Research Article

Alleviating the Intestinal Absorption of Rhein in Rhubarb through Herb Compatibility in Tiaowei Chengqi Tang in Caco-2 Cells

Ying Peng, Min Fan, Chongsheng Peng, Mengyue Wang, and Xiaobo Li

School of Pharmacy, Shanghai Jiao Tong University, Shanghai 200240, China

Correspondence should be addressed to Xiaobo Li; xbli@sjtu.edu.cn

Received 17 November 2017; Accepted 28 December 2017; Published 30 January 2018

1. Introduction

Tiaowei Chengqi Tang (TWCQT) is composed of rhubarb, processed liquorice, and Natrii Sulphas, which is used as a purgative in traditional Chinese medicine (TCM). This study focused on the intestinal absorption of rhein in disassembly of the TWCQT extracts through the Caco-2 cell monolayer model to explicate the possible detoxification mechanism of herb-herb compatibility in TWCQT. The results showed that the intestinal absorption of rhein occurred through active diffusion, and rhein might be composed of breast cancer resistance protein (BCRP) substrates. The extract of processed liquorice increased the exclusion rate and reduced intracellular uptake of rhein. The consistent results observed in TWCQT further implied that processed liquorice in TWCQT could suppress the absorption of rhein across the Caco-2 cell monolayer. It has therefore been concluded that the active ingredients of processed liquorice may play a critical role in reducing the intestinal absorption of rhein to alleviate the toxicity of rhubarb in TWCQT. Because of BCRP’s involvement in rhein transport, we conjectured that some components in processed liquorice could inhibit the transport of rhein, possibly by mediating BCRP. These results would provide new insight into this ancient drug combination in toxicity reduction and clinical use.
with rhubarb in rats [12]. Liquorice could inhibit the intestinal motility in rats [13] and induce CYP3A enzyme [14], which thereby could relieve the abdominal pain caused by rhubarb and accelerate the metabolism of anthraquinones. While the combination therapies of TWCQT have been validated and show potential clinical benefits, herb-herb interactions in prescriptions have not yet been fully clarified.

In the present study, we employed the Caco-2 cell monolayer model to investigate the transport difference of rhein alone and rhein in different decoctions, including rhubarb extract (RE), rhein with processed liquorice extract (rhein + PE), rhubarb with processed liquorice extract (RPE), and TWCQT and their mechanisms. The aim of this study was to clarify whether the intestinal transport of rhein is influenced by other components in rhubarb, the effects of prepared liquorice, and TWCQT on the intestinal absorption of rhein.

2. Materials and Methods

2.1. Herbs. Rhubarb, Natrii Sulphas, and processed liquorice were purchased from Leiyunshang Pharmacy of Leiyunshang Pharmaceutical Co., Ltd. (Shanghai, China). All herbs were authenticated by Professor M. Y. Wang, an experienced botanist specializing in medicinal herbs. Voucher specimen (2015II02-a, b, c) has been deposited at herbarium of School of Pharmacy, Shanghai Jiao Tong University, Shanghai, China.

The crude slices of rhubarb (8 g) were extracted twice with 8 times volume boiling water for 30 min each time. The filtrate was concentrated to prepare the stock solution of 1 mg/mL for rhubarb. Prepared liquorice (4 g) and TWCQT (rhubarb 8 g, Natrii Sulphas 6 g, and processed liquorice 4 g) were processed by the same approach to obtain the corresponding stock solution. The amounts of rhein in the 1 mg/ml RE, RPE, and TWCQT were determined by ultra-high-performance liquid chromatography-quadrupole time-of-flight-high-resolution mass spectrometry-mass spectrometry (UPLC-QTOF-HRMS-MS). All stock solutions were further diluted with D-Hanks to obtain a series of solutions with required concentrations.

2.2. Reagents and Chemicals. The human colon adenocarcinoma cell line, Caco-2, was purchased from the Cell Bank of the Academy of Science (Shanghai, China). Rhein was purchased from Man Site Biotechnology Co., Ltd. (Sichuang, China). Dulbecco’s Modified Eagle’s Medium (DMEM, high-glucose 4.5 g/l), nonessential amino acids (NEAA), and penicillin and streptomycin solution (10,000 U/ml penicillin and 10,000 μg/ml streptomycin) were purchased from Genome Bio-Medical Technology Co., Ltd. (Hangzhou, China). Fetal bovine serum (FBS) was purchased from Hangzhou Sijiqing Co., Ltd. (Hangzhou, China). Trypsin/ethylenediaminetetraacetic acid (EDTA) solution (0.25%) was obtained from Beijing Dingguo Changsheng Biotechnology Co., Ltd. (Beijing, China). Verapamil, Ko143, and MK-571 were purchased from Sigma-Aldrich (St. Louis, MO, USA). Millicell cell culture inserts were obtained from Millipore (USA). Twenty-four-well cell culture clusters were obtained from Costar (Corning Incorporated, USA).

2.3. Caco-2 Cell Culture and Cytotoxicity Assay. Caco-2 cells were maintained in DMEM containing 10% FBS (inactivation at 56°C for 30 min), 1% NEAA, and 1% penicillin and streptomycin solution in a humidified atmosphere with 5% CO2 at 37°C. The medium was replaced every 2 days, and the cells were passaged at 80 to 90% confluence using trypsin/phosphate-buffered saline (PBS, pH 7.4). The cytotoxicity of rhein, RE, PE, RPE, and TWCQT to Caco-2 cells was evaluated by MTT assay. Briefly, 200 μl of Caco-2 cells was seeded in a 96-well plate (Corning, NY, USA) at a density of 2 × 104 cells/ml. Following incubation for 24 h, different concentrations of test solution were added. Thereafter, 20 μl of 5 mg/mL MTT was added, the medium was removed after 4 h, and the remaining formazan crystals were solubilized with 150 μl of dimethyl sulfoxide (DMSO). The absorbance was measured at 490 nm on a microplate reader (Thermo Scientific, Tokyo, Japan). Cells incubated without the test samples were used as controls. In each MTT assay, every sample was tested in 6 replicates.

2.4. Transport of Analytes across the Caco-2 Monolayer. For transport experiments, the cells were seeded at a density of 1 × 105 cells/ml on polycarbonate membranes of cell culture inserts (6.5 mm membrane diameter, 0.4 μm pore size, and 0.33 cm2 surface area) and placed in 12-well cell culture clusters. The transepithelial electrical resistance (TEER) was assessed using a Millicell ERS-2 (Millipore, USA) to reflect the tightness of intercellular junctions. The Caco-2 cell monolayers were used for transport experiments on day 21 of postseeding with TEER values > 500 Ω cm².

The transports of rhein alone, rhein in RE, PE, RPE, and TWCQT across Caco-2 monolayers were investigated. Briefly, the cell monolayers were washed 3 times with D-Hanks. After each wash, the plates were incubated in fresh D-Hanks for 30 min at 37°C. The transport experiments were conducted by adding the test solution to either the apical (A, 0.5 ml) or basolateral side (B, 1.5 ml), while the receiving chamber contained the corresponding volume of prewarmed drug-free D-Hanks. Every experiment was repeated 3 times, and the plates were incubated in an orbital shaker at 37°C. Six sequential samples (50 μl) were taken at different times (30, 60, 90, 120, and 150 min) from both sides of the cell monolayer. The same volume of D-Hanks was immediately added to replace the samples obtained. Transport experiments of rhein were also performed in the presence of efflux transporter inhibitors (i.e., verapamil, Ko143, and MK-571). All inhibitors were loaded onto the A side of the monolayers. Verapamil is a P-glycoprotein (P-gp) inhibitor, Ko143 is a breast cancer resistance protein (BCRP) inhibitor, and MK-571 is a multidrug resistance protein 2 (MRP2) inhibitor. The concentrations of all the samples were analyzed by UPLC-QTOF-HRMS-MS.

2.5. Cellular Uptake of Analytes in Cultured Caco-2 Cell. Cell monolayers were prepared as described for the transport studies. Test solution were loaded onto the A side of the cell monolayers over 2 h at 37°C. Cells attached to the polycarbonate membranes were cut off from the inserts,
immersed in 1 ml blank D-Hanks, and sonicated for 15 min. The mixture was centrifuged at 10,000 × g for 5 min. The concentrations of all the samples were analyzed by UPLC-QTOF-HRMS-MS.

2.6. UPLC-QTOF-HRMS-MS Analysis. Analyte measurement was performed on a Waters ACQUITY UPLC system (Waters Corp., Milford, MA, USA). Chromatography was carried out on an ACQUITY UPLC BEH C18 column (100 mm × 2.1 mm i.d, 1.7 μm, Waters Corp., USA). The mobile phase consisted of 0.1% formic acid (A) and acetonitrile (B), using a gradient elution of 13% B at 0–8 min, 27.5% B at 8–14 min, 37.5% B at 14–25 min, and 100% B held for 2 min. The gradient was recycled back to 13% in 2.5 min for the next run. The flow rate was 0.6 ml/min. The detection wavelengths were set at 254, 270, and 340 nm. The injection volume was 3 μl. The temperature of the column oven was set to 35 °C.

Mass spectrometry was carried out using a Waters SYNAPT mass spectrometer (Waters Corp., Milford, MA, USA). Ionization was performed in the negative electrospray ionization (ESI) mode. The MS parameters were as follows: capillary voltage, 2.8 kV; cone voltage, 35 V; source temperature, 115 °C; desolvation temperature, 350 °C; gas flows of cone and desolvation, 50 and 700 l/h, respectively. For accurate mass measurement, leucine enkephalin was used as the lock mass. The MSE experiment in two scan functions was carried out as follows: function 1 (low energy), m/z 50–1000, 0.25 s per scan time, 0.02 s interscan delay, and 4 eV collision energy; function 2 (high energy), m/z 50–1000, 0.25 s per scan time, 0.02 s interscan delay, and 4 eV collision energy ramp of 55–70 eV.

2.7. Data Analysis. Data in the present study were presented as mean ± standard deviation (SD). Significant analysis was performed using Statistical Package for Social Sciences (SPSS) version 16.0 (SPSS Inc., Chicago, IL, USA). Statistical significance was determined by one-way ANOVA with Dunnett’s test and differences were considered significant when p < 0.05. In the Caco-2 cell model, the rate of transport was obtained from the amount transported versus time curve using linear regression. The apparent permeability (P_\text{app}) used as an expression of the absorption rate constant was calculated as follows: P_\text{app} = (dQ/dt)/(A \times C_0), where dQ/dt is the rate at which the compound appears in the receiver chamber (mg/s), A is the surface area of the filter membrane (1 cm²), and C_0 is the initial concentration in the donor chamber (mg/mL). Efflux ratio (ER) was calculated from the following equation: ER = P_\text{app}(B \rightarrow A)/P_\text{app}(A \rightarrow B), where P_\text{app}(B \rightarrow A) is the P_\text{app} value measured in the B to A direction and P_\text{app}(A \rightarrow B) is the P_\text{app} value measured in the A to B direction.

3. Result

3.1. Quantification of Rhein in RE, RPE, and TWCQT by UPLC-QTOF-HRMS-MS. The amounts of rhein in the 1 mg/ml RE, RPE, and TWCQT were determined by UPLC-QTOF-HRMS-MS (Figure 1). The results showed that the amounts of rhein in RPE (1 mg/ml) and TWCQT (1 mg/ml) were 27.7 μg/ml and 27.3 μg/ml, respectively; however, in RE (1 mg/ml) this amount was 8.61 μg/ml. This may be explained by the processed liquorice’s ability to improve the extraction rate of rhein in rhubarb during codecoction. Therefore, the concentrations of rhein in RE (8.61 μg/ml) and TWCQT (27.3 μg/ml) served as the references for uptake and transport experiments in cultured Caco-2 cells.

3.2. Cytotoxicity of Rhein, RE, PE, RPE, and TWCQT on Caco-2 Cell. To ensure cell viability during the permeability experiments, viability of cells was directly measured using an MTT test to evaluate the cytotoxicity of rhein, RE, PE, RPE, and TWCQT toward Caco-2 cells prior to transport experiments. Generally, a higher cell viability of more than 90% indicated that the compounds at the concentrations were nontoxic to the cells. The results showed that rhein at or below the concentration of 200 μg/ml was nontoxic to the Caco-2 cells after 48 h exposure (IC50 3.49 ± 0.96 mg/ml). RE, PE, RPE, and TWCQT at concentrations up to 4 mg/ml showed no toxic effects to Caco-2 cells (IC50 19.71 ± 0.96, 19.88 ± 1.34, 23.70 ± 2.52, and 21.87 ± 0.53 mg/ml, resp.).

3.3. Transport of Rhein through Caco-2 Cell Monolayers and Effects of Inhibitors. The transports of 8.61, 17.22, 34.44, 27.3, 54.6, and 109.2 μg/ml rhein across a Caco-2 cell monolayer from sides A to B and B to A were investigated (Figure 2).
and Table 1). The $P_{\text{app}}$ values of rhein from B to A were significantly ($p < 0.05$) higher than those from A to B. The ERs of rhein at different concentrations were all more than 1.5, which indicated that rhein could be absorbed across intestinal epithelial cells in active absorption patterns, and its transport process might be mediated by some transporters.

The effects of efflux transporters on rhein are shown in Figure 3(a). Verapamil (50 $\mu$mol/ml) reduced the amounts of rhein (27.3 $\mu$g/ml) transported from B to A, and induction of ER decreased to 2.12, but with no significant influence on the $P_{\text{app}}$ values of rhein from A to B or B to A. MK-571 (50 $\mu$mol/ml) showed significant influences on the $P_{\text{app}}$ values of rhein from B to A ($p < 0.05$) and resulted in decreased ER (1.98). The above results suggest that MRP2 might mediate the transport of rhein. After treatment of 10 $\mu$mol/ml of Ko143, the amounts of rhein transported from B to A significantly decreased ($p < 0.01$) and caused the decrease of ER (1.27), indicating that BCRP may be involved in the transport of rhein and perform an important role.

3.4. Transport of Rhein in RE, PE, RPE, and TWCQT through Caco-2 Cell Monolayers. The transport flux of rhein across the Caco-2 in RE, PE, RPE, and TWCQT is shown in Figure 2 and Table 1. No significant differences were observed in the $P_{\text{app}}$ values of rhein in 1, 2, and 4 mg/ml RE compared with corresponding concentrations of rhein alone, suggesting that other components in RE did not alter the membrane permeability of rhein in Caco-2 cells (Figure 2). However, the A to B flux and the B to A efflux of 27.3, 54.6, and 109.2 $\mu$g/ml rhein in Caco-2 cells were significantly inhibited ($p < 0.05$), causing an increase in ER (3.20–3.53, approximately 1.3-fold compared to rhein alone) in 1, 2, and 4 mg/ml of PE (Table 1). The $P_{\text{app}}$ values of rhein determined in RPE and TWCQT were consistent with those of PE, showing similar reduction in both directions with ER values of 3.16 to 3.66. These results indicated that processed liquorice in TWCQT might inhibit the absorption of rhein in Caco-2 cells (Table 1). Generally, transport flux of a compound with ER values more than 1.5 is considered as an active efflux. Results obtained from the study of transport of rhein in PE, RPE, and TWCQT were all greater than 3. Therefore, the mechanism of permeation for rhein in PE, RPE, and TWCQT in the translocation across Caco-2 cell monolayers was supposed to involve active efflux, and some components in processed liquorice, which may be the substrate of MRP or BCRP, would act as antagonists in the transport of rhein in Caco-2 cells.

3.5. Uptake of Rhein by Caco-2 Cells and Effects of Inhibitors. Results of the cellular uptake of rhein (27.3 $\mu$g/ml) into Caco-2 cell monolayers from the A compartment over 2 h are shown in Figure 3(b). After 2 h of incubation, the cellular uptake of rhein was 0.68 ng/$\mu$g protein. To investigate the involvement of active efflux in the uptake of rhein, the effects of P-gp, MRP2, and BCRP selective inhibitors on the uptake of rhein were performed. After treatment of Ko143, intracellular accumulation of rhein significantly increased, and no significant changes in the intracellular amounts of rhein were observed after treatment with verapamil or MK-571. With the treatment of Ko143, both the ER value and the intracellular accumulation were significantly altered, suggesting that BCRP may mediate the transport of rhein.

3.6. Uptake of Rhein in RE, PE, RPE, and TWCQT by Caco-2 Cells. Further, uptake studies were performed in the presence of RE, PE, RPE, and TWCQT to determine the effects of RE, PE, RPE, and TWCQT on the uptake of rhein (Figure 3(c)). The intracellular accumulation of rhein in RE adding 18.69 $\mu$g/ml rhein was approximately equal to that of 27.3 $\mu$g/ml rhein alone, suggesting that components in RE did not alter the uptake of rhein in Caco-2 cells. However, the uptake of rhein into Caco-2 cells was significantly reduced in PE, RPE, and TWCQT ($p < 0.05$) compared with rhein at

### Table 1: Effects of prepared licorice extract, rhubarb + prepared liquorice extract, and TWCQT on transport of rhein across the Caco-2 cell monolayer (mean ± SD, n = 6).

<table>
<thead>
<tr>
<th>Group</th>
<th>Concentration</th>
<th>$P_{\text{app}}$ ($\times 10^{-6}$ cm/s)</th>
<th>$P_{\text{BA}}$</th>
<th>$P_{\text{AB}}$</th>
<th>ER</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rhein</td>
<td>27.3 $\mu$g/ml</td>
<td>2.31 ± 0.13</td>
<td>6.54 ± 0.27</td>
<td>2.83</td>
<td></td>
</tr>
<tr>
<td></td>
<td>54.6 $\mu$g/ml</td>
<td>2.58 ± 0.26</td>
<td>7.02 ± 0.58</td>
<td>2.72</td>
<td></td>
</tr>
<tr>
<td></td>
<td>109.2 $\mu$g/ml</td>
<td>2.52 ± 0.19</td>
<td>6.72 ± 0.31</td>
<td>2.38</td>
<td></td>
</tr>
<tr>
<td>Rhein + PE</td>
<td>27.3 $\mu$g/ml + 1 mg/ml</td>
<td>1.28 ± 0.02*</td>
<td>4.32 ± 0.15*</td>
<td>3.38</td>
<td></td>
</tr>
<tr>
<td></td>
<td>54.6 $\mu$g/ml + 2 mg/ml</td>
<td>1.39 ± 0.09*</td>
<td>4.92 ± 0.23**</td>
<td>3.53</td>
<td></td>
</tr>
<tr>
<td></td>
<td>109.2 $\mu$g/ml + 4 mg/ml</td>
<td>1.92 ± 0.03*</td>
<td>6.14 ± 0.14*</td>
<td>3.20</td>
<td></td>
</tr>
<tr>
<td>RPE*</td>
<td>1 mg/ml</td>
<td>1.48 ± 0.11*</td>
<td>4.84 ± 0.14**</td>
<td>3.27</td>
<td></td>
</tr>
<tr>
<td></td>
<td>2 mg/ml</td>
<td>1.39 ± 0.07*</td>
<td>5.09 ± 0.27*</td>
<td>3.66</td>
<td></td>
</tr>
<tr>
<td></td>
<td>4 mg/ml</td>
<td>1.73 ± 0.15*</td>
<td>5.72 ± 0.14*</td>
<td>3.30</td>
<td></td>
</tr>
<tr>
<td>TWCQTb</td>
<td>1 mg/ml</td>
<td>1.56 ± 0.12*</td>
<td>4.94 ± 0.13**</td>
<td>3.16</td>
<td></td>
</tr>
<tr>
<td></td>
<td>2 mg/ml</td>
<td>1.48 ± 0.09*</td>
<td>5.15 ± 0.32*</td>
<td>3.48</td>
<td></td>
</tr>
<tr>
<td></td>
<td>4 mg/ml</td>
<td>1.82 ± 0.11*</td>
<td>5.82 ± 0.19*</td>
<td>3.19</td>
<td></td>
</tr>
</tbody>
</table>

PE: prepared licorice extract, RPE: rhubarb + prepared liquorice extract; TWCQT: Tiaowei Chengqi Tang. *The amounts of rhein in 1, 2, and 4 mg/ml RPE were 277, 55.4, and 110.8 $\mu$g/ml, respectively. **The amounts of rhein in 1, 2, and 4 mg/ml TWCQT were 27.3, 54.6, and 109.2 $\mu$g/ml, respectively; $*p < 0.05$; **$p < 0.01$ compared with rhein alone.
the same dosage, which is consistent with the permeability results. According to these results, we indicate that processed liquorice in TWCQT may inhibit the absorption of rhein in Caco-2 cells to alleviate the toxicity of rhubarb.

4. Discussion

Because of the toxicity of rhubarb and related anthraquinones [10], the US Food and Drug Administration (FDA) proposed limits on the dose and duration of usage of dietary supplements containing rhubarb for weight loss. In TCM theories, rhubarb is also considered to possess toxicity, and TWCQT, a multiherb prescription, which consists of rhubarb and other two drugs (processed liquorice and Natrii Sulfas), can be much safer than that of a single-herb rhubarb decoction when it is given at the same dose recorded in TCM [2]. It was pointed out that in the process of the codecoction of prescribed herbs, chemical constituents may change due to the solvent or heating [15]. In certain formulas, liquorice may reduce toxic intermediates when combined with some toxic herbs [16]; however, liquorice may increase the activity of components of some other herbs [17]. In our studies, we found that processed liquorice could increase the dissolution of rhein in RPE and TWCQT decoctions. Rhein is reported to be the major compound absorbed by the body as determined by human plasma analysis after oral administration of the water extract of rhubarb [6] and one of the major poisonous ingredients of rhubarb [18]. The pharmacokinetics of rhein in rats following oral administration of TWCQT showed that the concentration of rhein in plasma was reduced by herbs mixture [11]. These results suggested that some components in the TWCQT might reduce the absorption of rhein in rhubarb. Thus, it is meaningful to determine the potential role of herbs in TWCQT in the intestinal transport of rhein.
Aviello et al. [19] reported that rhein was devoid of cytotoxic and genotoxic effects in human colon adenocarcinoma cells (Caco-2) at 0.1–10 µg/ml; however, at concentrations present in the colon after a human therapeutic dosage of senna, rhein inhibited cell proliferation. In our study, we found that rhein (3.375–200 µg/ml) had no significant cytotoxic effect on proliferating Caco-2 cells with IC50 3.49 ± 0.96 mg/ml.

ATP-binding cassette transporters, including P-gp, MRP and BCRP abundantly located in the apical membrane of the intestinal epithelium, are crucial to limit toxicant absorption to prevent poisoning. Verapamil and MK-571 are well-known as the inhibitors of P-gp and MRP2, respectively [20]. Ko143, a potent and selective BCRP inhibitor, displays higher than 200-fold selectivity over P-gp and MRPI transporters [21]. Consistent with the previous study [22], we demonstrated that, in the Caco-2 cells, the permeability of rhein from B to A was significantly higher than that from A to B. In the presence of BCRP (Ko143) or MRP2 inhibitor (MK-571), the permeability of rhein significantly decreased from the B to A direction, suggesting that BCRP and MRP2 may participate in the efflux of rhein. However, MK-571 was reported to inhibit MRPs, P-gp, and BCRP with distinctive potencies. It was also a less effective inhibitor of BCRP [23]. Therefore, the role of MRP2 in the rhein transport requires further study. Our study revealed that the intracellular accumulations of rhein only significantly increased in the presence of Ko143, while they did not increase in the presence of MK-571 or verapamil. These results provided evidence that only BCRP was involved in rhein transport, which were consistent with Ye et al. [22]. Ye et al. found that rhein was more permeable in the B to A side than that in the opposite in the Madin Darby canine kidney (MDCK) II-BCRP cells; however, no significant differences of rhein permeability were observed in two directions in both MDCK II-MDR1 and MDCK II-MRP2 cells [21]. Nevertheless, Van Gorkom et al. reported that rhein was less cytotoxic in the MRPI overexpressing GLC4/ADR cell line compared to GLC4, MRPI inhibition with MK571 increased rhein cytotoxicity, and carboxyfluorescein efflux was blocked by rhein. Thus, they concluded that rhein was a substrate for the MRPI drug efflux pump and was a cytotoxic agent capable of inducing apoptosis [5]. These contradictory results may be explained by different cell lines and assessment methods.

It has been reported that there were no significant differences in \( P_{\text{app}} \) across the Caco-2 cell monolayer between the rhein alone and combinations of rhein, baikalin, and berberine [24]. Meanwhile, rhein has been proven to have no significant effect on the absorption of highly permeable drugs during coadministration, such as ketoprofen, paracetamol, propanolol, verapamil, digoxin, and rhodamine 123. However, furosemide permeability was enhanced by rhein, which may be partly due to the opening of the paracellular spaces and/or effects on active efflux [25]. In our study, we found that processed liquorice could increase the ER and decrease the intracellular accumulations of rhein, implying that the active ingredients from processed liquorice may play an important role in decreasing the absorption of rhein. Liquorice is a widely used herbal medicine native to southern Europe and parts of Asia and has beneficial applications in both the medicinal and the confectionery sectors. Unlike its usage in Europe, liquorice is commonly combined with other herbs in TCM prescriptions, to enhance the effectiveness of other ingredients, to reduce toxicity, and to improve flavor in almost half of Chinese herbal formulas [15]. The “mediation” effect of liquorice has been demonstrated to occur partially through modulation of drug transporter proteins. Liquorice was shown to inhibit P-gp in an in vivo ATPase assay [26] and in intestinal mucosa [27]. The inhibitory action of P-gp was enhanced by a combination of liquorice and Kansui [28] or Daphne genkwa [29]. However, He et al. reported that liquorice extraction and its main components (glycyrrhizin, glycyrrhetic acid, and liquiritin) could activate the P-gp and upregulate its expression [30]. Glycyrrhetic acid, the major metabolite of glycyrrhizin, was also found to activate the P-gp [31]. These discrepancies may be related to other chemicals in liquorice or different concentrations or inspection methods. In addition, glycyrrhizin has been demonstrated to increase hepatic glutathione content, possibly by inhibiting MRP2 [32]. As yet, no systematic studies have been published regarding the effects of liquorice and its active components on BCRP. The effect of liquorice and its active components on rhein transport in Caco-2 cells involved with BCRP requires further study. Further studies are also needed to elucidate how processed liquorice exerts its inhibitory action on the transport of rhein in Caco-2 cells.

5. Conclusions

In conclusion, our study provided evidence that processed liquorice in TWCQT could decrease the transport of rhein in Caco-2 cells. The active ingredients of processed liquorice may play a critical role in reducing the absorption of rhein to alleviate the toxicity of rhubarb in TWCQT. Because of the involvement of BCRP in rhein transport, we conjectured that some components in processed liquorice could inhibit the transport of rhein, possibly by mediating BCRP. These results would provide new insight into this ancient drug combination in toxicity reduction and clinical use.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Ying Peng and Min Fan contributed to this paper equally.

Acknowledgments

This work was financially supported by the National Science Foundation of China (no. 81274062).

References

Evidence-Based Complementary and Alternative Medicine


