

Research Article

Antileishmanial Activity of Medicinal Plants Used in Endemic Areas in Northeastern Brazil

**Aline Cavalcanti De Queiroz,¹ Thays de Lima Matos Freire Dias,¹
Carolina Barbosa Brito Da Matta,¹ Luiz Henrique Agra Cavalcante Silva,¹
João Xavier de Araújo-Júnior,² Givanildo Bernardino de Araújo,³
Flávia de Barros Prado Moura,³ and Magna Suzana Alexandre-Moreira¹**

¹ Laboratório de Farmacologia e Imunidade (LaFI), Instituto de Ciências Biológicas e da Saúde, Universidade Federal de Alagoas, 57072-970 Maceió, AL, Brazil

² Laboratório de Pesquisa em Recursos Naturais, Universidade Federal de Alagoas, Maceió, AL, Brazil

³ Laboratório de Plantas Tropicais (LPT), PPG-Dibict, Instituto de Ciências Biológicas e da Saúde, Universidade Federal de Alagoas, Maceió, AL, Brazil

Correspondence should be addressed to Magna Suzana Alexandre-Moreira; suzana.magna@gmail.com

Received 10 April 2014; Revised 7 June 2014; Accepted 23 June 2014; Published 13 July 2014

Academic Editor: Ulysses Paulino Albuquerque

Copyright © 2014 Aline Cavalcanti De Queiroz et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

This study investigates the leishmanicidal activity of five species of plants used in folk medicine in endemic areas of the state of Alagoas, Brazil. Data were collected in the cities of Colonia Leopoldina, Novo Lino, and União dos Palmares, Alagoas state, from patients with cutaneous leishmaniasis (*Leishmania amazonensis*) who use medicinal plants to treat this disease. Plants extracts were tested at a concentration of 1–100 $\mu\text{g}/\text{mL}$ in all experiments, except in an assay to evaluate activity against amastigotes, when 10 $\mu\text{g}/\text{mL}$ was used. All plants extracts did not show deleterious activity to the host cell evidenced by LDH assay at 100, 10, and 1 $\mu\text{g}/\text{mL}$ after 48 h of incubation. The plants extracts *Hyptis pectinata* (L.) Poit, *Aloe vera* L., *Ruta graveolens* L., *Pfaffia glomerata* (Spreng.) Pedersen, and *Chenopodium ambrosioides* L. exhibited direct activity against extracellular forms at 100 $\mu\text{g}/\text{mL}$; these extracts inhibited growth by 81.9%, 82.9%, 74.4%, 88.7%, and 87.4%, respectively, when compared with promastigotes. The plants extracts *H. pectinata*, *A. vera*, and *R. graveolens* also significantly diminished the number of amastigotes at 10 $\mu\text{g}/\text{mL}$, inhibiting growth by 85.0%, 40.4%, 94.2%, and 97.4%, respectively, when compared with control. Based on these data, we conclude that the five plants exhibited considerable leishmanicidal activity.

1. Introduction

The Trypanosomatidae comprise a large group of parasitic protozoa, some of which cause important diseases in humans [1]. For example, leishmaniasis is an infectious disease that is transmitted by insects and is prevalent in Europe, Africa, Asia, and the Americas. It causes significant morbidity and mortality and thus constitutes a serious public health problem [2]. Each year, the parasite kills thousands and debilitates millions of people; 2 million new cases are reported annually, and 350 million people are at risk [3]. Leishmaniasis is endemic in 98 countries [4], 82% of which are low-income countries.

Leishmania is an obligatory intracellular parasite of monocytes and macrophages and has a digenetic life cycle that alternates between two stages: flagellated promastigotes, which develop in the midgut of the insect vector, and amastigotes, which multiply in the host macrophage [5, 6]. The disease can be caused by nearly 21 *Leishmania* species and encompasses a spectrum of clinical manifestations, including cutaneous lesions, oropharyngeal mucosa inflammation, and visceral infection [1]. Thus, leishmaniasis can be categorized broadly into three types: (i) cutaneous leishmaniasis, in which parasites remain at the site of infection and cause localized long-term ulceration, (ii) mucocutaneous leishmaniasis, a chronic destruction of mucosal tissue that develops from the

cutaneous disease in less than 5% of affected individuals; and (iii) visceral leishmaniasis, the most serious form, in which parasites leave the inoculation site and proliferate in liver, spleen, and bone marrow, resulting in host immunosuppression and ultimately death in the absence of treatment [3].

Tegumentary leishmaniasis in New World is referred to as American tegumentary leishmaniasis (ATL) a serious zoonosis specially endemic in a lot of areas of Latin America. It is caused by *Leishmania* species of the subgenera *Leishmania* (*Viannia*) and (*Leishmania*) and is distributed from the south of the United States to the north of Argentina [2, 7]. In Brazil, ATL is detected in all states and has shown a high incidence over the last 20 years. There is wide genetic diversity among the *Leishmania* parasites; at least seven *Leishmania* species have been described as the etiological agent of human cutaneous disease, with most cases being caused by *Leishmania* (*Viannia*) *braziliensis*. However, *L. amazonensis* is also important as a causative agent of ATL, having been reported in the northeast, southeast, and west-central parts of Brazil [7–9].

Unfortunately, antileishmanial drugs, which are mainly based on antimonial therapy, are toxic, and recently developed and tested vaccines have shown relatively low protection under field conditions [6]. Pentavalent antimony (SbV) compounds such as sodium stibogluconate and meglumine antimoniate are the first choice of therapy for leishmaniasis. Despite their extensive clinical use for several decades, the mechanism of action remains unclear [10]. Other drugs used to treat leishmaniasis include pentamidine and amphotericin B, but the use of these drugs has been limited due to their high toxicity and cost [11]. Recently, the oral drug miltefosine was approved for the treatment of human visceral *Leishmania* infections, and fluconazole taken orally is also effective against cutaneous leishmaniasis [12].

Because of the adverse side effects of these treatment regimens, considerable attention has been given to the discovery and development of new, less toxic chemotherapeutic agents [13]. In an ongoing search for improved and cheaper leishmanicidal agents, plant-derived products represent an attractive option. According to the World Health Organization (WHO), approximately 80% of the world's inhabitants rely on traditional medicines for their health care [14]. The discovery of artemisinin, a sesquiterpene lactone produced by *Artemisia annua*, as a pharmaceutical for the treatment of malaria promoted interest in the discovery of new compounds from plants with antiprotozoal activity, especially those plants used against *Leishmania* parasites [15].

Because it is a very common disease in the Brazilian northeast leishmanioses has been traditionally treated with folk medicine using native and cultivated plants. Currently, even with access to conventional treatments, patients continue to use plants that are seen by them as having the capacity to heal sores resulting from cutaneous leishmaniasis. This study investigated the leishmanicidal activity against *Leishmania amazonensis* of five species of plants, *Ruta graveolens* L., *Aloe vera* L., *Chenopodium ambrosioides* L., *Pfaffia glomerata* (Spreng.) Pedersen, and *Hyptis pectinata* (L.) Poit, popularly known as arruda, babosa, mastruz, meracilina, and sambacaitá, respectively, used in folk medicine in endemic

areas of the state of Alagoas, Brazil, to treat cutaneous leishmaniasis [16]. These plants also have broad medicinal use on the part of other traditional peoples in the Brazilian northeastern region [17].

2. Materials and Methods

2.1. Ethnobotanical Survey and Plant Collection. The five species of plants examined are broadly used in Brazilian folk medicine for various purposes, including folk therapy to treat leishmaniasis. These species were selected based on the popular use of plants to treat this disease in Alagoas state. The ethnobotanical research was carried out after the community members were fully informed of its purpose. Aerial parts of *H. pectinata* (MUFAL 4050), *A. vera* (MUFAL 4052), and *R. graveolens* (MUFAL 4051) were collected in the city of Novo Lino. Aerial parts of *P. glomerata* (MUFAL 4053) were collected in the city of Colônia Leopoldina. *C. ambrosioides* (MUFAL 4054) was collected in the city of União dos Palmares. The samples were identified in Natural History Museum/UFAL.

2.2. Preparation of Aqueous Extracts. Aerial parts of *H. pectinata*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* were dried in an oven at 40°C for 96 h, pulverized, and processed with watch by infusing. Fresh succulent leaves of *A. vera* were crushed in an electric grinder, and the resultant slurry was used as the aqueous extract from this plant. The solutions were filtered and sterilized by filtering through sterile 0.22 μm membranes. For the experiments, the dry weight of each aqueous extract per mL was measured to determine the amount of solution required to achieve a given concentration in each well.

2.3. Biological Assays

2.3.1. Parasite Culture. *L. amazonensis* (IFLA/BR/1967/PH8 strain) promastigotes were grown in culture medium (Schneider's medium (Gibco, Life Technologies, São Paulo, Brazil)) containing 10% fetal bovine serum (FBS, Gibco, Life Technologies, São Paulo, Brazil), 2% human urine, and gentamicin (10 $\mu\text{g}/\text{mL}$) at 27°C in a BOD incubator. This strain was kindly provided by the *Leishmania* collection of the Oswaldo Cruz Institute, Rio de Janeiro, Brazil.

2.3.2. Culture of J774 Murine Macrophages. This adherent-phenotype macrophage line was cultured in Dulbecco's Modified Eagle's medium (DMEM, Sigma, São Paulo, Brazil) with 10% FBS and gentamicin (10 $\mu\text{g}/\text{mL}$) at 37°C under 95% humidity and 5% CO₂.

2.3.3. In Vitro Cytotoxicity Assays. The deleterious effects of the aqueous extracts of *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* were determined by assessing their cytotoxicity on murine macrophages (J774 cell line). Briefly, cell suspensions containing 7.5×10^5 cells/mL were placed in a 96-well plate in triplicate and incubated at 37°C for 1 h. Then, each aqueous extract was added at three

serial dilutions starting at 100 $\mu\text{g}/\text{mL}$ (final volume: 200 μL ; concentrations: 100, 10, and 1 $\mu\text{g}/\text{mL}$). Cell growth medium free from aqueous extracts was used as a basal growth control. Pentamidine was used as a drug standard and was added at three serial dilutions: 100, 10, and 1 μM . Cytotoxicity that was unrelated to pentamidine was assessed using the solvent DMSO (Sigma) as a vehicle control in this assay (DMSO was used to solubilize the pentamidine). For the vehicle control, the volume used to dissolve the pentamidine was considered as the highest concentration and also serially diluted. After cells had been exposed to plant aqueous extract, pentamidine or DMSO for 48 h at 37°C and 5% CO_2 , the assay was performed in an absorbance microplate reader to measure lactate dehydrogenase (Doles) [18].

2.3.4. Antileishmanial Assay against *L. amazonensis*. The Ethics Committee of Federal University of Alagoas (No. 014869/2006-86) approved all experimental protocols described in this study.

The cytotoxic effects of the aqueous extracts of *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* against promastigote forms were determined. Stationary phase *L. amazonensis* promastigotes were plated in 48-well vessels (Nunc) at 1×10^6 cells per well in Schneider's medium, supplemented with 10% FBS, 2% human urine, and gentamicin (10 $\mu\text{g}/\text{mL}$). The aqueous extracts were added at three serial dilutions (100, 10 and 1 $\mu\text{g}/\text{mL}$), and cell growth medium free from aqueous extracts was used as a basal growth control. Pentamidine was used as the drug standard and was added at three serial dilutions: 100, 10, and 1 μM . After 3 d, the extracellular load of *L. amazonensis* promastigotes was estimated by Neubauer chamber counting of the number of extracellular motile promastigotes in Schneider's medium [19].

The aqueous extracts of *P. glomerata* and *C. ambrosioides* were only evaluated in the experiments measuring cytotoxicity against promastigotes and macrophages because of the low amount of plant material obtained. Because the aim of this work was to mimic the popular use of plants commonly used in endemic areas of Alagoas (Brazil), we only obtained plants from the places in which the populace obtains them.

To evaluate the antileishmanial activity of the plants against extracellular replication, murine macrophages (the J774 cell line) were plated in 48-well vessels at 1.5×10^5 cells per well in complete culture medium. The cells immediately received 1×10^6 stationary phase *L. amazonensis* promastigotes and were incubated in complete medium supplemented with 10% FBS at 37°C. After 4 h, monolayers were extensively washed with warm HBSS (Sigma) to remove extracellular parasites and nonadherent cells, leaving approximately 1×10^5 adherent macrophages. All cultures were grown in DMEM supplemented with 10% FBS. The aqueous extracts were added at three serial dilutions from 100 to 1 $\mu\text{g}/\text{mL}$, and cell growth medium free from aqueous extracts was used as a basal growth control. Pentamidine was used as a drug standard and was added at three serial dilutions (100–1 μM). Extracellular parasites were absent throughout this period. After 3 d, the infected macrophage monolayers were extensively washed, and the medium was replaced by 0.5 mL

Schneider's medium, supplemented with 10% FBS and 2% human urine [19]. The monolayers were cultured at 26°C for an additional 3 d. The intracellular load of *L. amazonensis* amastigotes was estimated based on the production of proliferating extracellular motile promastigotes in Schneider's medium [20, 21].

To assess the activity of the plant aqueous extracts against the amastigote stage of the parasites, we utilized a model of infection on a cover glass [22]. The murine macrophages (the J774 cell line) were prepared in 24-well vessels (Corning) at 2×10^5 adherent cells/well and infected with 2×10^6 promastigotes in glass coverslips, inside which was placed 1 mL of culture medium. The cultures were cultured with or without the plant aqueous extracts (10 $\mu\text{g}/\text{mL}$) for 3 d under 37°C, 7% CO_2 . After 3 d, the coverslips were washed and stained using the Panoptic staining kit (LB), and the numbers of intracellular amastigotes were counted per 100 macrophages. The results are presented as the number of amastigotes per 100 macrophages and as a percentage of the infected macrophages.

2.4. Statistical Analysis. Data obtained from the *in vitro* experiments are expressed as the mean \pm standard error of the mean (mean \pm S.E.M.) of triplicate cultures. Significant differences between treated and control groups were evaluated using ANOVA and Dunnett post hoc tests. Differences with a *P* value of <0.05 were considered significant.

3. Results and Discussion

Determining the absence of toxic effects on host cells is an important criterion that must be evaluated when searching for active compounds with therapeutic potential against *L. amazonensis*. Results show the degree of toxicity of *C. ambrosioides*, *H. pectinata*, *R. graveolens*, *A. vera*, and *P. glomerata* (and pentamidine, the positive control) as measured in the J774 murine macrophage model (Figure 1). All plants used in this study exhibited no activity that was deleterious to the host cell, as evident from the viability of macrophages treated with 100, 10, and 1 $\mu\text{g}/\text{mL}$ after 48 h of incubation, whereas LDH values of cultures treated with media only or vehicle. This finding is very important for *A. vera*, in particular, because its leaf exudate contains saponin, which is toxic to monocytes. In a previous *in vitro* study, which was performed using murine peritoneal macrophages and a human monocyte cell line U937 that had been exposed to *A. vera* leaf exudate (300 $\mu\text{g}/\text{mL}$), minimal cell death was observed [23], in agreement with the present work. Moreover, the treatment with pentamidine did not show cytotoxic effect to host cells; however this standard drug at 100 μM presented optical density 25% greater than the vehicle group (DMSO 0.1%).

Furthermore, the determination of plants cytotoxicity against host cells is important in order to determine the selectivity of these against *Leishmania*, in particular, against amastigotes, as these are the evolutionary forms found in the mammalian hosts. Therefore, it is important that the plants have toxicity against *Leishmania* and not against the host cell. Moreover, this test is important to determine the

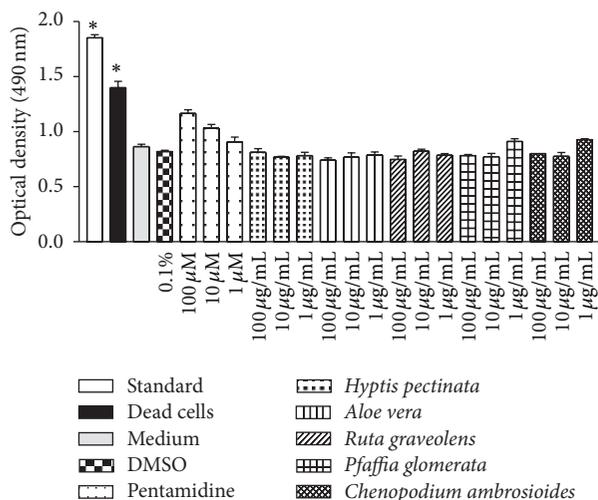


FIGURE 1: Cytotoxicity of *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* in mammalian cells. The experiments were performed three times independently, and each count was performed in triplicate. The data are reported as the means \pm S.E.M. Differences with P values < 0.05 were considered significant.

concentrations that can be used to evaluate activity against intracellular amastigotes, since macrophages are used as host cells in the assay.

The activities of the aqueous plant extracts against the extracellular replication of *L. amazonensis* were evaluated using *in vitro* assays (Figure 2), “which indirectly measure the effect against intracellular forms of *Leishmania*.” *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* exhibited direct activity against extracellular forms; the observed percentages of growth inhibition were 74.2%, 75.6%, 70.8%, 83.8%, and 82.1%, respectively, at 100 $\mu\text{g/mL}$, but were less potent than pentamidine (with maximum effect of 96.5% at 100 μM).

L. amazonensis is of particular importance because it is one of the most frequent species that causes human disease and specifically causes anergic diffuse leishmaniasis, a disfiguring cutaneous disease that is very difficult to cure [24]. In this study, *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* were shown to be active against promastigotes, and *H. pectinata*, *A. vera*, and *R. graveolens* were shown to be active against intracellular amastigotes of *L. amazonensis*. In addition, pentamidine was the most effective compound against promastigote and amastigote forms of *L. amazonensis*, presenting efficacy of 98.9% and 67.7% at 100 μM , respectively.

The activity of compounds against promastigotes and amastigotes can differ, depending on the targets of antileishmanial action, which may be selective for one of two developmental forms. Overall, promastigotes may be more sensitive than intracellular amastigotes, because amastigotes are adapted to survive in hostile intracellular environment as well as due to the fact the compounds have direct contact with promastigotes; in contrast, to have anti-amastigote activity, substance must be capable of crossing the membrane of the host cell.

Moreover, unfortunately, the concentration required to give 50% inhibition of the growth of *L. amazonensis* (IC_{50}) cannot be determined in the tests for evaluation of *in vitro* leishmanicidal activity because the plants extracts employed did not induce concentration-dependent killing. This effect can probably be a result of the large amount of substances present in the extracts of plants, which may have synergistic or antagonistic effects.

H. pectinata L. is known popularly in Brazil as sambacaitá and is used to treat inflammation, bacterial infections, and ache [25]. This plant exhibits antibacterial activity [26] and antinociceptive and antiedematogenic action, with low toxicity reported in the literature [27]. This study expands the scientific knowledge of this plant by demonstrating its promising activity against *L. amazonensis*. This plant inhibited promastigote growth by 81.9% at 100 $\mu\text{g/mL}$ (Figure 3) and inhibited amastigote growth by 55.3% at 10 $\mu\text{g/mL}$ (Figure 4), both with $P < 0.01$.

H. pectinata presents significant amounts of hyptolide and pectinolides A–C, among other 6-substituted-5,6-dihydro- α -pyrones [28] and pyrones exhibit antileishmanial activity [29]. Moreover, two pyrones isolated from the seeds of *Podolepis hieracioides* exhibited *in vitro* leishmanicidal activity against the promastigote forms of *L. donovani*, *L. major*, *L. infantum*, and *L. enriettii* and against the intracellular amastigote form of *L. donovani* [30]. The well-described constituents of *H. pectinata* include phytochemicals such as some terpenes, and its antileishmanial activity may be due to the presence of sesquiterpenes, such as β -caryophyllene and caryophyllene oxide, and to other terpenes that possess antimicrobial properties [26]. The chemical study of leaves from *Hyptis pectinata* resulted in the isolation of two new compounds, sambacaitaric acid (1) and 3-O-methyl-sambacaitaric acid (2), and nine known compounds, rosmarinic acid (3), 3-O-methyl-rosmarinic acid (4), ethyl caffeate (5), nepetoidin A (6), nepetoidin B (7), cirsiolol (8), cirsimaritin (9), 7-O-methyllyuteolin (10), and genkwanin (11). The EtOH extract, the hexane, EtOAc, and MeOH:H₂O fractions, and compounds 1, 2, and 4 exhibited antileishmanial activity [31].

Previous work using *A. vera* leaf exudate demonstrated its potential against promastigotes of *L. mexicana*, *L. tropica*, *L. braziliensis*, *L. major*, and *L. infantum* and against the axenic amastigotes of *L. donovani* [14]. Such studies are in agreement with the results observed here, which demonstrated strong *in vitro* leishmanicidal activity against promastigotes (82.9% growth inhibition at 100 $\mu\text{g/mL}$) and amastigotes (26.1% growth inhibition at 10 $\mu\text{g/mL}$), as such shown in Figures 3 and 4, respectively. The presence of alkaloids, triterpenes, cyanidins, proanthocyanidins, tannins, and saponins in *A. vera* leaf exudate was demonstrated. Alkaloids, triterpenes, and saponin-like compounds individually and synergistically have leishmanicidal activity [23].

Dutta et al. [32] demonstrated that the incubation of promastigotes with *A. vera* leaf exudate causes promastigote death through an apoptosis-like mechanism similar to metazoan apoptosis. However, the pathways of induction and/or execution differed at the molecular level because the *A. vera* leaf exudate-induced leishmanicidal effect did not involve

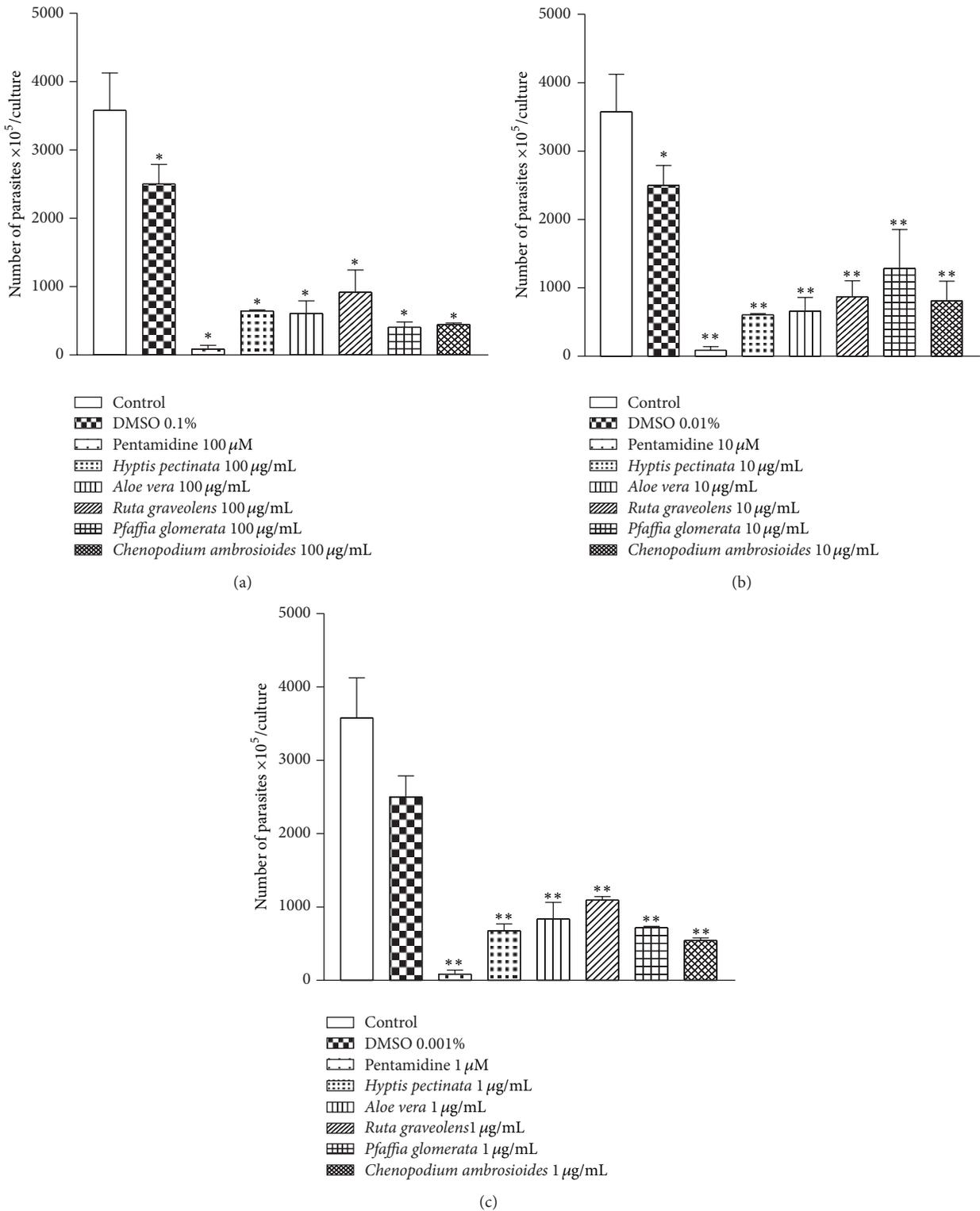


FIGURE 2: Direct antileishmanial effect of *H. pectinata*, *A. vera*, *R. graveolens*, *P. glomerata*, and *C. ambrosioides* against *L. amazonensis* promastigotes. The height of the bars indicates the number of parasites at each concentration of plant aqueous extract compared with the control experiment containing only the solvent DMSO. The plant aqueous extracts were added at concentrations of 100, 10, and 1 μg/mL in cultures of *L. amazonensis* promastigotes for 4 days. After 4 days, the extracellular load of *L. amazonensis* was measured. The experiments were performed three times independently, and each count was performed in triplicate. The data are reported as the means ± S.E.M. Differences with *P* values <0.05 were considered significant (**P* < 0.05; ***P* < 0.01).

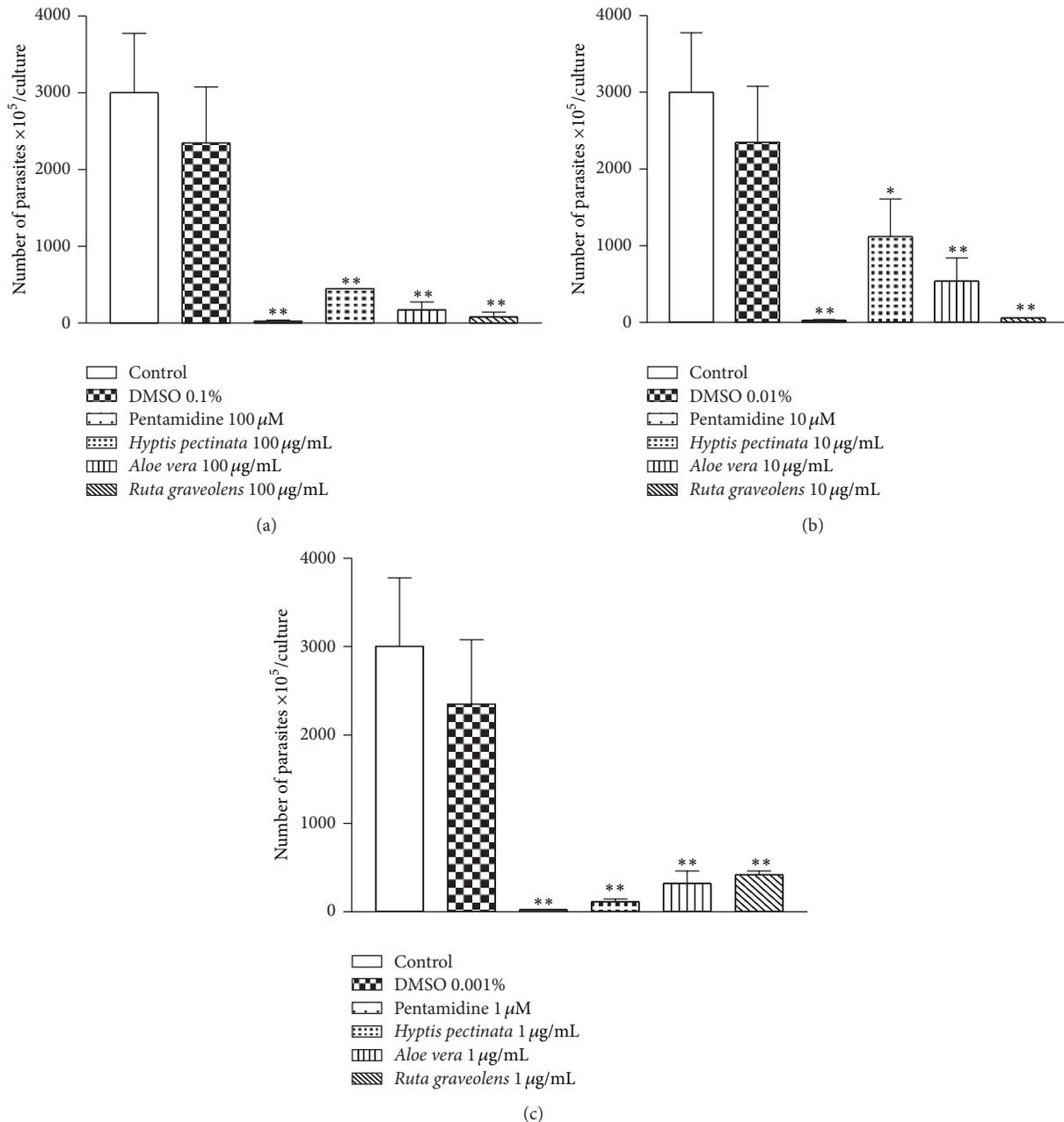


FIGURE 3: Antileishmanial effect of *H. pectinata*, *A. vera*, and *R. graveolens* against the replication of *L. amazonensis* promastigotes. The height of the bars indicates the parasite number at each concentration of plant aqueous extract compared with the control experiment containing only the solvent DMSO. Macrophages were infected with *L. amazonensis* promastigotes for 3 h. After infection, plant aqueous extracts were added at final concentrations of 100, 10, and 1 μ g/mL for 4 days. After 4 days, the extracellular load of *L. amazonensis* was measured. The experiments were performed three times independently, and each count was performed in triplicate. The data are reported as the means \pm S.E.M. Differences with P values < 0.05 were considered significant (* $P < 0.05$; ** $P < 0.01$).

caspases and major proteases, an increase in cytosolic Ca^{2+} , or the generation of reactive oxygen species. Additionally, *A. vera* leaf exudate also increases nitric oxide production. Nitric oxide causes extensive fragmentation of nuclear DNA in both axenic and intracellular amastigotes of *L. amazonensis*, and this fragmentation signal is regulated by non-caspase proteases of the proteasome [23].

R. graveolens (commonly known as rue) has been considered a medicinal plant since ancient times and is currently

used to treat various disorders, such as aching pain, eye problems, rheumatism, and dermatitis [33]. In this study, *R. graveolens* significantly diminished the number of promastigotes presenting percentages of inhibition of 74.4% at 100 μ g/mL (Figure 3) and exhibited anti-amastigote activity of 40.3% at 10 μ g/mL (Figure 4).

Although pharmacological data regarding the antileishmanial activity of *R. graveolens* has not been documented in the literature, some chemical constituents of this plant,

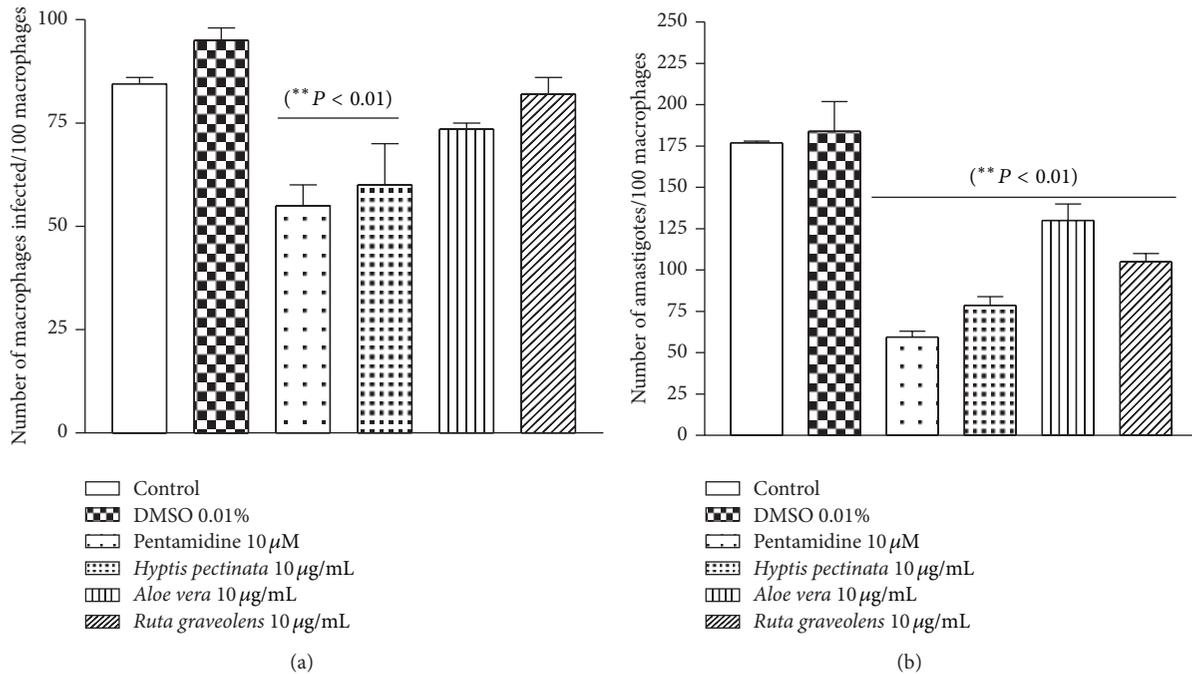


FIGURE 4: Antileishmanial effect of *H. pectinata*, *A. vera*, and *R. graveolens* against *L. amazonensis* amastigotes and the infection of macrophages. The height of the bars indicates the parasite number at each concentration of plant aqueous extract compared with the control experiment containing only the solvent DMSO. The numbers of infected macrophages (a) and intracellular amastigotes (b) were evaluated after the 3-day incubation with the plant extracts (10 µg/mL). Macrophages were infected with *L. amazonensis* promastigotes for 3 h. After infection, the plant aqueous extracts were added at a final concentration of 10 µg/mL for 3 days. Coverslips were washed, fixed, and stained, and the percentage of infected macrophages, noninfected macrophages, and intracellular amastigotes forms was determined by counting at least 100 cells/coverslip in triplicate on glass coverslips. Data are reported as the means ± S.E.M. Differences with P values <0.05 were considered significant.

such as rutacridone, gravacridonediol, and rhodesiacridone, possess antileishmanial activity [34]. The mechanism of action of some alkaloids has been attributed to their ability to intercalate DNA. In addition to these acridone alkaloids, *R. graveolens* is rich in coumarins [33, 35], chemical constituents that have been studied with respect to their cytotoxic activities against *Leishmania*; for example, coumarins of the mamea type have been purified from *Calophyllum brasiliense* and exhibit antileishmanial activities against *L. amazonensis* [36] and *L. braziliensis* [37].

The genus *Pfaffia* is a member of the Amaranthaceae family, and many of its Brazilian species have been commercialized as Brazilian ginseng; these species are used to treat diabetes and rheumatism and are used as a tonic and aphrodisiac. *P. glomerata* roots possess an *in vitro* cytotoxic effect against strains of *L. braziliensis*. A hydroalcoholic extract of *P. glomerata* is somewhat active against the promastigote forms of *L. braziliensis* (IC₅₀ 168.6 µg/mL) [38]. In the present study, *P. glomerata* exhibited direct activity against extracellular forms of the parasite, inhibiting their growth by 88.7% at 100 µg/mL (Figure 3). In addition, the mechanism of the antileishmanial action of this plant might involve the enhanced production of nitric oxide because it was able to protect the gastric mucosa by increasing the concentrations of nitric oxide in the stomach [39].

C. ambrosioides has been used to treat cutaneous leishmaniasis due to *L. braziliensis* among the rural population of a cocoa-producing coastal area of Bahia state, Brazil [40]. In this study, *C. ambrosioides* exhibited direct activity against extracellular forms of the parasite, inhibiting growth by 87.4% at 100 µg/mL (Figure 3).

C. ambrosioides also exhibited *in vitro* leishmanicidal effects against *L. amazonensis* promastigotes [41]. In the same way, Monzote et al. [42] showed that an essential oil from *C. ambrosioides* inhibits the progression of leishmanial infection both *in vitro* and *in vivo*. The essential oil had a minimal inhibitory concentration and EC₅₀ values of 27.82 and 3.74 µg/mL, respectively, against promastigotes of *L. amazonensis* [43]. Moreover, Patricio et al. [44] suggested that treatment with hydroalcoholic extracts of this plant by the intralésional route not only affects the regulatory mechanisms that control the dissemination of *L. amazonensis* but also appears to have a direct leishmanicidal effect. Previous work demonstrated that *C. ambrosioides* has the ability to recruit macrophages and further affects macrophage activation, which is fundamental to the control of *Leishmania*, by inducing nitric oxide [45]. The exact mechanism of action behind the direct antileishmanial effect exhibited by the essential oil of *C. ambrosioides* is unknown, but some authors have hypothesized that ascaridole (endoperoxide), an active

molecule, generates free radicals that act on parasitic DNA [43]. Some authors have reported the medical use of these plants in the northeast of Brazil against intestinal parasites [46, 47].

The results obtained here represent a worthwhile characterization of the antileishmanial activity of extracts from traditional medicinal plants from the Brazilian flora.

4. Conclusion

The five species of plants that are traditionally used to treat cutaneous leishmaniasis were active against promastigotes, and *H. pectinata*, *A. vera*, and *R. graveolens* were active against intracellular amastigotes of *L. amazonensis*. To our knowledge, these data corroborate, for the first time, the ethnopharmacological use of these plants for the treatment of Leishmaniasis.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The authors would like to thank Mr. Crispin Edivar Veras (Supervisor Hall Endemic of Novo Lino), Mr. Alfredo Emilio de Souza Filho (Nursing Assistant Municipal Secretary of Health of Novo Lino), Dr. Fernando de Araújo Pedrosa (Teacher of Parasitology UFAL), and Mr. José Pereira Venancio Junior (Military Police of Alagoas) for their collaboration with this work. The authors are also grateful to the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES, BR), the Ministério de Ciência e Tecnologia (MCT, BR), the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, BR), the Fundação de Amparo a Pesquisa de Alagoas (FAPEAL, BR), and the Ministério da Saúde (MS, BR) for fellowships and financial support. The authors wish to thank several of their colleagues at the Federal University of Alagoas for their constructive criticism and assistance in performing this study. The authors are grateful to CAPES, CNPq, INCT-INOVAR/CNPq (573.564/2008-6).

References

- [1] P. Cuervo, G. B. Domont, and J. B. De Jesus, "Proteomics of trypanosomatids of human medical importance," *Journal of Proteomics*, vol. 73, no. 5, pp. 845–867, 2010.
- [2] V. S. Amato, F. F. Tuon, H. A. Bacha, V. A. Neto, and A. C. Nicodemo, "Mucosal leishmaniasis. Current scenario and prospects for treatment," *Acta Tropica*, vol. 105, no. 1, pp. 1–9, 2008.
- [3] C. S. Peacock, K. Seeger, D. Harris et al., "Comparative genomic analysis of three *Leishmania* species that cause diverse human disease," *Nature Genetics*, vol. 39, no. 7, pp. 839–847, 2007.
- [4] J. Alvar, I. D. Vélez, C. Bern et al., "Leishmaniasis worldwide and global estimates of its incidence," *PLoS ONE*, vol. 7, no. 5, Article ID e35671, 2012.
- [5] M. C. C. Malta, H. P. Tinoco, M. N. Xavier, A. L. S. Vieira, É. A. Costa, and R. L. Santos, "Naturally acquired visceral leishmaniasis in non-human primates in Brazil," *Veterinary Parasitology*, vol. 169, no. 1-2, pp. 193–197, 2010.
- [6] M. Ouakad, N. Bahi-Jaber, M. Chenik, K. Dellagi, and H. Louzir, "Selection of endogenous reference genes for gene expression analysis in *Leishmania* major developmental stages," *Parasitology Research*, vol. 101, no. 2, pp. 473–477, 2007.
- [7] C. Baptista, A. O. Schubach, M. F. Madeira et al., "*Leishmania* (*Viannia*) *braziliensis* genotypes identified in lesions of patients with atypical or typical manifestations of tegumentary leishmaniasis: evaluation by two molecular markers," *Experimental Parasitology*, vol. 121, no. 4, pp. 317–322, 2009.
- [8] E. H. Gomes Rodrigues, M. E. Felinto de Brito, M. G. Mendonça et al., "Evaluation of PCR for diagnosis of American cutaneous leishmaniasis in an area of endemicity in Northeastern Brazil," *Journal of Clinical Microbiology*, vol. 40, no. 10, pp. 3572–3576, 2002.
- [9] A. C. T. Da Silva, E. Cupolillo, Â. C. Volpini, R. Almeida, and G. A. S. Romero, "Species diversity causing human cutaneous leishmaniasis in Rio Branco, state of Acre, Brazil," *Tropical Medicine and International Health*, vol. 11, no. 9, pp. 1388–1398, 2006.
- [10] Y. Chai, S. Yan, I. L. K. Wong, L. M. C. Chow, and H. Sun, "Complexation of antimony (SbV) with guanosine 5'-monophosphate and guanosine 5'-diphospho-D-mannose: formation of both mono- and bis-adducts," *Journal of Inorganic Biochemistry*, vol. 99, no. 12, pp. 2257–2263, 2005.
- [11] S. E. T. Borborema, H. F. D. A. Junior, J. A. O. Junior, and N. do Nascimento, "*In vitro* antileishmanial properties of neutron-irradiated meglumine antimoniate," *Brazilian Archives of Biology and Technology*, vol. 48, no. 2, pp. 63–68, 2005.
- [12] M. Ouellette, J. Drummelsmith, and B. Papadopoulou, "Leishmaniasis: drugs in the clinic, resistance and new developments," *Drug Resistance Updates*, vol. 7, no. 4-5, pp. 257–266, 2004.
- [13] K. K. Pitzer, K. A. Werbovetz, J. J. Brendle, and J. P. Scovill, "Synthesis and biological evaluation of 4-chloro-3, 5-dinitrobenzotrifluoride analogues as antileishmanial agents," *Journal of Medicinal Chemistry*, vol. 41, no. 24, pp. 4885–4889, 1998.
- [14] A. Dutta, G. Mandal, C. Mandal, and M. Chatterjee, "*In vitro* antileishmanial activity of *Aloe vera* leaf exudate: a potential herbal therapy in leishmaniasis," *Glycoconjugate Journal*, vol. 24, no. 1, pp. 81–86, 2007.
- [15] M. J. Chan-Bacab and L. M. Peña-Rodríguez, "Plant natural products with leishmanicidal activity," *Natural Product Report*, vol. 18, no. 6, pp. 674–688, 2001.
- [16] A. C. Queros, Levantamento Etnofarmacológico de Plantas Mediciniais Utilizadas no Tratamento de Leishmaniose Tegumentar em Novo Lino, Colônia Leopoldina e União dos Palmares, AL. Relatório de Iniciação Científica. CNPq, Universidade Federal de Alagoas, 2008.
- [17] U. P. de Albuquerque, V. A. da Silva, M. D. C. Cabral, N. Leal Alencar, and L. D. H. C. Andrade, "Comparisons between the use of medicinal plants in indigenous and rural caatinga (dryland) communities in NE Brazil," *Boletim Latinoamericano y del Caribe de Plantas Medicinales y Aromaticas*, vol. 7, no. 3, pp. 157–170, 2008.
- [18] R. F. Hussain, A. M. Nouri, and R. T. Oliver, "A new approach for measurement of cytotoxicity using colorimetric assay," *Journal of Immunological Methods*, vol. 160, no. 1, pp. 89–96, 1993.
- [19] N. A. Gomes, C. R. Gattass, V. Barreto-De-Souza, M. E. Wilson, and G. A. Dos Reis, "TGF-beta mediates CTLA-4 suppression of

- cellular immunity in murine kalaazar," *The Journal of Immunology*, vol. 164, no. 4, pp. 2001–2008, 2000.
- [20] C. M. D. C. Gomes, H. Goto, V. L. Ribeiro da Matta, M. D. Laurenti, M. Gidlund, and C. E. P. Corbett, "Insulin-like growth factor (IGF)-I affects parasite growth and host cell migration in experimental cutaneous leishmaniasis," *International Journal of Experimental Pathology*, vol. 81, no. 4, pp. 249–255, 2000.
- [21] F. L. Ribeiro-Gomes, A. C. Otero, N. A. Gomes et al., "Macrophage interactions with neutrophils regulate *leishmania major* infection," *Journal of Immunology*, vol. 172, no. 7, pp. 4454–4462, 2004.
- [22] M. P. Nunes, L. Cysne-Finkelstein, B. C. Monteiro, D. M. de Souza, N. A. Gomes, and G. A. DosReis, "CD40 signaling induces reciprocal outcomes in *Leishmania*-infected macrophages; roles of host genotype and cytokine milieu," *Microbes and Infection*, vol. 7, no. 1, pp. 78–85, 2005.
- [23] A. Dutta, D. Sarkar, A. Gurib-Fakim, C. Mandal, and M. Chatterjee, "In vitro and in vivo activity of *Aloe vera* leaf exudate in experimental visceral leishmaniasis," *Parasitology Research*, vol. 102, no. 6, pp. 1235–1242, 2008.
- [24] L. Monzote, A. M. Montalvo, R. Scull, M. Miranda, and J. Abreu, "Activity, toxicity and analysis of resistance of essential oil from *Chenopodium ambrosioides* after intraperitoneal, oral and intralesional administration in BALB/c mice infected with *Leishmania amazonensis*: a preliminary study," *Biomedicine & Pharmacotherapy*, vol. 61, no. 2-3, pp. 148–153, 2007.
- [25] M. F. Arrigoni-Blank, A. R. Antonioli, L. C. Caetano, D. A. Campos, A. F. Blank, and P. B. Alves, "Antinociceptive activity of the volatile oils of *Hyptis pectinata* L. Poit. (Lamiaceae) genotypes," *Phytomedicine*, vol. 15, no. 5, pp. 334–339, 2008.
- [26] P. O. Santos, M. D. J. C. Costa, J. A. B. Alves et al., "Chemical composition and antimicrobial activity of the essential oil of *Hyptis pectinata* (L.) Poit.," *Quimica Nova*, vol. 31, no. 7, pp. 1648–1652, 2008.
- [27] M. D. Bispo, R. H. V. Mourão, E. M. Franzotti et al., "Antinociceptive and antiedematogenic effects of the aqueous extract of *Hyptis pectinata* leaves in experimental animals," *Journal of Ethnopharmacology*, vol. 76, no. 1, pp. 81–86, 2001.
- [28] D. M. Boalino, J. D. Connolly, S. McLean, W. F. Reynolds, and W. F. Tinto, "α-Pyrone and a 2(5H)-furanone from *Hyptis pectinata*," *Phytochemistry*, vol. 64, no. 7, pp. 1303–1307, 2003.
- [29] D. Carmona, J. Sáez, H. Granados et al., "Antiprotozoal 6-substituted-5,6-dihydro-α-pyrone from *Raimondia cf. Monoica*," *Natural Product Research*, vol. 17, no. 4, pp. 275–280, 2003.
- [30] O. Kayser, A. F. Kiderlen, and S. L. Croft, "Antileishmanial activity of two γ-pyrone from *Podolepis hieracioides* (Asteraceae)," *Acta Tropica*, vol. 86, no. 1, pp. 105–107, 2003.
- [31] R. A. Falcao, P. L. A. do Nascimento, S. A. de Souza et al., "Antileishmanial phenylpropanoids from the leaves of *Hyptis pectinata* (L.) poit.," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 460613, 7 pages, 2013.
- [32] A. Dutta, S. Bandyopadhyay, C. Mandal, and M. Chatterjee, "Aloe vera leaf exudate induces a caspase-independent cell death in *Leishmania donovani* promastigotes," *Journal of Medical Microbiology*, vol. 56, no. 5, pp. 629–636, 2007.
- [33] S. K. Raghav, B. Gupta, A. Shrivastava, and H. R. Das, "Inhibition of lipopolysaccharide-inducible nitric oxide synthase and IL-1β through suppression of NF-κB activation by 3-(1*l*-dimethyl-allyl)-6-hydroxy-7-methoxy-coumarin isolated from *Ruta graveolens* L.," *European Journal of Pharmacology*, vol. 560, no. 1, pp. 69–80, 2007.
- [34] K. M. Ahua, J. R. Ioset, A. Ransijn, J. Mauël, S. Mavi, and K. Hostettmann, "Antileishmanial and antifungal acridone derivatives from the roots of *Thamnosma rhodesica*," *Phytochemistry*, vol. 65, no. 7, pp. 963–968, 2004.
- [35] F. G. Braga, M. L. M. Bouzada, R. L. Fabri et al., "Antileishmanial and antifungal activity of plants used in traditional medicine in Brazil," *Journal of Ethnopharmacology*, vol. 111, no. 2, pp. 396–402, 2007.
- [36] M. A. Brenzan, C. V. Nakamura, B. Prado Dias Filho, T. Ueda-Nakamura, M. C. M. Young, and D. Aparício Garcia Cortez, "Antileishmanial activity of crude extract and coumarin from *Calophyllum brasiliense* leaves against *Leishmania amazonensis*," *Parasitology Research*, vol. 101, no. 3, pp. 715–722, 2007.
- [37] M. A. Brenzan, I. C. Piloto Ferreira, M. V. Campana Lonardoní et al., "Activity of extracts and coumarins from the leaves of *Calophyllum brasiliense* on *Leishmania braziliensis*," *Pharmaceutical Biology*, vol. 46, no. 6, pp. 380–386, 2008.
- [38] A. G. Neto, A. A. da Silva Filho, J. M. L. C. Costa et al., "Evaluation of the trypanocidal and leishmanicidal in vitro activity of the crude hydroalcoholic extract of *Pfaffia glomerata* (Amaranthaceae) roots," *Phytomedicine*, vol. 11, no. 7-8, pp. 662–665, 2004.
- [39] C. S. Freitas, C. H. Baggio, J. E. da Silva-Santos et al., "Involvement of nitric oxide in the gastroprotective effects of an aqueous extract of *Pfaffia glomerata* (Spreng) Pedersen, Amaranthaceae, in rats," *Life Sciences*, vol. 74, no. 9, pp. 1167–1179, 2004.
- [40] F. Franca, E. L. Lago, and P. D. Marsden, "Plants used in the treatment of leishmanial ulcers due to *Leishmania (Viannia) braziliensis* in an endemic area of Bahia, Brazil," *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 29, no. 3, pp. 229–232, 1996.
- [41] J. L. Bezerra, G. C. Costa, T. C. Lopes et al., "Avaliação da atividade leishmanicida in vitro de plantas medicinais," *Revista Brasileira de Farmacognosia*, vol. 16, pp. 631–637, 2006.
- [42] L. Monzote, A. M. Montalvo, S. Almanonni, R. Scull, M. Miranda, and J. Abreu, "Activity of the essential oil from *Chenopodium ambrosioides* grown in Cuba against *Leishmania amazonensis*," *Chemotherapy*, vol. 52, no. 3, pp. 130–136, 2006.
- [43] L. Monzote, A. M. Montalvo, R. Scull, M. Miranda, and J. Abreu, "Combined effect of the essential oil from *Chenopodium ambrosioides* and antileishmanial drugs on promastigotes of *Leishmania amazonensis*," *Revista do Instituto de Medicina Tropical de Sao Paulo*, vol. 49, no. 4, pp. 257–260, 2007.
- [44] F. J. Patrício, G. C. Costa, P. V. S. Pereira et al., "Efficacy of the intralesional treatment with *Chenopodium ambrosioides* in the murine infection by *Leishmania amazonensis*," *Journal of Ethnopharmacology*, vol. 115, no. 2, pp. 313–319, 2008.
- [45] G. V. B. Cruz, P. V. S. Pereira, F. J. Patrício et al., "Increase of cellular recruitment, phagocytosis ability and nitric oxide production induced by hydroalcoholic extract from *Chenopodium ambrosioides* leaves," *Journal of Ethnopharmacology*, vol. 111, no. 1, pp. 148–154, 2007.
- [46] B. A. L. de Freitas, D. M. de Oliveira, D. G. de Oliveira, and L. J. Gomes, "Conhecimento e uso da flora pelos moradores do Assentamento São Judas Tadeu, Porto Da Folha, Sergipe," *Revista Ouricuri*, vol. 3, no. 2, pp. 19–36, 2013.
- [47] V. H. M. de Lima, J. P. Gomes-Neto, and M. D. S. Cavalcanti, "Recursos vegetais utilizados pela comunidade de Vila Velha, Ilha de Itamaracá, Pernambuco, Brasil," *Revista Ouricuri*, vol. 3, no. 1, 2013.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

