A Newly Authenticated Compound from Traditional Chinese Medicine Decoction Induces Melanogenesis in B16-F10 Cells by Increasing Tyrosinase Activity

Xiu Juan Xin, Jiahong Zou, Tao Zou, Huoli Shang, and Li Yun Sun

State Key Laboratory of Bioreactor Engineering, East China University of Science and Technology, 130 Mei-Long Road, Shanghai 200237, China

Correspondence should be addressed to Li Yun Sun; liyunsun@ecust.edu.cn

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Vitiligo is a kind of depigmenting skin disease with an estimated prevalence of 1% among population worldwide [1]. The disease seems to affect all ethnic groups equally [2], and vitiligo has no race and gender predilection. Most patients with vitiligo characteristics are prior to the age of 20 years; it is stubborn and disfigure on the patients which can derive serious social problems [3]. The goal of vitiligo treatment is to suppress depigmentation and stimulate repigmentation of the patients. There have reports that the melanin level could be improved by suppressing the inflammation (e.g., topical corticosteroids and immunomodulators) or oxidation (e.g., vitamin D3 analogues and pseudocatalase) in early to active lesions and stimulate melanocyte differentiation or migration; the therapies included phototherapy like PUVA, narrowband UVB, 308 nm excimer laser, heliotherapy, and or vitamin D3 analogues [4]. Unfortunately, not all patients respond to the available therapies and some remedies have shown side effects which would suppress further treatment [5]. The Traditional Chinese Medicine (TCM) decoctions that have been widely used as externally applied agent in China showed efficacious and less side effects on vitiligo cure.

The TCM decoction used here composed by nine kinds of herbs which has been prevented for more than 60 years in China for its safety and reliability and even used on children or pregnant woman. After two courses of treatment with this prescription, the lesions skin areas could be recovered with melanocyte. The skin color disorders could be improved after further three courses of treatment and recurrence does not happen in the following years [6]. According to the Chinese medicine theory, the decoction can modulate...
the essential deficiency by promoting blood circulation and
darken the skin color by balancing melanin cell proliferation
and pigmentogenesis [7]. However, the possible respond
compounds and the exact mechanisms of the melanogenesis
induced by the decoction remained ambiguous. In our
previous studies, a new compound authenticated as apigenin-
7-butylene glucoside which was one member of flavonoids
was extracted from the medicine formulas; the purpose of this
study is to uncover the possible pharmacological activity of
the compound using B16-F10 cells in vitro and to evaluate its
potential value on vitiligo treatment.

As for the etiopathogenesis of vitiligo, numerous patho-
genic factors have been reported, including autoimmunity
suppress, neurogenic dysregulation, autotoxicity, and
oxidative stress [8], and among those theories, the degener-
ation of melanin in melanocytes was thought to be the main
cause of vitiligo formation [9].

Melanin biosynthesis was modulated by various
cytokines, such as basic fibroblast growth factor, endothelin-
1, cAMP response binding protein, and melanin stimulating
hormone [10, 11]. Among these factors, tyrosinase (TYR) was
the most restrictive factor in melanin biosynthesis. Tyrosi-
nase is a rate-limiting enzyme, it catalyzes the oxidation of
L-tyrosine to 3,4-dihydroxyphenylalanine (L-DOPA), and
then L-DOPA is oxidized to produce dopaquinone and thus
affects the production of melanin and the complexity of human beings. The tyrosinase was thought to be the key
denzyme during melanogenesis [12, 13], so far, most of skin-
whitening agents in cosmetics business are TYR inhibitors
[14].

The following melanin biosynthetic processes after dopa-
quinone are catalyzed by tyrosinase-related protein 1 (TRP-1)
and tyrosinase-related protein 2 (TRP-2) [15]. The expression
level and activity of tyrosinase together with tyrosinase-
related proteins in the melanocyte are also affecting the
melanin formation process [16–18]. The protein levels of
the TYP and TRPs are regulated by the microphthalmia-
associated transcription factor (MITF) as reported [19–21].
MITF is an important transcription factor that regulates the
expression levels of tyrosinase family, such as tyrosine,
tyrosine-1, and tyrosine-2 and might indirectly participate in
the proliferation of melanocytes, cell survival, and melanogenesis [22–24].

The purpose of this study is to explore the possible pharma-
ocological activity of the newly authenticated compound from
the useful decoction which has been used for vitiligo
cure and to uncover the possible mechanisms of it on melanogen-
esis using B16-F10 cells in vitro. The decoction consists
of nine crude herbs, including Lithospermum Erythrorhizon,
Radix Salviae Miltiorrhizae, Cortex Lycii Radicis, Rhizoma
Bletillae, Divaricata Saposhnikovia Root, Radix Angelicae
Sinensis, Rhizoma Chuanxiong, Phryma Leptostachya, and
Radix Glycyrrhizae. After purification using affinity resin and
cation exchange column of the ethyl acetate fraction, a new
compound apigenin-7-butylene glucoside was extracted and
authenticated by High Performance Liquid Chromatography
(HPLC), mass spectrometry (MS), and nuclear magnetic
resonance (1H-NMR). The results showed that it had faint
cytotoxicity on B16-F10 cells but stimulated tyrosinase activity
and melanin mass to about 1.4-fold of the controls.

Moreover, it also promoted the transcription and expression
levels of Tyr, Trp-1, and Trp-2 genes in B16-F10 cells and
does not affect the transcription and expression levels of the
cytokine of MITF in B16-F10 cells. The results suggested that
the apigenin-7-butylene glucoside stimulated melanogenesis
activity of B16-F10 cells by activating the Tyr, Trp-1, and Trp-
2 melanin biosynthesis pathway and showed potential values
in vitiligo treatment.

2. Materials and Methods

2.1. Reagents and Cells Culture. RPMI 1640 with phenol
red, fetal bovine serum (FBS), penicillin/streptomycin, and
trypsin was purchased from HyClone (South Logan, UT,
USA). Dimethylsulfoxide (DMSO), thiazolyl blue tetra-
zolium bromide (MTT), Triton X-100, sodium hydroxide
(NaOH), and L-3,4-dihydroxyphenylalanine (L-DOPA) were
acquired from Shanghai Sangon Biotech (Shanghai, China).
PrimeScript™ 1st strand cDNA Synthesis Kit and SYBR
Premix Taq™ (Tli RNaseH) were purchased from TaKaRa
(Dalian, China). β-actin antibodies and goat anti-rabbit
peroxidase-conjugated secondary antibodies were purchased
from Sigma–Aldrich (St. Louis, MO, USA). Antibodies
against TRP-2, TRP-1, and MITF were purchased from Santa-
Cruz Biotechnology (Santa Cruz, CA, USA). Tyrosinase
antibody was purchased from Abcam (Cambridge, MA, USA).
The other chemicals and reagents used in the study were high-
grade commercial products.

The B16-F10 mouse melanoma cells line was purchased from
the Cell Bank of Chinese Academy of Sciences (Shang-
hai, China). Cells were grown in RPMI 1640 media with
phenol red, supplemented with 10% FBS, 100 U/ml
penicillin, and 100 μg/ml streptomycin in a dynamic incubation
system at 37°C in a 5% CO₂ (Thermo Fisher Scientific, USA),
humidified atmosphere.

2.2. Cell Viability Assay. Cell viability analysis uses MTT
assay. Briefly, B16-F10 cells inoculated into 96-well plates
(5×10⁴ cells/100 μl media/well) for 12 hrs. Different concen-
trations ofstocked apigenin-7-butylene glucoside (0.625,1.25,
2.5, 5.0, 10.0, and 50.0 μg/ml) were added and incubated
for another 48 hrs in the incubator. Then, 20μl MTT (5 mg/ml
in PBS) solution was added to each well and incubated at 37°C
in a 5% CO₂ humidified atmosphere for 4 hrs. MTT media
were subsequently collected from all cells and the produced
formazan crystals were solubilized with 150 μL DMSO per
well by shaking for 10 min at ~120 rpm. Then the absorbance
at 570 nm was performed in a microplate spectrophotometer
(Bio Rad, model 680). Absorbance of the cell samples without
apigenin-7-butylene glucoside added set as negative control,
and the cells survival was regarded as 100%. The viability of
the treatment cells was expressed as (A₅⁷⁰ treatment/A₅⁷⁰
control) × 100%. Each treatment group was performed in
triplicate and each experiment was repeated for three times.

2.3. Tyrosinase Activity and Melanin Content Assay. Tyrosi-
nase activity was determined by measuring the L-DOPA
oxidation rate. Briefly, the cells were treated with different
concentrations of apigenin-7-butylene glucoside for 48 hours, then washed twice with PBS (pH 7.2, 10 mM), and frozen at -80°C for 30 minutes in PBS (pH 7.2, 10 mM) containing 0.1% Triton X-100, after melting at room temperature. 100 μL freshly prepared substrate solution (0.1% v/v L-3,4-dihydroxyphenylalanine) was added to each lysate and incubated for 2 hours at 37°C, and the levels of dopachrome were monitored under 475 nm. Cells without the compound added were regarded as control, and the relative activity of tyrosinase was expressed as (A_{475} treatment/A_{475} control) × 100%.

As for the melanin content analysis, IN the NaOH solution (with 10% DMSO) was used to dissolve the melanin of the cells after treatment with different concentration of apigenin-7-butylene glucoside for 48 hours. The cells with alkaline were incubated at 80°C for 1 hour, the supernatant was collected, and the absorbance of melanin in the cell lysis was analyzed under 470 nm. The level of melanin was expressed as (A_{470} treatment/A_{470} control) × 100%. Each treatment was performed in triplicate and each experiment was repeated for at least three times.

2.4. Tyr, Trp-1, Trp-2, and Mitf mRNA Level Determination. Total RNA was isolated using RNAiso kit. qRT-PCR analysis was performed for mRNA level analysis. The qRT-PCR protocol was conducted as follows: SYBR Premix Ex Taq™ (TakaRa Bio Inc Takara, Japan) kit used for double-stranded cDNA amplification. The cycling parameters were 1 cycle PCR reaction at 95°C for 2 minutes, then 1 cycle at 95°C for 10 min, and 30 cycles at 95°C for 30 s and 60°C for 30 s. The C_{t} value of the target genes was measured during the exponential amplification phase. The relative expression levels (defined as fold change) of the target genes were determined using a 2^{-ΔΔCt} method, and β-actin was used as internal control. The expression level was normalized to the fold change corresponding to control cells, according to the levels of the housekeeping gene β-actin, defined as 100.0. In all cases, each sample was performed in triplicate and each experiment was repeated for at least three times.

2.5. Western Blot Analysis. The cells were incubated with 1.25 μg ml^{-1}, 2.5 μg ml^{-1}, and 5.0 μg ml^{-1} apigenin-7-butylene glucoside for 48 hours, then collected and washed with PBS (pH 7.2 10 mM) for twice, and then centrifuged 10 min at 3000rpm. Cells were lysed by RIPA kit (Beyotime Institute of Biotechnology, China). Proteins were quantified using BCA kit (Beyotime Institute of Biotechnology, China). Appropriate amounts of lysates (200 mg of total proteins per lane) were loaded onto SDS-polyacrylamide precast gels and ran at 120 V on ice. The samples were subsequently transferred from the gels onto polyvinylidene difluoride (PVDF) membranes. The membranes were then washed with TTBS and blocked with 5%w/v skimmed milk for 1 hour at 25°C. The membranes were then incubated overnight at 4°C with the selected primary antibodies, including tyrosinase (1/1000), MITF (1/200), TRP-1 (1/200), TRP-2 (1/200), and β-actin antibodies. The membranes were then washed three times in Tween-Tris-buffered Saline (TTBS, 5 min per wash) and incubated with the secondary antibodies and goat anti-rabbit IgG conjugated with HRP at a dilution of 1:2000 for 1 hour at room temperature. Immunoblots were visualized using ECL Plus western blotting detection Kit (Amersham International, Little Chalfont, UK) and the signal density was calculated using a DNR Bio-Image system (Oclaro Israel Ltd., Jerusalem, Israel).

2.6. Statistical Analysis. The data were expressed as mean ± SEM. Multiple comparison statistical analysis was performed with one-way analysis of variance (ANOVA) followed by Tukey’s post hoc test for correction.

3. Results

3.1. Cell Viability. The structure of apigenin-7-butylene glucoside was in Figure 1. The cell viability result showed that it had no cytotoxicity on B16-F10 cells within 50 μg mL^{-1} after 48 hours of treatment. The viability of the B16-F10 cells was 97.1%, 96.7%, 94.7%, and 93.3%, respectively, after being incubated with 0.625μg mL^{-1}, 1.25μg mL^{-1}, 2.5μg mL^{-1}, and 5.0μg mL^{-1} for 48 hrs. Increasing the concentration to 10 and 50μg mL^{-1}, the viability rate of the cells was 91.6% and 88.2% (>P=0.5) after 48 hours incubation, the inhibition rate was still lower than 15%, though the results showed a decreased trend with the increasing of compound concentration (Figure 2). The compound was safe to B16-F10 cells at a lower concentration and had minor cytotoxicity on B16-F10 cells even at higher concentration like 10 μg mL^{-1} and 50μg mL^{-1}. The results suggested the safety of the compound at lower compound concentration (<5.0μg mL^{-1}) for B16-F10 cells.

3.2. Apigenin-7-Butylene Glucoside Stimulates Cellular Tyrosinase Activity and Melanin Levels. The tyrosinase activity and melanin level of the B16-F10 cells increased obviously after being incubating with the apigenin-7-butylene glucoside. The compound obviously enhanced tyrosinase activity in a dose dependent manner within a concentration of 5.0μg mL^{-1}. The tyrosinase activity increased to about 12% at a concentration of 0.625 μg mL^{-1} compared with controls, and it increased to 17, 27, 32, and 22% at the concentrations of 1.25, 2.5, 5.0, and 10.0 μg mL^{-1}, respectively, when compared with the controls, as shown in Figure 3. Meanwhile, melanin levels of B16-F10 cells also promoted at all apigenin-7-butylene glucoside concentrations, the melanin level enhanced to 41.7%
Figure 2: Cell availability test of B16-F10 after subjecting to different concentrations of apigenin-7-butyleneglucoside (∗ implies differences at p<0.005; ∗∗ implies significant differences at p<0.001).

Figure 3: Tyrosinase activity evaluation of B16-F10 cells to different concentrations of apigenin-7-butyleneglucoside (∗ implies differences at p<0.005; ∗∗ implies significant differences at p<0.001).

Figure 4: Melanin level analysis of B16-F10 cells to different concentrations of apigenin-7-butyleneglucoside (∗ implies differences at p<0.005; ∗∗ implies significant differences at p<0.001).

Figure 5: mRNA level assay of Tyr, Trp-1, Trp-2 and Mitf gene in B16-F10 cells after apigenin-7-butyleneglucoside treatment (∗ implies differences at p<0.005; ∗∗ implies significant differences at p<0.001).

3.3. Apigenin-7-Butylen Glucoside Upregulates mRNA Level of Melanogenesis Pathway Genes. The transcription levels of Trp, MITF, Trp-1, and Trp-2 genes were detected in this study which have been reported to participate in the melanin biosynthesis process. qRT-PCR assay results showed that the mRNA levels of Trp, Trp-1, and Trp-2 genes were all increased, and the transcription levels of the detected genes showed a dose-dependent manner within 5.0 μg/mL as Figure 5 shown. Among the four genes, Tyr was much more sensitive to the stimulation of the apigenin-7-butyleneglucoside on transcription level; it was 3.38-fold higher than that of controls after being incubated with 5.0 μg/mL apigenin-7-butyleneglucoside compared with the control ones, as shown in Figure 4.

The tyrosinase activity and melanin level decreased after being incubated with higher compound concentration of 10 and 50 μg/mL compared with 5.0 μg/mL, this might be the inhibition effect of the compound on B16-F10 cells, and there were decreased number of cells at the higher compound concentrations.

After incubated with 5.0 μg/mL apigenin-7-butyleneglucoside compared with the control ones, as shown in Figure 4.

3.4. Protein Level Analysis of TYR, TRP-1, TRP-2, and MITF in B16-F10 Cells. The protein levels of TYR, TRP-1, TRP-2, and MITF were detected by western blotting analysis, the results showed that all the levels of detected protein increased compared with controls except MITF at a concentration of 5.0 μg/mL, and the protein levels increased in a
The TCM formula studied in our work has been used in clinical for many years on vitiligo treatment in Henan province, China. The decoction of the formula has a high cure rate, but it is difficult to popularize out of the region for lack understanding of compound ingredients and unclear pharmacological mechanism. According to previous LC-MS spectrum results, the decoction of the formula herbs was composed mainly by flavonoid under the detection conditions, as other reported prescription on depigment treatment [27], and some responsible compounds were identified for pharmacology studies [28]. However, the responsible ingredients in the formula were not revealed still and thus attracted our interesting. Our group focused on identifying active compounds on melanogenesis of this formula previously and have fortunately isolated and authenticated a new compound of apigenin-7-butylene glucoside using HPLC and LC-MS spectrum techniques, after gradual purification with the decoction of the formula, 112 mg purified compound was extracted from 96 g herb formula (the detailed procedure reported in another paper).

In this work, we mainly evaluated the effects of apigenin-7-butylene glucoside on pigmentation induction and the underlying mechanisms that responded to vitiligo remedy using B16-F10 cells in vitro. The results indicated that apigenin-7-butylene glucoside could significantly upregulate melanin synthesis and had no obvious cytotoxicity on B16-F10 cells within 5.0 μg/mL concentration. B16-F10 cells were incubated with 5.0 μg/mL apigenin-7-butylene glucoside for 48 hours, 93.3% B16-F10 cells were available in Figure 3 and almost did not inhibit the growth of cells. Increasing the concentration to 10 μg/mL or 50 μg/mL, the proliferation rate was inhibited compared with the controls but still remained at a higher survive rate of 91.6 and 88.2%; there was no statistical difference on cells inhibition (P>0.5). The lower cytotoxicity of the drugs is significant as potential medicine agents [29]. Actually, it is common characters for many clinical compounds which showed contrast effects on detected targets at different concentrations. Arsenic trioxide (ATO) was a kind of poison; the epidemiological studies reported that drinking water or foods contaminated with low concentration of ATO might increase cancer risk [30] or fetal to people, but the high level ATO drinking water reduced markedly overall breast cancer mortality by over 50% in the large patients during a 15-year contaminating period and in women under 60 by 70% [31]. The researcher found that, at therapeutic doses, ATO has an excellent safety profile for APL treatment even in children [32], although it has notorious toxicity at special doses due to its covalent binding to cellular targets [33].

In contrast to the lower cytotoxicity, the melanin levels increased obviously in B16-F10 cells, and it showed a dose-dependent manner within the concentration of 5 μg/mL, and the melanin level increased to approximately 141.7% at a concentration of 5 μg/mL compared with the controls (untreated B16-F10 cells, P<0.005). Moreover, at higher concentrations numerous skin dysfunctions [26]. Though most TCM prescriptions have attractive therapeutic effects, the application and popularization blocked for its magic composition and ambiguous pharmacological mechanisms.

4. Discussion

Vitiligo is a kind of intractable skin disease, with dysfunction of melanin synthesis, and the lesions were distributed on mammalian skin. The abnormal physiological features can cause psychological stress and heavy economic burden on patients. Although several treatments are available, vitiligo cannot be completely cured for many patients [25], and some treatments can induce serious side affection on patients. Traditional Chinese Medicine (TCM) is one of the effective agents for vitiligo treatment in clinical and has been used to cure various human diseases for over 4000 years, including dose depended manner for TYR, TRP-1, and TRP-2 within the lower compound concentrations as Figure 6 shown. Tyrosinase gene was more sensitive to the apigenin-7-butylene glucoside at the translational level than the others. The protein level of the tyrosinase after incubated with 5.0 μg/mL apigenin-7-butylene glucoside for 48 hours was 3.89-fold higher than the controls, TRP-2 and TRP-1 level were also enhanced compared to the controls, and they were 3.24-fold and 2.45-fold. As for the MITF, the protein levels also increased slightly after incubation with the compound, but it was not assensitive as TYR, TRP-1, and TRP-2 do.

![Expression level analysis of TYR, TRP-1, TRP-2 and MITF in B16-F10 cells after apigenin-7-butylene glucoside treatment](image)

**Figure 6: Expression level analysis of TYR, TRP-1, TRP-2 and MITF in B16-F10 cells after apigenin-7-butylene glucoside treatment (\* implies differences at p<0.005; \*\* implies significant differences at p<0.001).**
of 10 μg mL⁻¹ and 50 μg mL⁻¹, melanin level decreased slightly compared with 5 μg mL⁻¹ apigenin-7-butyleneglucosidetreatment, but they were still higher than controls, the melanin level was 133.6% and 128.7% (P < 0.005), individually. The reason might be the less cell number when incubated with higher compound concentrations. Although the inhibition effects of the compound on B16-F10 cells were not notable, they were still established in our experiments.

Moreover, the transcription and expression levels of melanogenic enzymes TYR, TRP1, and TRP2 were all increased in B16-F10 cells after incubated with apigenin-7-butyleneglucoside except the pigmentation associated transcription factor MITF.

The biosynthesis of melanin depended mainly on its precursor, tyrosine, in a common tyrosinase dependent way. The melanogenesis process was regulated by tyrosinase families, including tyrosinase (TYR), tyrosinase-related factors (TRP-1), and the cytokines MITF as reported [34]. Tyrosinase is the key enzyme in melanin biosynthesis and determines the color of skin and hair [35]. Tyrosinase modulates two key steps on melanin synthesis pathway, it catalyzes the hydroxylation of tyrosine to DOPA, and then DOPA is oxidized to dopaquinone; the two compounds are key products among melanin synthesis in the melanocytes and catalyzed by the same enzyme [34]. And so the tyrosinase active regulators have been clinically used for the treatment of several dermatologic disorders related to melanin extinction [36]. According to the melanogenesis theory, we speculated that the expression level and catalyst activity of tyrosinase families might have close relationship with melanin synthesis level. The present result showed that tyrosinase activity and melanin level in B16-F10 cells were enhanced after incubating within the detected conditions. The transcription and translation levels of Tyr, Trp-1, and Trp-2 genes in B16-F10 cells were also stimulated in this study (Figures 5 and 6). These results suggested that the apigenin-7-butyleneglucosidestimulated melanogenesis in B16-F10 cells by enhancing TYR-TRP-1 and TRP-2 expression.

Microphthalmia-associated transcription factor (MITF), which is one of the most important nuclear activation transcription factors of tyrosinase, TRP-1 and TRP-2, plays key role in melanocyte development regulation and survival [37, 38]. Our results showed that apigenin-7-butyleneglucoside had no effects on the MITF transcription and translation levels in B16-F10 cells as shown in Figures 5 and 6, though the MITF was reported to be the master regulator in the transcription of genes which were involved in melanin synthesis. Some reports believed that adiponectin and AICAR were well-known AMP-activatedprotein kinase (AMPK) activators; the compounds inhibited melanin synthesis through AMPK stimulating in melanocytes, but had no effects on MITF mRNA and translation levels [39]. Actually, besides MITF modulated melanin synthesis pathway, many other signal transduction pathways were also disclosed on the balancing of melanogenesis process, like α-melanocyte-stimulating hormone (α-MSH) pathway and cAMP response element-binding protein (CREB) related pathway [40]. Alpha-MSH which is a kind of peptide hormone which bonds to melanocortin 1 receptor, when induced by other elements, will trigger the activity of adenylate cyclase via G proteins. The increased intracellular cAMP level then actives the phosphorylation of response element-binding protein at Ser133 via protein kinase A and enhances the expression of the transcription factors, like Lymphoid enhancer-binding factor 1 (LEF-1). The transcription factor LEF-1 then binds on the promoter of Tyr gene and enhances the gene expression. The dysregulation of LEF-1 will induce hyperpigmentary disorders [41]. Taken together, our findings suggested that the functional signaling molecules on trigging the activity and expression levels of tyrosinase families needed further exploration.

There were many bioactive compounds derived from plants which have been used on regulating melanocytes growth and proliferation, and some agents have also been used for vitiligo treatment in clinical or for cosmetic purposes in commercially [42, 43]. This work focused on the bioactivities study of a newly authenticated compound of apigenin-7-butyleneglucoside, and the results suggested its stimulating functions on melanogenesis in B16-F10 cells. Apigenin-7-butyleneglucoside can stimulate the catalyse activity of tyrosinase and increase the transcription and expression level of Tyr familiar genes, including Tyr, Trp-1, and Trp-2. The enhancement effects of the compound on melanin synthesis were stimulated by the pathway of TYR-TRP, rather than the traditional MITF pathway. In conclusion, this study provided an interesting insight for potential mechanism studies of apigenin-7-butyleneglucoside in vitiligo treatment in the future.

Abbreviations

TYR: Tyrosinase
TRP-1: Tyrosinase-related protein 1
TRP-2: Tyrosinase-related protein 2
MITF: Microphthalmia-associated transcription factor.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors do not have any conflicts of interest.

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