain homeostasis and promote recovery (18). In the present study, the anti-stress and adaptogenic effect, if any, of L-arginine supplementation was studied in rats subjected to a comprehensive restraint (C-H-R) (19).

In the present study, two experiments were carried out, one for dose-dependent study of L-arginine supplementation in rats using the C-H-R animal model to evaluate the least effective dose for adaptogenic activity and another for studying biochemical parameters in animals administered the least effective single oral dose of L-arginine.

**Experimental Design**

In total, 42 overnight fasted Wistar strain male rats were used in this experiment. The L-arginine was dissolved in distilled water in the appropriate concentration. A single dose of L-arginine was given orally in a 0.5 ml volume through a gastric cannula to overnight fasted rats, 30 min prior to C-H-R exposure (19). The L-arginine doses studied were 12.5, 25, 50, 100, 200 and 500 mg/kg body weight. Six rats were used for each dose.

Six control rats were administered an equivalent amount of water orally 30 min prior to C-H-R exposure. The rats were exposed in a decompression chamber maintained at 5°C and a low atmospheric pressure of 428 mmHg pressure equivalent to an altitude of 4572 m. The rats were restrained and a rectal probe was inserted 2 cm past the rectum and kept there with the help of adhesive plaster. The rectal temperature ($T_{\text{rec}}$) of the rats was monitored once per minute, by using a 16-channel Isothermex Temperature Recorder (Columbus Instrument, Columbus, OH). When the rats attained a rectal temperature ($T_{\text{rec}}$) of 23°C, they were taken out of the chamber. The rats were allowed to recover to a normal $T_{\text{rec}}$ of 37°C at normal atmospheric pressure and ambient temperature 32 ± 1°C. The rats continued to be restrained during the recovery period.

Although the comfortable temperature for housing the animals was 25 ± 1°C, the temperature for recovery of $T_{\text{rec}}$ (37°C) was selected as 32 ± 1°C because this was close to the ambient temperature of the outside environment. A constant room temperature was maintained in all experiments as recovery time also depended on the ambient temperature. The time taken to attain $T_{\text{rec}}$ 23°C and its recovery to 37°C were used as a measure of endurance.

**Materials and Methods**

**Rats and Maintenance**

Randomly bred, healthy, adult albino rats of the Wistar strain, weighing 180 ± 20 g, from the animal colony of the Defence Institute of Physiology and Allied Sciences, Delhi, India, were used in this study. The experimental protocol was submitted to the animal ethics committee of the institute, and approval was obtained for conducting the experiments. The rats were kept in a room that was maintained at 25 ± 1°C with natural daylight. The room remained dark from 7 p.m. until 7 a.m. in the morning. The animals had free access to drinking water and food in pellet form (Lipton India Ltd., Calcutta, India). All the experiments were done on overnight fasted rats. The L-arginine used in the study was obtained from Sigma Chemical Company, St Louis, MO.

**Biochemical Analysis**

In the second experiment, 36 separate overnight fasted rats were used. Rats were divided into two groups, containing 18 rats in each. One group was given the least effective dose of L-arginine and the other served as control. Both arginine-treated and control rats were subdivided further into three groups, each group: (i) rats not exposed to C-H-R; (ii) rats exposed to C-H-R to a fall of $T_{\text{rec}}$ to 23°C; and (iii) rats exposed to C-H-R and recovered to $T_{\text{rec}}$ 37°C.

A single oral dose of L-arginine was administered orally to rats in a 0.5 ml volume, 30 min prior to C-H-R exposure. In control rats, the equivalent volume of water was administered orally 30 min prior to C-H-R exposure. About 4 ml of blood was collected from the orbital plexus of the eye, in heparinized tubes, of different group of rats both control and arginine
treated (exposed and unexposed) under mild ether anesthesia, on
attaining both $T_{rec} 23^\circ C$ and $T_{rec} 37^\circ C$. A portion of blood was
used to separate the plasma. Malondialdehyde (MDA), a marker
for lipid peroxidation, was measured in plasma by the method of
Douset et al. (20). Blood levels of lactate dehydrogenase (LDH), as a marker of cell membrane permeability, were esti-
mated (21). In plasma, the levels of nitric oxide (NO), as a
metabolite of nitrite and nitrate, were measured using diagnostic
kits obtained from Oxis International Inc., Portland, OR. Plasma
levels of testosterone, a stress marker (29), were estimated using
the radioimmunoassay kits obtained from Immunotech, Marseille,
Cedex, France. In blood, superoxide dismutase (SOD) was esti-
mated by the method of Beutler et al. (22), catalase (CAT) by the
method of Aebi (23) and reduced glutathione (GSH) by the
method of Beutler et al. (24). In blood and plasma, protein
was estimated by the method of Lowry et al. (25) for calculating the
specific activity of the enzymes.

**Statistics**

The results were analyzed using one-way analysis of variance
(ANOVA) for their statistical significance and expressed as the
mean ± SEM. The results of L-arginine-treated animals were
compared with the respective controls and $P$-values $<0.05$
were considered statistically significant.

**Results**

**Dose-dependent Studies**

The orally administered doses of 12.5, 25 and 50 mg/kg body
weight in rats were observed to have no significant effect on
the time taken to attain $T_{rec} 23^\circ C$ in comparison with controls.
However, L-arginine doses of 100, 200 and 500 mg/kg body
weight showed a significant increase in time taken to attain a
fall of $T_{rec}$ to $23^\circ C$. The time taken by the rats to recover $T_{rec}$
$37^\circ C$ from C-H-R-induced hypothermia was reduced signifi-
cantly at all the doses of L-arginine used, except the 500 mg/kg
body weight dose in comparison with control rats (Fig. 1).

The oral administration of L-arginine at a dose of 100 mg/kg body
weight showed a significant increase in time taken to attain a
fall of temperature to $37^\circ C$ during C-H-R exposure and recovery of rectal
温度 to $37^\circ C$. L-Arginine administration resulted in increased SOD levels in rats on attaining $T_{rec} 23^\circ C$, in
adaptogenic activity as evidenced by a significant increase
(53.5%) in the time taken to attain a fall of $T_{rec}$ to $23^\circ C$ and
significantly faster (39.6%) recovery ($T_{rec} 37^\circ C$) from C-H-R
exposure-induced hypothermia, in comparison with control rats (Fig. 1).

**Plasma MDA and Anti-oxidants**

The results regarding the oxidative stress, i.e. TBA-reactive
MDA levels and some of the circulating anti-oxidants (SOD,
CAT and GSH), of rats exposed to C-H-R stress with and without
prior intake of a single oral dose of L-arginine of 100 mg/kg body
weight (least effective dose) are shown in Table 1. Plasma MDA
showed a 19% increase in control rats on attaining $T_{rec} 23^\circ C$
during C-H-R exposure, but supplementation of L-arginine resulted
in MDA levels comparable with control values. In control rats
exposed to C-H-R, blood SOD levels showed a non-significant
decrease on attaining $T_{rec} 23^\circ C$, but increased significantly
on recovery ($T_{rec} 37^\circ C$). L-Arginine administration resulted in increased SOD levels in rats on attaining $T_{rec} 23^\circ C$, in

![Figure 1. Dose-dependent study of L-arginine in rats on time (in minutes)
taken to attain $T_{rec}$ $23^\circ C$ during C-H-R exposure and recovery of rectal
temperature to $37^\circ C$. *Significant in comparison with their respective control
values at $P < 0.05$.

<table>
<thead>
<tr>
<th>Dose in mg/kg body weight</th>
<th>Temperature in °C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.95 ± 0.1</td>
</tr>
<tr>
<td>12.5</td>
<td>1.13 ± 0.1*</td>
</tr>
<tr>
<td>25</td>
<td>1.02 ± 0.06</td>
</tr>
<tr>
<td>50</td>
<td>0.93 ± 0.04</td>
</tr>
<tr>
<td>100</td>
<td>0.76 ± 0.08*</td>
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<tr>
<td>200</td>
<td>0.73 ± 0.21</td>
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<td>500</td>
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</tbody>
</table>

**Table 1. L-Arginine (100 mg/kg) supplementation and plasma MDA and blood SOD, GSH and CAT levels**

<table>
<thead>
<tr>
<th></th>
<th>Unexposed</th>
<th>Exposed</th>
<th>Unexposed</th>
<th>Exposed</th>
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<tbody>
<tr>
<td></td>
<td>$T_{rec} 23^\circ C$</td>
<td>$T_{rec} 37^\circ C$</td>
<td>$T_{rec} 23^\circ C$</td>
<td>$T_{rec} 37^\circ C$</td>
</tr>
<tr>
<td>MDA (µmol/l)</td>
<td>6</td>
<td>0.95 ± 0.1</td>
<td>1.13 ± 0.1*</td>
<td>1.02 ± 0.06</td>
</tr>
<tr>
<td>SOD (U/mg protein)</td>
<td>6</td>
<td>4.72 ± 0.44</td>
<td>4.27 ± 0.49</td>
<td>6.4 ± 0.45*</td>
</tr>
<tr>
<td>CAT (U/mg protein)</td>
<td>6</td>
<td>15.4 ± 0.9</td>
<td>11.2 ± 1.3*</td>
<td>9.42 ± 0.8*</td>
</tr>
<tr>
<td>GSH (mg %)</td>
<td>6</td>
<td>21.67 ± 2.1</td>
<td>20.32 ± 1.9</td>
<td>21.46 ± 1.6</td>
</tr>
</tbody>
</table>

*Significant in comparison with their respective unexposed groups at $P < 0.05$. 
compared with the respective unexposed rats. The blood CAT activity in control rats exposed to C-H-R decreased on attaining $T_{\text{rec}}$ 23°C and on recovery ($T_{\text{rec}}$ 37°C), in comparison with unexposed rats. However, in rats administered L-arginine, blood CAT activity was comparable with the respective unexposed animals, both on attaining $T_{\text{rec}}$ 23°C and on recovery ($T_{\text{rec}}$ 37°C). No significant change was observed in the GSH levels of control rats on attaining $T_{\text{rec}}$ 23°C during C-H-R exposure and on recovery of $T_{\text{rec}}$ 37°C. The L-arginine administration had no significant effect on blood GSH levels.

**Blood LDH and Plasma NO and Testosterone**

The results of the blood LDH and plasma NO and testosterone levels are shown in Table 2. In control rats, blood LDH levels increased significantly (58%) on attaining $T_{\text{rec}}$ 23°C during C-H-R exposure and on recovery (105%) of $T_{\text{rec}}$ 37°C, in comparison with unexposed control rats. In rats administered L-arginine, the increase in blood LDH levels was less compared with controls (58% compared with 23.3 and 21.5 versus 105.2%). Both on attaining $T_{\text{rec}}$ 23°C and on recovery of $T_{\text{rec}}$ 37°C (Table 2). In control and L-arginine-treated rats, there was no change in the plasma NO levels on attaining $T_{\text{rec}}$ 23°C and recovery of $T_{\text{rec}}$ 37°C in comparison with the respective unexposed rats (Table 3). Plasma testosterone levels decreased significantly in both control and L-arginine-treated rats on attaining $T_{\text{rec}}$ 23°C and on recovery of $T_{\text{rec}}$ 37°C, in comparison with unexposed rats. The decrease was comparatively less in L-arginine-treated rats (Table 2).

**Discussion**

Dose-dependent studies were performed using a passive C-H-R animal model. In the C-H-R animal model, the time taken to attain $T_{\text{rec}}$ 23°C indicates exhaustion of the animal's energy resources or resistance to stress, and the time taken to recover $T_{\text{rec}}$ 37°C from C-H-R exposure-induced hypothermia ($T_{\text{rec}}$ 37°C), at normal atmospheric pressure and 32°C ambient temperature, indicates how fast the post-stress recovery of the animal is. The results of the dose-dependent study showed that for the two orally administered L-arginine doses (100 and 200 mg/kg body weight), the time taken to attain $T_{\text{rec}}$ 23°C and recovery ($T_{\text{rec}}$ 37°C) from C-H-R exposure-induced hypothermia was almost comparable. With the 100 mg dose, the stress resistance effect (i.e. the time taken to attain $T_{\text{rec}}$ 23°C) was slightly better than for the 200 mg/kg dose, while with the 200 mg/kg dose, the post-stress recovery effect was slightly better than with the 100 mg/kg body weight dose. This suggested that both the doses, i.e. 100 and 200 mg/kg, were effective. Hence out of these two doses of L-arginine, the lowest one, i.e. 100 mg/kg, was selected as the least effective anti-stress and adaptogenic dose. The L-arginine dose of 100 mg/kg body weight was used further to study the effect of L-arginine administration on some of the biochemical indices related to oxidative stress, NO levels and cell membrane permeability.

Hypoxia, cold and immobilization stressors present in the C-H-R animal model used are known to produce oxidative stress (2–4) and skeletal muscle fatigue (26). Free radicals cause oxidation of lipids, proteins and bases of nucleic acids. Free radicals also cause damage to biomembranes, reflected by lipid peroxidation, thereby compromising cell integrity and function. In the present study, plasma MDA levels also increased in control rats on attaining $T_{\text{rec}}$ 23°C during C-H-R exposure along with decreased blood CAT, suggesting oxidative stress. However, GSH and SOD levels showed no change on attaining $T_{\text{rec}}$ 23°C during C-H-R exposure; perhaps the stress was not sufficiently severe to cause changes in these parameters. On recovery of $T_{\text{rec}}$ 37°C from C-H-R exposure-induced hypothermia, the CAT values remained low but SOD showed an increase, which was reflected in normalized plasma MDA levels. The L-arginine (100 mg/kg body weight) administration resulted in decreased MDA levels, lower than in unexposed animals, both on attaining $T_{\text{rec}}$ 23°C during C-H-R exposure and on recovery of $T_{\text{rec}}$ 37°C. The observed decrease in oxidative stress was due to the better maintained CAT activity and increased SOD levels on attaining $T_{\text{rec}}$ 23°C and on recovery of $T_{\text{rec}}$ 37°C. This suggested an anti-oxidant effect of L-arginine administration. Lubec et al. (27) also reported similar findings of reduced lipid peroxidation product MDA in diabetic patients treated with L-arginine.

Exposure to stressful conditions increased cell membrane permeability, indicating cellular membrane damage. Herbert (28) observed altered cell membrane permeability as evidenced by a significant increase in circulating creatine phosphokinase (CPK) levels in rats restrained at 4°C for 2 h, and this increase correlated with the changes in rectal temperature. In the present study, blood LDH levels in control rats also increased, by 58 and 105% on attaining $T_{\text{rec}}$ 23°C and on recovery of

| Table 2. L-Arginine supplementation and circulating level of blood LDH, plasma NO and testosterone levels |
| --- | --- | --- | --- | --- |
| | n | Control Unexposed | Control Exposed $T_{\text{rec}}$ 23°C | Control Exposed $T_{\text{rec}}$ 37°C | L-Arginine-treated Unexposed | L-Arginine-treated Exposed $T_{\text{rec}}$ 23°C | L-Arginine-treated Exposed $T_{\text{rec}}$ 37°C |
| LDH (nmol/mg protein) | 6 | 24.8 ± 3.7 | 39.3 ± 4.7* | 50.9 ± 4.9* | 32.5 ± 6.7 | 47.3 ± 3.9* | 39.5 ± 4.8 |
| NO (μM) | 6 | 3.47 ± 0.7 | 2.92 ± 0.5 | 2.74 ± 0.4 | 3.57 ± 0.5 | 3.82 ± 0.6 | 2.90 ± 0.5 |
| Testosterone (ng/ml) | 6 | 2.62 ± 0.80 | 0.43 ± 0.02* | 0.23 ± 0.01* | 2.58 ± 0.70 | 0.53 ± 0.02* | 0.42 ± 0.02* |

*Significant in comparison with their respective unexposed groups at $P < 0.05$. 


discussion

L-arginine, the increase in blood LDH levels was less compared with controls (45.3 versus 58.5%, and 21.5 versus 105.2%) both on attaining $T_{\text{rec}}$ 23°C and on recovery of $T_{\text{rec}}$ 37°C (Table 2). In control and L-arginine-treated rats,

Discussion

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levels was also observed in control rats exposed to an organism’s loss of control or a defeat reaction during stress situations, while decreased testosterone levels are indicative of rats that were exposed to C-H-R stress.

Increased circulating testosterone levels have been reported in the organisms that are able to adapt successfully in stressful situations, while decreased testosterone levels are indicative of an organism’s loss of control or a defeat reaction during stress (29). In the present study, a significant decrease in testosterone levels was also observed in control rats exposed to Trec 25°C, which decreased further on recovery to Trec 37°C. l-Arginine supplementation showed no significant effect on testosterone levels on Trec 23°C during C-H-R exposure and recovery to Trec 37°C in comparison with control rats.

The result of the present study suggested that in rats the least effective anti-stress and adaptogenic dose of L-arginine supplementation was 100 mg/kg body weight. The observed anti-stress activity of L-arginine was due to its anti-oxidative effects, i.e. better maintained anti-oxidants (SOD and catalase), reduced lipid peroxidation (MDA) and a lower increase in blood LDH levels.

References

Received July 29, 2004; revised August 30, 2004; second revision September 30, 2004; accepted on November 4, 2004