

Research Article

Antihypertensive Effect of the GaMiSamHwangSaSimTang in Spontaneous Hypertensive Rats

Kyungjin Lee, Bumjung Kim, Heseung Hur, Khanita Suman Chinannai, Inhye Ham, and Ho-Young Choi

Department of Herbology, College of Korean Medicine, Kyung Hee University, 26 Kyungheedaero-ro, Dongdaemun-gu, Seoul 130-701, Republic of Korea

Correspondence should be addressed to Ho-Young Choi; hychoi@khu.ac.kr

Received 10 March 2015; Revised 29 April 2015; Accepted 4 May 2015

Academic Editor: Yanwei Xing

Copyright © 2015 Kyungjin Lee et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The present study was designed to evaluate the antihypertensive effect of GaMiSamHwangSaSimTang (HVC1), a 30% ethanol extract of a mixture comprising Pruni Cortex, Scutellariae Radix, Coptidis Rhizoma, and Rhei Rhizoma, on spontaneous hypertensive rats (SHRs). The systolic blood pressure (SBP) was measured every 4 or 7 days using the noninvasive tail cuff system. The vasorelaxant effects on isolated aortic rings were evaluated. Aortic rings were contracted using phenylephrine (PE) or KCl, and the changes in tension were recorded via isometric transducers connected to a data acquisition system. In this study, oral administration of HVC1 decreased the SBP of SHRs over the experimental period. HVC1 induced concentration-dependent relaxation in the aortic rings that had been precontracted using PE or KCl. The vasorelaxant effects of HVC1 on endothelium-intact aortic rings were inhibited by pretreatment with *N* ω -Nitro-L-arginine methyl ester (L-NAME) or methylene blue. HVC1 inhibited the contraction induced by extracellular Ca²⁺ in endothelium-denuded rat aortic rings that had been precontracted using PE or KCl. In conclusion, HVC1 reduced the SBP of SHR and relaxed isolated SHR aortic rings by upregulating NO formation and the NO-cGMP pathway and blocking the entry of extracellular Ca²⁺ via receptor-operative Ca²⁺ channel and voltage-dependent Ca²⁺ channel.

1. Introduction

Hypertension is a global public health issue and is associated with increased risk of cardiovascular disease, stroke, and kidney disease [1]. The disease is regarded as a “silent killer” as it rarely produces symptoms in its early stages and as a result many people go undiagnosed [1]. In 2008, the worldwide prevalence of high blood pressure was reported to be approximately 40% of adults aged 25 or older; every year, about 9.4 million deaths are estimated to be caused by hypertension, which accounts for 12.8% of the worldwide total [1]. Hypertension has a huge economic impact, and in higher-income countries, including those in Eastern Europe and Central Asia, the disease accounts for almost 23% of health care expenditure [2].

In 2013, in Korea, the prevalence of hypertension in men and women over 30 years of age was 32.4 and 22.2%, respectively; the prevalence increased with age and in adults over 70 years of age, the prevalence of hypertension in men was 59%

and in women was 64.3% [3]. The number of hypertensive patients is constantly increasing, and the cost of hypertension treatment is also steadily increasing in Korea. According to the National Health Insurance Statistical Yearbook 2013, the costs paid by national health insurance for the treatment of hypertension (\$1.9 billion) formed a larger proportion of total medical costs (\$46.3 billion) than any other disease, and—among 13.7 million patients—hypertension was the most common disease (5.5 million people) [4]. However, traditional medicines have not been widely used for the treatment of hypertension in Korea. There are several useful traditional medicines for the treatment of hypertension [5–7], but few patients choose to use them, and health insurance does not pay for them. This is because the herbal medicines for hypertension have not yet been developed throughout efficacy and safety studies in Korea.

Scutellariae Radix (SR), Coptidis Rhizoma (CR), and Rhizoma Rhei (RR) have been commonly used traditional medicines for cardiovascular diseases in China, Japan, and

Korea. A herbal prescription SanHuangXieXinTang (SamHwangSaSimTang in Korean) composed of SR, CR, and RR was reported to decrease U46619-induced increase in pulmonary arterial blood pressure [8]. And we found that Pruni Cortex (PC) has potent vasorelaxant activities in the previous study [9]. Therefore, these herbal materials are expected to be useful for the treatment of spontaneous hypertension. For the development of new antihypertensive herbal medicine, a new prescription GaMiSamHwangSaSimTang (HVCI) which consists of four kinds of traditional medicine including Pruni Cortex, *Scutellariae Radix*, *Coptidis Rhizoma*, and *Rhei Rhizoma* was developed based on SamHwangSaSimTang. In the previous animal study, we found that vasorelaxant effect of HVCI was better than SamHwangSaSimTang (data not shown).

In this study, we aimed to demonstrate the hypotensive effect and mechanisms of action of a HVCI on spontaneous hypertensive rats and we performed standardization of HVCI.

2. Materials and Methods

2.1. Preparation of GaMiSamHwangSaSimTang. The extract was prepared from a mixture of dried PC (200 g), SR (100 g), CR (100 g), and RR (200 g). PC and RR were purchased from Dongwoodang Co., Ltd. (Yeongcheon, Kyungpook, Republic of Korea). CR and SR were purchased from Dong Yang Herb Co., Ltd. (Seoul, Republic of Korea). Professor Hocheol Kim of Kyung Hee University identified these herbal medicines. The mixture was extracted with 30% ethanol for 2 h in a reflux apparatus. After reflux and filtration, the extract was evaporated using a rotary vacuum evaporator (N-N series, EYELA, Japan) at 60°C and lyophilized to yield 159.9 g of crude extract.

2.2. Chemicals and Drugs. Phenylephrine (PE), KCl, acetylcholine, *N* ω -Nitro-L-arginine methyl ester (L-NAME), methylene blue (MB), atropine, indomethacin, and ethylene glycol-bis-(2-aminoethylether)-N,N,N',N'-tetraacetic acid were purchased from Sigma Aldrich (St. Louis, MO, USA). Amlodipine besylate was purchased from Kolmar Korea Co., Ltd. (Yeongi-gun, Chungnam, Republic of Korea). All other reagents were of analytical purity.

2.3. Animals. Spontaneous hypertensive rats (SHR/lzm; male; weight: 200–250 g; age: 8 weeks) were purchased from Japan SLC Inc. (Hamamatsu, Shizuoka Prefecture, Japan). All procedures involving animals were conducted according to the animal welfare guidelines issued by the National Veterinary Research & Quarantine Service and World Organization for Animal Health (OIE), and this study was approved (KHUASP(SE)-13-018) by the Kyung Hee University Institutional Animal Care and Use Committee. The rats were housed under controlled conditions (22 \pm 2°C; lighting, 07:00–19:00), and food and water were available *ad libitum*.

2.4. Measurement of Blood Pressure. The systolic blood pressure of SHR was evaluated using the noninvasive tail cuff system (CODA 8-Channel High Throughput Non-Invasive

Blood Pressure system, Kent Scientific Co. Ltd., Torrington, CT, USA) [10]. We randomly divided SHR into four groups of six animals each. For 50 days, they were orally administered either distilled water, amlodipine, or HVCI. Amlodipine and HVCI were dissolved in distilled water. Amlodipine was not completely soluble in distilled water; therefore, an aqueous suspension of amlodipine was used in this experiment. Control rats were treated with distilled water (1 mL·kg⁻¹·day⁻¹), positive control rats were treated with amlodipine (10 mg·kg⁻¹·day⁻¹), and the rats of the two experimental groups were treated with HVCI (50 or 300 mg·kg⁻¹·day⁻¹).

2.5. Vasoactivity Measurement. Spontaneous hypertensive rat aortic rings were isolated and placed in organ chambers containing Krebs-Henseleit solution (K-H solution; 10 mL) at 37°C, and then the vasoactivity of HVCI was evaluated using previously described methods [9]. HVCI was dissolved in K-H solution. The endothelium-intact and endothelium-denuded aortic rings were contracted using PE (1 μ M) or KCl (60 mM) treatment. Endothelium-intact aortic rings were also preincubated with L-NAME (10 μ M), MB (10 μ M), indomethacin (10 μ M), or atropine (1 μ M) for 20 min before contraction with PE (1 μ M) treatment to investigate the vasorelaxant mechanisms of HVCI action. The presence of functional endothelium was verified by the ability of ACh (10 μ M) to induce more than 80% relaxation in rings that were precontracted by PE (1 μ M). The relaxant effect of HVCI on the aortic rings was calculated as a percentage of the contraction in response to PE or KCl.

2.6. High Performance Liquid Chromatography (HPLC) Analysis of HVCI. Precisely weighed HVCI (100 mg) was dissolved in methanol (10 mL; HPLC grade; J. T. Baker Co. Ltd., USA) and the solution was filtered through a 0.45 μ m syringe filter (poly(vinylidene difluoride), Milford, USA). The analytical standards used for the HPLC analysis of HVCI were as follows: sennoside A and sennoside B (*Rhei Rhizoma* standards, Sigma, USA), coptisine, berberine, wogonin (*Rhizoma Coptidis* standards, Sigma, USA), baicalin, baicalein (*Coptidis Rhizoma* standards, Sigma, USA), prunetin (*Pruni Cortex* standards, Sigma, USA), genistein-7-glucose, and prunetin-5-glucose (*Pruni Cortex* standards, isolated according to a previously published procedure [11]). Each standard (1 mg) was dissolved in 100 μ g/mL 50% methanol. Equal amounts of each standard mixture were combined and a HPLC chromatogram was obtained. The HPLC apparatus was a Gilson System equipped with a 234 Autosampler, a UV/VIS-155 detector, and a 321 HPLC Pump (Gilson, Seoul, Korea). A Luna 4.60 \times 264 mm C18 reverse-phase column with 5 μ m particles (Phenomenex, CA, USA) was used. The mobile phase consisted of 0.1% formic acid (A) and acetonitrile (HPLC grade, J. T. Baker Co. LTD., USA) (B) in a ratio specified by the following binary gradient with linear interpolation: 0 min, 20% B; 60 min, 30% B; 70 min, 60% B; 100 min, 70% B. The column eluent was monitored at 250 nm, and all solvents were degassed with a micro-membrane filter (poly(tetrafluoroethylene), Advantec, Tokyo, Japan). Chromatography was performed at room

temperature at a flow rate of 0.5 mL/min, using 10 μ L analyte, for 100 min.

2.7. Statistical Analysis. Data are expressed as mean \pm standard error of the mean (SEM). Statistical comparisons were made using Student's *t*-test or one-way analysis of variance (ANOVA) followed by Tukey's post hoc test. All statistical analyses were performed using SPSS v.13.0 statistical analysis software (SPSS Inc., USA). *P* values less than 0.05 were considered statistically significant.

3. Results

3.1. Effect of HVC1 on Blood Pressure in SHR. Before the experiment commenced (day 0), the systolic blood pressure (SBP) of the control group (198.8 ± 4.6 mmHg), the amlodipine $10 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ treated group (197.5 ± 1.4 mmHg), the HVC1 $50 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ treated group (201.7 ± 1.5 mmHg), and the HVC1 $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ treated group (203.3 ± 4.8 mmHg) were measured using the noninvasive tail cuff system. At the end of the experiment (day 50), the SBP of the control group had increased to 225.2 ± 1.9 mmHg. The SBP of the positive control group (i.e., the amlodipine-treated group) continuously decreased during the experimental period and thus this experiment was considered to be reliable.

Orally administered HVC1 doses of 50 and $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ also continuously decreased the SBP of SHR during the experimental period. On average, orally administered HVC1 doses of 50 and $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ decreased the SBP of SHR to 191.1 ± 2.5 and 186.6 ± 2.9 mmHg, respectively. The maximal hypotensive effect was recorded on day 36 and SBP was 179.8 ± 10.8 and 172.2 ± 5.8 mmHg for the 50 and $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ dose-treated groups, respectively (Figure 1).

3.2. Effects of HVC1 on PE- or KCl-Induced Contraction of Endothelium-Intact or Endothelium-Denuded Aortic Rings. HVC1 (3, 10, 30, 100, and $300 \mu\text{g}/\text{mL}$) caused concentration-dependent relaxation in both endothelium-intact and endothelium-denuded aortic rings that had been precontracted using PE ($1 \mu\text{M}$) treatment. However, endothelium-intact aortic rings were more relaxed than endothelium-denuded aortic rings (Figure 2(a)). HVC1 (3, 10, 30, 100, and $300 \mu\text{g}/\text{mL}$) also caused concentration-dependent relaxation in endothelium-intact and endothelium-denuded aortic rings that had been precontracted using KCl (60 mM) treatment. But there were no significant differences between endothelium-intact and endothelium-denuded aortic rings (Figure 2(b)).

3.3. Effect of HVC1 on Endothelium-Intact Aortic Rings Preincubated with L-NAME or MB. Incubation with L-NAME ($10 \mu\text{M}$) or MB ($10 \mu\text{M}$) significantly decreased HVC1-induced relaxation of endothelium-intact aortic rings that had been precontracted using PE ($1 \mu\text{M}$) treatment. However, the vasorelaxant effect of HVC1 $300 \mu\text{g}/\text{mL}$ was not affected by preincubation with MB (Figure 3).

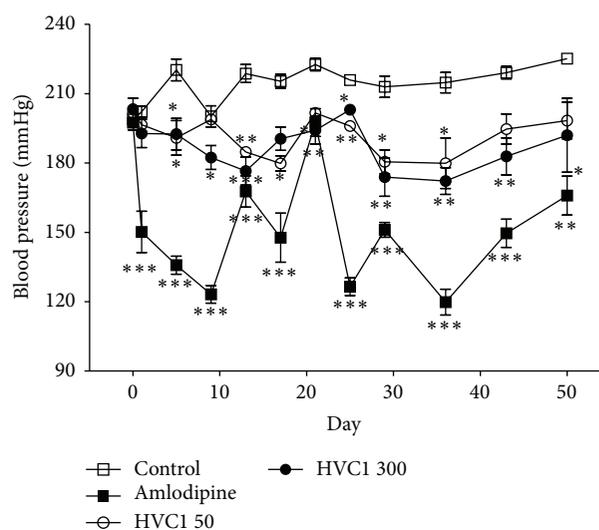


FIGURE 1: Hypotensive effect of HVC1 on the arterial systolic blood pressure of spontaneous hypertensive rats. Arterial systolic blood pressure was measured using the noninvasive tail cuff system. Values are expressed as mean \pm SEM ($n = 6$). * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$ versus control. Amlodipine = amlodipine $10 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ oral administration; HVC1 50 = HVC1 $50 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ oral administration; HVC1 300 = HVC1 $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ oral administration.

3.4. Effect of HVC1 on Endothelium-Intact Aortic Rings Preincubated with Indomethacin or Atropine. Incubation with indomethacin ($10 \mu\text{M}$) or atropine ($1 \mu\text{M}$) did not affect HVC1-induced relaxation of endothelium-intact aortic rings that had been precontracted using PE ($1 \mu\text{M}$) treatment (Figure 4).

3.5. Effect of HVC1 on Extracellular Ca^{2+} -Induced Contraction. In Ca^{2+} -free K-H solution, the cumulative addition of CaCl_2 (0.3–10 mM) induced progressively increased tension in rat aortic rings that had been precontracted using PE ($1 \mu\text{M}$; Figure 5(a)) or KCl (60 mM; Figure 5(b)) treatment. As shown in Figure 5, preincubation with HVC1 ($300 \mu\text{g}/\text{mL}$) for 20 min significantly inhibited the contraction induced by extracellular Ca^{2+} .

3.6. Standard Material Analysis. The retention times of the standards in the sample mixture were as follows: senno-side A, 3.49 min; genistein-7-glucose, 4.98 min; coptisine, 9.61 min; baicalin, 13.78 min; prunetin-5-glucose, 17.18 min; berberine, 21.22 min; baicalein, 59.76 min; wogonin, 72.53 min; prunetin, 74.12 min (Figure 6(a)). In the HPLC chromatogram of HVC1, the peaks for the standards were observed (Figure 6(b)).

4. Discussion

In this study, HVC1, a herbal prescription containing extracts of PC, SR, CR, and RR, decreased the SBP of SHR and relaxed aortic rings that had been contracted by treatment with PE or KCl.

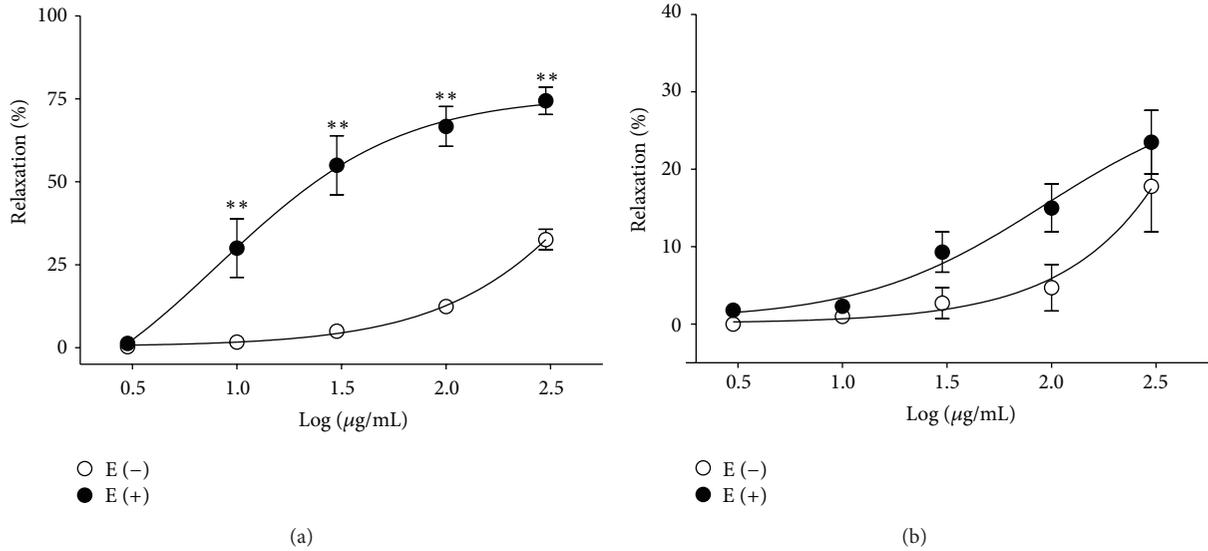


FIGURE 2: Concentration-dependent relaxant effects of HVC1 on precontracted spontaneous hypertensive rat aortic rings. Endothelium-intact [E (+)] or endothelium-denuded [E (-)] aortic rings were precontracted by phenylephrine (PE, 1 μ M) (a) or KCl (60 mM) (b). Values are expressed as mean \pm SEM ($n = 8$). ** $P < 0.01$ versus E (-).

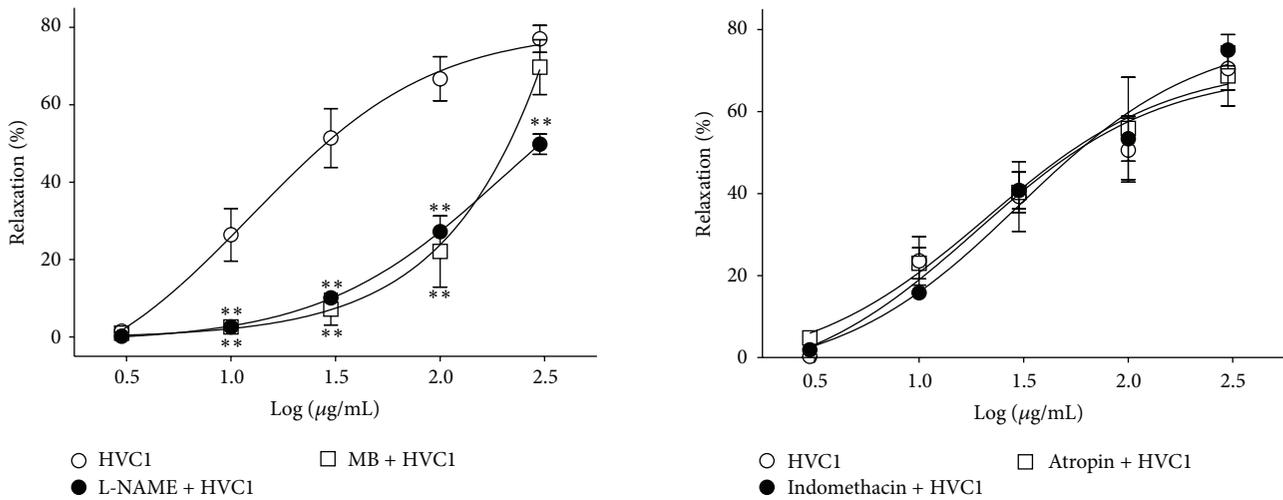


FIGURE 3: Effect of HVC1 on endothelium-intact aortic rings preincubated with *N* ω -Nitro-L-arginine methyl ester (L-NAME, 10 μ M) or methylene blue (MB, 10 μ M). The graph shows the relaxation responses induced by HVC1 in endothelium-intact spontaneous hypertensive rat aortic rings that had been precontracted with phenylephrine (PE, 1 μ M) in the presence or absence of L-NAME or MB in Krebs-Henseleit solution. The relaxant effects of HVC1 on isolated spontaneous hypertensive rat aortic rings were calculated as a percentage of the contraction in response to PE. Values are expressed as mean \pm SEM ($n = 4$). * $P < 0.05$ and ** $P < 0.01$ versus HVC1.

FIGURE 4: Effect of HVC1 on endothelium-intact aortic rings preincubated with indomethacin or atropine. Graph showing relaxation responses induced by HVC1 in endothelium-intact spontaneous hypertensive rat aortic rings that had been precontracted with phenylephrine (PE, 1 μ M) in Krebs-Henseleit solution in the presence or absence of indomethacin (10 μ M) or atropine (1 μ M). The relaxant effects of HVC1 on isolated spontaneous hypertensive rat aortic rings were calculated as a percentage of the contraction in response to PE. Values are expressed as mean \pm SEM ($n = 4$).

Over the 50-day long experimental period, the SBP of the control group (orally administrated distilled water) increased from 198.8 ± 4.6 mmHg on day 0 to 225.2 ± 1.9 mmHg on day 50. On the other hand, the SBP of the HVC1 50

and $300 \text{ mg}\cdot\text{kg}^{-1}\cdot\text{day}^{-1}$ treated groups significantly decreased during the experimental period. These results suggest that HVC1 has an antihypertensive effect.

Vascular tone is important for the regulation of blood pressure. In blood vessels, the vascular endothelium and smooth muscle play an important role in vasorelaxation. The

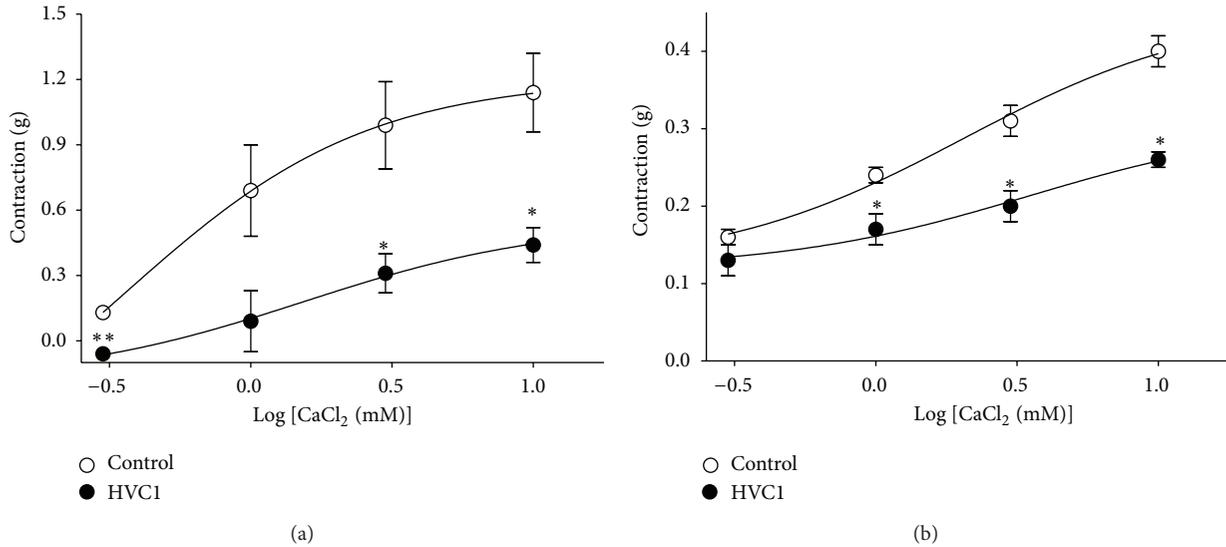


FIGURE 5: Effect of HVC1 on Ca²⁺-induced aortic ring contraction. Graph showing the inhibitory effect of HVC1 (300 μg/mL) on the contraction induced by extracellular Ca²⁺ addition (0.3–10 mM) in endothelium-denuded spontaneous hypertensive rat aortic rings that had been precontracted using phenylephrine (PE, 1 μM) (a) or KCl (60 mM) (b) treatment in Ca²⁺-free Krebs-Henseleit solution. Values are expressed as mean ± SEM (n = 4). *P < 0.05 and **P < 0.01 versus control.

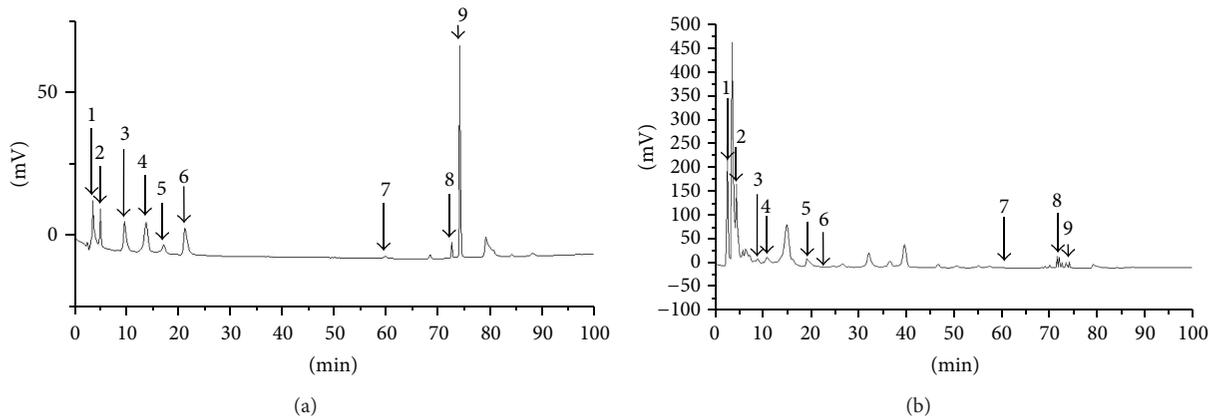


FIGURE 6: HPLC chromatogram of HVC1 standard mixtures (a) and HVC1 (b). 1: sennoside A; 2: genistein-7-O-β-glucopyranoside; 3: coptisine; 4: baicalin; 5: prunetin-5-O-β-glucopyranoside; 6: berberine; 7: baicalein; 8: wogonin; 9: prunetin.

vascular endothelium releases potent vasodilators such as nitric oxide (NO) and prostacyclin (PGI₂) [12].

NO is synthesized from L-arginine and when released from the vascular endothelium, it activates cyclic guanine monophosphate (cGMP), which leads to relaxation of vascular smooth muscles [12]. Thus, NO synthesis and the cGMP pathway are important factors in hypertension. In this study, preincubation with L-NAME (10 μM), an inhibitor of NO synthase, significantly decreased the HVC1-induced relaxation of endothelium-intact aortic rings that had been contracted using PE treatment. Preincubation with MB (10 μM), a soluble guanylate cyclase inhibitor, also significantly decreased HVC1-induced relaxation. These results suggested that the antihypertensive and vasorelaxant effects of HVC1 are partly related to NO synthesis and the NO-cGMP pathway.

PGI₂ is synthesized by cyclooxygenase and when released from the vascular endothelium, it activates adenylyl cyclase (AC). Activated AC increases the intracellular concentration of cyclic adenosine monophosphate (cAMP), which relaxes vascular smooth muscle by decreasing the intracellular calcium concentration [12]. In this study, preincubation with indomethacin (10 μM) did not affect the HVC1-induced relaxation of endothelium-intact aortic rings that had been contracted using PE treatment. These results suggested that PGI₂ might not be involved in the antihypertensive and vasorelaxant effect of HVC1.

Muscarinic receptors located on endothelial or smooth muscle cells also play an important role in vasorelaxation [13]. In this work, preincubation with atropine (1 μM), a nonselective muscarinic receptor antagonist, did not affect HVC1-induced relaxation of endothelium-intact aortic rings

that had been contracted using PE treatment. This result suggested that the muscarinic receptor might not contribute to the antihypertensive and vasorelaxant effects of HVCI.

The contraction and relaxation of vascular smooth muscle can be regulated by extracellular Ca^{2+} influx via the receptor-operative Ca^{2+} channel (ROCC) or the voltage-dependent Ca^{2+} channel (VDCC) without endothelial derived factors [14]. In the present study, preincubation with HVCI (300 $\mu\text{g}/\text{mL}$) for 20 min significantly inhibited the contraction induced by extracellular Ca^{2+} supplementation in endothelium-denuded aortic rings that had been contracted using PE or KCl treatment in Ca^{2+} -free K-H solution. These results suggested that the antihypertensive and vasorelaxant effects of HVCI are partly related to blockage of extracellular Ca^{2+} entry via the ROCC and VDCC.

Furthermore, HVCI (300 $\mu\text{g}/\text{mL}$) inhibited the PE-induced contractions to a greater extent than high K^{+} -induced contractions (Figure 5). Several reports have described that the involvement of the contractile elements is more related to agonist-induced contractions than high K^{+} . Phosphorylation of myosin light chain (MLC) is induced to a greater extent by receptor agonist like phenylephrine than by high K^{+} [15]. Moreover, drugs such as forskolin [16] and sodium nitroprusside [17] decrease the agonist-induced contractions to a greater extent than high K^{+} -induced contractions owing to the involvement of the contractile elements. The agonists like phenylephrine cause an initial spike in Ca^{2+} followed by small sustained rise in Ca^{2+} above the basal levels, thus increasing the Ca^{2+} sensitivity of MLC phosphorylation and leading to increased contraction. However, high K^{+} depolarization results in a maintained increase in the Ca^{2+} -induced contractions [18]. These findings suggest that HVCI involves the contractile elements of the aortic smooth muscle cells suggesting one of the possible mechanisms that HVCI selectively inhibits the receptor-linked Ca^{2+} channel or that it decreases the Ca^{2+} influx or Ca^{2+} sensitivity.

HVCI is a herbal prescription consisting of PC, SR, CR, and RR extracts. Sennoside A is a known standard component of RR, baicalin, baicalein, and wogonin are known standard components of SR, and berberine is a known standard component of CR [19]. However, the standard components of PC have not yet been established. In a previous study, we isolated genistein-7-*O*- β -glucopyranoside and prunetin-5-*O*- β -glucopyranoside from *Prunus* bark [11]. Prunetin is known as one of the major components of the *Prunus* species [20]. Therefore, we used these compounds as standards in the HPLC analysis of HVCI. Among these compounds, coptisine [21], baicalin [22, 23], berberine [24, 25], and wogonin [26] showed vasorelaxant activities and we found that prunetin also has vasorelaxant activities (data not shown). Therefore, these compounds might be the active compounds in HVCI that help treat hypertension.

5. Conclusions

In this study, HVCI reduced the SBP of SHRs and relaxed isolated SHR aortic rings by upregulating NO formation

and the NO-cGMP pathway and blocking the entry of extracellular Ca^{2+} via ROCC and VDCC. Therefore, HVCI could be a useful herbal medicine for the prevention and treatment of hypertension.

Conflict of Interests

The authors declare that they have no conflict of interests.

Acknowledgment

This study was supported by a Grant from the Korea Healthcare Technology R&D Project, Ministry of Health & Welfare, Republic of Korea (B110081).

References

- [1] World Health Organization, *A Global Brief on Hypertension; Silent Killer, Global Public Health Crisis*, World Health Organization, 2013.
- [2] T. A. Gaziano, A. Bitton, S. Anand, and M. C. Weinstein, "The global cost of nonoptimal blood pressure," *Journal of Hypertension*, vol. 27, no. 7, pp. 1472–1477, 2009.
- [3] Korea Centers for Disease Control and Prevention, *The Sixth Korea National Health and Nutrition Examination Survey (KNHANES VI-1)*, Korea Centers for Disease Control and Prevention, Daejeon, South Korea, 2013.
- [4] National Health Insurance Service, *National Health Insurance Statistical Yearbook 2013*, National Health Insurance Service, 2013.
- [5] Z. Chen, L. Wang, G. Yang, H. Xu, and J. Liu, "Chinese herbal medicine combined with conventional therapy for blood pressure variability in hypertension patients: a systematic review of randomized controlled trials," *Evidence-Based Complementary and Alternative Medicine*. In press.
- [6] X. Xiong, X. Yang, W. Liu, F. Chu, P. Wang, and J. Wang, "Trends in the treatment of hypertension from the perspective of traditional chinese medicine," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 275279, 13 pages, 2013.
- [7] J. Wang and X. Xiong, "Evidence-based Chinese medicine for hypertension," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 978398, 12 pages, 2013.
- [8] H.-H. Tsai, I.-J. Chen, and Y.-C. Lo, "Effects of San-Huang-Xie-Xin-Tang on U46619-induced increase in pulmonary arterial blood pressure," *Journal of Ethnopharmacology*, vol. 117, no. 3, pp. 457–462, 2008.
- [9] K. Lee, I. Ham, G. Yang et al., "Vasorelaxant effect of *Prunus yedoensis* bark," *BMC Complementary and Alternative Medicine*, vol. 13, article 31, 2013.
- [10] A. Daugherty, D. Rateri, L. Hong, and A. Balakrishnan, "Measuring blood pressure in mice using volume pressure recording, a tail-cuff method," *Journal of Visualized Experiments*, vol. 2009, no. 27, p. 1291, 2009.
- [11] J. Lee, G. Yang, K. Lee et al., "Anti-inflammatory effect of *Prunus yedoensis* through inhibition of nuclear factor- κB in macrophages," *BMC Complementary and Alternative Medicine*, vol. 13, article 92, 2013.
- [12] E. Stankevicius, E. Kevelaitis, E. Vainorius, and U. Simonsen, "Role of nitric oxide and other endothelium-derived factors," *Medicina*, vol. 39, no. 4, pp. 333–341, 2003.

- [13] L. Walch, C. Brink, and X. Norel, "The muscarinic receptor subtypes in human blood vessels," *Therapie*, vol. 56, no. 3, pp. 223–226, 2001.
- [14] H. Karaki, H. Ozaki, M. Hori et al., "Calcium movements, distribution, and functions in smooth muscle," *Pharmacological Reviews*, vol. 49, no. 2, pp. 157–230, 1997.
- [15] P. H. Ratz, "Receptor activation induces short-term modulation of arterial contractions: Memory in vascular smooth muscle," *The American Journal of Physiology—Cell Physiology*, vol. 269, no. 2, pp. C417–C423, 1995.
- [16] K. H. Han, G. J. Cheon, D. S. Yeon, and S. C. Kwon, "Forskolin changes the relationship between cytosolic Ca^{2+} and contraction in guinea pig ileum," *The Korean Journal of Physiology & Pharmacology*, vol. 13, no. 3, pp. 189–194, 2009.
- [17] H. Ozaki, T. Ohyama, K. Sato, and H. Karaki, " Ca^{2+} -dependent and independent mechanisms of sustained contraction in vascular smooth muscle of rat aorta," *Japanese Journal of Pharmacology*, vol. 52, no. 3, pp. 509–512, 1990.
- [18] R. A. Khalil, *Regulation of Vascular Smooth Muscle Function*, Morgan & Claypool Publishers, San Francisco, Calif, USA, 2010.
- [19] Ministry of Food and Drug Safety, *The Korean Pharmacopoeia*, Ministry of Food and Drug Safety, 10th edition, 2012.
- [20] H. A. Jung, A. R. Kim, H. Y. Chung, and J. S. Choi, "In vitro antioxidant activity of some selected *Prunus* species in Korea," *Archives of Pharmacal Research*, vol. 25, no. 6, pp. 865–872, 2002.
- [21] L.-L. Gong, L.-H. Fang, H.-L. Qin, Y. Lv, and G.-H. Du, "Analysis of the mechanisms underlying the vasorelaxant action of coptisine in rat aortic rings," *The American Journal of Chinese Medicine*, vol. 40, no. 2, pp. 309–320, 2012.
- [22] Y.-L. Lin, Z.-K. Dai, R.-J. Lin et al., "Baicalin, a flavonoid from *Scutellaria baicalensis* Georgi, activates large-conductance Ca^{2+} -activated K^+ channels via cyclic nucleotide-dependent protein kinases in mesenteric artery," *Phytomedicine*, vol. 17, no. 10, pp. 760–770, 2010.
- [23] Y. Huang, S. Y. Tsang, X. Yao, C. W. Lau, Y. L. Su, and Z. Y. Chen, "Baicalin-induced vascular response in rat mesenteric artery: role of endothelial nitric oxide," *Clinical and Experimental Pharmacology & Physiology*, vol. 29, no. 8, pp. 721–724, 2002.
- [24] D. G. Kang, E. J. Sohn, E. K. Kwon, J. H. Han, H. Oh, and H. S. Lee, "Effects of berberine on angiotensin-converting enzyme and NO/cGMP system in vessels," *Vascular Pharmacology*, vol. 39, no. 6, pp. 281–286, 2002.
- [25] W. H. Ko, X. Q. Yao, C. W. Lau et al., "Vasorelaxant and antiproliferative effects of berberine," *European Journal of Pharmacology*, vol. 399, no. 2-3, pp. 187–196, 2000.
- [26] M. Akinyi, X. M. Gao, Y. H. Li et al., "Vascular relaxation induced by *Eucommiae Ulmoides Oliv.* and its compounds Oroxylin A and wogonin: implications on their cytoprotection action," *International Journal of Clinical and Experimental Medicine*, vol. 7, no. 10, pp. 3164–3180, 2014.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

