Original Article

Isolation of Bioactive Compounds That Relate to the Anti-Platelet Activity of Cymbopogon ambiguus

I. Darren Grice,1 Kelly L. Rogers,2,3 and Lyn R. Griffiths3

1 Institute for Glycomics, Gold Coast campus, Griffith University, Queensland, 4222, Australia
2 Plate-forme d’imagerie dynamique, Institut Pasteur, Paris Cedex 15, France
3 Genomics Research Centre, Gold Coast campus, Griffith University, Queensland, 4222, Australia

Correspondence should be addressed to I. Darren Grice, d.grice@griffith.edu.au

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Infusions and decoctions of Cymbopogon ambiguus have been used traditionally in Australia for the treatment of headache, chest infections and muscle cramps. The aim of the present study was to screen and identify bioactive compounds from C. ambiguus that could explain this plant’s anti-headache activity. A dichloromethane extract of C. ambiguus was identified as having activity in adenosine-diphosphate-induced human platelet aggregation and serotonin-release inhibition bioassays. Subsequent fractionation of this extract led to the isolation of four phenylpropenoids, eugenol, elemicin, eugenol methylether and trans-isoelemicin. While both eugenol and elemicin exhibited dose-dependent inhibition of ADP-induced human platelet serotonin release, only eugenol displayed potent inhibitory activity with an IC50 value of 46.6 μM, in comparison to aspirin, with an IC50 value of 46.1 μM. These findings provide evidence to support the therapeutic efficacy of C. ambiguus in the non-conventional treatment of headache and inflammatory conditions.

1. Introduction

The native Australian lemongrass species, Cymbopogon ambiguus A. Camus. (Poaceae) is a strongly aromatic perennial grass found on rocky hillsides throughout the Northern Territory of Australia [1]. The leaves have been used traditionally to treat chest infections, sores, muscle cramps as well as headache and associated complaints (infusions and decoctions) [1–3]. Interestingly, a study on Australian medicinal plants identified very weak anti-viral (Ross River Virus) activity in a leaf extract of C. ambiguus [4], while another study found no anti-bacterial activity in extracts of Australian C. ambiguus against four Gram-positive and four Gram-negative bacterial species [5]. Little is known about the chemical constituents present in C. ambiguus apart from a GC-MS study by Barr et al. [3], which identified camphene, borneol, limonene, α-pinene, α-terpineol, camphor, isoborneol, 4-terpineol, myrcene, β-ocimene as being present in the essential oil. Nothing has been reported to date in relation to substantiating its use as a non-conventional traditional remedy for headache, apart from our original report showing that dichloromethane (DCM) (more potent extract) and methanolic (MeOH) extracts displayed potent inhibition of human platelet aggregation and serotonin (5-HT, 5-hydroxytryptamine) release [6]. To further investigate the basis for use of C. ambiguus as a remedy for headache, we report here the isolation and identification of principal bioactive constituents causing inhibition of human platelet aggregation and serotonin (5-HT) release.

Cymbopogon ambiguus is a headache remedy used in Australian traditional medicine for generalized headache disorders. Headaches associated with nausea, such as migraine are complex disorders, with abnormalities in platelet function reported. Altered 5-HT transport, decreased platelet 5-HT content, altered platelet cytosolic free-calcium concentrations and varying sensitivity to platelet agonists such as adenosine diphosphate (ADP) and collagen have been reported in association with headache/migraine episodes [7–14]. The serotonergic system is thus thought to play an important role in the pathophysiology of these disorders [15–20]. We and others have previously reported on the use of platelets as a model to assess the therapeutic serotonergic potential of tested chemicals in relation to potential headache/migraine treatment [6, 21, 22].
Cymbopogon ambiguus has been used as a traditional Australian aboriginal headache treatment and our studies have shown that it displays potent anti-platelet activity. To identify the bioactive constituent(s) responsible for this activity, lemongrass leaves were sequentially extracted with DCM then MeOH. Fractionation of the DCM extract led to the identification of four phenylpropanoids as the principal constituents. We describe herein the isolation and purification of these four phenylpropanoids from C. ambiguus along with pharmacological evaluation of two of these compounds. This is the first report identifying specific constituents that underpin the use of C. ambiguus as a non-conventional traditional Australian anti-headache medicine.

2. Methods

2.1. Plant Collection. Cymbopogon ambiguus (whole plant) was collected in 1998 in the vicinity of Alice Springs, Northern Territory, Australia. Identity of the material was confirmed by the Alice Springs Herbarium, with voucher specimens deposited at the Herbarium and also in the Genomics Research Centre, Gold Coast campus, Griffith University (sample ID: DN3081).

2.2. Extraction and Isolation. Dried whole plants (283 g) of C. ambiguus were powdered and exhaustively extracted (vigorous stirring in 2.5 l of solvent for 3 h at 22°C, then procedure repeated) with DCM followed by MeOH at room temperature. The DCM extract was then concentrated and dried under reduced pressure (50 mmHg, 30°C) to give a dark green aromatic solid residue (7.53 g) and the residue loaded onto a normal phase chromatography column (silica gel, 150 g, Merck, 60 μM, 4.3 × 11.7 cm). The column was then eluted by passing 600 ml volume through the column from 10% hexane in chloroform, to 100% hexane in 10% ethyl acetate (Banksia Scientific Co. Pty. Ltd.), hexane (Mallinckrodt chromAR HPLC), ethyl acetate (Banksia Scientific Co. Pty. Ltd.), hexane (Mallinckrodt chromAR HPLC) and methanol (Omnisolv, EM Science, Merck). 14C-5-hydroxytryptamine (14C-5-HT) (Amersham, specific activity 57 mCi mmol⁻¹, 50 μCi ml⁻¹). All solvents used were of HPLC grade, including acetonitrile (CH₃CN) (omnisolv. EM Science, Merck), chloroform (CHCl₃) (chromasolv. Riedel-de Haen), DCM (chromasolv. Riedel-de Haen), ethyl acetate (Banksia Scientific Co. Pty. Ltd.), hexane (Mallinckrodt chromAR HPLC) and methanol (Omnisolv, EM Science, Merck).

2.4. Characterization of Isolated Compounds. Structural characterization of the four phenylpropanoids was achieved by mass spectrometry (MS) (Bruker Daltonics BioAPEX 47e), extensive NMR experiments (Bruker AC-300, Karlsruhe, Germany) (1H, 3H-1H COSY, 1H-13C HMBC, 1JMOD) and comparison of 1H NMR (300 MHz, CDCl₃) data (see below) with previously reported assignments. Eugenol (1) [23]: 1H NMR δ 3.33 (d, 2H, J = 6.7 Hz, CH₂); 3.88 (s, 3H, OCH₃); 5.06 (dd, 1H, J = 2 Hz, 3J₁ = 10 Hz, HCHCH₂); 5.07 (dd, 1H, 3J₂ = 2 Hz, 3J₂ = 10 Hz, HCHCH₂); 5.50 (OH); 5.93 (ddd, 1H, 3J₁ = 10 Hz, Jtrans = 17 Hz, HCHCH₂); 5.50 (OH); 5.93 (ddd, 1H, 3J₁ = 10 Hz, Jtrans = 17 Hz, Jtrans = 6.7 Hz, HCHCH₂); 6.67–6.72 (m, 2H, CCHCH(OH), CCHCH(OH)); 6.86 (dd, 1H, Jortho = 8.5 Hz, Jmeta = 4.2 Hz, CCHCH(OH)). MS calculated for C₁₀H₁₃O₂ [M + H]⁺: 165.08. Found: 165.08.Elemicin (2) [24]: 1H NMR δ 3.33 (d, 2H, J = 6.6 Hz, CH₂); 3.82 (s, 3H, OCH₃); 3.84 (s, 6H, (H₂O)COCC(OH)₃C(OH)); 5.08 (dd, 1H, J = 1.8 Hz, 3J₁ = 10 Hz, HCHCH₂); 5.11 (dd, 1H, J = 1.8 Hz, 3J₁ = 10 Hz, HCHCH₂); 5.95 (ddd, 1H, J = 10 Hz, Jtrans = 17 Hz, HCHCH₂); 5.97 (dd, 1H, J = 10 Hz, Jtrans = 17 Hz, HCHCH₂); 6.40 (2H, 2 × aromatic H). MS calculated for C₁₁H₁₅O₃Na [M + Na]⁺: 231.11. Found: 231.11. Eugenol methylether (3) [23]; 1H NMR δ 3.31 (d, 2H, J = 6.6 Hz, HC = CH₂); 3.84 (3H, OCH₃); 3.85 (s, 3H, OCH₃); 5.04 (dd, 1H, J = 1.5 Hz, 3J₁ = 10.1 Hz, HCHCH₂); 5.05 (dd, 1H, J = 1.5 Hz, 3J₁ = 10.1 Hz, HCHCH₂); 5.95 (ddd, 1H, 3J₁ = 10.1 Hz, Jtrans = 16.8 Hz, HCHCH₂); 6.75 (m, 3H, Jortho = 8.4 Hz, Jmeta = 1.4 Hz, 3 × aromatic H). MS calculated for C₁₁H₁₅O₃ [M + H]⁺: 179.09. Found: 179.09. trans-Isoelemicin (4) [25]; 1H NMR δ 1.86 (dd, 3H, J = 6.4 Hz, 4J = 1.5 HCHCH₂); 3.81 (s, 3H, OCH₃); 3.83 (s, 6H, 2 × OCH₃); 6.13 (dd, 1H, Jtrans = 15.6 Hz, J = 6.4 Hz, HCHCH₂); 6.31 (dd, 1H, J = 1.5 Hz, Jtrans = 15.6 Hz, HCHCH₂); 6.54 (s, 2H, 2 × aromatic H). MS calculated for C₁₂H₁₇O₃ [M + H]⁺: 209.11. Found: 209.11.

2.5. Platelet Aggregation and [¹⁴C]-5HT Release Bioassays. The bioassays utilised are based on a modified published method, described previously by our group [26] and by Groenevegen and Heptinstall [27]. Fresh blood (45 ml) was collected from healthy human volunteers (20- to 35-year old, not taking aspirin or other drugs likely to interfere with platelet function for at least 2 weeks prior to blood sampling). Platelet aggregation and 5-HT release were measured simultaneously. [¹⁴C]-5HT (6 μl) (specific activity 50 μCi μl⁻¹) was added to 9 ml of citrated blood to label intracellular storage granules. Platelet-poor plasma (PPP) and platelet-rich plasma (PRP) fractions were prepared, with the PRP being adjusted to 300 × 10⁹ platelets/ml by dilution with PPP. A sample consisting of 450 μl of PPP and 100 μl of either, PBS with 1% ethanol or DMSO (control), or
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Cymbopogon ambiguus (283 g dry weight)

DCM extract (7.53 g)
Silica flash column hexane \(\rightarrow\) CHCl3
10% gradient elution

MeOH extract (16.84 g)

50% MeOH fraction (5.73 g)

C18 flash column H2O \(\rightarrow\) MeOH

Acetonitrile soluble portion of combined 50% + 60% hexane fraction (1.16 g)

50% aq. MeOH fraction (5.73 g)

100% H2O fraction active (601 mg)

Eugenol (1) (75 mg, 0.03%), 33.8 min
Elemicin (2) (520 mg, 0.18%), 49.1 min
Eugenol methyl ether (3) (322 mg, 0.11%), 50.7 min
trans-Iso-elemicin (4) (162 mg, 0.06%), 53.3 min

Figure 1: Bioassay-guided fractionation of C. ambiguus (whole plant) to isolate compounds (1–4).

Figure 2: HPLC chromatogram of the combined 50 : 50 and 60 : 40 hexane/CHCl3 fractions: eugenol (33.8 min); elemicin (49.1 min); eugenol methylether (50.7 min); trans-Iso-elemicin (53.3 min).

the test sample, was initially measured in order to set 100% aggregation (a decrease in optical density reflects an increase in platelet aggregation). The final concentration of ethanol or DMSO did not exceed 0.2% (v/v). Labeled PRP (450 μl) was then stirred (1000 r.p.m. at 37°C for 3 min in the presence of 100 μl of the C. ambiguus extract (or isolated compound) or PBS containing either 1% ethanol or DMSO (control). ADP (50 μl) was then added to samples, and platelet aggregation was monitored for 6 minutes. Platelet aggregation was measured in a four-channel platelet aggregometer (Monitor IV plus, Helena Laboratories, Beaumont, USA). Samples of PRP were incubated without the addition of ADP to measure the amount of [14C] 5-HT that was not taken up by platelets, and which may have been spontaneously released during the procedure (blank). ASA (50 μl, 14 mm) was then added to irreversibly inhibit further 5-HT release, and samples were placed on ice for ~5 min. Samples were then centrifuged (6000 r.p.m.) for 8 min, and duplicate 50 μl aliquots were taken from the supernatant and counted (Wallac 1450, MicroBeta counter) for determination of % [14C] 5-HT release. Percentage 5-HT release was calculated as follows:

\[
\text{Amount of } [14C] 5-HT - \text{blank} \times 100% \quad (1)
\]

Total counts – blank

IC50 values were generated using the software of Graphpad prism (3.0).

2.6. Bioethics Clearance. Blood samples for the platelet studies were collected by a qualified phlebotomist and the study had full ethical clearance from the Griffith University Ethics Committee for experimentation on human subjects. Informed written consent was obtained from all participants.

2.7. Statistical Analysis. Platelet aggregation and 5-HT release response, was expressed as a percentage of the control value. IC50 values (concentration producing 50% inhibition of the maximum response) were calculated by non-linear regression analysis of the dose-response curves using the software of Graphpad Prism (3.0). Results are presented as the means ± SEM of n experiments.
3. Results

3.1. Extraction. Both DCM and MeOH extracts of *C. ambiguus* produced a dark green residue that was strongly aromatic (lemon scented). The total weight of the DCM extract after drying was 7.53 g, and for the MeOH extract, 16.84 g.

3.2. Isolation and Structural Characterization. Silica column chromatography (10% gradient increments from 10:90 hexane:CHCl₃ through to 100:0 hexane:CHCl₃ elution) of the DCM fraction eluted an active 50:50–60:40 hexane fraction. The dried residue (weighing ~1.16 g) was resuspended in CH₃CN and fractionation of the soluble components utilising HPLC, afforded five compounds (Figures 1 and 2). Four of these compounds were purified further (purity was ascertained by HPLC analysis) and then subjected to MS and NMR analysis. The fifth compound (eluting at 54.11 min) (Figure 2) was not successfully purified and was therefore not structurally characterised in these studies.

The assignment of the structures for eugenol, elemicin, eugenol methylether and trans-iso-elemicin (Figure 3), which are known compounds, was established on the basis of the ¹H NMR and MS data. Chemical shifts were identical to those previously reported (see Methods section—Characterization of Isolated Compounds). The chemical structure and identity of each compound is shown in Figure 3. The isolation of these compounds was guided by the activity of the active hexane fraction (50% and 60%) in the ADP-induced platelet 5-HT release bioassay. Both eugenol and elemicin were investigated pharmacologically in further studies.

3.3. Platelet Aggregation and [¹⁴C]-5-HT Release Bioassay. For the platelet aggregation and 5-HT release studies, a submaximal concentration of ADP was used to stimulate the platelet response. Elemicin’s effect on inhibiting ADP-induced platelet aggregation was less potent than eugenol in our results and therefore we focussed our interests on eugenol. Effects of increasing eugenol concentrations (0, 46, 61 and 152 μM) on the ADP-induced platelet aggregation clearly showed that concentrations above 61 μM did not inhibit platelet aggregation further. Interestingly, eugenol and elemicin exhibited dose-dependent inhibition of ADP (2 μM)-induced platelet 5-HT release, although eugenol at a much lower concentration than elemicin (see Table 1 and Figure 4 (eugenol data only)). Concentration ranges of eugenol, elemicin and aspirin chosen to examine their inhibitory activity on human platelet 5-HT release were 2.8 × 10⁻⁶ to 2.8 × 10⁻⁴ M, 1.7 × 10⁻⁴ to 3.5 × 10⁻³ and 2.5 × 10⁻⁶ to 2.5 × 10⁻⁴ M, respectively. Despite structural similarities, there were remarkable differences in the level of activity between eugenol and elemicin [1]. Eugenol was up to 50 times more potent than elemicin, with IC₅₀ values calculated to be 46.6 μM and 1729.8 μM, respectively. ASA is an irreversible inhibitor of platelet cyclooxygenase (COX) activity, and still remains the standard to which other anti-platelet drugs are often compared [28]. The dose-dependent effect of eugenol compared favorably to ASA (Figure 4), which exhibited an IC₅₀ value of 46.1 μM in these studies.

4. Discussion

Bioassay-directed fractionation of the DCM extract of *C. ambiguus* afforded the isolation and structural characterization of four compounds, eugenol, elemicin, eugenol methylether and trans-iso-elemicin. Like 1,8-cineole (the major active constituent of *Melaleuca, Eucalyptus* and
previous studies using rabbit platelets, eugenol was found to be up to 1000 times more potent than elemicin and was found to compare favorably with indomethacin [34]. An additional study found eugenol to be 29 times more potent than aspirin at inhibiting arachidonic acid-induced human platelet aggregation [35]. Based on reports in the literature and the structural characteristics of eugenol methylether and trans-iso-elemicin, which were also isolated in the active fraction, these two compounds were not investigated further in our pharmacological studies [34]. Although, interestingly it has been demonstrated that eugenol methylether has anti-nociceptive effects on formalin-induced hyperalgesia [36]. However, our interests and discussion here focus on eugenol due to its principal activity in our bioassays.

Studies have demonstrated that eugenol has anti-inflammatory, anti-platelet (using rabbit platelets), anti-nociceptive and anti-ulcerogenic effects [37–42]. Furthermore, this drug has also been shown to potentiate GABA\(_A\) transmission [43, 44]. Eugenol is reported to cause inhibition of thromboxane \(A_2\) and \(B_2\) formation, without any influence on the lipoxygenase pathway and appears to be an inhibitor of COX activity similarly to aspirin. In addition, more recent data indicates that this drug inhibits the rise in intracellular Ca\(^{2+}\) caused by collagen, adrenalin, ADP and AA [38]. Hence, eugenol appears to have pharmaceutical features that might be considered ideal characteristics of a migraine treatment. In support of these earlier studies [29], our results provide evidence that eugenol does not have significant effects on the primary phase of ADP induced platelet aggregation. Concentrations of eugenol above 61 \(\mu\)M did not inhibit platelet aggregation further (Figure 5) and this result strongly suggests that eugenol selectively targets the second phase of platelet aggregation. This second phase of aggregation involves the “release reaction,” which is dependent on cytosolic increases in Ca\(^{2+}\) and the activation of PLC (phospholipase C) and the COX pathway [45], which mediate platelet dense granule secretion. Therefore, 5-HT release could be completely inhibited even when aggregation is not. In line with this, high concentrations of eugenol that inhibited 5-HT release only inhibited the second phase of platelet aggregation (Figures 4 and 5). The modulation of 5-HT release was the principal pathway we were interested in investigating.

Aspirin is a potent anti-inflammatory analgesic, known to cause inhibition of the COX pathway, namely the COX-1 and COX-2 isoforms of this enzyme. In general, aspirin is used to assist in prevention of heart attacks, strokes, arthritis, diabetes and migraines and may also slow the mental decline of old age or provide neuroprotection in a mouse model of Parkinson’s disease [46, 47]. Previous studies have also shown that a single concentration of eugenol, inhibits both COX-1 and COX-2 enzymes [48]. Our results show that eugenol exhibits pharmacological effects that resemble those of aspirin and this data supports previous evidence that eugenol is a COX inhibitor and acts as a potent anti-platelet drug. Interestingly, eugenol is reported to exhibit analgesic properties [38] and has also been used to treat gastrointestinal upsets and chronic diarrhoea and is approved by the Food and Drug Administration of the USA. It is noteworthy that other traditional medicines

### Table 1: IC\(_{50}\) Values for Components of \(C.\) ambiguus and aspirin against ADP-induced platelet 5-HT release.

<table>
<thead>
<tr>
<th>Compound</th>
<th>IC(_{50}) ((\mu)M)</th>
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<tr>
<td>Eugenol</td>
<td>46.6 ± 3.7</td>
</tr>
<tr>
<td>Elemicin</td>
<td>1729.8 ± 147.5</td>
</tr>
<tr>
<td>Acetylsalicylic acid (ASA, Aspirin)</td>
<td>46.1 ± 3.1</td>
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Results represent IC\(_{50}\) ± SEM (\(n = 4\)) as determined by the software of Graphpad prism (3.0).

**Prostanthera** sp., eugenol and elemicin are volatile monoterpenoids (i.e., C\(_{10}\) compounds) with anti-inflammatory affects [29]. Indeed, other similar compounds have previously been isolated from *Cymbopogon* sp., including, limonene and \(\alpha\)-terpineol, which are generally common components of citrus oils [30, 31]. Eugenol and elemicin have also been isolated from several other plants including *Myristica fragrans* (nutmeg) and *Syzygium aromaticum* (clove oil), where they are believed to be some of the major constituents responsible for the biological activities of these plants [29, 32, 33]. Elemicin has been identified previously in another Australian native, *Cymbopogon procerus* [3]; however, this is the first report of eugenol in the *Cymbopogon* sp. and indeed the first evidence for the presence of these compounds in the Australian native lemongrass species, *C. ambiguus*.

In these studies, eugenol was found to be up to 50-fold more active than elemicin at inhibiting 5-HT release from human platelets. From dose-dependant responses IC\(_{50}\) values of 46.6 \(\mu\)M and 1729.8 \(\mu\)M were determined, respectively, in our 5-HT release inhibition assay (Figure 4 and Table 1). In previous studies using rabbit platelets, eugenol was found to be up to 50-fold more potent than elemicin and was found to compare favorably with indomethacin [34]. An additional study found eugenol to be 29 times more potent than aspirin at inhibiting arachidonic acid-induced human platelet aggregation [35]. Based on reports in the literature and the structural characteristics of eugenol methylether and trans-iso-elemicin, which were also isolated in the active fraction, these two compounds were not investigated further in our pharmacological studies [34]. Although, interestingly it has been demonstrated that eugenol methylether has anti-nociceptive effects on formalin-induced hyperalgesia [36]. However, our interests and discussion here focus on eugenol due to its principal activity in our bioassays.

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known to modulate 5-HT activity have also been reported as treatments for gastrointestinal tract [49] and allergic disorders [50]. Eugenol is also likely to have potential in the treatment of other diseases related to platelet aggregation, such as, thrombosis, transient ischemia, inflammation, tumor growth and promotion of atherosclerosis [51, 52]. However, there is only limited data relating to its potential use as an anti-headache/migraine or anti-platelet drug [29]. Few side effects have been reported in relation to the use of eugenol. One report on its use as a traditional dental material does however document local irritative and cytotoxic effects along with hypersensitivity reactions when used in contact with soft oral tissues [53].

5. Conclusions

In conclusion, four known compounds (eugenol, elemicin, eugenol methyl ether and trans-isoelemicin) were isolated from the DCM extract of *C. ambiguus*. Eugenol and elemicin were identified as contributing the principal activity of this extract and demonstrated dose-dependent inhibition of ADP-induced human platelet $[^{14}C]$-5-HT release, with $IC_{50}$ values of 46.6 $\mu$M and 1729.8 $\mu$M, respectively. Eugenol was also identified as having potent affects on the second-phase of platelet activation. The results of these studies provide evidence identifying specific constituents (principally eugenol) that have anti-platelet activity (Figure 6) that establish the basis of the therapeutic activity and traditional use of *C. ambiguus* as a non-conventional remedy for headache conditions.

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