

Research Article

Preliminary Studies of the Immunomodulator Effect of the *Bougainvillea xbuttiana* Extract in a Mouse Model

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Received 20 October 2014; Revised 2 February 2015; Accepted 4 February 2015

Academic Editor: Sarang Bani

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Bougainvillea xbuttiana is used as an analgesic in folk medicine in Mexico. The purpose of the present study was to determine the effects of the ethanolic extract from *B. xbuttiana* on macrophages activities. The phytochemical screening was performed for determine the presence of alkaloids, flavonoids, triterpenes, and saponins. The effects of *B. xbuttiana* were analyzed using the macrophages activities as determined by the H₂O₂ release, spreading and phagocytic index, vacuoles formation percentage, and mediators production. The viability percentage was determined in live cells after fixing and staining with crystal violet. The presence of H₂O₂ in macrophages was performed by using the peroxidase-phenol red solution. The cytokine production was determined by two assays, ELISA for detection of IL-6, IL-10, and IFN- γ and biological assay for TNF detection. The results showed that the *Bxb* extract dose-dependent manner produces (a) an increase in levels of H₂O₂ and spreading and vacuoles formation percentages, (b) a decrease in phagocytic index and in the amounts of TNF, IL-6, and IFN- γ , and (c) an increase significant in IL-10 and NO production. This study indicates that the ethanolic extract from *Bougainvillea xbuttiana* was able to activate macrophages. The combination of these results suggests that this extract has an immunomodulator effect.

1. Introduction

There are a lot of archaeological evidences that humans used medicinal plants for better living, reducing disease, and improving quality of life. Several studies have described the use of plants in traditional medicine for many years and have recently gained much importance in the field of pharmacological industries. The knowledge and assessments of the biological properties of extracts from plants can serve as a source of future drug candidates in many areas of health [1].

The relevant literature shows that many plants or plant products have been used as an alternative source for the treatment of different diseases. Several of these herbs have been used to exert immunomodulatory effect in the treatment of various diseases [2]. Medicinal plants with immunomodulatory activity are used for cases of organ transplant rejection or for treatment of the various autoimmune diseases [3, 4].

Immunomodulation is a process used to describe an increase or decrease in the magnitude of immune response. The immunostimulants are those compounds that act in an enhancement of immune reactions and are involved in nonspecific system stimulation [5]. In contrast, the immunosuppressive compounds are those capable of decreasing the resistance against infections and stress caused by environmental factors or chemotherapy [6].

Medicinal plants can inhibit or stimulate immune response which could be useful in the treatment of various human diseases. In particular, medicinal plants capable of inhibiting the cellular and humoral responses could have useful applications in the treatment of immunological disorders [7].

The state of macrophages activation and T and B lymphocytes plays a major role in the pathogenesis of immune-mediated disorders [8]. Macrophages are essential members

of the innate immune system and, together with neutrophils, eosinophils and natural killer cells are a first line of defense to identify, eliminate or contain invading microorganisms and toxic macromolecules. Macrophages are pivotal key in the maintenance of tissue homeostasis and are responsible for detecting, engulfing, and destroying pathogens [9]. In response to injury, macrophages bind to invading pathogens and deliver them to other components of adaptive immunity, which is constituted by antibody and cell mediated responses that are performed by different lymphocyte cells, B cells, and T cells, respectively [9–11]. During macrophage activation, several compounds are released such as cytokines, reactive oxygen species, nitric oxide, and lipid inflammatory mediators which are implicated in the inflammatory response [12]. With respect to cytokines, they regulate the intensity and duration of the immune response by stimulating or inhibiting the activation, proliferation, and differentiation of various cells and by regulating their secretion of antibodies or other cytokines [12–14]. Inflammatory cytokines can be classified into two groups: those involved in acute inflammation, named proinflammatory cytokines, and those responsible for reducing inflammation, named anti-inflammatory cytokines [15–17]. The proinflammatory cytokines are cytokines which promote systemic inflammation and include interleukin-1 (IL-1), interferon-gamma (IFN- γ), and tumor necrosis factor (TNF). By contrast, anti-inflammatory cytokines are those responsible for regulating and maintaining homeostasis. Major anti-inflammatory cytokines include IL-4, IL-6, IL-10, and IL-13 [17–19].

Nitric oxide (NO) plays an important role in various functions including vasoregulation, neurotransmission, regulation of immune responses and cytotoxic activity against tumor cells and a variety of pathogens by macrophages mid. It is involved in the pathogenesis and control of infectious diseases [20]. NO is derived from L-arginine in a reaction catalyzed by nitric oxide synthase (NOS). The NOS has different isoforms: the constitutive NOS (cNOS) and inducible NOS (iNOS) [21, 22]. Endotoxins or cytokines induce the expression of NOS that is capable of generating an amount of NO greater than the other types of NOS [21, 22]. The increment of NO production contributes to tissue damage inflammatory and infectious diseases including septic shock and stroke [21–23].

For a long time, the compounds derived from medicinal plants were the basis for use as drugs. Various compounds of plants have been described for their anti-inflammatory or immunomodulator activity.

Bougainvillea is from family Nyctaginaceae which has 30 genera distributed mainly in tropical climates. It is reported in literature the existence of 18 different species. In Cuernavaca, Morelos and other states of Mexico the predominant species is the *B. xbutiana*. *Bougainvillea* is widely used as a medicinal plant in the states of central and southern Mexican territory. Infusions prepared from flowers or leaves are used for gastrointestinal and respiratory disorders. With respect to gastrointestinal disorders, the infusion is used as antidiarrheal and to reduce the acid content of the stomach. In the case of respiratory disorders, the infusion is used to treat cough, asthma, bronchitis, and whooping cough. For

the treatment of these diseases, the infusions are prepared from flowers or leaves, which are administered orally. For other parts of the plant, root decoction is used for fevers and as a laxative [24, 25]. For the many attributes found by the use of *Bougainvillea*, we decided to examine its effects on macrophage activation, which was determined by assays phagocytic index, vacuoles formation, peroxide hydrogen, cells expansion, and mediators production.

2. Materials and Methods

2.1. Chemicals, Reagents, and Buffers. Fetal calf serum (FCS) and RPMI-1640 medium were purchased from Sigma (St. Louis, MO, USA). Capture and detection antibodies and recombinant cytokines were purchased from BD Biosciences Pharmingen (USA), and recombinant TNF was purchased from Boehringer Mannheim (Mannheim, Germany).

2.2. Plant Material and Extraction of *Bougainvillea xbutiana*. The plant specimens were collected from Cuernavaca (Morelos, Mexico). *Bougainvillea xbutiana* was identified from a voucher specimen (23683) in Herbarium HUMO, CIByC-UAEM. Extraction method is the crucial first step in the analysis of medicinal plants, because it is necessary to extract the desired chemical components from the plant materials for further characterization. Patent MX/a/2011/813522 provides details in extraction method of *Bougainvillea xbutiana* flowers named as *Bxb* [25].

2.3. Phytochemical Tests. Phytochemicals screening methods for detecting the presence of secondary metabolites in *Bxb* were (a) Dragendorff test for detecting alkaloids; (b) Shinoda test [26] for detecting flavonoids; (c) the foam test [26, 27] for determining saponins; (d) Liebermann-Buchard test for detecting triterpenes; (e) Folin-Ciocalteu Method to quantify total phenols contents [28] and (f) reaction with $AlCl_3$ in base medium for quantifying total flavonoic contents [28].

2.4. Animals. Female mice of CD1 strain weighing 20–25 g were purchased from Facultad de Medicina, UAEM. The animals were kept under strict ethical conditions according to international guidelines for the care and use of laboratory animals. The experiments designed for this study were approved by the Committee of Experimental Animal Administration of the university with protocol number 13MM/2011, [29].

2.5. Peritoneal Macrophages. Female mice of the CD1 strain were sacrificed and peritoneal cells were collected by peritoneal lavage as previously described by Cohn and Benson, 1965 [30]. The obtained cell suspension was centrifuged and the cells were resuspended in RPMI-1640 supplemented with 10% FCS and adjusted at a final concentration of 1×10^6 cells/mL and distributed in 96-well microplates. Cell cultures were incubated for 2 hours at 37°C with an atmosphere of 5% CO_2 . Then nonadherent cells were removed and adherent cells of different concentrations of the *Bxb* were

added. At different times of incubation, the supernatants were collected and frozen at -20°C until assessment of mediators.

2.6. Viability Assay. Culture supernatants from macrophages treated or not treated with *Bxb* incubated for 24, 48, 72, and 96 hours were discarded and the live cells were fixed and stained with crystal violet for 10 minutes at room temperature. Excess stain was removed through washing and absorbance was determined in a microplate reader with a filter of 620 nm. To calculate the viability percentage, the following formula was used: $[(A_{\text{sample}}/A_{\text{control}}) \times 100]$ [31].

2.7. Phagocytosis Assays. The probe to determine the phagocytic index was performed as described by Zebedee et al., 1994 [32]. Briefly, cultures of macrophages at a concentration of 1×10^6 cells/mL treated or not treated with *Bxb* were incubated at 37°C with an atmosphere of 5% CO_2 . At the end of 24, 48, 72, and 96 hours of incubation, the supernatants were collected and a solution consisting 1:5 of opsonized yeast was added. After 1 hour of incubation, the supernatants were removed and safranin solution was added for 40 seconds. The cells were examined by using optical microscope at 40x. The phagocytic index (PI) was determined as follows: number of macrophages with internalized yeast/100.

2.8. Vacuole Formation Assay. Macrophage cultures were incubated with RPMI-1640 medium plus 5% FCS and 1 mM NH_4Cl /mL and then were treated with 2.9, 29, and 290 $\mu\text{g}/\text{mL}$ *Bxb* at 37°C with 5% CO_2 [33]. After 24, 48, 72, and 96 hours of incubation, cells were stained by 0.05% neutral red for 5 minutes. Cells were then washed with PBS containing 0.2% BSA, 70% ethanol, and 0.37% HCl. The absorbance was determined using the microtiter plate reader with a filter of 540 nm. Vacuole formation percentage was calculated by the following formula: $[(A_{\text{sample}} - A_{\text{control}})/A_{\text{control}}] \times 100$.

2.9. Spreading Index. Spreading index was determined by the method described by Arruda et al. (2004) [34]. In brief, the suspension of $1 \times 10^6/\text{mL}$ macrophages was distributed in microplate which contained slides to assess cell adhesion. Cultures treated or not treated with 2.9, 29, and 290 $\mu\text{g}/\text{mL}$ *Bxb* were incubated at 37°C with 5% CO_2 . After 24, 48, 72, and 96 hours of incubation, the slides were removed and immersed in methanol and then stained with crystal violet. After drying, the cells were observed with an optical microscope of 40x magnification. The spreading index (SI) corresponds to a percentage value of 100 macrophages.

2.10. Hydrogen Peroxide. Hydrogen peroxide production in macrophage was determined as described by Pick and Mizel, in 1981 [35]. Macrophage cultures treated or not treated with 2.9, 29, and 290 $\mu\text{g}/\text{mL}$ *Bxb* were incubated at 37°C with 5% CO_2 . After 24, 48, 72, and 96 hours of incubation, the supernatants were removed. Directly on the cells was added a solution of phenol with 140 mM NaCl; 10 mM K_2PO_4 ; 5.5 mM dextrose; and 5.5 mM horseradish peroxidase. Absorbance was determined in the microplate reader with 620 nm filter and the hydrogen peroxide concentration was determined by comparison with a standard curve.

TABLE 1: Qualitative identification of phytochemical compounds in *Bxb*.

Alkaloids	+++
Flavonoids	+++
Saponins	+
Triterpenes	++

Absent (-), present (+), moderate (++), and abundant (+++).

2.11. Cytokines ELISA and Biological Assays

2.11.1. ELISA Assay. Cytokines such as IL-6, TNF, and IFN- γ present in the supernatant of macrophages from CD1 mice treated or not treated with *Bxb* were detected by ELISA assay [36]. All assays were performed according to manufacturer's instructions. Minimal detection for IL-6 and IL-10 was 10 pg/mL and for IFN- γ 100 pg/mL.

2.11.2. Biological Assay. It was used to measure TNF present in the supernatant of macrophages treated or untreated with *Bxb* [37]. The percentage of TNF cytotoxicity on L929 cells was calculated by the following formula: $(A_{\text{control}} - A_{\text{sample}}/A_{\text{control}}) \times 100$. TNF amounts are expressed as pg/mL estimated from the ratio of a 50% cytotoxic dose of the test to that of standard mouse recombinant TNF.

2.12. Nitrite Determination. Nitrite in the presence of macrophage supernatants of treated and untreated mice with *Bxb* was determined as previously described by Keller et al., 1990 [38]. Briefly, macrophages supernatants from treated or untreated *Bxb* for different times were mixed with freshly prepared Griess reagent (0.1% naphthalenediamine hydrochloride, sulphonylamide 1%, and 3% H_3PO_4). Absorbance was measured in a microplate reader with 540 nm filter. The amount of nitrite was determined by a standard curve of NaNO_2 . Minimal detection for nitrite was 1.25 nM.

2.13. Statistical Analyses. The data were statistically analyzed by Student's *t*-test. A value of $P < 0.01$ was considered significant.

3. Results

3.1. Qualitative and Quantitative Measurements. The standardized extracts were as follows: one is based on identifying and the second based on quantifying a *Bxb* to a characteristic chemical. The phytochemical test was positive for secondary metabolites such as alkaloids, triterpenes, flavonoids, and saponins (Table 1). We have previously reported the quantitative identification of phytochemical compounds in *Bxb* extract [28].

3.2. Effects of *Bxb* on Macrophages Viability. To determine the effect of *Bxb* extract on viability of peritoneal macrophages, these cells were obtained by peritoneal lavage, cultured and treated *in vitro* with different concentrations of *Bxb*, and incubated for distinct time. The effect of *Bxb* was determined by measuring viability percentage. The *Bxb* induced

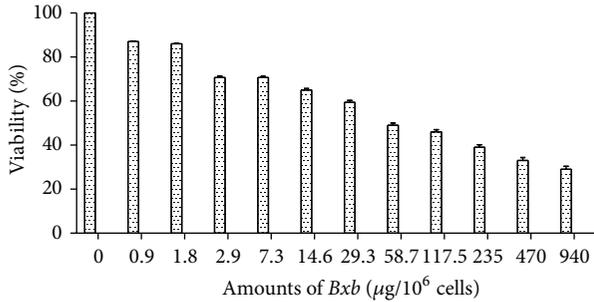


FIGURE 1: Effect of *Bxb* on peritoneal macrophage viability. Peritoneal macrophages were obtained and treated *in vitro* with different concentrations of *Bxb* as described above. The viability percentage was observed at 48 h after extract exposition. Each bar represents the mean value of samples from three experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

dose-dependent inhibition on cell viability (Figure 1). Peritoneal macrophages treated during 48 hours with 2.9 up to 14.6 µg/mL of *Bxb* showed viability percentage decreased by 30% (Figure 1). For groups of cells exposed to 29.3 up to 237 µg/mL, the viability decreased by 51% up to 60%. The lowest viability percentage was obtained in macrophages treated with 470 and 940 µg/mL of *Bxb* (Figure 1). We also determined that a concentration of *Bxb* extract that induces the decreasing in 50% macrophage viability was 29 µg/mL. After determining the optimal interaction between cell and *Bxb*, we designed the further analysis using the concentrations as follows: 0, 2.9, 29, and 290 µg/mL.

The kinetics of cytotoxicity percentage of peritoneal macrophages treated or not treated with different concentrations of *Bxb* during distinct time periods is shown in Figure 2. In cultures of macrophages treated with 2.9 µg/mL of *Bxb* the cytotoxicity percentage was 5%, 24%, and 29% for 24, 48, and 72 hours, respectively (Figure 2). The cytotoxicity percentage in macrophages treated with 29 µg/mL of *Bxb* was 5%, 40%, and 50% for 24, 48, and 72 hours, respectively (Figure 2). In macrophages treated with 290 µg/mL of *Bxb*, the cytotoxicity percentages were 7%, 61%, and 69% for 24, 48, and 72 hours, respectively (Figure 2).

3.3. Effects of *Bxb* on Macrophages Activities. To evaluate the activities of macrophages, the cells were obtained by peritoneal lavage, cultivated and treated *in vitro* with different concentrations of *Bxb*, and incubated for distinct time. The effects of *Bxb* on macrophages were performed by measuring H_2O_2 , phagocytic index, spreading index, and percentage of vacuoles formation.

As shown in Figure 3, the macrophage groups treated with 2.9, 29 and 290 µg/mL of *Bxb* during different periods of time the phagocytic index was significantly lower than those obtained from untreated macrophages cultures ($P < 0.001$).

The vacuoles formation was quantified by a period of 24 up to 96 hours, using the neutral red assay. The vacuolating percentage in macrophages treated with 2.9 µg/mL of *Bxb* was similar when compared with untreated macrophages

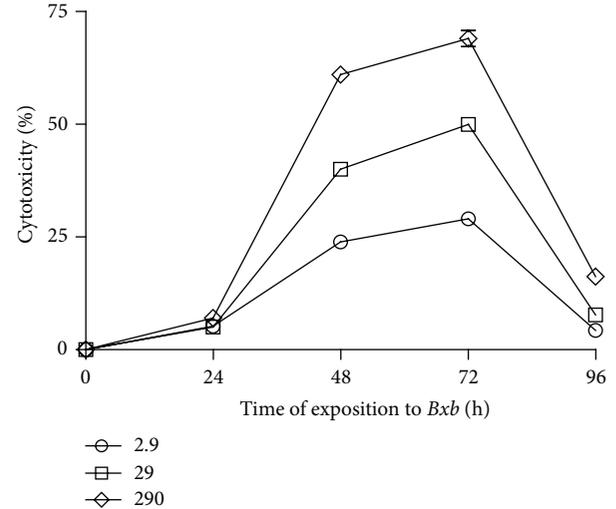


FIGURE 2: Cytotoxic activity of *Bxb*. Peritoneal macrophages were obtained and exposed *in vitro* to 2.9, 29, and 290 µg/mL of *Bxb* as described above. The cytotoxic percentage was observed at different times after *Bxb* exposition. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

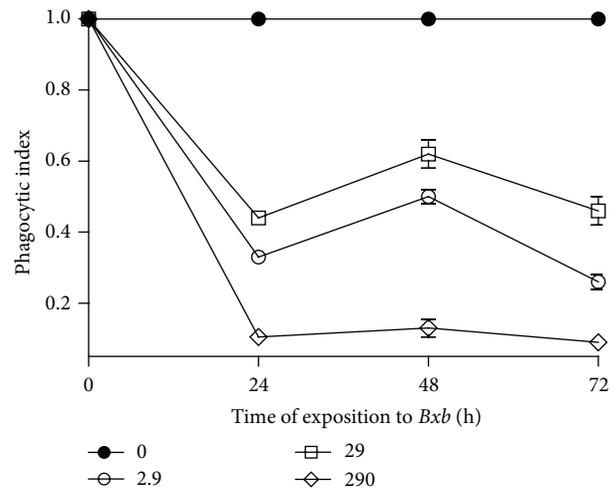


FIGURE 3: Phagocytosis index. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 µg/mL of *Bxb* and the phagocytic index was determined as described above. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P > 0.05$.

(Figure 4). In contrast, when the macrophages were treated with 29 µg/mL of *Bxb* extract during 24 and 48 hours, the vacuolating percentage was significantly higher when compared with the results obtained in control macrophages ($P < 0.001$), decaying thereafter. In case of macrophages treated with 290 µg/mL of *Bxb* for 24 hours, the vacuolating percentage was significantly higher than those obtained in untreated macrophages ($P < 0.001$), decaying thereafter.

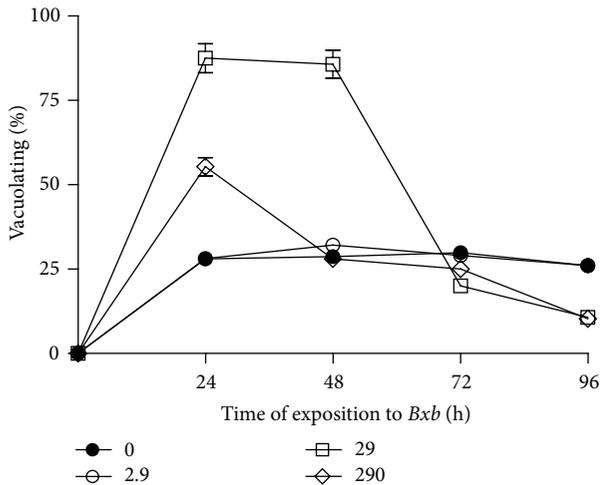


FIGURE 4: Vacuole formation. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 μg/mL of *Bxb* as described above. After different times of incubation at 37°C in an atmosphere of 5% CO₂, the cells were stained with neutral red for 5 min. The absorbance was determined at 540 nm, and the results were expressed as described above. Each point represents the mean value of samples from four experiments in different groups of four mice. Statistical differences between the treatments were $P < 0.05$.

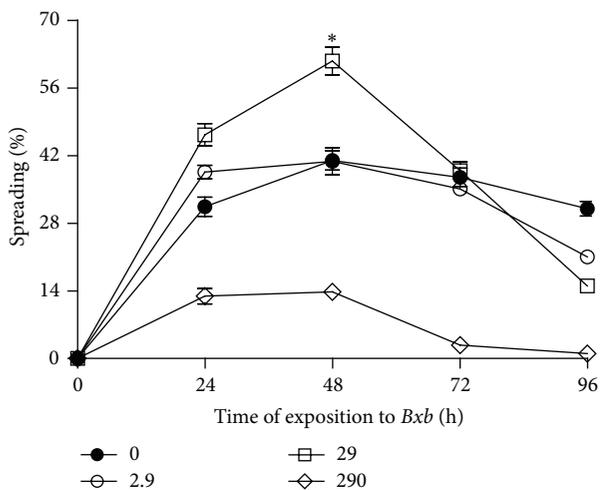


FIGURE 5: Spreading index. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 μg/mL of *Bxb* and the spreading percentage was determined as described above. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

In Figure 5 is shown the spread index. In macrophages treated with 2.9 μg/mL of *Bxb*, the percentage of spreading was similar to that obtained from untreated macrophages. For groups of macrophages treated with 29 μg/mL of *Bxb* during 48 hours, the percentage of spreading presented was significantly higher when compared with those results observed in untreated macrophages ($P < 0.001$). In contrast, in macrophages treated with 290 μg/mL of *Bxb*, the spreading

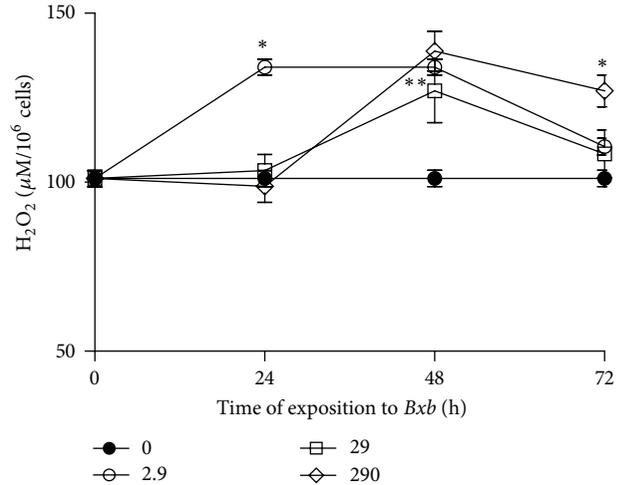


FIGURE 6: Hydrogen peroxide production. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 μg/mL of *Bxb* and the hydrogen peroxide production was determined directly on cells as described above. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

percentages were significantly lower than those observed for the untreated macrophages ($P < 0.001$).

The levels of hydrogen peroxide present in supernatants of macrophages treated or not treated with *Bxb* are shown in Figure 6. The highest levels of H₂O₂ were observed among peritoneal macrophages treated with 2.9 μg/mL of *Bxb* for 24 and 48 hours. For groups of macrophages treated with 29 and 290 μg/mL of *Bxb* during 48 hours, the levels of H₂O₂ were significantly higher than those obtained in untreated macrophages ($P < 0.001$).

3.4. Effect of *Bxb* on Cytokines and NO Production. To determine the capacity of *Bxb* to stimulate the production of cytokines and NO, groups of mice were sacrificed and their macrophages were collected by peritoneal lavage and they were *in vitro* treated with different concentrations of *Bxb* for distinct time periods.

The kinetics of proinflammatory cytokines production is shown in Figure 7. TNF production was significantly lower in macrophages treated with 2.9, 29, and 290 μg/mL of *Bxb* for all time periods studied here than that obtained in untreated macrophages ($P < 0.001$). Treatment of macrophages with 2.9, 29 and 290 μg/mL of *Bxb* caused significant decrease in the production of IL-6 when compared with untreated macrophages ($P < 0.01$). Figure 7 also shows that the levels of IFN-γ, from macrophages treated with 2.9, 29, and 290 μg/mL of *Bxb*, were significantly decreased when compared with those levels obtained from untreated macrophages cultures ($P < 0.01$).

In Figure 8 is shown the kinetics of anti-inflammatory cytokines production. The *in vitro* treatment of macrophages with *Bxb* resulted in IL-10 production. For macrophages cultures treated with 2.9, 29, and 290 μg/mL of *Bxb*,

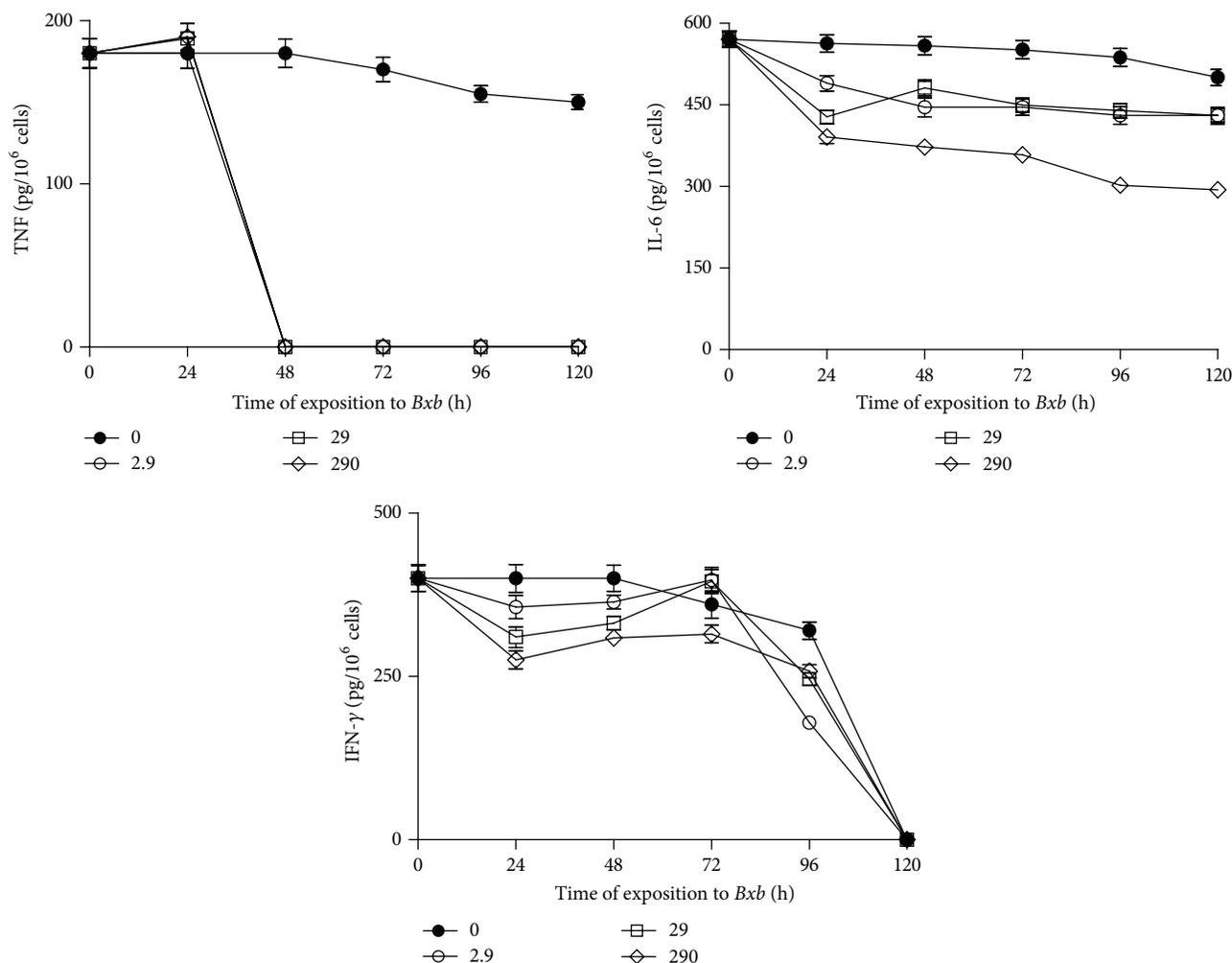


FIGURE 7: Proinflammatory cytokines production. Peritoneal macrophages were obtained and exposed *in vitro* to 2.9, 29, and 290 μg/mL of *Bxb* as described above. At different times, the levels of IL-6 and IFN-γ were determined by ELISA assay using monoclonal antibodies as the probe. The levels of TNF were determined by standard assay with L929 cells. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

the IL-10 levels were significantly higher than those obtained from untreated macrophages cultures ($P < 0.01$).

As shown in Figure 9, the *Bxb* increased the NO production in macrophages in a concentration-dependent manner. For macrophages cultures treated with 2.9, 29, and 290 μg/mL of *Bxb*, the NO production was significantly higher than those obtained from untreated macrophages ($P < 0.01$), decaying thereafter.

4. Discussion

Many natural products have played a significant role in maintaining and improving the quality of human life. The modulation of immune response to alleviate many diseases has been one mechanism of interest for pharmacology studies. A great number of medicinal plants have been shown to stimulate or inhibit immune response. In the present study, ethanolic extract obtained from *Bxb* native from Mexico was investigated for its effects in macrophage

activities and mediators production. For this purpose, we firstly evaluated the cytotoxic activity of the *Bxb* extract in peritoneal macrophages. The *Bxb* showed cytotoxic activity dose-dependent manner.

The phytochemical screening of secondary metabolites in *Bxb* revealed the presence of alkaloids, flavonoids, triterpenes, and saponins. These compounds have been shown to contain molecules with potent effects on the host immune system by either stimulant or suppressor of immune response.

Exposure of organisms to bacterial infection can result in the activation of defense mechanisms which include phagocytosis, cell motility, and spreading. In the present study, it has been clearly demonstrated that *Bxb* exerts inhibitory effect on the phagocytic ability by macrophages *in vitro* culture. Similar results have been reported with extracts from other plant families [39]. Particles internalization by macrophages results in the generation of vacuoles. In this study, we also observed that peritoneal macrophages exposed to 29 μg/mL of *Bxb* presented high presence of vacuoles. In

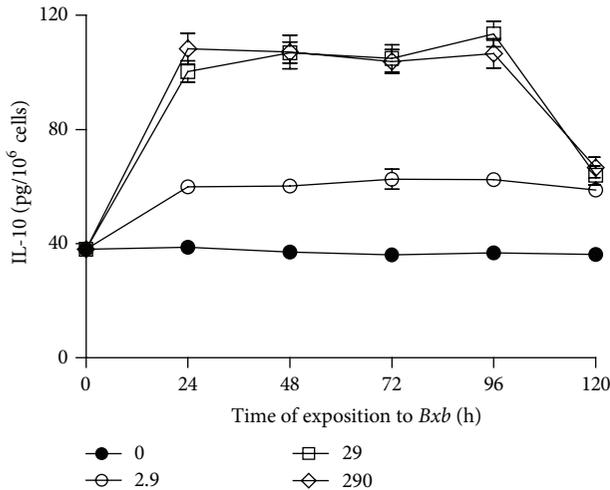


FIGURE 8: Anti-inflammatory cytokines production. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 $\mu\text{g/mL}$ of *Bxb* and at different times the levels of IL-10 were assayed by ELISA assay using monoclonal antibodies as the probe. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

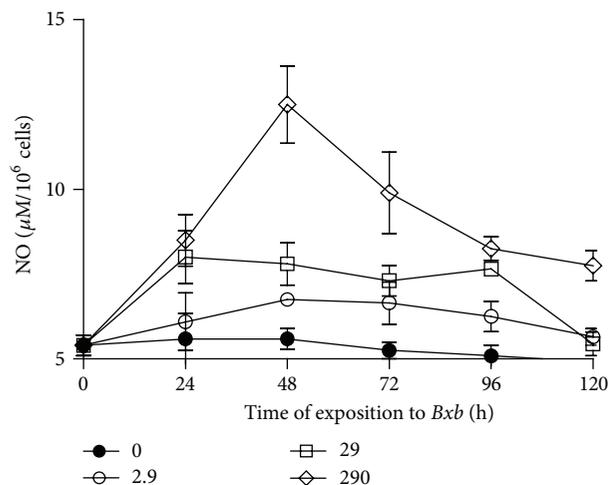


FIGURE 9: NO production. Peritoneal macrophages were obtained and exposed to 2.9, 29, and 290 $\mu\text{g/mL}$ of *Bxb* and at different times the levels of NO were determined by Griess colorimetric reaction. Each point represents the mean value of samples from four experiments in different groups of five mice. Statistical differences between the treatments were $P < 0.05$.

these cells, the plasma membrane was well-preserved which is characteristic of cell viability. The pronounced spreading has been considered a marker of macrophages and is a typical morphological characteristic of activated macrophages [39, 40]. In this study, we observed that 29 $\mu\text{g/mL}$ of *Bxb* extract was capable to induce the spreading in macrophages. The spreading provides morphological evidence of considerable cellular modifications in cytoskeleton reorganization and changes of membrane protein activities and expression [41].

Therefore, the phagocytosis index observed in *Bxb* treatment of macrophages was coordinated with macrophage activity. These parameters thus indicate that *Bxb* has an efficient capacity to activate macrophages. Hydrogen peroxide is important in cell signaling and it is an effector molecule for microbicidal and cytotoxic response of macrophages after stimulation and the development of inflammation. In our study, it was also found that *Bxb* induces the hydrogen peroxide production in peritoneal macrophages. Taken together, these data demonstrated that the *Bxb* is able to generate the macrophage activation.

Macrophages activated produce and release products including several cytokines, inorganic reactive radicals, reactive oxygen intermediates, and reactive nitrogen intermediates with biological activities [9, 10]. In this study, we exposed peritoneal macrophages to *Bxb* to demonstrate the production of cytokines. The proinflammatory cytokines are essential for initiating the inflammation process leading to tissue destruction. These cytokines induce tissue destruction and reduce the capacity to repair damaged tissue by stimulating the production of other mediators [42]. Among the proinflammatory cytokines, IL-6 is one of most important mediators of fever and acute-phase response. TNF has cytostatic and cytotoxic effects and the ability to cause apoptosis [43]. The production of IFN- γ and IL-10 provides important protection to cells and tissues against deleterious effects of free radicals [44]. The reduction of these cytokines may reflect the anti-inflammatory activity of plant extracts. Various studies have shown the immunomodulatory effects from plants extracts. In our study, it was found that the *Bxb* was capable of decreasing the proinflammatory cytokines such as TNF, IL-6, and IFN- γ production in macrophages and promoted the production of anti-inflammatory cytokines, such as IL-10. These combined results suggest an immunomodulatory activity of the extract *Bxb*. The reduction in the biological activity of IL-1 or TNF is accomplished by highly specific strategies for the treatment of patients with inflammatory diseases [9, 10].

Nitric oxide is a mediator related to cell activation which contributes to the death or inhibition of a variety of pathogens [20, 45]. In our study, it was found that 29 $\mu\text{g/mL}$ of *Bxb* was able to induce the NO production. These results are in agreement with other studies that have demonstrated that NO has an important regulatory role in the various types of inflammatory processes. Nitric oxide is synthesized in large quantities by activated macrophages and has been demonstrated to be involved in the pathogenesis of acute and chronic inflammatory conditions.

In conclusion, the *B. xbutiana* extract is able to suppress the production of mediators, such as TNF, IFN- γ , and IL-6, and enhanced the production of NO and IL-10 in a dose-dependent manner. Further detailed studies are required to identify the active constituents and their mechanism for this effect.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

This work was supported by Secretaría de Educación Pública (SEP-PROMEP), México.

References

- [1] U. M. Acuña, K. Jancovski, and E. J. Kennelly, "Polyisoprenylated benzophenones from *Clusiaceae*: potential drugs and lead compounds," *Current Topics in Medicinal Chemistry*, vol. 9, no. 16, pp. 1560–1580, 2009.
- [2] J. D. Phillipson and L. A. Anderson, "Ethnopharmacology and western medicine," *Journal of Ethnopharmacology*, vol. 25, no. 1, pp. 61–72, 1989.
- [3] S. Murakami, M. Takeno, Y. Kirino et al., "Screening of tuberculosis by interferon- γ assay before biologic therapy for rheumatoid arthritis," *Tuberculosis*, vol. 89, no. 2, pp. 136–141, 2009.
- [4] V. Ramgolam, S. G. Ang, Y. H. Lai, C. S. Loh, and H.-K. Yap, "Traditional Chinese medicines as immunosuppressive agents," *Annals of the Academy of Medicine Singapore*, vol. 29, no. 1, pp. 11–16, 2000.
- [5] A. R. Bafna and S. H. Mishra, "Protective effect of bioactive fraction of *Sphaeranthus indicus* Linn. against cyclophosphamide induced suppression of humoral immunity in mice," *Journal of Ethnopharmacology*, vol. 104, no. 3, pp. 426–429, 2006.
- [6] C. K. Atal, M. L. Sharma, A. Kaul, and A. Khajuria, "Immunomodulating agents of plant origin. I. Preliminary screening," *Journal of Ethnopharmacology*, vol. 18, no. 2, pp. 133–141, 1986.
- [7] A. Mirshafiey, B. H. A. Rehm, A. A. Sahmani, A. Naji, and A. Razavi, "M-2000, as a new anti-inflammatory molecule in treatment of experimental nephrosis," *Immunopharmacology and Immunotoxicology*, vol. 26, no. 4, pp. 611–619, 2004.
- [8] F. Rieux-Laucat, A. Fischer, and F. Le Deist, "Cell-death signaling and human disease," *Current Opinion in Immunology*, vol. 15, no. 3, pp. 325–331, 2003.
- [9] C. F. Nathan, "Secretory products of macrophages," *The Journal of Clinical Investigation*, vol. 79, no. 2, pp. 319–326, 1987.
- [10] C. F. Nathan and J. B. Hibbs Jr., "Role of nitric oxide synthesis in macrophage antimicrobial activity," *Current Opinion in Immunology*, vol. 3, no. 1, pp. 65–70, 1991.
- [11] A. Aderem, "Role of Toll-like receptors in inflammatory response in macrophages," *Critical Care Medicine*, vol. 29, no. 7, pp. 16–18, 2001.
- [12] A. K. Abbas, M. E. Williams, H. J. Burstein, T. L. Chang, P. Bossu, and A. H. Lichtman, "Activation and functions of CD4⁺ T-cell subsets," *Immunological Reviews*, vol. 123, pp. 5–22, 1991.
- [13] R. L. Coffman, "Origins of the T_H1-T_H2 model: a personal perspective," *Nature Immunology*, vol. 7, no. 6, pp. 539–541, 2006.
- [14] T. Jo, N. Terada, Y. Takauchi et al., "Cytotoxic actions of cytokines on cultured mouse luteal cells are independent of nitric oxide," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 55, no. 3-4, pp. 291–296, 1995.
- [15] T. R. Mosmann, L. Li, H. Hengartner, D. Kagi, W. Fu, and S. Sad, "Differentiation and functions of T cell subsets," *Ciba Foundation Symposia*, vol. 204, pp. 148–158, 1997.
- [16] M. Stein and S. Gordon, "Regulation of tumor necrosis factor (TNF) release by murine peritoneal macrophages: role of cell stimulation and specific phagocytic plasma receptors," *European Journal of Immunology*, vol. 21, no. 2, pp. 431–437, 1991.
- [17] P. H. Van Der Meide and H. Schellekens, "Cytokines and the immune response," *Biotherapy*, vol. 8, no. 3-4, pp. 243–249, 1996.
- [18] C. Gérard, C. Bruyins, A. Marchant et al., "Interleukin 10 reduces the release of tumor necrosis factor and prevents lethality in experimental endotoxemia," *Journal of Experimental Medicine*, vol. 177, no. 2, pp. 547–550, 1993.
- [19] M. Howard, T. Muchamuel, S. Andrade, and S. Menon, "Interleukin 10 protects mice from lethal endotoxemia," *Journal of Experimental Medicine*, vol. 177, no. 4, pp. 1205–1208, 1993.
- [20] C. Nathan and Q. W. Xie, "Nitric oxide synthases: roles, tolls, and controls," *Cell*, vol. 78, no. 6, pp. 915–918, 1994.
- [21] Z. Amirghofran, S. Malek-Hosseini, H. Golmoghaddam, F. Kalantar, and M. Shabani, "Inhibition of Nitric Oxide production and proinflammatory cytokines by several medicinal plants," *Iranian Journal of Immunology*, vol. 8, no. 3, pp. 159–169, 2011.
- [22] J. D. MacMicking, C. Nathan, G. Hom et al., "Altered responses to bacterial infection and endotoxic shock in mice lacking inducible nitric oxide synthase," *Cell*, vol. 81, no. 4, pp. 641–650, 1995.
- [23] T. J. Guzik, R. Korbut, and T. Adamek-Guzik, "Nitric oxide and superoxide in inflammation and immune regulation," *Journal of Physiology and Pharmacology*, vol. 54, no. 4, pp. 469–487, 2003.
- [24] E. Edwin, E. Sheeja, E. Toppo, V. Tiwari, and K. R. Dutt, "Antidiarrhoeal, anti ulcer and antimicrobial activities of leaves of *Bougainvillea glabra* Choisy," *Ars Pharmaceutica*, vol. 48, no. 2, pp. 135–144, 2007.
- [25] A. L. Alvarez Perez Gil, L. Barbosa Navarro, M. Patipo Vera, and V. L. Petricevich, "Anti-inflammatory and antinociceptive activities of the ethanolic extract of *Bougainvillea xbuttiana*," *Journal of Ethnopharmacology*, vol. 144, no. 3, pp. 712–719, 2012.
- [26] X. A. Dominguez, *Metodos de Investigación Fitoquímica*, Limusa, Mexico City, Mexico, 1985.
- [27] J. H. Doughari, A. M. Elmahmood, and S. Manzara, "Studies on the antibacterial activity of root extracts of *Carica papaya* L.," *African Journal Microbiology Research*, vol. 1, pp. 37–41, 2007.
- [28] L. Arteaga Figueroa, L. Barbosa Navarro, M. Pati, M. Patiño Vera, and V. L. Petricevich, "Antioxidant activity, total phenolic and flavonoids contents, and cytotoxicity evaluation of *Bougainvillea xbuttiana*," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 6, no. 5, 2014.
- [29] "International society on toxinology: membership application," *Toxicon*, vol. 30, no. 12, pp. 1–12, 1992.
- [30] Z. A. Cohn and B. Benson, "The differentiation of mononuclear phagocytes: morphology, cytochemistry, and biochemistry," *The Journal of Experimental Medicine*, vol. 121, pp. 153–170, 1965.
- [31] M. A. Hamilton, "Trimmed Spearman-Kärber method for estimating median lethal concentrations in toxicity bioassays," *Environmental Science Technology*, vol. 12, no. 4, p. 417, 1978.
- [32] S. L. Zebedee, R. K. Koduri, J. Mukherjee et al., "Mouse-human immunoglobulin G1 chimeric antibodies with activities against *Cryptococcus neoformans*," *Antimicrobial Agents and Chemotherapy*, vol. 38, no. 7, pp. 1507–1514, 1994.
- [33] T. L. Cover, W. Puryear, G. I. Perez-Perez, and M. J. Blaser, "Effect of urease on HeLa cell vacuolation induced by *Helicobacter pylori* cytotoxin," *Infection and Immunity*, vol. 59, no. 4, pp. 1264–1270, 1991.
- [34] M. S. P. Arruda, V. B. Richini, S. M. A. Oliveira, and F. R. Vilani-Moreno, "Experimental murine mycobacteriosis: evaluation of the functional activity of alveolar macrophages in thalidomide-treated mice," *Brazilian Journal of Medical and Biological Research*, vol. 37, no. 4, pp. 485–492, 2004.

- [35] E. Pick and D. Mizel, "Rapid microassays for the measurement of superoxide and hydrogen peroxide production by macrophages in culture using an automatic enzyme immunoassay reader," *Journal of Immunological Methods*, vol. 46, no. 2, pp. 211–226, 1981.
- [36] J. R. Schumacher, A. O'Garra, and P. Schrader, "Characterization of monoclonal antibodies to mouse interleukin-5 and development of mouse and human IL-5 ELISA assay," *Journal of Immunology*, vol. 141, no. 5, pp. 1576–1581, 1988.
- [37] M. R. Ruff and G. E. Gifford, "Purification and physicochemical characterization of rabbit tumor necrosis factor," *Journal of Immunology*, vol. 125, no. 4, pp. 1671–1677, 1980.
- [38] R. Keller, R. Keist, A. Wechsler, T. P. Leist, and P. H. van der Meide, "Mechanisms of macrophage-mediated tumor cell killing: a comparative analysis of the roles of reactive nitrogen intermediates and tumor necrosis factor," *International Journal of Cancer*, vol. 46, no. 4, pp. 682–686, 1990.
- [39] F. Benencia, M. C. Courrèges, and F. C. Coulombié, "Anti-inflammatory activities of *Trichilia glabra* aqueous leaf extract," *Journal of Ethnopharmacology*, vol. 71, no. 1-2, pp. 293–300, 2000.
- [40] J. A. Cleary, G. E. Kelly, and A. J. Husband, "The effect of molecular weight and beta-1,6-linkages on priming of macrophage function in mice by (1,3)-beta-D-glucan," *Immunology and Cell Biology*, vol. 77, no. 5, pp. 395–403, 1999.
- [41] R. B. Johnston Jr., "Current concepts: immunology. Monocytes and macrophages," *The New England Journal of Medicine*, vol. 318, no. 12, pp. 747–752, 1988.
- [42] B. Wójciak-Stothard, M. Denver, M. Mishra, and R. A. Brown, "Adhesion, orientation, and movement of cells cultured on ultrathin fibronectin fibers," *In Vitro Cellular and Developmental Biology - Animal*, vol. 33, no. 2, pp. 110–117, 1997.
- [43] P. M. Preshaw and J. J. Taylor, "How has research into cytokine interactions and their role in driving immune responses impacted our understanding of periodontitis?" *Journal of Clinical Periodontology*, vol. 38, no. 11, pp. 60–84, 2011.
- [44] C. Gabay, "Interleukin-6 and chronic inflammation," *Arthritis Research & Therapy*, vol. 8, supplement 2, article S3, 2006.
- [45] P. Kourilsky and P. Truffa-Bachi, "Cytokine fields and the polarization of the immune response," *Trends in Immunology*, vol. 22, no. 9, pp. 502–509, 2001.



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