Research Article

Antimycobacterial Activity and Safety Profile Assessment of Alpinia galanga and Tinospora cordifolia

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Tuberculosis (TB) remains a common deadly infectious disease and worldwide a major health problem. The current study was therefore designed to investigate the in vitro antimycobacterial activity of different extracts of Alpinia galanga and Tinospora cordifolia. Moreover, a safety assessment for both plants was carried out. Dichloromethane and ethanolic extracts of each plant were examined against H37Rv INH-sensitive and resistant INH strain of Mycobacterium tuberculosis. The safety assessment of both plants has been performed through in vivo acute and chronic toxicity studies in animal model. Body weight, food consumption, water intake, organ’s weight, and haematological and biochemical parameters of blood and serum were evaluated. The extracts of A. galanga and T. cordifolia produced significant and dose-dependent inhibitory activity with maximum effect of 18-32% at 50 μg/ml against both strains of M. tuberculosis. No effect on the body weight or food and water consumption was observed but A. galanga caused significantly an increase in the relative weight of the heart, liver, spleen, and kidney. Haematological studies of both plants revealed a slight but significant fall in the RBC and WBC level as well as haemoglobin and platelets. In addition, A. galanga extracts increased significantly liver enzymes and bilirubin and glucose.

1. Introduction

Tuberculosis (TB) is one of the global health problems and leading causes of human morbidity and mortality [1, 2]. TB is recognized as one of the top 10 causes of death worldwide. The World Health Organization (WHO) estimated 10.4 million people fell ill with TB in 2016 [3]. In 2016, there were an estimated 1.3 million TB deaths among HIV-negative people and an additional 374 000 deaths among HIV-positive people. Over 95% of TB deaths occur in low- and middle-income countries [3]. Although the current treatment of TB with drug combination of isoniazid, rifampicin, ethambutol, and pyrazinamide has a high success rate, drug-resistant TB is a continuing threat [4, 5]. In 2016, there were 600 000 new cases with resistance to rifampicin (RRTB), the most effective first-line drug, of which 490 000 had multidrug-resistant TB (MDR-TB) [3]. Consequently, the need to develop new novel tuberculosis drugs which are less toxic and more effective against resistant Mycobacterium strains is imperative. Natural products, either as pure isolated compounds or as extracts, represent a promising source of novel drugs for the treatment of various diseases including TB [5, 6]. Therefore, we were encouraged to investigate two well-known medicinal plants, namely, Alpinia galanga and Tinospora cordifolia.

Alpinia galanga (L.) Willd. (Zingiberaceae) known as Galangal, a member of the ginger family and native to Southern China and Thailand, is primarily used as a flavoring especially in the preparation of fresh Thai curry paste and Thai soup [7]. It is widely cultivated in Southeast Asian countries, e.g., China, Indonesia, Thailand, India, and Philippines [8]. Galangal exhibited different pharmacological activities such as antimicrobial, anti-inflammatory, carminative, antipyretic, aphrodisiac, and emmenagogue and traditionally has been used for the treatment of various diseases such as kidney disorders, diabetes, cough, tuberculosis, bronchitis, rheumatism, asthma, and heart diseases [8–12].
**Tinospora cordifolia** (Willd.) Miers which belongs to the family Menispermaceae is a climbing shrub widely distributed in tropical areas in South East Asia, e.g., India, China, Myanmar, Sri Lanka, Indonesia, Malaysia, Thailand, and Philippines [13]. It is introduced in Saudi Arabia from Pakistan and grown as ornamental. In Indian colloquial, this plant is known as Giloya, meaning elixir of paradise, which kept celestial beings young and saved from aging [14]. The plant has significant medicinal importance and is widely used in the Indian folk medicine for increasing the lifespan, promoting intelligence, and improving memory and as antiaging agent [13–15]. Recent studies on *T. cordifolia* reported antioxidant, radical scavenging, hepatoprotective, anticancer, antiallergic, immunomodulatory, and anti-inflammatory effects [16–21]. The current study was therefore designed to investigate the antimycobacterial activity of different extracts of *Alpinia galanga* and *Tinospora cordifolia*. Moreover, a safety profile assessment of both plants has been carried out through in vivo acute and chronic toxicity studies in animal models.

### 2. Materials and Methods

#### 2.1. Plant Materials

The plant materials, namely, *Alpinia galanga* (roots and rhizomes) and *Tinospora cordifolia* (leaves and stems) were purchased from India, Dawa Khana Tibbiya College, Aligarh Muslim University, Aligarh. The plants were authenticated by Professor S.H. Afaq, Department of Ilmul Advia, Ajmal Khan Tibbiya College, Aligarh Muslim University, Aligarh. A voucher specimen for each plant was preserved in the lab for further documentation.

#### 2.2. Extraction of the Plants

The air-dried and powdered plant materials were defatted with petroleum ether. Then the plant materials were percolated separately with dichloromethane in 10 L percolator for one day. The process was repeated till the exhaustion of the plant materials. Then the obtained extracts were collected, combined, filtered, and evaporated using a rotary evaporators (Buchi® evaporator, Switzerland) at 40 °C under vacuum. Then, the same extraction process was done for both plant materials using ethanol 96%. The extracts were then preserved at 4 °C until testing.

#### 2.3. Phytochemical Screening

Thin layer chromatography was developed for each plant's extract using different mixtures of organic solvents as mobile phases, visualized under UV (254 and 366 nm), and sprayed with various chemical reagents, e.g., anisaldehyde-sulfuric acid for terpenoids, Dragendorff's reagent for alkaloids, Borntrager reagent for anthraquinones, etc. according to previously published methodology [22].

#### 2.4. Determination of the Antimycobacterial Activity

**2.4.1. Mycobacterial Strains/Isolates.** The *in vitro* antimycobacterial activity of the extracts was carried out against two *Mycobacterium tuberculosis* strains, that are H37Rv INH-sensitive and resistant INH strains. Samples were prepared at 5 mg/ml in DMSO by sonicating and vortexing as needed and then added to Middlebrook 7H12 media and serially diluted 2-fold within 96-well plates in a volume of 100 μl per well. *M. tuberculosis* H37Rv was then added in a volume of 100 μl 7H12 to achieve a bacterial density of approximately 1 x 10⁵ CFU/ml. The highest final concentration of the samples was therefore 50 μg/ml with a maximum final DMSO concentration of 1% v/v.

**2.5. Acute Toxicity Study.** For acute studies, Swiss albino mice (home bred) aged 6–7 weeks, weighing about 24–28 g, were taken from Animal Care Centre (College of Pharmacy, King Saud University) and fed on Purina Chow diet and water ad libium, were used in this study. The animals were maintained under controlled temperature, humidity, and automated light cycles (12 h light, 12 h dark). The protocol of the current study (CBR 4537) was approved by the Ethics Committee of the Experimental Animal Care Society, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia. According to the test guideline of Organization for Economic Cooperation and Development (OECD), the toxicity tests were carried out [23].

**2.6. Toxicity Study Design.** For the determination of the acute toxicity, mice (males) were randomly divided into different groups (N=6–10). Different doses of each test extract (0.5, 1, 2, 5, 8, and 10 g/kg) were administered intraperitoneally. The extracts were suspended in 0.2% aqueous Tween 80 or 0.25% carboxymethyl cellulose. The animals were observed for 72 h for signs of toxicity and mortality and LD₅₀ was calculated according to published method [24].

**2.7. Chronic Toxicity Study.** A total of 40 male and 40 female Swiss albino mice were randomly allocated to the control and test groups. The extract in each case was mixed with drinking water for feasibility of administration due to long treatment duration. The dose selected was 1/4₀₀₀ of the LD₅₀. The treatment was continued for a period of 12 weeks [24]. The food consumption and water intake recorded weekly. The body weights of animals recorded shortly before the administration of the tested extracts and at the end of each week. The animals were then observed for all external general symptoms of toxicity, body weight changes, and mortality. The average pre- and posttreatment body weights, vital organ weights of the treated animals, were compared with the control group. Ten male and ten female rats were used in each group having one control and two treated groups. One group of treated female rats were mated with treated males and pregnancy outcomes were studied. Urine was collected 1-2 days before the end of the treatment. The treated animals were fasted for 12 h and then anesthetized. Blood samples were collected via heart puncture and centrifuged at 3000 rpm for 10 min. The plasma was then stored at -20 °C pending for analysis of the biochemical parameters. Vital organs were removed, weighed, and investigated for apparent signs of toxicity and stored in 10% formalin for histological studies. The percentage of each organ relative to the body weight of the animal was calculated.

**2.8. Determination of the Antimycobacterial Activity.** BD Bactec™ MGIT 960 kit (Becton, Dickinson and Company, USA) was used for antymycobacterial susceptibility testing of *Mycobacterium tuberculosis*. The Bactec MGIT 960 kit was used with Bactec MGIT System.
2.8. Haematological Studies. Whole noncentrifuged blood was used for determination of some haematological values. The blood was analyzed for WBC and RBC count, haemoglobin, platelets, neutrophils, and lymphocytes measurement using Contraves Digicell 3100H (Zurich).

2.9. Serum Analysis of Biochemical Parameters. A colorimetric method was used for the determination of the biochemical parameters (AST, ALT, GGT, ALP, bilirubin, glucose, lipid profile, and total protein) in plasma. The enzyme activity was quantified spectrophotometrically using commercial enzymatic kits (Crescent Diagnostics Test Kits, SA) [25].

2.10. Statistical Analysis. The results were presented as mean ± standard error of the mean (SEM). Statistical differences were analyzed using ANOVA with Dunnett as posttest. A value of p < 0.05 was considered statistically significant [26].

3. Results

As presented in Table 1, the yield of the dichloromethane extracts is more than the ethanolic extracts of both plant species. The results of the phytochemical screening in Table 1 indicated different types of active constituents in Alpinia galanga such as essential oil with terpenoids and flavonoids, while Tinospora cordifolia indicated the presence of terpenoids, alkaloids, and flavonoids.

### Table 1: Results of extraction of the plants and phytochemical screening.

<table>
<thead>
<tr>
<th>Plants</th>
<th>Part used</th>
<th>Solvent</th>
<th>Yield in % (g)</th>
<th>Phytochemical screening</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alpinia galanga</td>
<td>roots &amp; rhizomes</td>
<td>Dichloromethane</td>
<td>4.2 (115)</td>
<td>terpenoids, essential oil, phenolics</td>
</tr>
<tr>
<td></td>
<td>roots &amp; rhizomes</td>
<td>Ethanol</td>
<td>1.0 (26)</td>
<td>terpenoids, flavonoids, phenolics</td>
</tr>
<tr>
<td>Tinospora cordifolia</td>
<td>leaves &amp; stems</td>
<td>Dichloromethane</td>
<td>2.2 (54)</td>
<td>terpenoids, flavonoids</td>
</tr>
<tr>
<td></td>
<td>leaves &amp; stems</td>
<td>Ethanol</td>
<td>1.5 (35)</td>
<td>flavonoids, alkaloids</td>
</tr>
</tbody>
</table>

3.1. Antimycobacterial Activity. As depicted in Figure 1, whereas ethanolic and dichloromethane extracts of Alpinia galanga (AGET and AGDC) produced significant (p < 0.001) and dose-dependent inhibitory activity against sensitive strains of Mycobacterium tuberculosis (MT), the negative control (DMSO) did not show any effect on the bacterial growth. AGET exhibited a significant (p < 0.001) and dose-dependent inhibition on sensitive strains of MT. Maximum inhibitory effect was shown at 50 μg/ml (22.3%, Figure 1). Moreover, AGET produced a significant (p < 0.001) inhibitory effect on resistant strains of MT only at the highest concentration 50 μg/ml (12.7%, Figure 1). In addition to that, Figure 1 demonstrated that AGDC showed significant (p < 0.001) dose-dependent inhibitory effect only on sensitive strains of MT with maximum effect at 19.7% at 50 μg/ml.

The results of the inhibitory effect of Tinospora cordifolia ethanolic and dichloromethane extracts (TCET and TCDC) against sensitive and resistant MT are shown in Figure 2. Figure 2 demonstrated that TCET produced a significant (p < 0.001) and dose-dependent inhibitory effect against both sensitive and resistant strains of MT with maximum effect of 32.3% and 22.7% at 50 μg/ml, respectively. Furthermore, TCDC showed a significant (p < 0.001) antimycobacterial effect against the sensitive strain of MT with maximum effect of 23% at 50 μg/ml (Figure 2). It also produced significant (p < 0.001) and dose-dependent inhibitory effect against the resistant strain of MT with maximum effect of 18.3% at 50 μg/ml (Figure 2).

3.2. Effect of the Extracts in Acute Toxicity Test. The administration of a dose lower than 5 g/kg of all extracts did not show any mortality or observable symptoms. However, gross behavioural changes such as increased heart rate, convulsion, twitches, itching and excitation, mortality, and other signs of toxicity manifestations were observable with the highest doses (5, 8 and 10 g/kg). The treatment of the animals with doses ≥ 5 g/kg of A. galanga dichloromethane extract (AGDC) produced increased respiration, pilo erection, Straub tail, tremors, increased muscle tone, and sedation, while A. galanga ethanolic extract (AGET) produced defecation, writhing, sedation, and calmness. The administration of T. cordifolia ethanolic extract (TCET) increased heart rate and CNS excitation (convulsion, twitches, and itching). In addition, both extracts of T. cordifolia produced defecation and writhing. The mortality rate and LD₅₀ values are demonstrated in Table 2. As depicted in Table 2, the most toxic extract was AGDC (6.6 g/kg) followed by the TCET (6.83 g/kg). The rest of the extracts were relatively safer with LD₅₀ values of 7.5–77 g/kg (Table 2).
3.3. Effect of the Extracts on Body Weight, Food, and Water Consumption and Organ’s Weight in Chronic Oral Toxicity Test. During the treatment period of 12 weeks, the body weight has increased gradually in the control and extracts treated mice compared to the control mice (data are not shown). The percentage of increase in body weight of the extracts treated female and male mice groups (data are not shown).

3.4. Effect of the Extracts on Haematological Parameters in Chronic Oral Toxicity Test. The haematological analysis from control and treated animal groups for the chronic toxicity study is shown in Table 4. Treatment with A. galanga extracts AGDC at 166 mg/kg and AGET at 193 mg/kg showed variable changes in the haematological parameters, while AGDC and AGET decreased significantly ($P < 0.01$) red and white blood cells count and only AGET decreased significantly ($P < 0.01$) haemoglobin and number of platelets. Treatment with T. cordifolia extract TCET at 170 mg/kg showed a significant decrease ($P < 0.01$) in red and white blood cells count, haemoglobin, and lymphocytes (Table 4), whereas TCDC only decreased number of platelets significantly ($P < 0.05$).

3.5. Effect of the Extracts on Biochemical Parameters in Chronic Oral Toxicity Test. For the biochemical parameters, the administration of A. galanga extract AGDC at 166 increased significantly AST ($P < 0.05$), ALT, GGT, ALU, bilirubin, and blood glucose level ($P < 0.001$) (Table 5). In addition, the treatment with AGET at 193 mg/kg showed a significant increase of ALT, ALT and bilirubin ($P < 0.01$), GGT ($P < 0.01$), and ALU ($P < 0.05$). Furthermore, as demonstrated in Table 5, the administration of T. cordifolia extract TCDC caused a significant decrease of GGT ($P < 0.001$) and blood glucose level ($P < 0.01$), whereas TCET showed a significant increase of AST and bilirubin ($P < 0.01$) and ALU ($P < 0.05$) (Table 5). In addition to that, both plants exhibited a significant increase ($P < 0.01$) of sodium, potassium, and creatinine level (Table 5).

3.6. Effect of the Extracts on Lipid Profile and Total Protein in Chronic Oral Toxicity Test. As shown in Table 6, the treatment with A. galanga extract AGDC at 166 increased significantly ($P < 0.001$) the level of cholesterol, triglycerides, HDL, and VLDL and decreased significantly ($P < 0.001$) the total protein compared to the normal control group. On the contrary, the administration of AGET significantly ($P < 0.05$) decreased the level of lipid profile compared to the normal control group (Table 6). In addition, the administration of TCDC significantly ($P < 0.01$) reduced the level of all lipid profile compared to the normal control group, whereas TCET caused a significant ($P < 0.01$) elevation of cholesterol, triglycerides, HDL, and VLDL and a significant ($P < 0.05$) reduction of LDL and total protein (Table 6).
Table 3: Effect of the extracts on the relative organ weight of mice after 12 weeks treatment.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>AGDC 166 mg/kg</th>
<th>% change</th>
<th>AGET 193 mg/kg</th>
<th>% change</th>
<th>TCDC 189 mg/kg</th>
<th>% change</th>
<th>TCET 170 mg/kg</th>
<th>% change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart</td>
<td>0.11 ± 0.006</td>
<td>0.15 ± 0.004 **</td>
<td>36 †</td>
<td>0.15 ± 0.01***</td>
<td>36 †</td>
<td>0.13 ± 0.006</td>
<td>18 †</td>
<td>0.12 ± 0.008</td>
<td>9 †</td>
</tr>
<tr>
<td>Liver</td>
<td>1.29 ± 0.07</td>
<td>1.47 ± 0.02 **</td>
<td>14 †</td>
<td>1.48 ± 0.01***</td>
<td>15 †</td>
<td>1.14 ± 0.32</td>
<td>11 †</td>
<td>1.46 ± 0.02**</td>
<td>13 †</td>
</tr>
<tr>
<td>Lungs</td>
<td>0.39 ± 0.01</td>
<td>0.45 ± 0.001*</td>
<td>15 †</td>
<td>0.44 ± 0.02</td>
<td>12 †</td>
<td>0.36 ± 0.006</td>
<td>8 †</td>
<td>0.40 ± 0.01</td>
<td>2 †</td>
</tr>
<tr>
<td>Spleen</td>
<td>0.11 ± 0.007</td>
<td>0.15 ± 0.01*</td>
<td>36 †</td>
<td>0.12 ± 0.01*</td>
<td>9 †</td>
<td>0.13 ± 0.006</td>
<td>18 †</td>
<td>0.13 ± 0.007</td>
<td>18 †</td>
</tr>
<tr>
<td>Kidney</td>
<td>0.32 ± 0.01</td>
<td>0.38 ± 0.01*</td>
<td>18 †</td>
<td>0.44 ± 0.01***</td>
<td>37 †</td>
<td>0.33 ± 0.003</td>
<td>3 †</td>
<td>0.44 ± 0.008**</td>
<td>37 †</td>
</tr>
<tr>
<td>Testis</td>
<td>0.23 ± 0.007</td>
<td>0.26 ± 0.01</td>
<td>13 †</td>
<td>0.28 ± 0.01*</td>
<td>21 †</td>
<td>0.23 ± 0.02</td>
<td>0.27 ± 0.02</td>
<td>17 †</td>
<td></td>
</tr>
</tbody>
</table>

All values represent mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001; ANOVA, followed by Dunnett's multiple comparison test. Test value compared with control group.
<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>AGDC 166 mg/kg</th>
<th>% change</th>
<th>AGET 193 mg/kg</th>
<th>% change</th>
<th>TCDC 189 mg/kg</th>
<th>% change</th>
<th>TCET 170 mg/kg</th>
<th>% change</th>
</tr>
</thead>
<tbody>
<tr>
<td>RBC (x 10^6/mm³)</td>
<td>7.8 ± 0.12</td>
<td>6.80 ± 0.12**</td>
<td>13↓</td>
<td>6.35 ± 0.15***</td>
<td>18↓</td>
<td>7.00 ± 0.20</td>
<td>10↓</td>
<td>6.92 ± 0.08**</td>
<td>11↓</td>
</tr>
<tr>
<td>WBC (x 10^3/mm³)</td>
<td>9.10 ± 0.17</td>
<td>8.45 ± 0.06**</td>
<td>7↓</td>
<td>6.97 ± 0.11***</td>
<td>23↓</td>
<td>9.17 ± 0.12</td>
<td>-</td>
<td>7.77 ± 0.19**</td>
<td>15↓</td>
</tr>
<tr>
<td>Hemoglobin (g/dl)</td>
<td>11.75 ± 0.19</td>
<td>11.0 ± 0.20</td>
<td>6↓</td>
<td>9.05 ± 0.19***</td>
<td>23↓</td>
<td>11.45 ± 0.42</td>
<td>2↓</td>
<td>10.32 ± 0.19**</td>
<td>12↓</td>
</tr>
<tr>
<td>Platelets (x 10^5/μL)</td>
<td>276.7 ± 4.32</td>
<td>263.0 ± 3.24</td>
<td>5↓</td>
<td>244.7 ± 3.37**</td>
<td>12↓</td>
<td>255.0 ± 2.58*</td>
<td>8↓</td>
<td>260.25 ± 2.68*</td>
<td>6↓</td>
</tr>
<tr>
<td>Neutrophils (x 10^3/mm³)</td>
<td>3.70 ± 0.09</td>
<td>3.15 ± 0.14</td>
<td>15↓</td>
<td>3.35 ± 0.06</td>
<td>9↓</td>
<td>3.45 ± 0.15</td>
<td>7↓</td>
<td>3.30 ± 0.25</td>
<td>11↓</td>
</tr>
<tr>
<td>Lymphocytes (x 10^3/mm³)</td>
<td>5.95 ± 0.11</td>
<td>5.52 ± 0.23</td>
<td>7↓</td>
<td>5.67 ± 0.18</td>
<td>5↓</td>
<td>5.37 ± 0.25</td>
<td>10↓</td>
<td>4.87 ± 0.12***</td>
<td>18↓</td>
</tr>
</tbody>
</table>

All values represent mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001; ANOVA, followed by Dunnett’s multiple comparison test. Test value compared with control group.
### Table 5: Effect of the extracts on biochemical parameters of mice in chronic toxicity test.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>AGDC 166 mg/kg</th>
<th>% change</th>
<th>AGET 193 mg/kg</th>
<th>% change</th>
<th>TCDC 189 mg/kg</th>
<th>% change</th>
<th>TCET 170 mg/kg</th>
<th>% change</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/l)</td>
<td>102.75 ± 5.21</td>
<td>128.25 ± 6.10*</td>
<td>26†</td>
<td>134.0 ± 4.14**</td>
<td>31†</td>
<td>98.45 ± 5.18</td>
<td>4†</td>
<td>131.0 ± 5.36**</td>
<td>28†</td>
</tr>
<tr>
<td>ALT (U/l)</td>
<td>30.62 ± 2.14</td>
<td>47.97 ± 1.42***</td>
<td>56†</td>
<td>53.92 ± 4.45**</td>
<td>76†</td>
<td>26.7 ± 1.64</td>
<td>13†</td>
<td>39.62 ± 2.93</td>
<td>35†</td>
</tr>
<tr>
<td>GGT (U/l)</td>
<td>4.75 ± 0.15</td>
<td>6.72 ± 0.23***</td>
<td>42†</td>
<td>5.75 ± 0.29*</td>
<td>21†</td>
<td>3.12 ± 0.08***</td>
<td>34†</td>
<td>4.6 ± 0.28</td>
<td>3†</td>
</tr>
<tr>
<td>ALP (U/l)</td>
<td>320.75 ± 9.64</td>
<td>414.0 ± 6.83***</td>
<td>29†</td>
<td>425.0 ± 6.64***</td>
<td>32†</td>
<td>347.50 ± 7.55</td>
<td>8†</td>
<td>385.5 ± 6.06*</td>
<td>12†</td>
</tr>
<tr>
<td>Bilirubin (mg/dl)</td>
<td>0.52 ± 0.01</td>
<td>0.77 ± 0.01***</td>
<td>47†</td>
<td>0.71 ± 0.02**</td>
<td>36†</td>
<td>0.50 ± 0.01</td>
<td></td>
<td>0.68 ± 0.02**</td>
<td>30†</td>
</tr>
<tr>
<td>Glucose (mg/dl)</td>
<td>122.0 ± 4.88</td>
<td>196.25 ± 7.18***</td>
<td>61†</td>
<td>136.25 ± 6.26</td>
<td>11†</td>
<td>779.0 ± 0.02**</td>
<td>36†</td>
<td>123.50 ± 8.45</td>
<td>26†</td>
</tr>
<tr>
<td>Sodium (mEq/L)</td>
<td>84.18 ± 1.97</td>
<td>10714 ± 2.8***</td>
<td>27†</td>
<td>99.10 ± 2.36**</td>
<td>17†</td>
<td>117.47 ± 3.9***</td>
<td>40†</td>
<td>105.99 ± 1.9 ***</td>
<td>26†</td>
</tr>
<tr>
<td>Potassium (mEq/L)</td>
<td>5.0 ± 0.02</td>
<td>7.22 ± 0.28***</td>
<td>45†</td>
<td>6.97 ± 0.11***</td>
<td>40†</td>
<td>6.6 ± 0.12***</td>
<td>32†</td>
<td>5.60 ± 0.12</td>
<td>12†</td>
</tr>
<tr>
<td>Calcium (mg/dl)</td>
<td>7.42 ± 0.59</td>
<td>5.61 ± 0.66</td>
<td>24†</td>
<td>5.52 ± 0.65</td>
<td>26†</td>
<td>5.23 ± 0.42**</td>
<td>29†</td>
<td>6.95 ± 0.47</td>
<td>6†</td>
</tr>
<tr>
<td>Urea (mmol/l)</td>
<td>39.1 ± 1.78</td>
<td>38.95 ± 1.41</td>
<td>29†</td>
<td>29.92 ± 3.50</td>
<td>23†</td>
<td>59.35 ± 4.26**</td>
<td>52†</td>
<td>65.0 ± 3.53***</td>
<td>66†</td>
</tr>
<tr>
<td>Uric acid (mg/dl)</td>
<td>6.97 ± 0.25</td>
<td>4.95 ± 0.09***</td>
<td>29†</td>
<td>8.87 ± 0.30***</td>
<td>27†</td>
<td>6.87 ± 0.30</td>
<td></td>
<td>10.27 ± 0.45***</td>
<td>47†</td>
</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>0.92 ± 0.04</td>
<td>1.01 ± 0.04</td>
<td>9†</td>
<td>1.27 ± 0.02***</td>
<td>38†</td>
<td>1.44 ± 0.02***</td>
<td>56†</td>
<td>1.52 ± 0.09***</td>
<td>65†</td>
</tr>
</tbody>
</table>

*All values represent mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001; ANOVA, followed by Dunnett's multiple comparison test. Test value compared with control group.
Table 6: Effect of the extracts on lipid profile and total protein of male and female mice in chronic toxicity test.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>AGDC 166 mg/kg</th>
<th>% change</th>
<th>AGET 193 mg/kg</th>
<th>% change</th>
<th>TCDC 189 mg/kg</th>
<th>% change</th>
<th>TCET 170 mg/kg</th>
<th>% change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cholesterol</td>
<td>108.0 ± 3.43</td>
<td>170.75 ± 4.4***</td>
<td>58†</td>
<td>85.7 ± 4.37*</td>
<td>21†</td>
<td>77.9 ± 4.95**</td>
<td>28†</td>
<td>133.75 ± 3.42**</td>
<td>24†</td>
</tr>
<tr>
<td>Triglycerides (mg/dl)</td>
<td>60.6 ± 2.29</td>
<td>147.0 ± 5.36***</td>
<td>143†</td>
<td>39.92 ± 1.38***</td>
<td>34†</td>
<td>43.71 ± 2.24**</td>
<td>28†</td>
<td>119.25 ± 6.1***</td>
<td>97†</td>
</tr>
<tr>
<td>HDL (mg/dl)</td>
<td>51.0 ± 0.87</td>
<td>87.55 ± 5.4***</td>
<td>72†</td>
<td>37.82 ± 2.21***</td>
<td>26†</td>
<td>39.25 ± 1.2***</td>
<td>23†</td>
<td>82.1 ± 3.3***</td>
<td>61†</td>
</tr>
<tr>
<td>VLDL (mg/dl)</td>
<td>12.12 ± 0.45</td>
<td>29.4 ± 107***</td>
<td>143†</td>
<td>79.8 ± 0.27***</td>
<td>34†</td>
<td>8.75 ± 0.44**</td>
<td>28†</td>
<td>23.85 ± 1.2***</td>
<td>97†</td>
</tr>
<tr>
<td>LDL (mg/dl)</td>
<td>44.64 ± 4.35</td>
<td>60.3 ± 6.66</td>
<td>35†</td>
<td>35.34 ± 1.53</td>
<td>20†</td>
<td>26.47 ± 1.33**</td>
<td>41†</td>
<td>28.67 ± 4.98*</td>
<td>36†</td>
</tr>
<tr>
<td>Total Protein (g/dl)</td>
<td>5.3 ± 0.14</td>
<td>3.05 ± 0.11***</td>
<td>42†</td>
<td>3.7 ± 0.18***</td>
<td>30†</td>
<td>6.25 ± 0.40</td>
<td>18†</td>
<td>3.67 ± 0.320***</td>
<td>31†</td>
</tr>
</tbody>
</table>

All values represent mean ± SEM. *p < 0.05; **p < 0.01; ***p < 0.001; ANOVA, followed by Dunnett's multiple comparison test. Test value compared with control group.
4. Discussion

Tuberculosis (TB) remains a common deadly infectious disease and worldwide a major health problem. The World Health Organization (WHO) estimated 10.4 million people fell ill with TB in 2016 [3]. In 2016, WHO reported about 1.5 million TB deaths. TB ranks as the second leading cause of deaths among infectious diseases after HIV [27]. The current combination therapy with antimycobacterial drugs like isoniazid, rifampicin, streptomycin, and ethambutol causes various side effects mainly hepatotoxicity. Moreover, the development of multidrug-resistant bacterial strains made the treatment more difficult and complicated. Consequently, there is an urgent need for novel, more effective, with lower side effects, and less expensive drugs. Keeping these aforementioned facts in mind, medicinal plants have received more attention as a potential source in drug discovery against TB [28]. In the current study, we examined the in vitro antimycobacterial activity of four extracts obtained from two medicinal plants namely Alpinia galanga and Tinospora cordifolia against two Mycobacterium tuberculosis strains, that are H37Rv INH-sensitive and resistant INH strains. The present work was further extended to evaluate the safety profile of both A. galanga and T. cordifolia whereas in vivo assessment of the acute and chronic toxicity in animal model has been performed. Criteria for selection these plant species for investigation are the traditional uses to treat cough and other respiratory tract conditions including tuberculosis [8, 11, 13, 15] as well as the previously reported antimicrobial activity against various bacterial strains [8, 9, 15, 29, 30].

In general the H37Rv INH-sensitive M. tuberculosis strain showed more susceptibility than the resistant INH strain to the investigated extracts. Our obtained results revealed that the ethanolic and dichloromethane extracts of A. galanga (AGET and AGDC) possessed a considerable significant antimycobacterial activity at the highest concentration tested (50 μg/ml). Our data are in agreement with previously published reports on A. galanga [8, 31]. In the study carried out by Soundhari and Rajarajan [31], it was shown that A. galanga has an antimycobacterial activity against isoniazid-resistant strain with MIC-value of 250 μg/ml, whereas Gupta and coworkers [8] described a bactericidal activity against M. tuberculosis under axenic aerobic conditions at 50-100 μg/ml. The variation in the active concentrations may be attributed to differences in the methods used in extraction and the assay used for the evaluation of the antimycobacterial activity. A recent published study done by Warit and coworkers [32] reported the evaluation of the antituberculosis activity of one of the major compounds in A. galanga, namely, 1'-acetoxychavicol acetate and its enantiomers. It was shown that the S-enantiomer of 1'-acetoxychavicol acetate has a remarkable antimycobacterial activity against H37Ra and H37Rv strains with MIC values of 0.2 and 0.7 μg/ml, respectively. There is little data in the literature about antimycobacterial activity of Tinospora cordifolia. To the best of our knowledge this is the first report on the antimycobacterial activity of T. cordifolia against H37Rv INH-sensitive and resistant INH strains. The results of the inhibitory effect of T. cordifolia ethanolic and dichloromethane extracts (TCET and TCDC) against sensitive and resistant M. tuberculosis revealed a considerable significant antimycobacterial activity at the highest concentration tested (50 μg/ml). Our obtained result was indirectly in agreement with a recently published study by Gupta and coworkers [33] who reported the isolation of a polysaccharide from T. cordifolia which inhibited the survival of M. tuberculosis by controlling influence on host immune responses. It is assumed that this modulation would improve the therapeutic efficacy of currently used antituberculosis drugs and offer an interesting strategy for the development of other choice of treatments to control this disease.

Our phytochemical screening showed the presence of terpenoids, essential oils, phenolic compounds (phenylpropanoids), and flavonoids in A. galanga. These results are in agreement with data reported on the chemistry of A. galanga [34–37]. Moreover, our data revealed the presence of terpenoids, alkaloids, and flavonoids in T. cordifolia. These results are also in agreement with reports on preliminary phytochemical screenings of this plant [19, 38, 39]. We assume that the displayed antimycobacterial activity could be attributed to such classes of natural compounds which may contribute together in growth inhibition of M. tuberculosis.

Flavonoids isolated from Erythrina schliebenii were studied for antimycobacterial activity against M. tuberculosis (H37Rv strain) and exhibited MIC values 36.9-101.8 μM [40]. A recent study demonstrated that isorhamnetin possessed antimycobacterial activity against multidrug- and extensively drug-resistant clinical isolates of H37Rv strain of M. tuberculosis, with MIC values of 158 and 316 μM, respectively [41]. In addition, 3-cinnamoyltribuloside, tribuloside, afzelin, and astilbin which were isolated from Heritiera littoralis showed antimycobacterial activity against Mycobacterium madagas- cariense and Mycobacterium indicus pranii, with MIC values in the range of 0.8-1.6 mg/ml [42, 43].

Furthermore, terpenoids, which were isolated from various natural sources, e.g., medicinal plants, fungi, and marine organisms also displayed a promising and interesting antitubercular activity against different strains of M. tuberculosis [43–47]. Recently, Isaka and coworkers [44, 45] reported a potent antitubercular activity of several lanostane triterpenoids isolated from different cultures of Ganoderma species with MIC values ranging between 0.78 and 12.5 μg/ml. In addition, a lot of isolated alkaloids belonging to various classes, e.g., indole, pyrrole, indoloquinoline, carbazole, manzamine, quinoline, isoquinoline, and pyrrolidine alkaloids were investigated for their antimycobacterial activities against M. tuberculosis, where many of them exhibited a notable and potent efficacy and may be regarded as lead molecules for the treatment of tuberculosis [48].

In the present study, we investigated the acute and chronic toxicity of both plant species in animal model. The treatment of animals with doses lower than 5 g/kg of all extracts in acute toxicity test did not show any mortality or observable symptoms. The mortality rate and LD50 values were calculated. Surprisingly, A. galanga showed more toxicity than T. cordifolia. In the chronic toxicity test, we evaluated the extracts effects on body weight, food and water consumption, organ weight, haematological and biochemical parameters, and lipid profile. After 12 weeks of treatment, it was observed
that both plants have no effect on the body weight or food and water consumption but A. galanga caused significantly an increase in the relative weight of the heart, the liver, the spleen, and kidney as compared to the control group. The rise in organ weight could be referred to stimulation of xenobiotic enzymes promoting to raise in proteins synthesis. Regularly the inducement of these enzymes leads to a raise of relative organ weight following an exposure to xenobiotic [49, 50]. It could be argued that these alterations might be toxicological significant particularly for heart, liver, and kidney but the raise in the relative weight of spleen might be due to the high unevenness of the weight of this organ [51]. Haematological studies of both plants revealed a significant fall in the RBC and WBC level as well as haemoglobin and platelets. Our results were not in agreement with data reported previously by Qureshi and coworkers [52] who demonstrated a significant rise in the RBC level of A. galanga-treated animals. In addition, A. galanga extracts increased significantly all liver enzymes (AST, ALT, GGT, and ALU) which could indicate a liver damage or injury. The significant increase of blood glucose level by A. galanga extracts could be attributed to insufficiency in the storage of glucose or to demolition of β−cells of the pancreas. In our study, the prolonged administration of the extracts showed a large variation in the lipid profile which could be attributed to the nature of the constituents in those extracts and might be connected to liver damage.

5. Conclusion

The results reported in this study are quite interesting showing considerable antimycobacterial activity of A. galanga and T. cordifolia. The antimycobacterial activity is consistent with traditional use in the treatment of cough-diseases including tuberculosis. Moreover, our results have exhibited that A. galanga and T. cordifolia might be toxic to heart, liver, and kidney after long-term administration. Thus, some caution should be taken into consideration when these plant species are administered for long periods.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Mohamed F. Alajmi carried out the study design and experimental work and corrected the manuscript for publication. Ramzi A. Mothana performed data collection and interpretation and literature search and wrote the manuscript. Adnan J. Al-Rehaily and Jamal M. Khaled supported carrying out the acute and chronic toxicity assays and correcting the manuscript. All authors read and approved the final manuscript.

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