Salvianolate Protects Hepatocytes from Oxidative Stress by Attenuating Mitochondrial Injury

Qiang Zhao,1 Yuan Peng,1 Kai Huang,1,2 Yang Lei,1 Hong-Liang Liu,1 Yan-Yan Tao,1,2 and Cheng-Hai Liu1,2,3

1 Institute of Liver Diseases, Shuguang Hospital Affiliated to Shanghai University of Traditional Chinese Medicine, 528 Zhangheng Road, Shanghai 201203, China
2 Shanghai Key Laboratory of Traditional Chinese Clinical Medicine, 528 Zhangheng Road, Shanghai 201203, China
3 E-Institute of TCM Internal Medicine, Shanghai Municipal Education Commission, 1200 Cailun Road, Shanghai 201203, China

Correspondence should be addressed to Yan-Yan Tao; taoyanyan1023@126.com and Cheng-Hai Liu; chenghailiu@hotmail.com

Received 6 January 2016; Revised 1 April 2016; Accepted 11 April 2016

Academic Editor: Chang-Gue Son

Copyright © 2016 Qiang Zhao et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Salvianolate is widely used to treat angiocardiopathy in clinic in China, but its application in liver diseases remains unclear. Our study aims to investigate the effect of Salvianolate on rat hepatic injury by protecting hepatocyte mitochondria. To evaluate the effects of Salvianolate on injured hepatocytes, alpha mouse liver 12 (AML-12) cells were induced with hydrogen peroxide (H$_2$O$_2$) and treated with Salvianolate. Cell viability and MitoTracker Green for mitochondria and 5,5′,6,6′-tetrachloro-1,1′,3,3′-tetraethylbenzimidazole-carbocyanide iodine (JC-1) levels and cytochrome C (Cyto-C) expressions were detected in vitro. To identify the effect of Salvianolate on protecting against mitochondria injury, male Wistar rats were injected with carbon tetrachloride (CCl$_4$) and treated with Salvianolate (40 mg·kg$^{-1}$). Serum liver function, parameters for peroxidative damage, hematoxylin and eosin (H&E) staining, and transmission electron microscope (TEM) of hepatocyte mitochondria were assayed. Our results showed that Salvianolate effectively protected hepatocytes, increased mitochondria vitality, and decreased Cyto-C expressions in vitro. Besides, Salvianolate alleviated the liver function, attenuated the indicators of peroxidation, and relieved the mitochondria injury in vivo. In conclusion, Salvianolate is effective in protecting hepatocytes from injury in vitro and in vivo, and the mechanism might be related to its protective effect on hepatocyte mitochondria against oxidative stress.

1. Introduction

Hepatocyte injury could be caused by a wide range of risk factors, including viruses, drugs, metabolic disorders, and immune attacks [1]. When hepatocytes are damaged, a large number of cytokines will be released, which further aggravates liver injury and eventually leads to liver fibrosis. Thus, protecting hepatocytes is of vital importance for curing varieties of acute and chronic liver diseases. And searching for an appropriate drug with strong protective effect against hepatocyte injury is a key point in healing live diseases.

Mitochondria are the main organelle producing alanine aminotransferase (ATP) and providing majority of the energy for cells physiological needs [2]. Moreover, mitochondria also play important roles in intracellular signal transduction and differentiation and apoptosis of cells [3–5]. Oxidative stress, related to mitochondria injury, is a common pathological mechanism for hepatic injury caused by etiologic agents [6, 7]. Therefore, antagonizing mitochondrial damage could effectively protect hepatocytes from injury [8–10].

Salvianolate, the main water soluble compound extracted from Radix Salvia miltiorrhiza, is widely used in clinic in treating cardiovascular diseases by inhibiting reactive oxygen species (ROS) production [11]. However, its pharmacological effect on liver diseases remains unclear.

In this study, we aimed to observe the effects of Salvianolate on hepatocyte mitochondria injury. To clarify our aim, several questions should be formulated. Specifically, they are as follows. (1) Could Salvianolate inhibit hepatic injury? (2) If so, could Salvianolate protect hepatocytes by inhibiting mitochondria injury? Our results demonstrated that Salvianolate could reduce hepatocyte mitochondria injury.
These findings might suggest a cytoprotective therapeutic potential of Salvianolate in the context of mitochondria injury-mediated liver diseases.

2. Materials and Methods

2.1. Drug. Salvianolate (Lot. number 120418) was provided by Shanghai Green Valley Pharmaceutical Co., Ltd.; N-acetyl-L-cysteine (NAC) (Lot. number WXBB2161V) was purchased from Sigma Chemical Co., Ltd.

2.2. Reagents. CCl₄ and H₂O₂ were obtained from Sinopharm Group Co., Ltd. All the other reagents were of the highest grade commercially available.

2.3. Cell Culture. Alpha mouse liver 12 (AML-12) cell line was purchased from Shanghai Life Science Research Institute, Chinese Academy of Sciences Institute of Biochemistry and Cell Biology. AML-12 cells were cultured in a 1:1 mixture of Dulbecco's Modified Eagle's Medium (DMEM) and Ham's F12 medium supplemented with 0.005 mg/mL insulin, 0.005 mg/mL transferrin, 5 ng/mL selenium, 40 ng/mL dexamethasone, 10% fetal bovine serum (FBS), 100 units/mL of penicillin, and 100 units/mL of streptomycin. All the cells were grown in a humidified incubator at 37°C in a 5% CO₂ atmosphere.

2.4. Drug Treatments. Salvianolate was initially dissolved as concentrated stock solution in 1 mg/mL sterile phosphate-buffered saline (PBS). For evaluating the effect of Salvianolate on AML-12 cells in vitro, AML-12 cells were cultured in a 96-well plate (Corning, NY, USA) at a density of 5,000 cells/well and were incubated with Salvianolate at the concentration of 3.125–400 μg/mL for 24 hours.

2.5. Cell Viability Assay. After the cells were incubated with Salvianolate at the concentration of 3.125–400 μg/mL for 24 hours, the cell viability assays were performed using the Cell Counting Kit 8 (CCK8) (Beyotime Biotechnology, Jiangsu, China), according to the manufacturer's protocol.

2.6. MitoTracker Staining. For live cell analysis, AML-12 cells were cultured in a 96-well plate at a density of 5,000 cells/well and were incubated with Salvianolate at the concentration of 6.25 μg/mL, 12.5 μg/mL, and 25 μg/mL for a period of 24 hours. Then mitochondrial membrane potential (MMP) was assessed by JC-1 (Beyotime, Haimen, China) staining. Briefly, AML-12 cells were stained with JC-1 for 15 min at 37°C. Cells treated with 10 μM carbonyl cyanide m-chlorophenylhydrazone (CCCP) were used as negative control. CCCP is a protonophore which can cause dissipation of MMP. Cells were finally scanned with the fluorescence microplate reader (Molecular Devices, Inc., Sunnyvale, California, USA) and the fluorescence was analyzed. Red emission of the dye represented a potential-dependent aggregation in the mitochondria, reflecting MMP. Green fluorescence represented the monomeric form of JC-1, appearing in the cytosol after mitochondrial membrane depolarization.

2.7. JC-1 Staining. AML-12 cells were cultured in a 96-well plate at a density of 5,000 cells/well and were incubated with Salvianolate at the concentration of 6.25 μg/mL, 12.5 μg/mL, and 25 μg/mL for a period of 24 hours. Then mitochondrial membrane potential (MMP) was assessed by JC-1 (Beyotime, Haimen, China) staining. Briefly, AML-12 cells were stained with JC-1 for 15 min at 37°C. Cells treated with 10 μM carbonyl cyanide m-chlorophenylhydrazone (CCCP) were used as negative control. CCCP is a protonophore which can cause dissipation of MMP. Cells were finally scanned with the fluorescence microplate reader (Molecular Devices, Inc., Sunnyvale, California, USA) and the fluorescence was analyzed. Red emission of the dye represented a potential-dependent aggregation in the mitochondria, reflecting MMP. Green fluorescence represented the monomeric form of JC-1, appearing in the cytosol after mitochondrial membrane depolarization.

2.8. Intracellular Cyto-C Measurement. AML-12 cells were cultured in a 96-well plate at a density of 5,000 cells/well and treated with 0.5 mM H₂O₂ for 20 min. Then H₂O₂ was discarded and the cells were incubated with Salvianolate at the concentration of 6.25 μg/mL, 12.5 μg/mL, and 25 μg/mL for a period of 24 hours. After 24 hours, cells were washed with cold PBS twice and fixed in 4% formaldehyde. After washing with PBS, the cells were then blocked with 5% bovine serum albumin in PBS buffer for 30 min at room temperature before being incubated with the Cyto-C (1:400) primary antibodies (Beyotime, Haimen, China). Cells were then stained with FITC-conjugated secondary antibodies. After washing, cells were double-stained with Hoechst 33342 (Beyotime, Haimen, China) to visualize the nuclei. Images were taken by Cellomics ArrayScan VTi HCS Reader.

2.9. Animals and Experimental Design. For the experiment, 43 heads of specific pathogen-free male Wistar rats weighing 150 ± 10 g were obtained from Shanghai Laboratory Animal Center, Chinese Academy of Science (Shanghai, China), and acclimated for at least 1 week before the experiment. All animals were housed in an environmentally controlled room at 22 ± 2°C and 60 ± 5% relative humidity under 12/12 h light/dark cycle. All rats were housed under controlled temperature (22°C), humidity (55%), and lighting (12-hour artificial light and dark cycle), with free access to tap water and rat chow. The standard diet pellets contained not less than 20% protein, 5% fibers, 3.5% fats, and 6.5% ash and vitamins mixture. To establish hepatocyte injury model in vivo, 43 male Wistar rats were initially divided into two groups as the normal control group (n = 10, olive oil injection) and model group (n = 33, CCl₄ injection). Firstly, rats in model group received 100% CCl₄ at dose of 3 mL kg⁻¹ with a single subcutaneous injection. After 6 h later, the model rats were further divided into 3 groups as follows: the model control group (n = 11, distilled water and CCl₄ injection), Salvianolate group (n = 11, 40 mg kg⁻¹ of Salvianolate and CCl₄ injection), and NAC group (n = 11, 500 mg kg⁻¹ of NAC and CCl₄ injection). Then, except for the rats in normal control group, which received only pure olive oil, rats in other groups were injected with 50% CCl₄-olive oil solution at dose of 2 mL kg⁻¹ subcutaneously once a week for 4 times. Subsequently, rats in Salvianolate group were injected intraperitoneally with
Salvianolate at a daily dose of 40 mg·kg\(^{-1}\) for 2 weeks. Rats in NAC group were treated with NAC orally at the dose of 500 mg·kg\(^{-1}\) in the same period of time. All rats were starved for 12 hours after the last treatment, and then they were sacrificed. The serum and liver samples were harvested under pentobarbital sodium anesthesia.

Our experiments conformed to the ethical guidelines outlined in the Guide for the Care and Use of Laboratory Animals by the Laboratory Animal Center, Shanghai University of Traditional Chinese Medicine.

2.10. Measurements of Serum Indicators of Liver Function. Activities of serum alanine transaminase (ALT), aspartate aminotransferase (AST), and total bile acid (TBA) were quantitated by using commercial kits following the instructions provided by the manufacturer (Nanjing Jian Cheng Bioengineering Institute, Nanjing, China), including the use of standardization.

2.11. Parameters for Peroxidative Damage in Liver Tissue. Hepatic homogenates were centrifuged at 3,000 rpm for 20 min at 4°C. Supernatants were immediately collected and assayed for enzyme activities. Levels of malondialdehyde (MDA), glutathione (GSH), and antisynergized free radical (ASA FR) were assayed according to the protocols of kits purchased from Nanjing Jian Cheng Bioengineering Institute. All these parameters were expressed by gram protein which was determined by the BCA protein assay kit (Pierce, Thermo Scientific, Rockford, USA) according to the manufacturer's protocol using bovine serum albumin as a standard.

2.12. Histopathological and Immunohistological Analysis of Liver Tissue. Liver tissues were fixed in buffered formalin and embedded in paraffin wax. After routine processing, liver sections of 4 𝜇m thickness underwent staining with H&E. Semiquantitative analysis of liver injury was performed according to the nonalcoholic fatty liver disease activity score (NAS): (1) hepatic steatosis: 0 (<5%); 1 (5%–33%); 2 (34%–66%); and 3 (>66%) and (2) ballooning degeneration of liver cell: 0, none; 1, rare; and 2, many [12]. Immunohistological analyses for Cyto-C and 4-hydroxynonenal (4-HNE) were performed by using anti-Cyto-C and anti-4-HNE antibodies (Abcam, ab46545) and visualized by ChemMate™ Envision™ Detection Kit (Dako). The section was pretreated using heat mediated antigen retrieval with sodium citrate buffer (pH 6, epitope retrieval solution 1) for 20 min. The section was then incubated with the first antibody, 1 μg/mL, for 15 minutes at room temperature and detected using HRP conjugated compact polymer system. DAB was used as the chromogen. The section was then counterstained with hematoxylin. All the images were analyzed with a light microscope (Olympus BX40, Japan).

2.13. Liver Perfusion and Processing for Ultrastructural Analysis. For ultrastructural analysis, the rat liver was cleared by perfusion with PBS of room temperature through the portal vein at a flow rate of 3 mL·min\(^{-1}\). One minute later, 2.5% glutaraldehyde was perfused for an additional one minute at the same flow rate. Whereafter, livers were quickly removed and thoroughly immersed in 2.5% glutaraldehyde for 48 hours at 4°C. Subsequently, the liver tissues were cut from the fixative for TEM, respectively, as described previously [13].

2.14. Statistical Analysis. All data were analyzed by using PASW Statistics 18 software. For measurement data, differences between the groups were assessed by nonparametric one-way analysis of variance. Values in the text are presented as mean ± SD. For enumeration data, the results were performed by ridit-analysis. \(p < 0.05\) was considered statistically significant.

3. Results

3.1. Salvianolate Inhibited H\(_2\)O\(_2\) -Induced Hepatocyte Injury In Vitro. H\(_2\)O\(_2\) was widely used to induce hepatocyte injury for the classical oxidative stress injury model in vitro [14, 15]. Our previous study found that the proper working concentration of H\(_2\)O\(_2\) was 0.5 mM, which caused cell viability decrease by about 40% in 30 minutes [16]. Therefore, we exposed AML-12 cells to concentration of 0.5 mM H\(_2\)O\(_2\) for 30 minutes to establish an oxidative stress injury model in this study. To evaluate the most effective concentration of Salvianolate, H\(_2\)O\(_2\)-induced AML-12 cells were treated with different concentrations of Salvianolate from 3.125 to 400 𝜇g/mL progressively. We found that the survival rate was 100% in the normal control group and Salvianolate presented the nontoxic concentration from 3.125 to 100 𝜇g/mL (Figure 1(a)). To examine the cytoprotective effects of Salvianolate on oxidative stress-induced hepatocyte injury, AML-12 cells were exposed to 0.5 mM H\(_2\)O\(_2\) for 20 min to induce hepatocyte injury. Subsequently, cells were incubated with 3.125–12.5 𝜇g/mL Salvianolate or 10 mM NAC as positive drug for 24 h. Then the cell viability was detected by the CCK8 assay. As shown in Figure 1(b), 0.5 mM H\(_2\)O\(_2\) induced a significant level of cell damage (\(p < 0.05\)). In contrast, both Salvianolate and NAC effectively protected against hepatocyte injury induced by H\(_2\)O\(_2\) in vitro (\(p < 0.05\)), and NAC had a stronger protective effect than Salvianolate (\(p < 0.05\)) (Figure 1(b)).

3.2. Salvianolate Ameliorated H\(_2\)O\(_2\)-Induced Mitochondrial Injury in Hepatocytes In Vitro. Based on the results above, we found that Salvianolate could protect AML-12 cells from H\(_2\)O\(_2\)-induced injury at the concentrations of 6.25 𝜇g/mL, 12.5 𝜇g/mL, and 25 𝜇g/mL. To confirm whether Salvianolate ameliorated hepatocytes by protecting mitochondria against injury, we further evaluated the effects of treatment with Salvianolate for 24 h on expressions of viable mitochondria, MMP dissipation, and expression of Cyto-C in cytoplasm following the exposure to 0.5 mM H\(_2\)O\(_2\) for 20 min in vitro. Firstly, we quantified the viable mitochondria expression upon H\(_2\)O\(_2\)-induced oxidative stress with or without Salvianolate by quantifying the fluorescence intensity of mitochondria probe with MitoTracker Green kits. As shown in Figure 2(a), the fluorescence intensity of mitochondria probe in model control group was obviously weakened...
compared with that in normal control group. Notably, mitochondria probe fluorescence intensity was enhanced by incubation with NAC and dose-dependent Salvianolate. Simultaneously, we examined the fluorescence intensity ratio of JC-1 aggregation/JC-1 monomer. As the results revealed in Figure 2(b), the ratio of JC-1 aggregation/JC-1 monomer was sharply decreased in the model control group, while treatment of Salvianolate or NAC significantly attenuated the condition. To further confirm the protective effect of Salvianolate on hepatocyte mitochondria, we investigated the effect of Salvianolate on the expression of Cyto-C in AML-12 cells cytoplasm. As shown in Figure 2(c), expression of Cyto-C in cytoplasm was higher in the model control group compared with that in the normal control group, and treatment with Salvianolate significantly attenuated the levels of Cyto-C.

3.3. Salvianolate Attenuated CCl₄-Induced Hepatic Inflammation in Rats. Next, we investigated the effects of Salvianolate on CCl₄-induced liver injury to further confirm the role of Salvianolate in protecting hepatocyte mitochondria injury in vivo. As shown in Figures 3(a)–3(d), hepatopancreas somatic indices (HSI) and levels of serum indicators of liver function such as ALT, AST, and TBA were all upregulated after CCl₄ administration. Salvianolate, as well as NAC, could significantly downregulate the serum liver functions. Furthermore, we investigated the effects of Salvianolate on liver inflammation by H&E staining. As shown in Figure 3(e), intact lobe structure with tidy hepatic cords and no degeneration, necrosis, or inflammatory cells infiltration were observed in normal control group. Meanwhile, a considerable mass of fat vacuoles in hepatic tissue accompanied with mononuclear cells infiltration alongside the portal area and central vein was shown in model group, as well as ballooning and hydropic degeneration of hepatocytes. After treated with Salvianolate or NAC, those pathological changes were alleviated notably. Additionally, ridit-analysis showed that treatment with CCl₄ led to fatty and ballooning degeneration significantly, while those pathological changes in Salvianolate 40 mg/kg group were strikingly reversed. However, the alleviative effect of NAC was observed only on inhibiting hepatocyte fatty degeneration (Figure 3).

3.4. Salvianolate Alleviated CCl₄-Induced Liver Oxidative Stress Injury in Rats. Besides, tissue oxidative stress parameters including MDA, GSH, and ASAFR were also tested to observe the effect of Salvianolate on liver oxidative stress injury. As shown in Figures 4(a)–4(c) and 3(e), after injury by CCl₄, higher levels of MDA and 4-HNE and lower GSH and ASAFR were presented in the model control group compared with those in the normal control group. Treatment of Salvianolate could downregulate the level of MDA and upregulate the levels of GSH, ASAFR, and 4-HNE (Figures 4(a), 4(b), and 3(e)).

3.5. Salvianolate Relieved Hepatocyte Mitochondrial Injury In Vivo. To further evaluate the effect of Salvianolate on CCl₄-induced hepatocyte injury in vivo, the morphology of liver mitochondria was observed by TEM. As shown in Figure 5, normal hepatocytes were observed in normal group and mitochondrial configuration differed with sphericity, rod or filament. Besides, mitochondrial cristae, resulting from inner mitochondrial membrane (IMM) sinking into matrix, were arranged closely. However, after stimulating with CCl₄, pathological characteristics of hepatocytes such as cellular nucleus concentrating, mitochondria swelling, and mitochondria cristae fragmentation or disappearance were commonly detected. However, all those pathological changes

![Figure 1](image_url) Cytotoxicity and proliferation of Salvianolate in AML-12 cell lines. (a) AML-12 cells were cultured in a 96-well plate at a density of 5,000 cells/well and incubated with Salvianolate with concentrations of 3.125–400 μg/mL for 24 h. (b) AML-12 cells were cultured in a 96-well plate at a density of 5,000 cells/well and exposed to 0.5 mM H₂O₂ for 20 min; then the cells were incubated with Salvianolate with concentrations of 6.25–25 μg/mL or NAC with concentration of 10 mM for 24 h. Both cytotoxicity of Salvianolate and proliferation of Salvianolate or NAC were detected by CCK8. * p < 0.05 and ** p < 0.01 versus normal control; # p < 0.05 and ## p < 0.01 versus model control.
Figure 2: Protective effects of Salvianolate on H₂O₂-induced mitochondrial injury in AML-12 cells in vitro. Hepatocytes injury model was established with 0.5 mM H₂O₂, and then the cells were incubated with Salvianolate with concentrations of 6.25, 12.5, or 25 μg/mL or NAC with concentration of 10 mM for 24 h. (a) Semiquantification data for expressions of viable mitochondria in AML-12 cells by quantifying the fluorescence intensity of mitochondria probe with MitoTracker Green kits. (b) Semiquantification data for expression of MMP in AML-12 cells by examining the fluorescence intensity ratio of JC-1 aggregation/JC-1 monomer. (c) Expression of Cyto-C (green fluorescence marked with red arrows) in cytoplasm was revealed by the images taken by Cellomics ArrayScan VTI HCS Reader. **p < 0.01 versus normal control; p < 0.05 and ## p < 0.01 versus model control.

were ameliorated after the rats were given Salvianolate or NAC.

4. Discussion

Oxidative stress is an important determinant in the pathophysiology of many hepatic diseases, for example, alcoholic hepatitis, viral hepatitis, liver fibrosis, and even liver cancer [17]. A growing body of evidences suggest that oxidative stress due to excessive production of ROS is associated with hepatocyte mitochondria injury [18]. Mitochondria are important organelles in eukaryotic cells, playing a significant role in cellular energy production. Once mitochondria are damaged by excessive ROS, the process of cell apoptosis and death can be mediated and accelerated [19]. Therefore, attempts to target ROS have been based on increasing antioxidant bioavailability and reducing ROS generation. Because of their cytotoxicity, CCl₄ and H₂O₂ have been widely used in various experiments in vivo or vitro with their activities to induce oxidative stress, resulting in the imbalance between oxidation and antioxidation [20, 21]. Thus, we employed H₂O₂ and CCl₄ in our study to investigate the effect of Salvianolate on hepatocytes oxidative stress injury in vitro and in vivo, respectively.
Figure 3: Protective effects of Salvianolate on CCl₄-induced liver injury in rats. Male Wistar rats were treated as described in the legend. Serum ALT (b), AST (c), and TBA (d) were assessed by liver function tests kits, respectively. Levels of serum liver functions were decreased, and HSI (a) was alleviated by Salvianolate treatment with dose of 40 mg/kg in CCl₄-induced liver injury mice. (e) Histologic evaluation of liver tissues was stained with H&E, and the expression of Cyto-C and 4-HNE was assayed by IHC (×200). Semiquantitative analysis of liver injury was performed according to the nonalcoholic fatty liver disease activity score (NAS): (f) hepatic steatosis: 0 (<5%); 1 (5%–33%); 2 (34%–66%); and 3 (>66%) and (g) ballooning degeneration of liver cell: 0, none; 1, rare; and 2, many. **p < 0.001, versus normal control; #p < 0.05 and ##p < 0.001, versus model control.
Salvia miltiorrhiza, a traditional Chinese medicine, has been used as a common clinical medicine for thousands of years. Salvianolate, as the main water soluble component of *Salvia miltiorrhiza*, has been proved to exert strong protective effects on vascular disorders with its antioxidant function [11]. However, the effects of Salvianolate on liver diseases via protecting mitochondria from oxidative stress remain not entirely clear yet.

In order to explore the effect of Salvianolate against liver injury, we investigated the role of Salvianolate in hepatocyte injury *in vitro* and *in vivo*, respectively. Firstly, toxicological and pharmacological studies on Salvianolate were performed on mouse hepatocytes (AML-12 cell line) *in vitro*, which implied that the Salvianolate could inhibit H\textsubscript{2}O\textsubscript{2}-induced hepatocyte injury at multiple concentrations without obvious cytotoxicity on hepatocytes. Secondly, we investigated if Salvianolate could protect the hepatocytes by inhibiting mitochondria injury. As is known to us, it is important for mitochondria to maintain physiological function with normal MMP, while MMP dissipation is considered to be a common early event of apoptosis [22]. Transformation of JC-1 fluorescence that turns from red to green indicates the descent of MMP, so it can be adopted to determine whether the cells are in the early apoptosis stage. The JC-1 staining showed that hepatocytes MMP decreased significantly after stimulating by H\textsubscript{2}O\textsubscript{2}, while treatment with Salvianolate for 24h could increase MMP remarkably in a dose-dependent manner. Cyto-C is an initiation factor of cell apoptosis locating in intermembrane space between inner and outer mitochondrial membrane. When hepatocytes are stimulated by excessive ROS, permeability transition of mitochondria changes and Cyto-C will be released into plasma from
mitochondria. Our study revealed that Salvianolate could decrease the expression of Cyto-C and 4-HNE in damaged hepatocyte plasma significantly compared with model group, which indicated that Salvianolate could keep mitochondrial membrane integrality via antagonizing H$_2$O$_2$-induced oxidative stress.

Finally, to confirm the effect of Salvianolate in vivo, we investigated the effect of Salvianolate on hepatocyte mitochondria in CCl$_4$-induced liver injury model with rats. Salvianolate could alleviate levels of serum liver function in CCl$_4$-induced liver injury rats. It was obvious that Salvianolate could relieve the infiltration of inflammatory cells and degeneration of hepatocytes. Besides, level of the indicator of membrane peroxidation, MDA, was decreased, while the main nonenzymatic-ROS scavengers in liver cells such as GSH and ASAFl were increased obviously by Salvianolate treatment. However, the results of serum liver function and histopathological assessment indicated that the antioxidant effect of Salvianolate was observed only when it was used at dose of 40 mg/kg. These results suggested that Salvianolate had potential anti-injury effect on CCl$_4$-induced liver injury.

5. Conclusions

Above all, Salvianolate is effective in alleviating hepatocytes injury via protecting mitochondria in vitro and in vivo, which might provide alternative treatment options for hepatic injury.

Abbreviations

ALT: Alanine transaminase  
AMF-12: Alpha mouse liver 12  
ASAFl: Antisuperoxide anion free radical  
AST: Aspartate aminotransferase  
ATP: Alanine aminotransferase  
CCCP: Carbonyl cyanide m-chlorophenylhydrazone
CCK: Cell Counting Kit  
CCl4: Carbon tetrachloride  
Cyto-C: Cytochrome C  
FBS: Fetal bovine serum  
GSH: Glutathione  
H&E: Hematoxylin and eosin  
HIS: Hepatopancreas somatic indices  
4-HNE: 4-Hydroxynonenal  
H2O2: Hydrogen peroxide  
IHC: Immunological Histological Chemistry  
JC-1: 5,5′,6,6′-Tetrachloro-1,1′,3,3′-tetraethylbenzimidazole-carbocyanide iodine  
MDA: Malondialdehyde  
MMP: Mitochondrial membrane potential  
NAC: N-Acetyl-L-cysteine  
NAS: Nonalcoholic fatty liver disease activity score  
PBS: Phosphate-buffered saline  
ROS: Reactive oxygen species  
TBA: Total bile acid  
TEM: Transmission electron microscope.

Competing Interests

The authors declare no conflict of interests.

Authors’ Contributions

Yan-Yan Tao and Yuan Peng designed the project; Qiang Zhao, Hong-Liang Liu, and Yuan Peng performed experiments; Kai Huang and Yang Lei analyzed the data; Qiang Zhao, Yuan Peng, and Yan-Yan Tao prepared the paper; Cheng-Hai Liu made critical revision for paper. Qiang Zhao and Yuan Peng equally contributed to the work as the first author.

Acknowledgments

This work was supported by the following grants: National Natural Science Foundation of China (nos. 81173405, 81102701, and 81503322), National Science and Technology Major Project (2014ZX10005001), the International S&T Cooperation Program of China (2014DFA31440), E-Institute (E03008), the Program for Outstanding Medical Academic Leader of Shanghai Municipalities (LJH0005), China Postdoctoral Science Foundation (no. 2015M571595), “Three-Year Action Plan” for Development of TCM in Shanghai (ZY3-CCCX-2-1003), and Innovative Research Team in Universities of Shanghai Municipal Education Commission.

References

[19] X. Zhang and J. J. Lemasters, “Translocation of iron from lysosomes to mitochondria during ischemia predisposes to...


